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1 **Respective contribution of Appl to mushroom body axon growth and long-term** 2 **memory in *Drosophila***

3
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17

18 **Summary**

19 The Amyloid Precursor Protein (APP) is linked to Alzheimer's disease. *Appl* is the single *Drosophila*
20 APP ortholog and is expressed in all neurons throughout development. *Appl* was previously
21 shown to modulate, cell-autonomously, axon outgrowth in the mushroom bodies (MBs), the fly
22 olfactory memory center. Furthermore, *Appl* knockdown in the MBs results in loss of memory
23 following the association of odorants with electric shocks. Nevertheless, the memory defects of
24 flies devoid of *Appl* remains unknown because *Appl^d* flies, carrying the only known null allele,
25 show an abnormal response to electric shock. We found that *Appl^d* affects the normal function of
26 *vnd*, the gene just proximal to *Appl*. We report here that *vnd* is required for MB β -branch axon
27 outgrowth. Moreover, *vnd* is expressed in neurons close to, but not within, the MB during
28 development and is required non-cell-autonomously for MB axon outgrowth. To decipher
29 developmental and memory defects specifically due to a loss of only *Appl* function, we generated
30 a precise *Appl* null allele (*Appl^{C2.1}*) by CRISPR/Cas9 genomic engineering. With *Appl^{C2.1}*, we
31 confirmed the partial requirement for *Appl* in MB axon outgrowth but found no defects of electric
32 shock avoidance, allowing us to test for potential memory defects. *Appl^{C2.1}* flies showed a
33 complete loss of long-term memory which was fully rescued by MB-restricted expression of *Appl⁺*
34 only during the adult stage. Therefore, we demonstrate that the complete lack of *Appl* affects
35 memory independently from structural developmental defects.

36

37 **Introduction**

38 Alzheimer's disease (AD) is associated with extracellular accumulation of amyloid fibrils derived
39 from the Amyloid Precursor Protein (APP). APPs have therefore been intensely investigated,
40 however their physiological function in the brain remains unclear and controversial [1-3].
41 Although there are three paralogues in mammals (APP, APLP1 and APLP2), *Drosophila* encodes a

42 single APP homologue, called *Appl*, that is expressed in all neurons throughout development. It
43 has been shown that the type I transmembrane protein coding gene *Appl* is a conserved neuronal
44 modulator of a Wnt planar cell polarity (Wnt/PCP) pathway, a regulator of cellular orientation
45 within the plane of an epithelium [4]. It has been proposed that *Appl* is part of the membrane
46 complex formed by the core PCP receptors *Fz* (*frizzled*) and *Vang* (*Van Gogh*) [4, 5].

47 The mushroom bodies (MBs) are two bilaterally symmetric structures in the central brain
48 that are required for learning and memory [6]. Each MB is comprised of 2000 neurons that arise
49 from 4 neuroblasts. Three types of neurons appear sequentially during development: the
50 embryonic/early larval γ , the larval $\alpha'\beta'$, and the late larval/pupal $\alpha\beta$ type. Each $\alpha\beta$ neuron
51 projects an axon that branches to send an α branch dorsally, which contributes to the formation
52 of the α lobe, and a β branch medially, which contributes to the formation of the β lobe [7]. So
53 far, *Appl^d* is the only reported null *Appl* allele and it results from a synthetic genomic deletion
54 removing the *Appl* locus [8]. The *Appl^d*-bearing chromosome was selected, after γ irradiation, as
55 a translocation of a partial duplication of the X chromosome on the Y chromosome to a X
56 chromosome terminal deficiency (S1 Fig). *Appl^d* flies are viable, fertile and display no gross
57 structural defects in the brain [8]. However, the *Appl* signaling pathway is required for proper
58 axon outgrowth in the MBs since *Appl^d* flies display modestly-penetrant axonal defects in $\alpha\beta$
59 neurons. In particular, *Appl* is required cell-autonomously for β -branch axon outgrowth [4, 9].
60 Using *in vivo* inducible RNA interference strategies, it was shown that *Appl* knockdown in the MBs
61 results in loss of memory following the association of odorants with electric shocks [10].
62 Nevertheless, the memory defect of *Appl* null flies cannot be assessed for aversive memory
63 because *Appl^d* flies, unlike wild-type flies, show an abnormal response to electric shock [8].

64 The *ventral nervous system defective* (*vnd*) gene, which is immediately proximal to *Appl*,
65 encodes a Nk2-class homeodomain transcription factor, that acts in a context-dependent manner
66 as an activator or repressor and is essential for the development of the nervous system. The *vnd*
67 gene encodes two proteins: Vnd-A and Vnd-B whose mRNAs arise from two different promoters.
68 These two proteins differ in their aminoterminal domains and are identical in the remainder of
69 their sequences. While Vnd-A is a transcription repressor from promoters containing Nk-2 binding
70 sites, Vnd-B directly activates transcription [11]. Flies bearing a null allele of *vnd* in hemi- or
71 homozygous condition are embryonic lethal [12].

72 Interestingly, we found that the *Appl^d* chromosome also genetically affects *vnd* function.
73 To genetically dissect *Appl* and *vnd* functions, we generated CRISPR alleles that precisely deletes
74 the *Appl* gene without affecting *vnd* function (*Appl^{C2.1}*) and that precisely deletes each one of the
75 two *vnd* transcripts without affecting *Appl* (*vnd^{CΔA}* and *vnd^{CΔB}*). We showed here that *vnd-A*, but
76 not *vnd-B*, is also required for MB β -branch axon outgrowth. Unexpectedly, *vnd* is expressed in
77 neurons close to, but not within, the MB during development and is required non-cell-
78 autonomously, likely by promoting the production of one (or several) secreted factor(s) involved
79 in the MB axon outgrowth. To analyze the exact developmental and memory defects due to the
80 specific loss of the *Appl* function only, we used *Appl^{C2.1}* and first confirmed the partial requirement
81 of *Appl* in MB β -branch axon outgrowth. Interestingly, *Appl^{C2.1}* flies, in contrast to *Appl^d* flies,
82 showed a normal response to electric shock but a complete loss of long term memory
83 independently from potential defects during brain development.

84

85 Results

86

87 **Generating new *AppI* CRISPR alleles**

88 The complex chromosome structure associated with *AppI^d* complicates its subsequent genetic
89 manipulation (S1 Fig). Therefore, we intended to produce new null *AppI* alleles by removing the
90 entire *AppI* transcriptional unit via CRISPR/Cas9-mediated deletion [13]. We recovered *AppI^{C1.4}*
91 and *AppI^{C2.1}*, two CRISPR *AppI* null alleles (Fig 1 and S2 Fig), following an adaptation of a published
92 protocol in order to produce circa 50kb deletions [14]. *AppI^d* MBs display a modestly-penetrant
93 cell-autonomous axon growth defect of the β -branches which manifests as an absence of the β
94 lobe [4, 5]. Surprisingly, we found a strong difference in the penetrance of the absence of the β
95 lobe phenotype in the two *AppI* CRISPR alleles. Although 9% of *AppI^{C2.1}* MBs lacked the β lobe ,
96 slightly lower than the 14.5% described for *AppI^d* [15], *AppI^{C1.4}* MBs displayed a much higher
97 penetrance (66%) of this phenotype (Fig 1). We determined the precise deletions present in the
98 two alleles via sequencing and found that while the *AppI^{C2.1}* deletion precisely removed the *AppI*
99 transcriptional unit, the *AppI^{C1.4}* allele also removed a part of the *vnd* transcriptional unit (Fig 1).
100 We then similarly mapped the *AppI^d* deletion and found that, unlike *AppI^{C2.1}*, it removes most of
101 the intergenic region between *AppI* and *vnd* which may influence *vnd* function (Fig 1). We then
102 assessed, by genetic complementation tests, if the different *AppI* null alleles possibly affected *vnd*
103 function. We used *vnd^A* a molecularly-characterized lethal allele which impacts both *vnd-B* and
104 *vnd-A* transcripts (Fig 2) [16]. Although *AppI^{C2.1}/vnd^A* MBs displayed no anatomical MB
105 phenotypes, *AppI^d/vnd^A* and *AppI^{C1.4}/vnd^A* MBs displayed 14% and 69% of β lobe absence,
106 respectively (Fig 2). Flies heterozygous for any of the four mutations displayed no MB phenotypes:
107 *vnd^A/+* (n = 72), *AppI^{C1.4}/+* (n = 60), *AppI^{C2.1}/+* (n = 42) and *AppI^d/+* previously described [4].
108 Therefore, *AppI^d* and *AppI^{C1.4}* but not *AppI^{C2.1}* affect *vnd* functions in MB development. Taken
109 together, these data strongly suggest that *vnd*, the gene just proximal to *AppI*, is also involved in
110 the MB β -branch axon growth and likely to a much greater extent than *AppI* itself.

111

112 **Generation of new *vnd* CRISPR alleles deleted either for the *vnd-B* or for the *vnd-A* function**

113 There are two *vnd* transcripts, *vnd-A* and *vnd-B* that produce two different Vnd proteins,
114 respectively Vnd-A and Vnd-B (Fig 2). It was proposed that, while Vnd-A has a main role during
115 embryogenesis, Vnd-B is associated with metamorphosis [11]. In order to determine which Vnd
116 isoform is required for MB β -branch axon growth, we produced the new *vnd* alleles *vnd^{CDA}* and
117 *vnd^{CDB}* by CRISPR/Cas9 genomic engineering that eliminate either the *vnd-A* or the *vnd-B* isoform,
118 respectively (Fig 3 and S3 Fig). Males bearing a *vnd^{CDB}* mutant allele (*vnd^{CDB}/Y*), as well as *vnd^{CDB}/*
119 *AppI^{C1.4}* females, are viable and have essentially wild-type MBs (Fig 3). Males bearing a *vnd^{CDA}*
120 mutant allele (*vnd^{CDA}/Y*) are embryonic lethal similarly to other *vnd* null alleles (*vnd^A*, *vnd^{A38}* and
121 *vnd⁶*) which affect both isoforms. Also, *AppI^{C1.4}/vnd^{CDA}* female MBs displayed a strong phenotype
122 of β lobe absence with a penetrance (64%) similar to those of *AppI^{C1.4}/vnd^A* female MBs (Fig 4).
123 Genetically, *AppI^{C1.4}/vnd^{CDA}; Dp-AppI⁺/+* females have two wild-type doses of *AppI⁺* and the β lobe
124 absence phenotype (69%) was not distinguishable from that observed in *AppI^{C1.4}/vnd^{CDA}; +/-*
125 females (64%) and therefore be entirely due to *vnd* lack-of-function. Supporting this hypothesis,
126 we observed a complete rescue when an extra dose of *vnd⁺* is supplied in *AppI^{C1.4}/vnd^{CDA}; Dp-*
127 *vnd⁺/+* females (Fig 4). *AppI^{C1.4}/Y* males have no *AppI⁺* function (null allele) and also display
128 reduced *vnd⁺* function. Moreover, *AppI^{C1.4}/Y* male MB phenotype is moderately rescued by the
129 presence of an extra dose of *AppI⁺* and strongly rescued by an extra dose of *vnd⁺* (Fig 4). Taken

130 together, these results strongly indicate that the *vnd-A* transcript but not the *vnd-B* transcript is
131 specifically required for MB β -branch axon growth.

132

133 **Vnd is expressed around the MBs and is not required within the MBs**

134 In order to know where *vnd* is expressed, we employed a *vnd-T2A-GAL4* line where GAL4 is under
135 the control of endogenous *vnd* regulatory sequences via CRISPR gene targeting so that GAL4 and
136 *vnd* are translated from the same mRNA transcript [17, 18]. UAS-GFP labelling revealed a pattern
137 in the ventral ganglion of the third larval instar central nervous system (L3 CNS) similar to that
138 described with immunostaining with antibodies against Vnd [11] thus validating the use of the
139 *vnd-T2A-GAL4* line as a *bona fide vnd* reporter (S4 Fig). GFP was detected, in the developing brain,
140 close to the MBs visualized by anti-Fas2 staining, from L3 to 24 h after puparium formation (APF)
141 (Fig 5A,A' - 5D,D'). Staining reveals two different morphological structures: one organized in a
142 honeycomb pattern and one containing filamentous structures with neurite appearance. To
143 determine the identity of the honeycomb structures, we performed DAPI staining at the L3 stage.
144 We observed a significant DAPI staining in these structures indicating that they likely correspond
145 to cell nuclei (Fig 5E,E'). Moreover, these cell nuclei are Vnd positive and Repo negative (Fig 5F,F'-
146 5G,G') indicating that these cells correspond to neurons, rather than glia. The neurites
147 corresponding to these cell nuclei are very close to the developing MB medial lobe from which
148 the adult β lobe develops (Fig 5). Noticeably, we could not observe any GFP labeling within the
149 MBs themselves. This unexpected expression pattern suggests a non-cell-autonomous role for
150 Vnd in the MB β -branch axon growth. We tested this hypothesis by MARCM mosaic analysis which
151 allows the generation of homozygous *vnd* loss-of-function MB clones in an otherwise
152 heterozygous genetic background and overcome organismal lethality [7, 19]. Mitotic
153 recombination was induced in late-stage embryos/early first instar larvae and the clones were
154 analyzed at the adult stage. We obtained 20 MB *vnd*^{A/A} clones that include the $\alpha\beta$ neurons. All the
155 20 *vnd* mutant clones displayed a β -branch axon growth that looked identical to that in wild-type
156 clones (S5 Fig). Moreover, we obtained similar results, namely a normal β -branch axon growth,
157 with two additional different null alleles, *vnd*^{A38} (8 clones) and *vnd*⁶ (4 clones) (S5 Fig). Taken
158 together, these results demonstrate that, although *vnd* function is strongly required for MB β -
159 branch axon growth, *vnd* is neither required nor expressed in the MBs themselves. Therefore, it
160 is most likely that Vnd regulates MB axon growth by a non-cell-autonomous mechanism.

161

162 ***Appl*-null flies have no long-term memory strictly due to the absence of *Appl* in adult MBs**

163 Although it is clear that *Appl* function is required for aversive olfactory memory, it has not yet
164 been possible to assess the memory of *Appl*-null flies, because *Appl*^d flies exhibit impaired electric
165 shock avoidance [8, 20]; In such flies a defect in the aversive memory assay could result from an
166 impairment in electric shock perception during conditioning and therefore not represent true
167 memory defects. We showed here that *Appl*^d also affects *vnd* function (Fig 2) and hence wondered
168 if this impairment in electric shock avoidance was really due to the lack of the *Appl* function itself.
169 In order to test this hypothesis, we evaluated the long-term memory (LTM) performance of
170 *Appl*^{C2.1/Y} flies which are null for *Appl* but wild-type for *vnd*. The electric shock avoidance and
171 olfactory acuity appears wild-type in *Appl*^{C2.1} flies (Fig 6). Interestingly, *Appl*^{C2.1} flies showed a
172 complete loss of LTM, and this memory defect was fully rescued by the restricted expression of
173 an UAS-*Appl* transgene in adult MBs (Fig 6). No rescue was detected in the non-induced 18°C

174 controls (S6 Fig). These results conclusively demonstrate that the specific and complete loss of
175 Appl affects long-term memory independently from any potential defects during brain
176 development.

177

178 Discussion

179

180 It has been proposed that Appl is part of the membrane complex formed by the core PCP proteins
181 Fz and Vang [4]. *Appl^d* was, previous to this report, the only *Appl* null allele described. *Appl^d*
182 mutant flies are completely lacking Appl function and exhibit 14.5 % MB β -lobe loss [15] in
183 accordance with what has been reported [4, 5]. This modestly-penetrant axonal phenotype due
184 to the lack of Appl could be due to partially redundant function provided by the other
185 transmembrane receptors of the complex. In particular, flies homozygous for the loss of function
186 allele *Vang^{stbm-6}* exhibit 50% β -lobe loss [5]. As the *Appl^d* allele involves complex chromosomal
187 rearrangements [8], we decided to leverage CRISPR in order to produce a definitive deletion of
188 around 50 kb to specifically eliminate the *Appl* transcriptional unit. We recovered two *Appl*
189 complete deletions: *Appl^{C1.4}* and *Appl^{C2.1}* removing around 52 kb and 48 kb and exhibiting 66 %
190 and 9 % β -lobe loss respectively. DNA sequences analysis revealed that, while the *Appl^{C2.1}* deletion
191 removes only *Appl* sequences, the *Appl^{C1.4}* deletion additionally removes a part of the proximally
192 located transcriptional unit corresponding to the *vnd* gene (Fig 1). This indicates that in addition
193 to Appl, *vnd* is likely involved in the β -branch axon outgrowth. Moreover, as can be expected from
194 the extent of the deletions and based on genetic complementation tests, *Appl^{C1.4}* deletion, but
195 not *Appl^{C2.1}* deletion, affects *vnd* function. Most importantly, the *Appl^d* deletion, which removes
196 most of the Appl coding sequence and also the intergenic region between *Appl* and *vnd*, also
197 affects *vnd* function (Fig 1 and 2). The role of *vnd* in β -branch axon outgrowth is complicated by
198 the fact that the *vnd* viable mutant alleles (*Appl^d* and *Appl^{C1.4}* deletions), are associated with the
199 lack of *Appl* function. Nevertheless, employing a small (less than 100 kb) genomic *Appl⁺*
200 duplication, we found that flies lacking only *vnd* function exhibit 69% β -lobe loss that is completely
201 rescued by a small genomic *vnd⁺* duplication (Fig 4). This demonstrates the requirement of *vnd*
202 function for MB β -branch axon outgrowth. To better understand the roles of both Vnd isoforms
203 in the MB β -branch axon outgrowth, we produced two mutations, *vnd^{C Δ B}* and *vnd^{C Δ A}*, which
204 remove the *vnd-B* and the *vnd-A* transcript respectively (Fig 3 and S3 Fig). Surprisingly, although
205 Vnd-A has a significant role in MB axon growth, apparently Vnd-B plays little to no role in this
206 process. Noticeably, MB phenotype of *Appl^{C1.4}/vnd^{C Δ A}* flies is similar in penetrance to the one of
207 *Appl^{C1.4}/vnd^A*, indicating that the lack of *vnd-A* function seems to have the same effect than the
208 complete lack of both *vnd* functions.

209 It may have been anticipated that Vnd, a transcription factor, was required within the
210 MBs and was probably being part of the Appl signaling pathway in order to act on axon growth.
211 Consequently, Vnd should have been expressed within the MBs and the *vnd* function was
212 expected to be cell-autonomous. However, endogenous *vnd* transcription monitored by GAL4
213 expression that is translated from the same mRNA transcript and validated by an anti-Vnd
214 antibody show no sign of expression within the MBs. Rather, *vnd* is transcribed within a neuronal
215 brain structure in close proximity to the developing MB medial lobe from which the adult β lobe
216 develops (Fig 5). Also, using MARCM mosaic analysis, we show that β axons extended from MB
217 clones null for *vnd* function exhibit wild-type growth patterns (S5 Fig) demonstrating a non-cell-

218 autonomous requirement for *Vnd*. Therefore, we favor the hypothesis where *vnd* expressed in a
219 MB surrounding brain structure is generating, directly or indirectly, one or several secreted
220 factors that will act on the axon growth of the developing β medial lobe.

221 Prior to this study, it was not possible to evaluate the effects of a complete lack of *App1* in
222 aversive olfactory memory due to the failure of *App1^d* mutants to show normal shock reactivity
223 [8, 20]. It was therefore hypothesized that it was the lack of *App1 per se* which was the cause of
224 this impairment in electric shock avoidance. We reevaluated this hypothesis with the *App1^{C2.1}*
225 allele which removes only the *App1* function. Flies bearing only a deletion of *App1* react normally
226 to electric shock and olfactory cues similarly to control flies but show a total lack of 24h memory
227 after a long-term conditioning (Fig 6). Therefore, it is not the lack of *App1* function in the *App1^d*
228 mutants which is the origin of the impairment in electric shock avoidance. Importantly, the
229 memory defect can be completely rescued by the expression of *App1⁺* solely in the post-
230 development adult MBs. This demonstrates that a complete lack of *App1* results in a specific
231 memory defect independently of any role that *App1* might have in the developing brain.
232 Consequently, the modest mutant MB axonal growth morphological phenotype displayed by the
233 *App1^{C2.1}* allele likely has no measurable impact on long-term memory. This is in accordance with
234 the requirement of the vertical lobes but not the median lobes in aversive long-term memory
235 [21].

236 The accumulation of amyloid- β (A β) is a hallmark feature of AD [22, 23]. Nevertheless, the
237 production of A β peptides is at the expense of the full-length APP protein levels, whose roles in
238 AD are, as yet, unclear. The *Drosophila* memory defect due to the complete lack of *App1* may
239 therefore be relevant to the memory defects associated with human AD. Therefore, our
240 observation that the memory defect resulting from the complete lack of *App1* could be completely
241 rescued by the expression of *App1⁺* in a specific adult brain region, thus bypassing any potential
242 role of APP1 in development could be of significance to our understanding of the molecular basis
243 of AD.

244

245 **Materials and methods**

246

247 ***Drosophila* stocks**

248 All crosses were performed on standard culture medium at 25° C. Except where otherwise stated,
249 all alleles and transgenes have been described previously (<http://flystocks.bio.indiana.edu/>). The
250 following alleles were used: *App1^{C1.4}*, *App1^{C2.1}*, *vnd^{CAA}* and *vnd^{CAB}* were generated in this study. *App1-
251 y⁺* (BL #56073), *App1^d* (BL #43632), *Vang^{stbm-6}* (BL #6918) and *vnd^A* (BL #57139), *vnd^{A38}* [24](Chu
252 1998), *vnd⁶* [12](Jimenez 1995). The following transgenes were used: *UAS-mCD8::GFP* (BL #5137),
253 *UAS-App1* (BL #38403) and the duplications resulting from genomic transgenes *Dp-App1⁺* (BL
254 #32288) and *Dp-vnd⁺* (BL #30219) both inserted into the same attP site at 65B2. We used three
255 GAL4 lines: *vnd-T2A-GAL4* which reveals *vnd* transcription, *c739-GAL4* (BL #7362) expressed in
256 adult $\alpha\beta$ MB neurons [25] and VT30559 expressed in adult MB neurons [26]. Recombinant
257 chromosomes were obtained by standard genetic procedures and were molecularly verified.

258

259 **CRISPR-Cas9 strategy**

260 All guide RNA sequences (single guide RNA (sgRNA)) were selected using the algorithm
261 targetfinder.flycrispr.neuro.brown.edu/ containing 20 nucleotides each (PAM excluded) and are

262 predicted to have zero off-targets. We selected one pair of sgRNA for the *App1* gene and two pairs
263 of sgRNA for the *vnd* gene. For *App1*, the pair is targeting the all transcriptional unit (S2 Fig). For
264 *vnd*, each pair is targeting either the A specific region of *vnd* or the B specific region of *vnd* (S3
265 Fig). We used the following oligonucleotide pairs: CRISPR-1 *App1* fwd and CRISPR-1 *App1* rev to
266 target the all *App1* region, CRISPR-1 *vnd* A fwd and CRISPR-1 *vnd* A rev to target the A region of
267 *vnd*, CRISPR-1 *vnd* B fwd and CRISPR-1 *vnd* B rev to target the B region of *vnd* (see the
268 corresponding oligonucleotide sequences in S2 and S3 Fig.). We introduced two sgRNA sequences
269 into pCFD4 [14], a gift from Simon Bullock (Addgene plasmid # 49411) by Gibson Assembly (New
270 England Biolabs) following the detailed protocol at crisprflydesign.org. For PCR amplification, we
271 used the protocol described on that website. Construct injection was performed by Bestgene
272 (Chino Hills, CA) and all the transgenes were inserted into the same attP site (VK00027 at 89E11).
273 Being aware that the DNA excision of nearly 50 kb might be rare, we have decided to use a positive
274 screen before validation by genomic PCR. We have used a stock where the *App1* gene is marked
275 by a y^+ construct. The deletion of the *App1* gene should therefore be associated to y^- phenotype.
276 Transgenic males expressing the *App1* sgRNAs and bearing an isogenized X chromosome (y *App1*-
277 y^+ *Act-Cas9 w^**) were crossed to *FM7c/ph⁰ w* females. 180 crosses were set up and only two
278 crosses gave rise to y *App1^{deletion?} Act-Cas9 w^*/FM7c* females indicating 1% of CRISPR efficacy.
279 From each of these two crosses, a single y *App1^{deletion} Act-Cas9 w^*/FM7c* female was crossed with
280 *FM7c* males to make a stock (*App1^{C1.4}* and *App1^{C2.1}* where Cas9 was removed). The deletion were
281 then validated by genomic PCR using two couples of primers: *App1^{C1.4}* fwd
282 (GAGCCAGATACACAAGCACA) / *App1^{C1.4}* rev (GGCTTTGTTTACTTCTGGC) and *App1^{C2.1}* fwd
283 (TCCTACTACGTTCCACAATC) / *App1^{C2.1}* rev (TAATGCCCAACATATCCAAC). The precise endpoints of
284 the deletion were determined by sequencing (Genewiz, France). Transgenic males expressing the
285 different *vnd* sgRNAs were crossed to y *nos-Cas9 w^** females bearing an isogenized X
286 chromosome. 100 crosses were set up for each sgRNA pair, with up to 5 males (because of poor
287 viability) containing both the sgRNAs and *nos-Cas9*, and 5 *FM7c/ph⁰ w* females. From each
288 selected cross, a single y *vnd^{deletion?} nos-Cas9 w^*/FM7c* female was crossed with *FM7c* males to
289 make a stock which was validated for the presence of an indel by genomic PCR with primers
290 flanking the anticipated deletion and the precise endpoints of the deletion were determined by
291 sequencing (Genewiz, France) using *vnd*-specific primers: *vnd^{CΔA}* fwd (ccaacaaagccgagagtctct) /
292 *vnd^{CΔA}* rev (cgggaatttctaagccagggt) and *vnd^{CΔB}* fwd (cgatttggggcgtgtgagta) / *vnd^{CΔB}* rev
293 (gttgggctttaatccgggagt). A CRISPR efficacy of at least 5% was obtained in both cases. We kept a
294 single stock from each set of CRISPR experiment: *vnd^{CΔA}* and *vnd^{CΔB}* where Cas9 was removed.
295

296 **Brain dissection, immunostaining and MARCM mosaic analysis**

297 Adult fly heads and thoraxes were fixed for 1 h in 3.7% formaldehyde in phosphate-buffered
298 saline (PBS) and brains were dissected in PBS. For larval and pupal brains, brains were first
299 dissected in PBS and then fixed for 15 min in 3.7% formaldehyde in PBS. They were then treated
300 for immunostaining as described [19, 27]. Antibodies, obtained from the Developmental Studies
301 Hybridoma Bank, were used at the following dilutions: Mouse monoclonal anti-Fas2 (1D4) 1:10,
302 mouse monoclonal anti-Repo (8D1.2) 1:10 and guinea pig polyclonal anti-vnd (1/1000). Goat
303 secondary antibodies conjugated to Cy3 against mouse or guinea pig IgG (Jackson
304 Immunoresearch laboratory) were used at 1:300 for detection. DAPI (Sigma) was used after
305 secondary antibody washes. Tissues were incubated 10 minutes at room temperature in a 1/1000

306 solution from a stock containing 1 mg/ml of DAPI solution, then washed. To generate clones in
307 the MB, we used the MARCM technique [19]. First instar larvae were heat-shocked at 37°C for 1
308 h. Adult brains were fixed for 15 min in 3,7% formaldehyde in PBS before dissection and GFP
309 visualization.

310
311 **Production of the anti-Vnd antiserum**
312 Full length vnd-RA was amplified via PCR from cDNA and cloned into a pRSET bacterial expression
313 vector (ThermoFisher) in frame with an N-terminal 6x-His tag, amplified in DH5alpha and
314 transformed into BL21 competent cells for protein production. His-tagged VndA was purified over
315 a Nickel-NTA agarose resin (Qiagen) according to manufacturer's recommendation. Purified
316 protein was injected into guinea pigs by Squarix GmbH (Marl, Germany, www.squarix.de),
317 selected by assuring that pre-bleeds did not produce signal in Western blots of fly extracts.
318 Successive bleeds were screened for distinct signal in Western blots using embryo extracts and
319 by immunohistochemistry, where signal was expected post 2.5hrs AED.

320
321 **Behavior experiments**
322 For behavior experiments, flies were raised on standard medium at 18°C and 60% humidity in a
323 12-h light/dark cycle. We used the TARGET system [28] to inducibly express UAS-*Appl*⁺ construct
324 exclusively in adult flies and not during development. 0-3 day old flies were transferred to fresh
325 bottles containing standard medium and kept at 30.5°C for 2 days before conditioning. For non-
326 induced experiments, flies were transferred to fresh bottles containing standard medium and
327 kept at 18°C before conditioning. All the behavior experiments, including the sample sizes, were
328 conducted similarly to other studies [29]. Groups of 20-50 flies were subjected to olfactory
329 conditioning protocol consisting of five associative cycles spaced by 15min inter-trial intervals (5x
330 spaced training). Conditioning was performed at 25°C using previously described barrel-type
331 machines that allow the parallel training of up to 6 groups [21]. Throughout the conditioning
332 protocol, each barrel was plugged into a constant air flow at 2 L/min. For a single cycle of
333 associative training, flies were first exposed to an odorant (the CS+) for 1 min while 12 pulses of
334 5s long 60 V electric shocks were delivered; flies were then exposed 45 s later to a second odorant
335 without shocks (the CS-) for 1 min. The odorants 3-octanol (Fluka 74878, Sigma-Aldrich) and 4-
336 methylcyclohexanol (Fluka 66360, Sigma-Aldrich), diluted in paraffin oil to a final concentration
337 of 2.79·10⁻¹ g/L, were alternately used as conditioned stimuli. During unpaired conditionings, the
338 odor and shock stimuli were delivered separately in time, with shocks occurring 3 min before the
339 first odorant. Flies were kept at 18°C on standard medium between conditioning and the memory
340 test. The memory test was performed at 25°C in a T-maze apparatus [30](Tully 1985), 24 h after
341 spaced training. Each arm of the T-maze was connected to a bottle containing 3-octanol and 4-
342 methylcyclohexanol, diluted in paraffin oil to a final concentration identical to the one used for
343 conditioning. Flies were given 1 min to choose between either arm of the T-maze. A performance
344 score was calculated as the number of flies avoiding the conditioned odor minus the number of
345 flies preferring the conditioned odor, divided by the total number of flies. A single performance
346 index value is the average of two scores obtained from two groups of genotypically identical flies
347 conditioned in two reciprocal experiments, using either odorant (3-octanol or 4-
348 methylcyclohexanol) as the CS⁺. The indicated 'n' is the number of independent performance
349 index values for each genotype. The shock response tests were performed at 25°C by placing flies

350 in two connected compartments; electric shocks were provided in only one of the compartments.
351 Flies were given 1 min to move freely in these compartments, after which they were trapped,
352 collected, and counted. The compartment where the electric shocks were delivered was
353 alternated between two consecutive groups. Shock avoidance was calculated as for the memory
354 test. Because the delivery of electric shocks can modify olfactory acuity, our olfactory avoidance
355 tests were performed at 25°C on flies that had first been presented another odor paired with
356 electric shocks. Innate odor avoidance was measured in a T-maze similar to those used for
357 memory tests, in which one arm of the T-maze was connected to a bottle with odor diluted in
358 paraffin oil and the other arm was connected to a bottle with paraffin oil only. Naive flies were
359 given the choice between the two arms during 1 min. The odor-interlaced side was alternated for
360 successively tested groups. Odor concentrations used in this assay were the same as for the
361 memory assays. At these concentrations, both odorants are innately repulsive.

362
363 **Microscopy and image processing**
364 Images were acquired at room temperature using a Zeiss LSM 780) equipped with a 40x PLAN
365 apochromatic 1.3 oil-immersion differential interference contrast objective lens and a Leica SP8
366 laser scanning confocal microscopes (MRI Platform, Institute of Human Genetics, Montpellier,
367 France. The immersion medium used was Immersol 518F. The acquisition software used was Zen
368 2011 (black edition) for the Zeiss and LasX for the Leica. Contrast and relative intensities of the
369 green (GFP), of the red (Cy3) and of the DAPI channels were processed with ImageJ (Fiji) software.
370 Settings were optimized for detection without saturating the signal.

371
372 **Statistics**
373 Comparison between two groups expressing a qualitative variable was analyzed for statistical
374 significance using the Chi² test. (BiostaTGV: <http://biostatgv.sentiweb.fr/?module=tests>). For
375 behavior experiments, all data are presented as mean ± SEM. 2 groups of about 30 flies were
376 reciprocally conditioned, using respectively octanol or methylcyclohexanol as the CS⁺. The
377 memory score was calculated from the performance of two groups as described above, which
378 represents a single experimental replicate. Comparisons of the data series between two
379 conditions were achieved by a two-tailed unpaired t-test. Comparisons between more than two
380 distinct groups were made using a one-way ANOVA test, followed by Tukey pairwise comparisons
381 between the experimental groups and their controls. ANOVA results are presented as the value
382 of the Fisher distribution F(x,y) obtained from the data, where x is the number of degrees of
383 freedom between groups and y is the total number of degrees of freedom for the distribution. In
384 the figures, asterisks illustrate the significance level of the t-test, or of the least significant
385 pairwise comparison following an ANOVA. Values of p < 0.05 were considered to be significant.
386 Statistical significance was defined as: ns = not statistically different, **p<0.01 and****p<0.0001.

387
388

389 **Figure legends**

390
391 **Fig 1. Generating new *App1* CRISPR alleles.**
392 (A) Schematic representation of *App1* (blue) and *vnd* (green) genes and transcripts. White boxes
393 represent 5'UTR and 3'UTR, blue and green boxes represent *App1* and *vnd* coding sequences

394 respectively. Schematic representations of the *AppI^d*, *AppI^{C1.4}* and *AppI^{C2.1}* mutant alleles where
395 the deleted sequences are represented in light blue. The hatched black segment represents the
396 uncertainty where the deletion of *AppI^d* starts in 5'. (B) Representation of the 3' limit of the *AppI^d*,
397 *AppI^{C1.4}* and *AppI^{C2.1}* mutant alleles (light blue). The hatched black segment represents the
398 uncertainty where the *AppI^d* deletion ends in 3'. The orange star represents the position of the 3'
399 sgRNA used to create the two CRISPR deletions. (C) Wild-type (WT) MB revealed by an anti-Fas2
400 staining. The pink and yellow arrowheads show respectively the α and β lobe (n=216 MBs). (D-E)
401 Anti-Fas2 staining reveals the loss of β lobe on a representative *AppI^{C1.4}* brain (n=147 MBs) (D)
402 and on *AppI^{C2.1}* brain (n=253 MBs) (E). The % represents the proportion of loss of β lobe for each
403 genotype. Scale bar = 50 μ m.

404

405 **Fig 2. *AppI^d* and *AppI^{C1.4}*, but not *AppI^{C2.1}*, are impaired for *vnd* function.**

406 A) Schematic representation of *vnd* gene and transcripts. White boxes represent 5' UTR and 3' UTR,
407 green boxes represent coding sequences. Red star represents point mutation on *vnd^A* allele.
408 Schematic representation of *vnd-B* and *vnd-A* transcripts. The *vnd^A* mutation affects both
409 transcripts. (B-D) Anti-Fas2 staining reveals the loss of β lobe on a representative *AppI^d/vnd^A* brain
410 (n=214 MBs) (B), *AppI^{C1.4}/vnd^A* brain (n=147 MB) (C) but not on *AppI^{C2.1}/vnd^A* brain (n=248 MBs)
411 (D). The % represents the proportion of loss of β lobe for each genotype. Scale bar = 50 μ m.

412

413 **Fig 3. Generating new *vnd* CRISPR alleles deleted either for the *vnd-B* or the *vnd-A* function.**

414 (A) Schematic representation of *vnd* gene and transcripts. White boxes represent 5' UTR and
415 3' UTR, green boxes represent coding sequences. The hatched black boxes represent the deleted
416 region of *vnd^{CDA}* and *vnd^{CDB}* (B-C) Anti-Fas2 staining reveals essentially wild-type looking MBs on
417 a representative *vnd^{CDB}* brain (n=146 MBs) (B) and *AppI^{C1.4}/vnd^{CDB}* brain (n=159 MB) (C). The %
418 represents the proportion of loss of β lobe for each genotype. Scale bar = 50 μ m.

419

420 **Fig 4. *Vnd-A* is required for MB β -branch axon outgrowth.**

421 (A) Quantitation of the rescue of MB β lobe absence of *AppI^{C1.4}/Y* (66%; n= 147 MBs, see a
422 representative brain in Fig.1D) and *AppI^{C1.4}/vnd^{CDA}* (64%; n=198 MBs) (light blue) by an *AppI*
423 duplication (dark blue) (50%; n=150 MBs, 69%; n=89 MBs) and by a *vnd* duplication (green) (3%;
424 n=191 MBs ; 0%; n=164 MBs). **p<0.01 and ****p<0.0001. (B-D) Anti-Fas2 staining on a
425 representative rescued *AppI^{C1.4}/Y; Dp-vnd^{+/+}* brain (B), *AppI^{C1.4}/vnd^{CDA}; +/-* brain (C) and on
426 *AppI^{C1.4}/vnd^{CDA}; Dp-vnd^{+/+}* brain (D). The % represents the proportion of loss of β lobe for each
427 genotype. Scale bar = 50 μ m.

428

429 **Fig 5. *vnd* is expressed close to the developing MB medial lobe.**

430 (A-D) Visualization of *vnd* expression in brain using *vnd-GAL4* driven *UAS-GFP* at L3, 0h, 6h, and
431 24h APF. GFP was visualized on green and anti-FasII staining labelling of MB axons on red. A-D are
432 confocal z stacks taken the whole MB and A'-D' are stacks of five plans from A-D respectively
433 comprising GFP⁺ regions surrounding MB. Arrowheads point to *vnd-GAL4* driven *UAS-GFP*
434 filamentous structures that surround MB medial lobes, which correspond to γ lobes from L3 to 6
435 h APF (A'-C') and β lobes at 24 h APF (D'). n \geq 10 for each time point. (E-G) show single confocal
436 plans showing *vnd* expression using *vnd-GAL4* driven *UAS-GFP* (single channel) at L3. E' to G'
437 are merges of single channel E to G respectively and DAPI staining (red on E'), anti-*vnd* (red on F') and

438 anti-Repo (red on G'). DAPI labelled nuclei in honeycomb GFP⁺ structures are pointed by arrows
439 in E'. These nuclei were also vnd⁺ (arrows in F') and they were not labelled by an anti-repo (white
440 arrow in G') which labels glia nuclei (red dots pointed by a yellow arrow in G'). Scale bars are 50
441 μm . n \geq 5 brains for each type of staining.

442
443 **Fig 6. The long-term memory defect of *Appl*^{C2.1}/Y flies is rescued by *Appl*⁺ expression in the adult**
444 **mushroom body.**

445 (A) Olfactory acuity was normal in *Appl*^{C2.1}/Y flies. For octanol, data presented as mean \pm SEM. *t*-
446 test n=12, $t_{22} = 1.12$, $P = 0.26$, ns: not significant. For methylcyclohexanol, data presented as mean
447 \pm SEM. *t*-test n=12, $t_{22} = 1.51$, $P = 0.15$, ns: not significant. (B) Shock reactivity was normal in
448 *Appl*^{C2.1}/Y flies. Data presented as mean \pm SEM. *t*-test. n=12, $t_{22} = 1.47$, $P = 0.16$, ns: not significant.
449 (C) Flies only null for *Appl* have no measurable LTM which is fully rescued by the restricted
450 expression of *Appl* in adult MBs. Data presented as mean \pm SEM. n = 22-23, $F_{(4, 108)} = 16,41$, $P <$
451 0.0001 . Significance level of *t*-test or least significant pairwise comparison following ANOVA:
452 ****p < 0.0001, ns: not significant.

453

454 Supporting information

455

456 S1 Fig. Scheme of *Appl*^d chromosomal organization

457 (A) Schematic representation of *Appl*^d mutant allele. The upper panel represents the wild-type
458 (WT) X (black) and Y (orange) chromosomes. The *elav*, *Appl* and *vnd* genes are represented
459 respectively in yellow, blue, and green boxes. The middle panel represents the deficiency Df
460 (1)RT518 on the X chromosome deleting the distal part of the chromosome including the *elav* and
461 *Appl* genes. The X Dup Y chromosome represents a duplication of the X chromosome linked to
462 the Y chromosome including the distal part of the X chromosome including the *elav* gene. The
463 lower panel represents a proposed structure of the *Appl*^d chromosome where the deficiency
464 bearing X chromosome had been linked by gamma rays to the X Dup Y chromosome (adapted
465 from [8]).

466

467 S2 Fig. Structure of the *Appl*^{C1.4} and *Appl*^{C2.1} CRISPR alleles

468 (A) Schematic representation of *Appl* (blue) and *vnd* (green) genes and transcripts. White boxes
469 represent 5'UTR and 3'UTR, blue and green boxes represent *Appl* and *vnd* coding sequences
470 respectively. The yellow stars in *Appl* 5' region and the orange star in *Appl* 3' region represent the
471 position of the sgRNA used to create *Appl*^{C1.4} and *Appl*^{C2.1} mutants. The red and blue boxes
472 represent respectively the regions magnified in B and C. (B) Representation of the 5' sequence
473 targeted by the sgRNA (yellow) and the PAM domain (magenta) in the WT chromosome *Appl*
474 (blue) (upper line). The lower line is the sequence of the *Appl*^{C1.4} mutant showing the beginning
475 of the deletion about 1kb after the region targeted by the sgRNA. (C) Representation of the 3'
476 sequence targeted by the sgRNA (orange) and the PAM domain (magenta) in the WT chromosome
477 *Appl* (blue) and *vnd* (green) (upper line). The lower line is the sequence of the *Appl*^{C1.4} mutant
478 showing the end of the deletion about 6kb after the region targeted by the sgRNA. (D)
479 Representation of the 5' and 3' sequences targeted by the sgRNA (yellow in 5' and orange in 3')
480 and the PAM domain (magenta) in the WT chromosome *Appl* (blue) (upper line). The lower line

481 is the sequence of the *Appl*^{C2.1} mutant showing the beginning of the deletion in the sequence
482 targeted by the 5' sgRNA (yellow) and the end of the deletion in the sequence targeted by the 3'
483 sgRNA (orange).

484

485 **S3 Fig. Structure of the *vnd*^{CAB} and *vnd*^{CDA} CRISPR alleles**

486 (A) Schematic representation of *vnd* (green) gene and transcripts. White boxes represent 5'UTR
487 and 3'UTR, green boxes represent *vnd* coding sequences. The yellow stars represent the position
488 of the sgRNA used to create *vnd*^{CAB} mutant. The orange stars represent the position of the sgRNA
489 used to create *vnd*^{CDA} mutant. (B) Representation of the sequences targeted by the sgRNA (yellow)
490 and the PAM domain (blue) in the WT chromosome *vnd* (green) (upper line). The lower line is the
491 sequence of the *vnd*^{CAB} mutant showing the beginning of the deletion in the sequence targeted
492 by the 5' sgRNA (yellow) and the end of the deletion in the sequence targeted by the 3' sgRNA
493 (yellow). (C) Representation of the sequences targeted by the sgRNA (orange) and the PAM
494 domain (blue) in the WT chromosome *vnd* (green) (upper line). The lower line is the sequence of
495 the *vnd*^{CDA} mutant showing the beginning of the deletion at one nucleotide from the sequence
496 targeted by the 5' sgRNA (orange) and the end of the deletion within the sequence targeted by
497 the 3' sgRNA (orange).

498

499 **S4 Fig. Vnd L3 CNS labelling**

500 (A-C) Visualization of *vnd* expression in VNC using both *vnd*-*GAL4* driven *UAS*-*GFP* (green) and an
501 anti-*vnd* antibody (red) at L3 stages. Images A-C and A'-C' are the same confocal z-projection.
502 Laser intensities leading to the green color were increased from B, C to B', C' in order to illustrate
503 the perfect colocalization between the two markers. Low laser intensity allowed us to show red
504 nuclear *vnd* staining in only some GFP⁺ cell bodies (white arrowheads in A- C). Soma containing
505 *vnd*⁺ positive nuclei that do not seem to be surrounded by GFP in A-C, can be visualized by
506 increased green laser intensity A'- C' (yellow arrowheads), which saturates somas visualized in A-
507 C. Scale bars are 50 μ m. n \geq 5 brains.

508

509 **S5 Fig. *vnd* null mutant clones**

510 (A-B) A representative neuroblast MARCM clone in a wild-type (WT) brain (A) and in a *vnd*^A brain
511 (B). Scale bar = 50 μ m. (C) Number of clones obtained in WT, *vnd*^A, *vnd*⁶ and *vnd* ^{Δ 38}. Full
512 genotypes: (A) *w*^{*} *tubP*-*GAL80* *hsFLP122* *FRT19A*/*w*¹¹¹⁸ *sn* *FRT19A*; *c739*-*GAL4* *UAS*-*mCD8*-*GFP*/+
513 (also control *hsFLP122* in C). (B) *w*^{*} *tubP*-*GAL80* *hsFLP122* *FRT19A*/*y* *vnd*^A *w* *FRT19A*; *c739*-*GAL4*
514 *UAS*-*mCD8*-*GFP*/+ (also *vnd*^A in C). (C) control *hsFLP1*: *w* *tubP*-*GAL80* *hsFLP1* *FRT19A* (from the
515 Bloomington stock 5133)/*w* *UAS*-*mCD8*-*GFP* *sn* *FRT19A*; *c739*-*GAL4* *UAS*-*mCD8*-*GFP*/+. *vnd*⁶: *w*
516 *tubP*-*GAL80* *hsFLP1* *FRT19A*/*y* *vnd*⁶ *w* *UAS*-*mCD8*-*GFP* *sn* *FRT19A*; *c739*-*GAL4* *UAS*-*mCD8*-*GFP*/+.
517 *vnd* ^{Δ 38}: *w* *tubP*-*GAL80* *hsFLP1* *FRT19A*/*vnd* ^{Δ 38} *w* *UAS*-*mCD8*-*GFP* *FRT19A*; *c739*-*GAL4* *UAS*-*mCD8*-
518 *GFP*/+.

519

520 **S6 Fig. In absence of *Appl*⁺ induction, *Appl*^{C2.1}/*Y* flies display a long-term memory defect.** Data
521 presented as mean \pm SEM. n = 15, $F_{(4, 70)} = 17.62$, $P < 0.0001$. Significance level of *t*-test or least
522 significant pairwise comparison following ANOVA: ****p < 0.0001.

523

524

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544 References

- 547 1. Soldano A, Hassan BA. Beyond pathology: APP, brain development and Alzheimer's
548 disease. *Curr Opin Neurobiol.* 2014;27:61-7. Epub 2014/03/19. doi: 10.1016/j.conb.2014.02.003.
549 PubMed PMID: 24632309.
- 550 2. Nicolas M, Hassan BA. Amyloid precursor protein and neural development. *Development.*
551 2014;141(13):2543-8. Epub 2014/06/26. doi: 10.1242/dev.108712. PubMed PMID: 24961795.
- 552 3. Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol*
553 *Med.* 2016;8(6):595-608. Epub 20160601. doi: 10.15252/emmm.201606210. PubMed PMID:
554 27025652; PubMed Central PMCID: PMC4888851.
- 555 4. Soldano A, Okray Z, Janovska P, Tmejova K, Reynaud E, Claeys A, et al. The *Drosophila*
556 Homologue of the Amyloid Precursor Protein Is a Conserved Modulator of Wnt PCP Signaling.
557 *PLoS biology.* 2013;11(5):e1001562. Epub 2013/05/22. doi: 10.1371/journal.pbio.1001562.
558 PubMed PMID: 23690751; PubMed Central PMCID: PMC3653798.
- 559 5. Liu T, Zhang T, Nicolas M, Boussicault L, Rice H, Soldano A, et al. The amyloid precursor
560 protein is a conserved Wnt receptor. *Elife.* 2021;10. Epub 20210909. doi: 10.7554/eLife.69199.
561 PubMed PMID: 34515635; PubMed Central PMCID: PMC48437438.
- 562 6. Lin S. The making of the *Drosophila* mushroom body. *Front Physiol.* 2023;14:1091248.
563 Epub 20230113. doi: 10.3389/fphys.2023.1091248. PubMed PMID: 36711013; PubMed Central
564 PMCID: PMC9880076.
- 565 7. Lee T, Lee A, Luo L. Development of the *Drosophila* mushroom bodies: sequential
566 generation of three distinct types of neurons from a neuroblast. *Development.*
567 1999;126(18):4065-76. PubMed PMID: 10457015.

- 568 8. Luo L, Tully T, White K. Human amyloid precursor protein ameliorates behavioral deficit of
569 flies deleted for *Appl* gene. *Neuron*. 1992;9(4):595-605. Epub 1992/10/01. PubMed PMID:
570 1389179.
- 571 9. Cassar M, Kretschmar D. Analysis of Amyloid Precursor Protein Function in *Drosophila*
572 *melanogaster*. *Front Mol Neurosci*. 2016;9:61. Epub 2016/08/11. doi:
573 10.3389/fnmol.2016.00061. PubMed PMID: 27507933; PubMed Central PMCID:
574 PMC4960247.
- 575 10. Goguel V, Belair AL, Ayaz D, Lampin-Saint-Amaux A, Scaplehorn N, Hassan BA, et al.
576 *Drosophila* amyloid precursor protein-like is required for long-term memory. *J Neurosci*.
577 2011;31(3):1032-7. Epub 2011/01/21. doi: 10.1523/JNEUROSCI.2896-10.2011. PubMed PMID:
578 21248128.
- 579 11. Stepchenko AG, Pankratova EV, Doronin SA, Gulag PV, Georgieva SG. The alternative
580 protein isoform NK2B, encoded by the *vnd/NK-2* proneural gene, directly activates transcription
581 and is expressed following the start of cells differentiation. *Nucleic Acids Res*. 2011;39(13):5401-
582 11. Epub 2011/03/21. doi: 10.1093/nar/gkr121. PubMed PMID: 21422076; PubMed Central PMCID:
583 PMC3141242.
- 584 12. Jimenez F, Martin-Morris LE, Velasco L, Chu H, Sierra J, Rosen DR, et al. *vnd*, a gene
585 required for early neurogenesis of *Drosophila*, encodes a homeodomain protein. *EMBO J*.
586 1995;14(14):3487-95. doi: 10.1002/j.1460-2075.1995.tb07355.x. PubMed PMID: 7628450;
587 PubMed Central PMCID: PMC394416.
- 588 13. Doudna JA, Charpentier E. Genome editing. The new frontier of genome engineering with
589 CRISPR-Cas9. *Science*. 2014;346(6213):1258096. doi: 10.1126/science.1258096. PubMed PMID:
590 25430774.
- 591 14. Port F, Chen HM, Lee T, Bullock SL. Optimized CRISPR/Cas tools for efficient germline and
592 somatic genome engineering in *Drosophila*. *Proc Natl Acad Sci U S A*. 2014;111(29):E2967-76.
593 Epub 2014/07/09. doi: 10.1073/pnas.1405500111. PubMed PMID: 25002478; PubMed Central
594 PMCID: PMC4115528.
- 595 15. Marquilly C, Busto GU, Leger BS, Boulanger A, Giniger E, Walker JA, et al. *Htt* is a repressor
596 of *Abl* activity required for APP-induced axonal growth. *PLoS Genet*. 2021;17(1):e1009287. Epub
597 2021/01/20. doi: 10.1371/journal.pgen.1009287. PubMed PMID: 33465062; PubMed Central
598 PMCID: PMC7845969.
- 599 16. Haelterman NA, Jiang L, Li Y, Bayat V, Sandoval H, Ugur B, et al. Large-scale identification
600 of chemically induced mutations in *Drosophila melanogaster*. *Genome Res*. 2014;24(10):1707-18.
601 doi: 10.1101/gr.174615.114. PubMed PMID: 25258387; PubMed Central PMCID:
602 PMC4199363.
- 603 17. Lee YJ, Yang CP, Miyares RL, Huang YF, He Y, Ren Q, et al. Conservation and divergence of
604 related neuronal lineages in the *Drosophila* central brain. *Elife*. 2020;9. Epub 2020/04/07. doi:
605 10.7554/eLife.53518. PubMed PMID: 32255422; PubMed Central PMCID: PMC7173964.
- 606 18. Chen HM, Huang Y, Pfeiffer BD, Yao X, Lee T. An enhanced gene targeting toolkit for
607 *Drosophila*: *Golic+*. *Genetics*. 2015;199(3):683-94. Epub 2015/01/02. doi:
608 10.1534/genetics.114.173716. PubMed PMID: 25555988; PubMed Central PMCID:
609 PMC4349064.
- 610 19. Lee T, Luo L. Mosaic analysis with a repressible cell marker for studies of gene function in
611 neuronal morphogenesis. *Neuron*. 1999;22(3):451-61. PubMed PMID: 10197526.

- 612 20. Bourdet I, Preat T, Goguel V. The full-length form of the *Drosophila* amyloid precursor
613 protein is involved in memory formation. *J Neurosci*. 2015;35(3):1043-51. doi:
614 10.1523/JNEUROSCI.2093-14.2015. PubMed PMID: 25609621; PubMed Central PMCID:
615 PMCPMC6605540.
- 616 21. Pascual A, Preat T. Localization of long-term memory within the *Drosophila* mushroom
617 body. *Science*. 2001;294(5544):1115-7. PubMed PMID: 11691997.
- 618 22. Palop JJ, Mucke L. Amyloid-beta-induced neuronal dysfunction in Alzheimer's disease:
619 from synapses toward neural networks. *Nat Neurosci*. 2010;13(7):812-8. doi: 10.1038/nn.2583.
620 PubMed PMID: 20581818; PubMed Central PMCID: PMCPMC3072750.
- 621 23. Musiek ES, Holtzman DM. Three dimensions of the amyloid hypothesis: time, space and
622 'wingmen'. *Nat Neurosci*. 2015;18(6):800-6. doi: 10.1038/nn.4018. PubMed PMID: 26007213;
623 PubMed Central PMCID: PMCPMC4445458.
- 624 24. Chu H, Parras C, White K, Jimenez F. Formation and specification of ventral neuroblasts is
625 controlled by *vnd* in *Drosophila* neurogenesis. *Genes Dev*. 1998;12(22):3613-24. doi:
626 10.1101/gad.12.22.3613. PubMed PMID: 9832512; PubMed Central PMCID: PMCPMC317241.
- 627 25. Aso Y, Grubel K, Busch S, Friedrich AB, Siwanowicz I, Tanimoto H. The mushroom body of
628 adult *Drosophila* characterized by GAL4 drivers. *J Neurogenet*. 2009;23(1-2):156-72. Epub
629 2009/01/14. doi: 907683862 [pii]
630 10.1080/01677060802471718. PubMed PMID: 19140035.
- 631 26. Placais PY, de Tredern E, Scheunemann L, Trannoy S, Goguel V, Han KA, et al. Upregulated
632 energy metabolism in the *Drosophila* mushroom body is the trigger for long-term memory. *Nat*
633 *Commun*. 2017;8:15510. Epub 20170605. doi: 10.1038/ncomms15510. PubMed PMID:
634 28580949; PubMed Central PMCID: PMCPMC5465319.
- 635 27. Boulanger A, Clouet-Redt C, Farge M, Flandre A, Guignard T, Fernando C, et al. *ftz-f1* and
636 *Hr39* opposing roles on *EcR* expression during *Drosophila* mushroom body neuron remodeling.
637 *Nat Neurosci*. 2011;14(1):37-44. Epub 2010/12/07. doi: nn.2700 [pii]
638 10.1038/nn.2700. PubMed PMID: 21131955.
- 639 28. McGuire SE, Le PT, Osborn AJ, Matsumoto K, Davis RL. Spatiotemporal rescue of memory
640 dysfunction in *Drosophila*. *Science*. 2003;302(5651):1765-8. PubMed PMID: 14657498.
- 641 29. de Tredern E, Rabah Y, Pasquer L, Minatchy J, Placais PY, Preat T. Glial glucose fuels the
642 neuronal pentose phosphate pathway for long-term memory. *Cell Rep*. 2021;36(8):109620. doi:
643 10.1016/j.celrep.2021.109620. PubMed PMID: 34433052; PubMed Central PMCID:
644 PMCPMC8411112.
- 645 30. Tully T, Quinn WG. Classical conditioning and retention in normal and mutant *Drosophila*
646 *melanogaster*. *J Comp Physiol [A]*. 1985;157(2):263-77. PubMed PMID: 3939242.
- 647

Figure 1

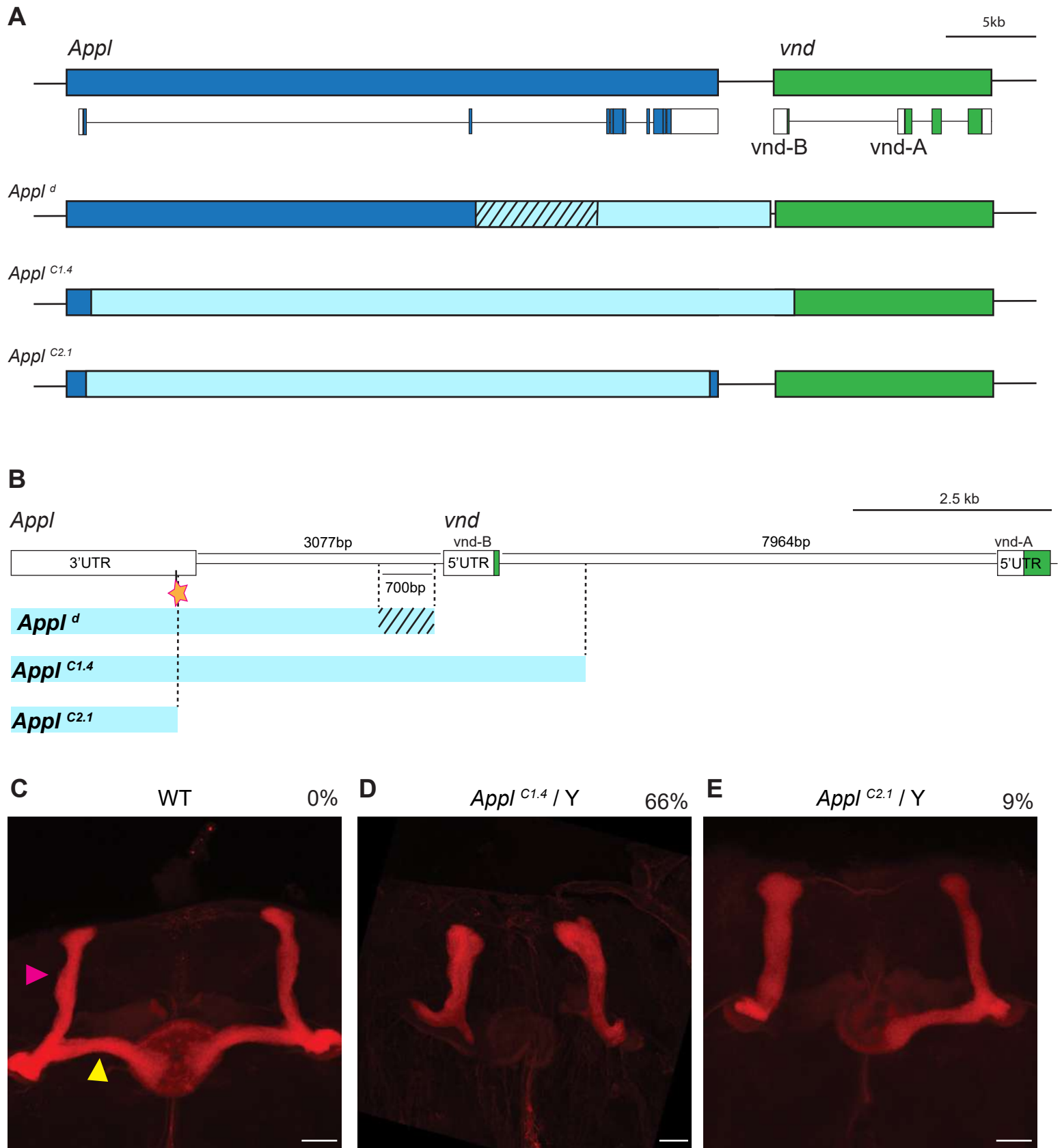


Figure 2

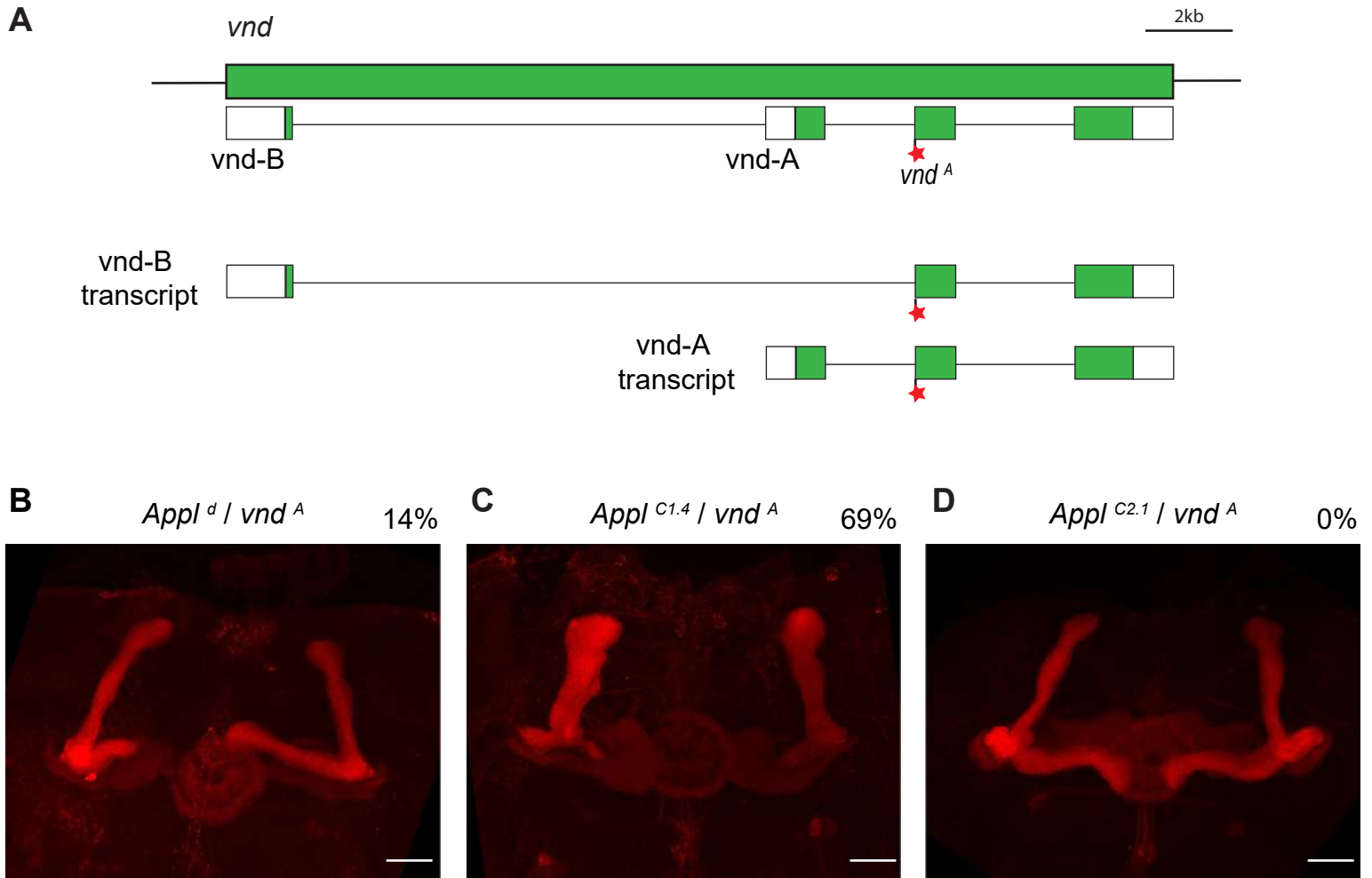
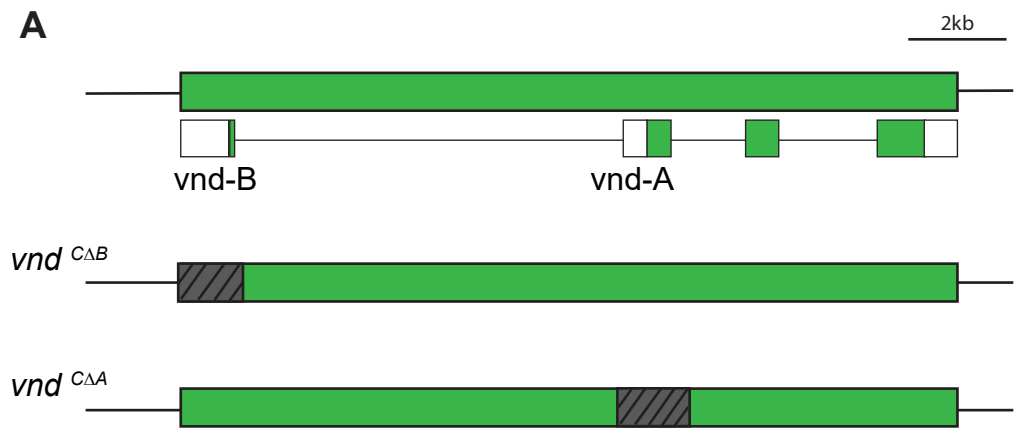
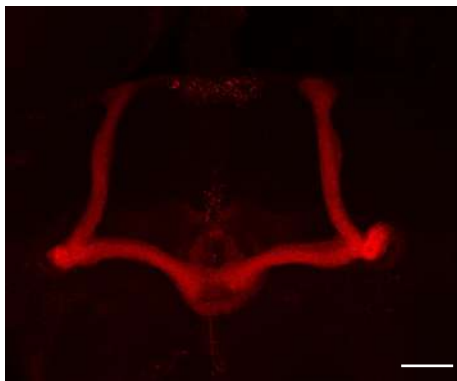


Figure 3



B *vnd*^{CΔB} / Y 1%



C *Appl*^{C1.4} / *vnd*^{CΔB} 0%



Figure 4

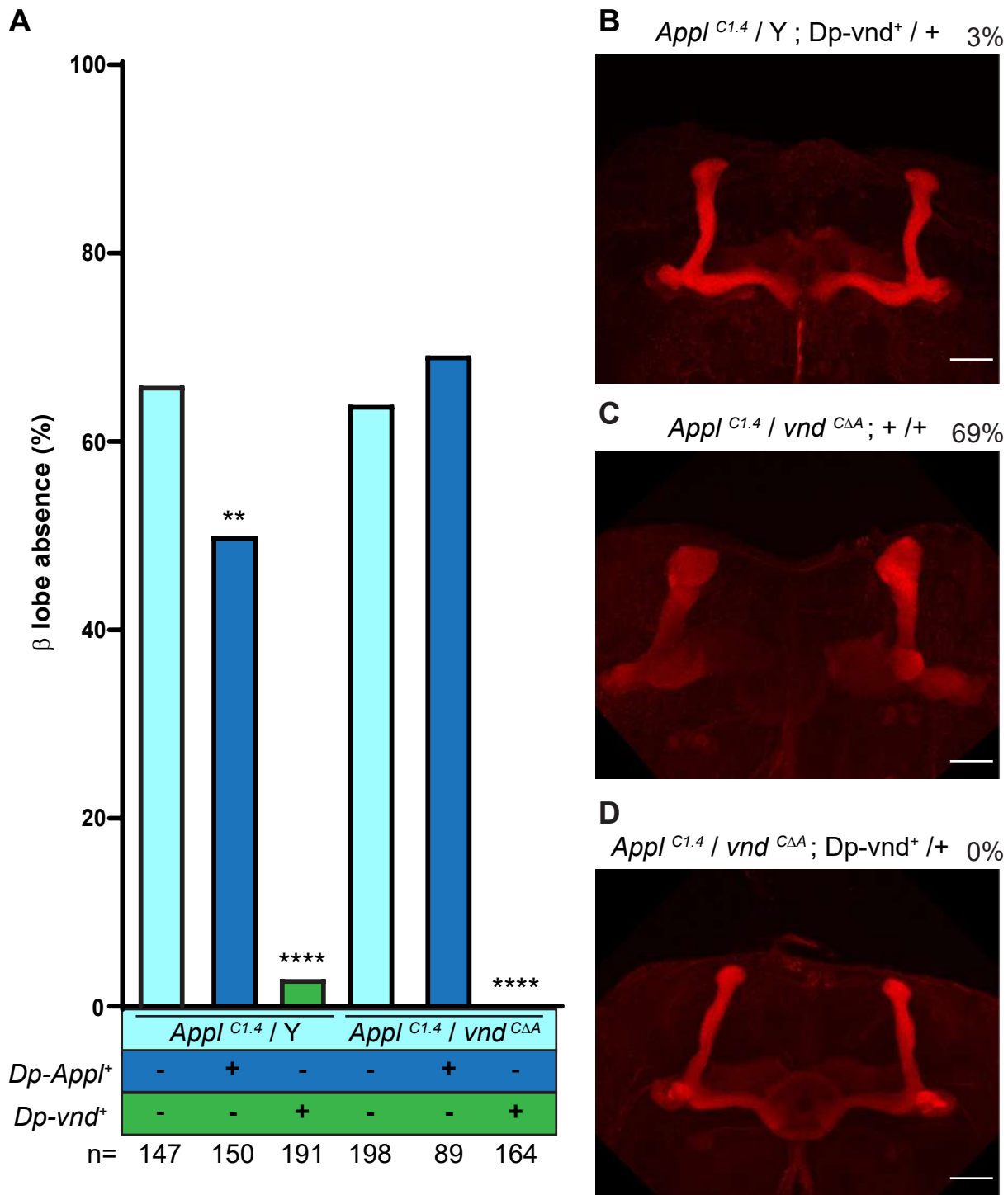


Figure 3

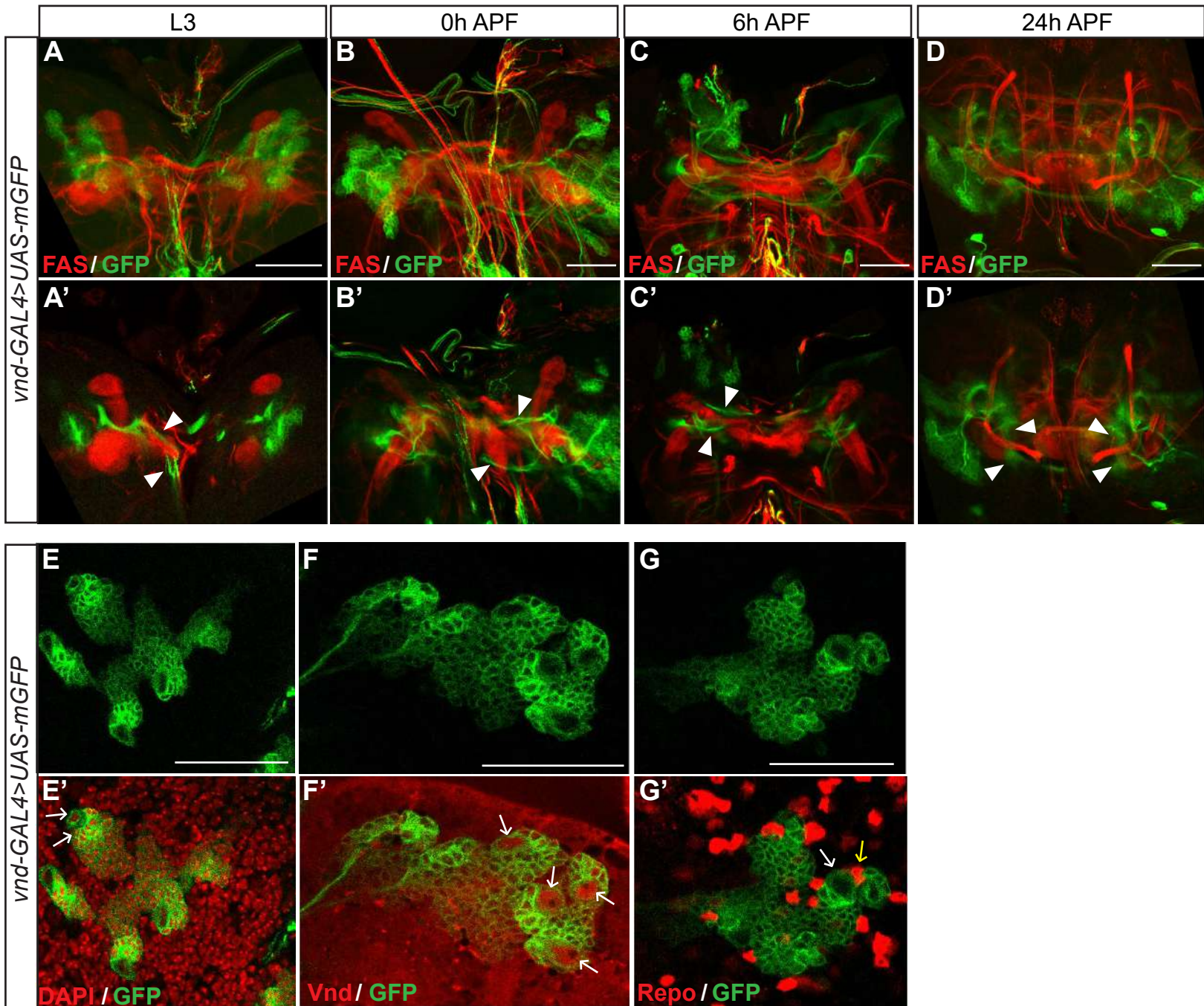


Figure 6

