



HAL
open science

The ROSA-Like Prophage Colonizing *Staphylococcus aureus* Promotes Intracellular Survival, Biofilm Formation, and Virulence in a Chronic Wound Environment

Nour Ahmad-Mansour, Lucile Plumet, Cassandra Pouget, Karima Kissa, Catherine Dunyach-Remy, Albert Sotto, Jean-Philippe Lavigne, Virginie Molle

► To cite this version:

Nour Ahmad-Mansour, Lucile Plumet, Cassandra Pouget, Karima Kissa, Catherine Dunyach-Remy, et al.. The ROSA-Like Prophage Colonizing *Staphylococcus aureus* Promotes Intracellular Survival, Biofilm Formation, and Virulence in a Chronic Wound Environment. *Journal of Infectious Diseases*, 2023, 10.1093/infdis/jiad218 . hal-04250564

HAL Id: hal-04250564

<https://hal.umontpellier.fr/hal-04250564v1>

Submitted on 19 Oct 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

The ROSA-like prophage colonizing *Staphylococcus aureus* promotes intracellular survival, biofilm formation and virulence in a chronic wound environment

Nour Ahmad-Mansour¹, Lucile Plumet¹, Cassandra Pouget², Karima Kissa¹, Catherine Dunyach-Remy², Albert Sotto³, Jean-Philippe Lavigne² and Virginie Molle¹

¹VBIC, INSERM U1047, University of Montpellier, Montpellier, France.

²VBIC, Department of Microbiology and Hospital Hygiene, CHU Nîmes, University of Montpellier, INSERM U1047, 30029 Nîmes, France.

³VBIC, Department of Infectious Diseases, CHU Nîmes, University of Montpellier, INSERM U1047, 30029 Nîmes, France.

Abstract

The transition from colonization to invasion is critical in diabetic foot ulcer (DFU). *S. aureus* can colonize DFU, or invade the underlying tissues, causing serious infections. The ROSA-like prophage has previously been implicated in strain colonization characteristics of *S. aureus* isolates in uninfected ulcers. Here, we investigated this prophage in the *S. aureus* colonizing strain using an *in vitro* chronic wound medium (CWM) mimicking the chronic wound environment. CWM reduced bacterial growth and increased biofilm formation and virulence in a zebrafish model. Moreover, the ROSA-like prophage promoted intracellular survival of *S. aureus* colonizing strain in macrophages, keratinocytes and osteoblasts.

Keywords: Diabetic foot ulcer, colonizing *Staphylococcus aureus*, ROSA-like prophage, chronic wound medium

INTRODUCTION

Chronic wounds such as pressure ulcers, venous/arterial ulcers and diabetic foot ulcers (DFU) cause high morbidity and mortality [1]. *Staphylococcus aureus* is the most frequently isolated bacteria in polymicrobial DFU, leading to delayed wound healing [2]. This species colonizes approximately 30% of the human population and causes various clinical infections [3]. Therefore, it is crucial to differentiate between colonizers and pathogens causing infection in DFU [4–6]. One major difference between colonizing and infecting *S. aureus* strains isolated from DFUs is the presence of a stable prophage, named ROSA-like [7]. The integrated ROSA-like phage increased biofilm formation, reduced virulence, cytotoxicity and the number of viable intracellular bacteria in infected osteoblasts compared to ROSA-like-negative strains [7,8]. However, these studies were conducted under conditions not reproducing the chronic wound environment, which could affect *S. aureus* colonizing strain adaptation. In this study, we assessed whether the ROSA-like prophage was involved in the persistence and adaptive ability of *S. aureus* to the stressful conditions of DFU using an *in vitro* medium that closely mimics the chronic wound microbiological, cellular, and inflammatory environment [1].

METHODS

Bacterial strains, media and growth conditions. *S. aureus* DFU clinical isolate NSA1385 and its isogenic ROSA-like (Δ rosa) excision variant were previously published [5]. Briefly, NSA1385 was identified in an uninfected DFU, which did not progress to infection. The ROSA-like ejection variant (Δ rosa) was produced after iron-restricted growth. Sequencing demonstrated that the only difference between the two strains was the deletion of this phage [7]. *S. aureus* strains were grown in Tryptic Soy Broth (TSB) medium or Chronic Wound Medium (CWM) at 37°C and 225 rpm. We have previously described the *in vitro* CWM [1], comprising 79.5% Bolton broth, 0.5% hemolyzed human blood and 20% heat inactivated human serum, with 1.10⁶/mL of human

45 keratinocyte debris (HaCaT cells), buffered to pH 8.0. Bacterial growth was observed using a microplate reader
46 (Tecan, Lyon, France).

47 **Kinetics of early biofilm formation.** Biofilm Ring Test® (BioFilm Control, Saint Beaulieu, France) was used
48 according to the manufacturer's instructions to evaluate early biofilm formation. *S. aureus* strains were grown
49 at 37 °C for 6 h in BHI or in CWM. Standardized to an OD600 of 1.00 ± 0.05 , the bacterial suspension was diluted
50 1:250 to 4.10^6 CFU/mL. 1% magnetic beads were added to the bacterial suspension in BHI or CWM. 200 μ L were
51 added to a 96-well microplate in triplicate and incubated for 3 h at 37°C. The microplate was placed on a
52 magnetic block for 1 min and scanned with a custom plate reader. The BFCE3 software calculated adhesion
53 strength from images before and after magnetic attraction. A low BFI value (<2) indicates complete
54 immobilization of beads due to sessile cells, while a high value (>7) indicates high mobility of beads under
55 magnet action, designating no biofilm formation.

56 **Zebrafish killing assay.** The zebrafish model was employed for bath immersion infection to assess *S. aureus*
57 virulence. NSA1385 and Δ rosa strains were grown in TSB or CWM overnight at 37°C. Cultures were diluted,
58 centrifuged, and reconstituted in fish water at 2.10^7 bacteria/mL. Embryos dechorionated at 48 hpf were bath
59 immersed and infection was performed as previously described [9]. Bath immersion was performed on injured
60 embryos 2 days after fertilization while the mouth was closed and the viability monitored for 24 h.

61
62 **Cells infecting and viability assay.** Intracellular bacterial growth and cytotoxicity during macrophage (RAW
63 264.7), keratinocyte (HaCaT) and osteoblast (MC3T3-E1) infections were quantified as previously described [9].
64 Briefly, *S. aureus* strains were grown in TSB or CWM broth to mid-exponential phase growth prior to infection.
65 In 24-well plates, RAW 264.7 cells (MOI 20:1), HaCaT cells (MOI 100:1), and MC3T3-E1 (MOI 100:1) were
66 infected with *S. aureus* NSA1385 and Δ rosa strains, then incubated for 1 h for RAW 264.7 cells, or 1h30 for HaCaT
67 and MC3T3-E1 cells, at 37°C and 5% CO₂. Cells were washed and the extracellular bacteria killed by gentamicin
68 treatment (100 μ g/mL) for 60 min and cells were washed twice with PBS (T0) and incubated in fresh medium
69 with 5 μ g/mL lysostaphin for 5, 24, and 48 h. Intracellular bacteria were counted by lysing infected cells with
70 0.1 Triton X-100 in PBS. Lactate dehydrogenase (LDH) activity in cell culture supernatant was measured to
71 quantify cytotoxicity using the CyQuant LDH Cytotoxicity Assay Kit (Thermo Fisher Scientific, Rockford, IL,
72 USA) according to the manufacturer's instructions.

73
74 **Statistical Analysis.** GraphPad software package Prism 9.5.5 was used to estimate the statistical significance of
75 differences across groups, which is indicated in the relevant figure legends.

76 **Ethical statement.** All animal experiments were conducted in conformity with European Union guidelines for
77 the care and use of laboratory animals at the University of Montpellier as previously described [9].

79 RESULTS

80 ROSA-like prophage affects *S. aureus* growth, early biofilm formation and virulence in Chronic Wound 81 Media

82 The bacterial growth of *S. aureus* NSA1385 and Δ rosa strains was compared in CWM versus TSB standard
83 medium. *S. aureus* NSA1385 and Δ rosa growth curves showed no significant differences in TSB (Figure 1A).
84 However, both strains grew slower in CMW in comparison to TSB, with a significant delay for Δ rosa in CWM
85 compared to NSA1385 ($p < 0.001$) (Figure 1A). Therefore, the ROSA-like prophage is directly involved in the
86 growth rate delay in a chronic wound environment.

87 Next, we assessed the impact of the ROSA-like prophage on early biofilm formation using the Biofilm Ring test.
88 *S. aureus* NSA1385 formed a biofilm faster than the Δ rosa variant in both media and even faster in CWM than
89 in the classical BHI medium (BFI = 9 ± 0.2 vs. 11 ± 0.1 and 12 ± 0.2 vs. 16 ± 0.3 , for NSA1385 and Δ rosa, respectively)
90 ($p < 0.01$). The difference remained significant at 2 h and 3 h ($p < 0.001$), confirming the role of the ROSA-like
91 prophage in promoting biofilm formation and especially in a chronic wound environment (Figure 1B).

92 The impact of the ROSA-like prophage on bacterial virulence after growth in CWM was studied in the zebrafish
93 model [9]. Strains grown in TSB showed over 90% mortality with Δ rosa as early as 10 hpi, while 70% of embryos
94 survived following NSA1385 infections at 21 hpi. Interestingly, opposite results were observed after growth in
95 CWM, as larvae infected with NSA1385 were all dead at 10 hpi, while embryos infected with Δ rosa survived
96 until 21 hpi. The control group showed no mortality (Fig 1C). Therefore, the ROSA-like prophage increases
97 virulence in a chronic wound environment.

99 **ROSA-like phage is involved in the intracellular persistence of *S. aureus* in chronic wound environment.**

100 *S. aureus* survival within phagocytic and non-phagocytic cells was analyzed in RAW 264.7 murine
101 macrophages, MC3T3-E1 murine osteoblasts, and HaCat human keratinocytes grown in TSB or CWM.

102 RAW 264.7 macrophages were used to phagocytose *S. aureus* and viable bacteria were recovered at 5 h and 24
103 h post-Gentamicin treatment (pGt). At T0, *S. aureus* NSA1385 and Δ rosa had similar numbers of phagocytosed
104 bacteria, indicating that neither the ROSA-like prophage nor the CWM affected macrophage phagocytosis
105 (Figure 2A). However, the Δ rosa strain had a higher viable intracellular bacterial count than NSA1385 in both
106 media at T0, 5 and 24 h pGt suggesting that the ROSA-like prophage was involved in decreasing *S. aureus*
107 intramacrophagic survival. Interestingly, intracellular *S. aureus* NSA1385 significantly enhanced its
108 intramacrophagic survival when cultured in CWM versus TSB ($p < 0.01$), indicating that the ROSA-like prophage
109 was critical for *S. aureus* colonizing strain adaptation to macrophage survival in chronic wound environment
110 (Figure 2A).

111 We used the LDH assay to determine whether the higher survival of *S. aureus* NSA1385 in CWM was related to
112 a lower cytotoxicity of the infected macrophage cells. Infection with *S. aureus* NSA1385 cultured in TSB or CWM
113 enhanced LDH release over time in infected Raw 264.7 macrophages (Figure 2B). However, NSA1385 generated
114 less LDH when grown in CWM versus TSB ($p < 0.01$). This suggested that *S. aureus* NSA1385 grown in CWM
115 had a distinct intracellular fate versus TSB growth, as bacteria survived for extended periods within
116 macrophages, associated with lower levels of cytotoxicity. Therefore, the inability of macrophages to eliminate
117 intracellular *S. aureus* NSA1385 in a chronic wound environment may be a serious defect of the host innate
118 immunity, leading to intracellular reservoirs of colonizing *S. aureus*. Moreover, the ROSA-like prophage
119 affected macrophage cytotoxicity by decreasing LDH over time in Δ rosa-infected cells grown in CWM versus
120 TSB ($p < 0.001$) (Figure 2B).

121 In non-phagocytotic cells, *S. aureus* is known to proliferate and persist intracellularly [10]. To test whether the
122 chronic wound environment influenced survival with ROSA-like prophage in infected osteoblasts and
123 keratinocytes, we allowed osteoblast cells to phagocytose *S. aureus* and viable bacteria were recovered at T0, 5
124 and 24 h pGt. At T0, *S. aureus* NSA1385 had significantly more phagocytosed bacteria compared to Δ rosa in
125 both media ($p < 0.001$), indicating that the ROSA-like prophage increased osteoblast phagocytosis independently
126 of the CWM (Figure 2C). In comparison to the NSA1385 strain, the Δ rosa strain had a higher percentage of
127 infecting inoculum that persisted within osteoblasts at T5h pGt ($p < 0.01$), with the difference becoming more
128 pronounced at T24 ($p < 0.001$) (Figure 2C). Interestingly, this behavior was associated with impairment of
129 NSA1385 to multiply and persist within osteoblasts over time, despite higher phagocytosis. In contrast, the
130 Δ rosa variant was unaffected, and internalized Δ rosa CFUs grew normally [8]. However, growth of *S. aureus*
131 NSA1385 in CWM increased its survival at T5h versus in TSB ($p < 0.05$), but by T24h no difference was observed
132 in intracellular counts (Figure 3C). However, intracellular bacteria of the Δ rosa strain grown in CWM were
133 significantly lower over time in comparison to TSB ($p < 0.01$), suggesting that the CWM reduced Δ rosa
134 intracellular survival. Overall, these results indicated that the ROSA-like prophage decreased the intracellular
135 survival of the NSA1385 colonizing strain independently of the chronic wound environment, but the Δ rosa
136 variant was sensitive to this environment. Moreover, an increased cytotoxic response over time against
137 osteoblasts was observed with NSA1385 strain grown in TSB and CWM, in accordance with their increased
138 intracellular survival (Figure 2D). The non-phagocytic human keratinocytes cell line HaCaT showed similar
139 results (Figure 2E,F). Overall, our data indicated that in non-phagocytic cells, the CWM had no impact on
140 phagocytosis defect but promoted intracellular survival of the NSA1385 strain and increased its related
141 cytotoxicity.

142

143 Discussion

144 In DFU, *S. aureus* experiences stress conditions including increased glucose concentration, lower
 145 temperature, lower tissue oxygenation, and exposure to antibiotics [11]. Environmental stresses can cause
 146 cellular damage in bacteria and impair macromolecule function, essential for survival and growth [12].
 147 Prophages provide numerous advantages to endure these challenging conditions [13]. Here, we reported that
 148 the ROSA-like prophage of *S. aureus* colonizing strain, isolated from DFU, promoted bacterial growth, biofilm
 149 formation, persistence and virulence in chronic wound environment.

150 Most *S. aureus* isolates carry multiple prophages, affecting virulence, toxin production, immune evasion, and
 151 host preference. These genetic elements carry environmentally regulated adaptive genes [14]. For instance,
 152 prophages in *Vibrio parahaemolyticus* improve ultraviolet sensitivity, DNA methylase activity, quorum sensing,
 153 and resistance to environmental stress [12]. Integration of phage Min27 into *Escherichia coli* genome increases
 154 swimming motility, allowing bacteria to grow faster, move to high-nutrient areas and avoid harmful
 155 environments [15]. Moreover, removing all nine prophages from *E. coli* reduced resistance to oxidative, osmotic,
 156 and acid stress [13]. Here, ROSA-like prophage improved bacterial growth in CWM, as well as rapid biofilm
 157 formation, consistent with the NSA1385 colonizing phenotype. In addition, larvae infected with NSA1385
 158 grown in CWM had increased mortality rate, confirming the impact of DFU environment on ROSA-like
 159 prophage's role in bacterial virulence. Furthermore, studies have shown that prophages help negative
 160 regulation of cellular inflammatory response, improving bacterial survival in the host through phagocytosis
 161 evasion [12]. Interestingly, our findings showed no difference for the colonizing strain's phagocytosis. We thus
 162 hypothesized that the mechanisms for *S. aureus* proliferation are either to multiply and hide in cells without
 163 killing them, or to improve other infecting strains' ability to evade the immune system and infiltrating deeper
 164 tissues. However, while these data point to virulence factors being influenced by environmental stress, a study
 165 on the colonizing strain pre-cultivated in a wound-like medium versus infecting strains isolated from DFU
 166 found no difference in any of the studied genes [11]. Only after a long exposure did the colonizing strain
 167 NSA1385 increase *fnbpA* gene expression and decrease *hla* gene expression [11]. Better understanding the role
 168 of the ROSA-like prophage in *S. aureus* behavior under stress will help develop new strategies to prevent and
 169 treat infections.

171 Notes

172 *Financial support.* N.A.M. received a PhD grant from InfectioPôle Sud Méditerranée 2020; grant number AO-
 173 IHU-2020-3. C.P., C.D.R., A.S. and J.P.L. were supported by the University Hospital of Nîmes. This University
 174 hospital provided its structural, human, and financial support through the award obtained by our team during
 175 the internal call for tenders "Thématiques phares". We thank Sarah Kabani for editing the manuscript.

176 *Potential conflicts of interest.* The authors declare no conflict of interest.

178 References

- 179 1. Pouget C, Dunyach-Remy C, Bernardi T, et al. A Relevant Wound-Like in vitro Media to Study Bacterial
 180 Cooperation and Biofilm in Chronic Wounds. *Front Microbiol* [Internet]. 2022 [cited 2023 Jan 10]; 13. Available
 181 from: <https://www.frontiersin.org/articles/10.3389/fmicb.2022.705479>
- 182 2. Durand BARN, Pouget C, Magnan C, Molle V, Lavigne J-P, Dunyach-Remy C. Bacterial Interactions in
 183 the Context of Chronic Wound Biofilm: A Review. *Microorganisms*. Multidisciplinary Digital Publishing
 184 Institute; 2022; 10(8):1500.
- 185 3. Lavigne J-P, Hosny M, Dunyach-Remy C, et al. Long-Term Intrahost Evolution of *Staphylococcus aureus*
 186 Among Diabetic Patients With Foot Infections. *Front Microbiol* [Internet]. 2021 [cited 2023 Jan 14]; 12. Available
 187 from: <https://www.frontiersin.org/articles/10.3389/fmicb.2021.741406>
- 188 4. Sotto A, Richard J-L, Jourdan N, et al. Miniaturized oligonucleotide arrays: a new tool for
 189 discriminating colonization from infection due to *Staphylococcus aureus* in diabetic foot ulcers. *Diabetes Care*.
 190 2007; 30(8):2051–2056.
- 191 5. Sotto A, Lina G, Richard J-L, et al. Virulence Potential of *Staphylococcus aureus* Strains Isolated From
 192

- 193 Diabetic Foot Ulcers: A new paradigm. *Diabetes Care*. **2008**; 31(12):2318–2324.
- 194 6. Sotto A, Richard J-L, Messad N, et al. Distinguishing colonization from infection with *Staphylococcus*
195 *aureus* in diabetic foot ulcers with miniaturized oligonucleotide arrays: a French multicenter study. *Diabetes*
196 *Care*. **2012**; 35(3):617–623.
- 197 7. Messad N, Prajsnar TK, Lina G, et al. Existence of a Colonizing *Staphylococcus aureus* Strain Isolated in
198 Diabetic Foot Ulcers. *Diabetes*. **2015**; 64(8):2991–2995.
- 199 8. Rasigade J-P, Dunyach-Rémy C, Sapin A, et al. A Prophage in Diabetic Foot Ulcer–Colonizing
200 *Staphylococcus aureus* Impairs Invasiveness by Limiting Intracellular Growth. *J Infect Dis*. **2016**; 214(10):1605–
201 1608.
- 202 9. Ahmad-Mansour N, Plumet L, Huc-Brandt S, et al. Investigating Pathogenicity and Virulence of
203 *Staphylococcus pettenkoferi*: An Emerging Pathogen. *Int J Mol Sci*. Multidisciplinary Digital Publishing Institute;
204 **2021**; 22(24):13614.
- 205 10. Josse J, Velard F, Gangloff SC. *Staphylococcus aureus* vs. osteoblast: relationship and consequences in
206 osteomyelitis. *Front Cell Infect Microbiol* [Internet]. **2015** [cited 2021 Mar 17]; 5. Available from:
207 <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4660271/>
- 208 11. Pouget C, Gustave C-A, Ngba-Esebe C, et al. Adaptation of *Staphylococcus aureus* in a Medium
209 Mimicking a Diabetic Foot Environment. *Toxins*. Multidisciplinary Digital Publishing Institute; **2021**; 13(3):230.
- 210 12. Pazhani GP, Chowdhury G, Ramamurthy T. Adaptations of *Vibrio parahaemolyticus* to Stress During
211 Environmental Survival, Host Colonization, and Infection. *Front Microbiol*. **2021**; 12:737299.
- 212 13. Wang X, Kim Y, Ma Q, et al. Cryptic prophages help bacteria cope with adverse environments. *Nat*
213 *Commun*. Nature Publishing Group; **2010**; 1(1):147.
- 214 14. Wendling CC, Refardt D, Hall AR. Fitness benefits to bacteria of carrying prophages and prophage-
215 encoded antibiotic-resistance genes peak in different environments. *Evol Int J Org Evol*. Wiley-Blackwell; **2021**;
216 75(2):515.
- 217 15. Li D, Qian X, Liu X, et al. orf6 and orf10 in Prophage phiv142-3 Enhance the Iron-Acquisition Ability
218 and Resistance of Avian Pathogenic *Escherichia coli* Strain DE142 to Serum. *Front Vet Sci* [Internet]. **2020** [cited
219 2023 Feb 27]; 7. Available from: <https://www.frontiersin.org/articles/10.3389/fvets.2020.588708>
- 220

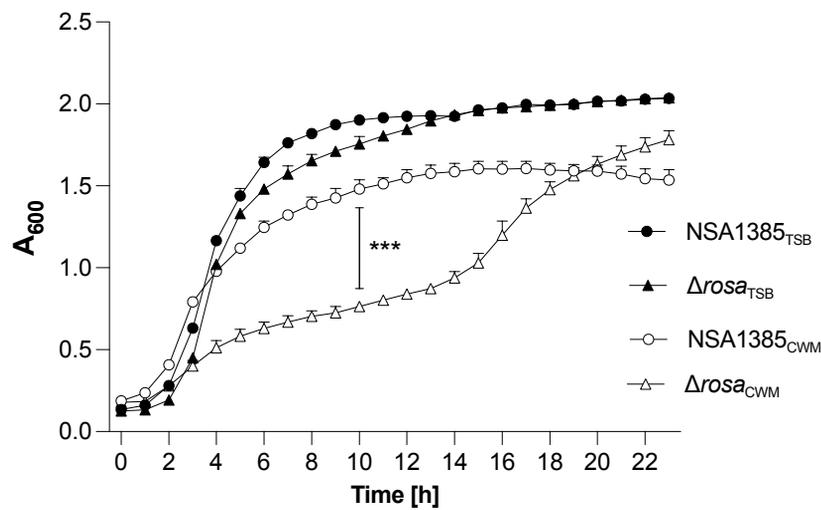
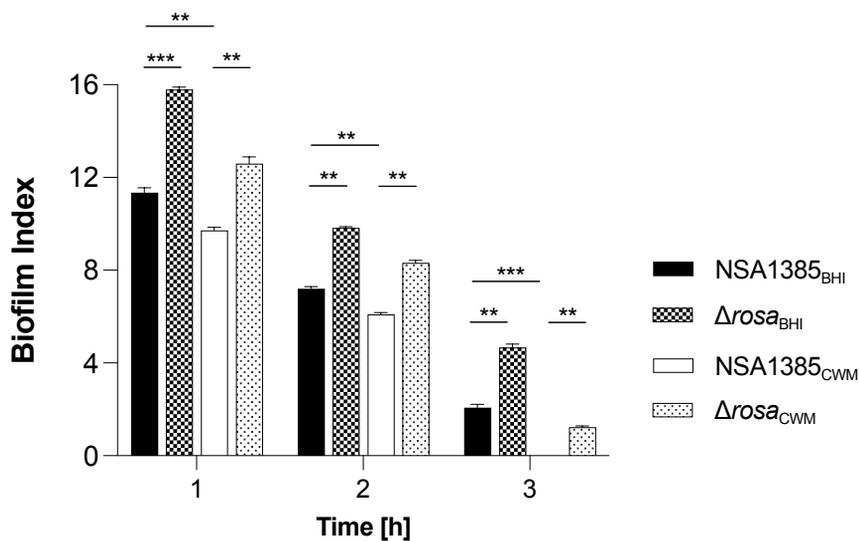
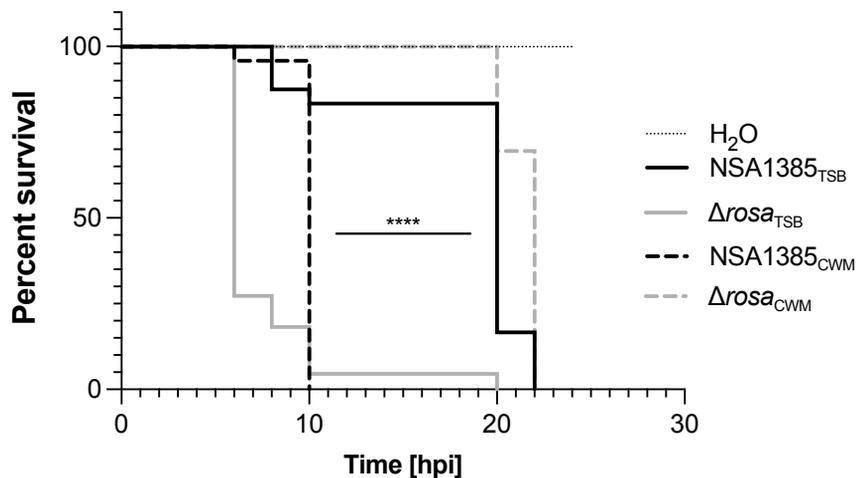
A**B****C**

Figure 1. Growth, biofilm formation and virulence of *S. aureus* in chronic wound media (CWM) (A) Growth kinetics of NSA1385 (black circles) and Δ *rosa* (black triangles) strains in TSB; NSA1385 (white circles) and Δ *rosa* (white triangles) strains in CWM. At each time point ($n=3$), the data show the mean A_{600} readings SD. Welsh's t test, *** $p < 0.001$. (B) Kinetics of biofilm formation determined by BioFilm Ring Test[®]. Comparisons were performed with t -test, *** $p < 0.001$, ** $p < 0.01$. (C) Kaplan–Meier representation of the survival of embryos injured in the tail fin at 48 h post-fertilization (hpf) in a bath infected with *S. aureus* strains at 2.10^7 CFU/mL grown in TSB or CWM or “fish water” (negative control). The proportion of surviving embryos ($n=24$ for each, indicative of five separate experiments) is used to express the results.

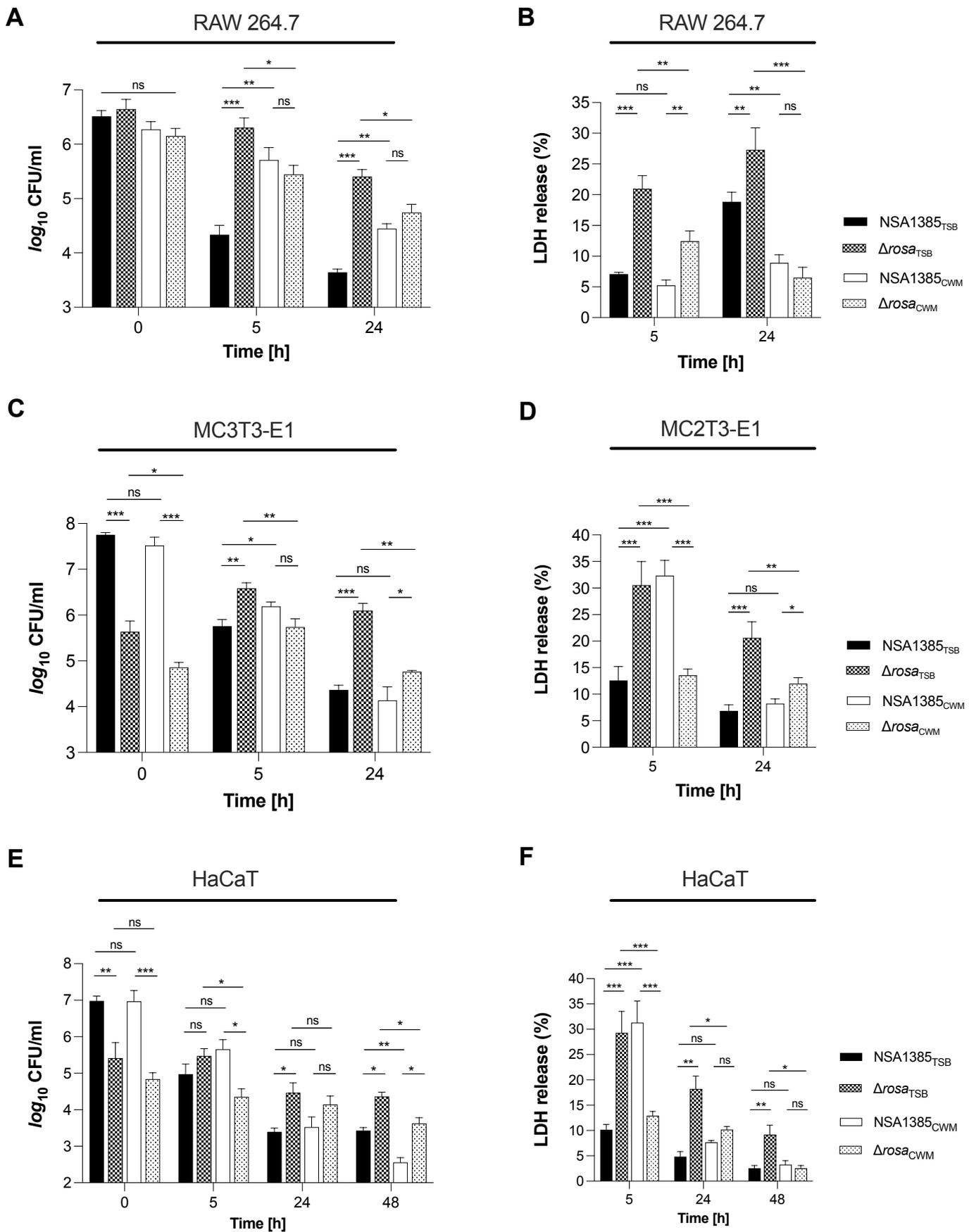


Figure 2. *S. aureus* survival in infected cells and influence of chronic wound environment. NSA1385_{TSB} (black), Δ rosa_{TSB} (dotted black), NSA1385_{CWM} (white) and Δ rosa_{CWM} (dotted white) bacteria were used to infect RAW 264.7 macrophages (A), MC2T3-E1 osteoblasts (C) and HaCaT keratinocytes cells (E). The average and standard deviation (SD) of five different experiments are represented. (B, D, and F) LDH release was measured using the CyQUANT assay kit. Data are expressed relative to the 100% positive control ($n=3$ biological repeats). A two-way ANOVA test was used to establish statistical significance, with * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; ns, not significant.