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Hydrophobic Dye Solubilization via Hybrid Imogolite Nanotube probed by Second Harmonic Scattering

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Abstract: This article explores the organization and interactions of Disperse Orange 3 (DO3) hydrophobic dye molecules within hybrid organic-inorganic imogolite nanotubes. In pure water, the DO3 dye molecules self assemble into large insoluble 2D nanosheet whose structure is also explored by molecular dynamic simulations. The dye molecules are however efficiently solubilized when in presence of hybrid imogolite nanotubes. The filling of the internal hydrophobic cavity of the nanotubes is quantified. The organization of the molecules inside the nanotube is probed by the polarization resolved Second Harmonic Scattering (SHS) technique coupled with simulation. At the highest loading, the dyes fill the nanotube with their principal axis parallel to the nanotube walls showing a strong SHS signal due to this encapsulation.

1 Introduction

Nonpolar molecules in an aqueous solvent create perturbation in the water-water cohesion forces. An effective force acts to minimize this perturbation *i.e.* the amount of surface exposed to water. This force, often called hydrophobic effect is of fundamental importance in chemical and biological system¹⁻³. At the molecular level, the interaction between hydrophobic molecules in an aqueous solution drives their relative solubility and their ability

to self-assembly. The self-assembly process leads to dimer formation and sometimes even higher order aggregates⁴. Different ways can be used to increase the solubility of hydrophobic molecules: (i) the addition of micellar amphiphilic molecules i.e., surfactants⁵, (ii) the use of a water soluble organic co-solvent like alcohol⁶ and (iii) the use of host-guest interaction by supramolecular structure like macrocycles⁷. Among the various hollow nanostructures like carbon nanotubes, halloysite, zeolite and so on, hybrid Imogolite Nanotube (IMO-CH₃), is one of the rare, if not the only, inside-out janus nanotubular structure obtained in a one pot synthesis procedure. IMO-CH₃, has a composition from the outside to the inside of the nanotube given by the formula (OH)₃Al₂O₃SiCH₃. It has thus an internal surfaces covered by Si-CH₃ groups⁸ while the external surface remains hydrophilic. IMO-CH₃ nanotubes are thus forming dispersion of monodisperse single digit hydrophobic pores (~1.8 nm) in water. This material has demonstrated its ability to encapsulate organic molecules^{9, 10} thanks to the hydrophobic effect which pushes insoluble molecules inside the hydrophobic cavity. IMO-CH₃ may thus act as an hydrophobic nano-reactor¹¹ for various molecules.

Recently, a new optical type of measurement has shown promising results to probe at the molecular level supramolecular organization¹²⁻¹⁴ and self-assembly process^{15, 16}. This technique is based on the nonlinear optical process of Second Harmonic Scattering (SHS), a nonlinear optical phenomenon involving the conversion of two photons at the fundamental frequency ω into one photon at the harmonic frequency 2ω . This nonlinear optical technique combines the ability to probe in situ a colloidal suspension^{17, 18} and a high sensitivity about the geometry of a supramolecular organization. We proposed in this work to apply this technique in combination with molecular dynamics calculations to probe the solvation, organization and self-assembly process of a hydrophobic azo dye named Disperse Orange 3 and referred in the following as DO3. This molecule exhibits strong intermolecular van der Waals like attractive forces because of its large permanent dipoles¹⁹ and also presents a large second harmonic response because of its push-pull character²⁰. The main objective of the present paper is to present how SHS technique is able to bring microscopic information about the organization of this dye in aqueous solution. This study is divided in two parts. First, we present the study of the dye self-assembly in different water-ethanol mixture. Second, the encapsulation of the dye within IMO-CH₃ nanotubes suspension is presented and discussed.

2 Materials and methods

2.1 Solution preparation

The Disperse Orange 3 (DO3) is obtained from Sigma Aldrich at a purity of 96%. Absolute ethanol is from Sigma Aldrich. 18 M Ω .cm water (MilliQ) is used. A stock solution of DO3 in ethanol at 1mM is prepared and equilibrated during 24 h. From this stock solution, different samples containing 0.05 mM of DO3 are prepared and equilibrated at various ethanol-water ratio prior to perform SHS measurements. For the solution in pure water, the preparation slightly differs: a fraction of the stock solution in ethanol is evaporated and the residue was redispersed in water.

The IMO-CH₃ nanotube are synthesized according to the following procedure^{8, 21, 22} : around 14 g of aluminum-tri-sec-butoxide (ASB) was added into a Teflon reactor containing 800 ml of an HCl solution to obtain an Al/H molar ratio of 2. The mixture was left under stirring for around one hour. The Si precursor, trimethoxy(methyl)silane (TMMS), was added drop by drop to reach a 0.6 molar ratio of Si/Al. The slight excess of Si precursor was shown to hinder the formation of aluminum hydroxide particles during synthesis. The whole mixture was stirred for a few more hours before it was placed in an oven at 90°C for 5 days. The resulting product was dialyzed using 10 kDa dialysis membranes against MilliQ water for several days to reach an external conductivity below 10 $\mu\text{S}\cdot\text{cm}^{-1}$. IMO-CH₃ concentration, in the resulting solution, is around 6.5 g/L. The suspension was diluted and added to DO3 solution to reach a final concentration of 0.25 g/L.

2.2 SHS measurements

The polarization resolved SHS experimental setup is detailed elsewhere²³. The SHS intensity is monitored in the right-angle direction as a function of the input polarization angle α , see Figure 1, which was selected with a rotating half-wave plate. The second harmonic light was selected in the Γ state by an analyzer placed in front of the spectrometer either in the vertical or horizontal state referred respectively as Vout and Hout states. All experimental data were recorded for 10 s under stirring conditions; a magnetic stirrer is operated in the cell measurement. In all the experimental polar plots, the SHS Intensity of the solvent has been removed according:

$$I_{SHS}(\alpha, \Gamma) = I_{suspension}(\alpha, \Gamma) - I_{solvent}(\alpha, \Gamma) \quad (1)$$

Where $I_{suspension}$ is the intensity of the suspension, ie the dye in the solvent with or without imogolite, and $I_{solvent}$ is the intensity of the solvent mixture alone. The experimental polarization plots are analyzed and fitted with the Fourier model²⁴:

$$I_{SHS}(\alpha, \Gamma) = K_{SHS}^{\Gamma} (1 + I_{2,SHS}^{\Gamma} \cos(2\alpha) + I_{4,SHS}^{\Gamma} \cos(4\alpha)) \quad (2)$$

Here K_{SHS}^{Γ} is a constant, $I_{2,SHS}^{\Gamma}$ and $I_{4,SHS}^{\Gamma}$ are the amplitudes of the second and fourth harmonics in the series, respectively. The parameter $I_{2,SHS}^V$ is related to the local microscopic structure, i.e. the first hyperpolarizability of the emitter and $I_{2,SHS}^H$, $I_{4,SHS}^V$ to the long-range correlations. In the case of uncorrelated species, the SHS signal is a purely incoherent phenomenon and these amplitudes vanishes²⁴. On the contrary, when dye orientations are correlated, the scattered photons have a well-defined phase relationship and these coefficients differ from zero.

2.3 Molecular Dynamics simulations

Force fields were based on the GROMOS 54A7 parameter set²⁵. Topology files for DO3 and ethanol were created using the Automated Topology Builder²⁶ and are publicly available under the molecule ID 1197414 and ID 902261 respectively. The Extended Single Point Charge SPC/E water model was used. Simulations were carried out using GROMACS software²⁷ in the NPT ensemble using a Nose-Hoover thermostat and a Parrinello-Rahman barostat. Temperature was set to 298.15 K and pressure was maintained at 1 atmosphere. The LINear Constraint Solver (LINCS) method²⁸ was applied to constrain all bond lengths. The time step was 2fs, and electrostatic interactions were calculated using Particle-Mesh-Ewald summation. Each Radial Distribution Function (RDF) presented in Figure 2 is calculated on 20 ns production runs by the gmx rdf function available in the GROMACS package. The RDF functions are calculated between the N-N intermolecular atom pairs as depicted in Figure 2. A first set of MD calculation is performed on 8 nm sized box with 6 molecules of DO3 and different ethanol-water ratio. A second calculation has been done at high dye density with 40 dyes molecules in the same 8 nm sized box filled with water solvent.

2.4 Polarization Resolved SHS modelling

The computational program PySHS^{29, 30} has been used to calculate the polarization plots of different supramolecular organizations. The inputs of this program are the hyperpolarizability of the DO3 molecule and the position and orientation of each molecules in the supramolecular aggregate. The DO3 molecule is assumed to be a 1D molecule, which means that only the hyperpolarizability component β_{zzz} is considered. Here the z axis corresponds to the principal axis of the molecule. The position and orientation of dye in the aggregate are described for each kind of aggregate considered. All the input parameters required to perform a computation are given in the supporting informations (SI). In order to establish a direct comparison with the experimental data, the predicted polarization plots are normalized with the number of dyes in the aggregate and multiply by the experimental $I(0^\circ, V_{out})$ intensity of the 0.05 mM DO3 in ethanol solution. In such a way, the different predicted polarization plots can be put in regards and directly compared to the experimental data.

3 Results and Discussion

3.1 The DO3 behavior in water ethanol mixture

Figure 1 presents the evolution of the polarization resolved SHS Intensity as a function of the input polarization angle α for a solution of DO3. The different colors are related to increasing the water ratio in the solvent mixture. The blue curves, obtained with ethanol as mono solvent, exhibits the classical shape of randomly oriented molecules, well dispersed in the medium, with all the parameters I_4^V , I_2^H , I_4^H close to zero. The depolarization ratio define as the ratio of $I(0^\circ, V_{out})$ to $I(0^\circ, H_{out})$, is found to be 5.1 confirming the assumption that only the hyperpolarizability component β_{zzz} have to be considered^{31, 32}. When the water ratio increases in the mixture, the patterns of the polarization plots changes with the emergence of a peak on the H_{out} polar plots. This peak signs the apparition of a coherent signal coming from correlated molecules arranged in a supramolecular aggregate^{33, 34}. At high ethanol ratio (100 and 50%), no large aggregate are observed, while at low ethanol ratio (5% and 0%), large aggregates are present. All the coefficient of the fits are given in Table 1.

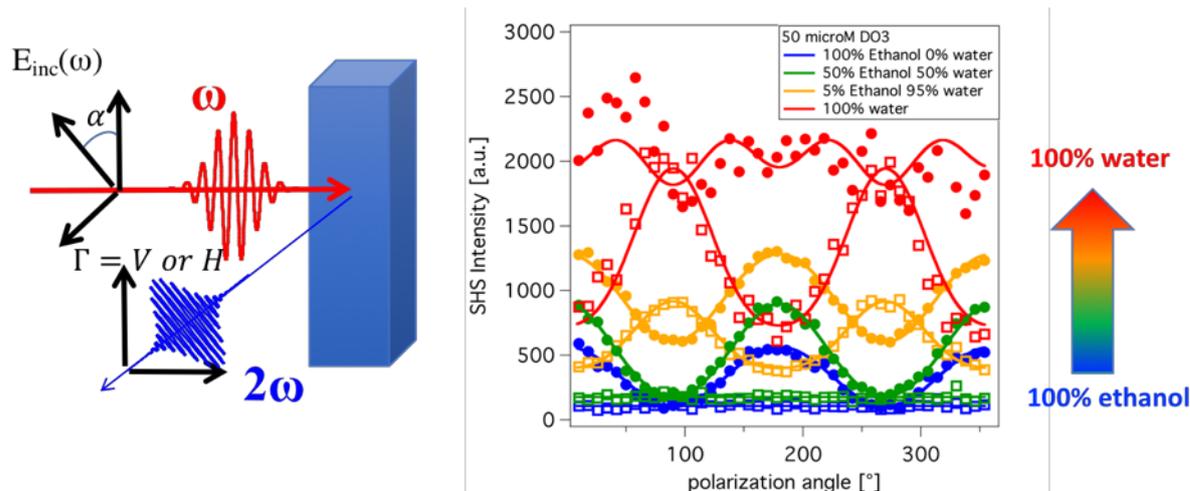


Figure 1. Left, the polarization resolved SHS experimental setup. Right, the SHS polarization plots obtained with $[DO3]=0.05$ mM for different ethanol-water ratio. The filled circle are $I(\alpha, V_{out})$ polar plot and empty square are $I(\alpha, H_{out})$ polar plot. The solid lines are a fit using the equation (2).

ratio	100% ethanol	50 % ethanol	5% ethanol	0% ethanol
I_2^V	0.68	0.69	0.38	0.04
I_4^V	0.01	0.03	-0.01	-0.07
I_2^H	0.00	-0.01	-0.42	-0.51
I_4^H	0.00	0.00	0.06	0.07

Table 1. The coefficients obtained by the fit equation 2 on the Figure 1 data.

In order to go further and to give a microscopic description of the organization in the aggregates, molecular dynamics simulations are performed for three ethanol-water ratio: 100%, 50% and 0%. Figure 2 left, shows the radial distribution functions obtained for these different mixtures. In the case of pure ethanol, the RDF given in insert confirms that the dye is fully solvated and that no aggregate is observed. For a 50-50 mixture, a peak at around 0.5 nm is observed and shows the formation of dimers. In the case of pure water, this peak is always observed with oscillations showing that several molecules are packed together. In order to get structural information about the microscopic organization of the dyes inside the aggregate, a simulation with 40 dyes in a water box has been performed. The RDF of this simulation is

presented in Figure 2 right. The radial distribution function shows that a well-defined structure is obtained with the dye packed in a layer arrangement as depicted in the snapshot taken at the end of the simulation and given in insert, Figure 2. This organization can be seen as a nanosheet with dye distanced by 0.44 nm and oriented in a mixed “up” and “down” direction. This structure is used as a starting point to simulate the polarization plots using the PySHS package^{29, 30}.

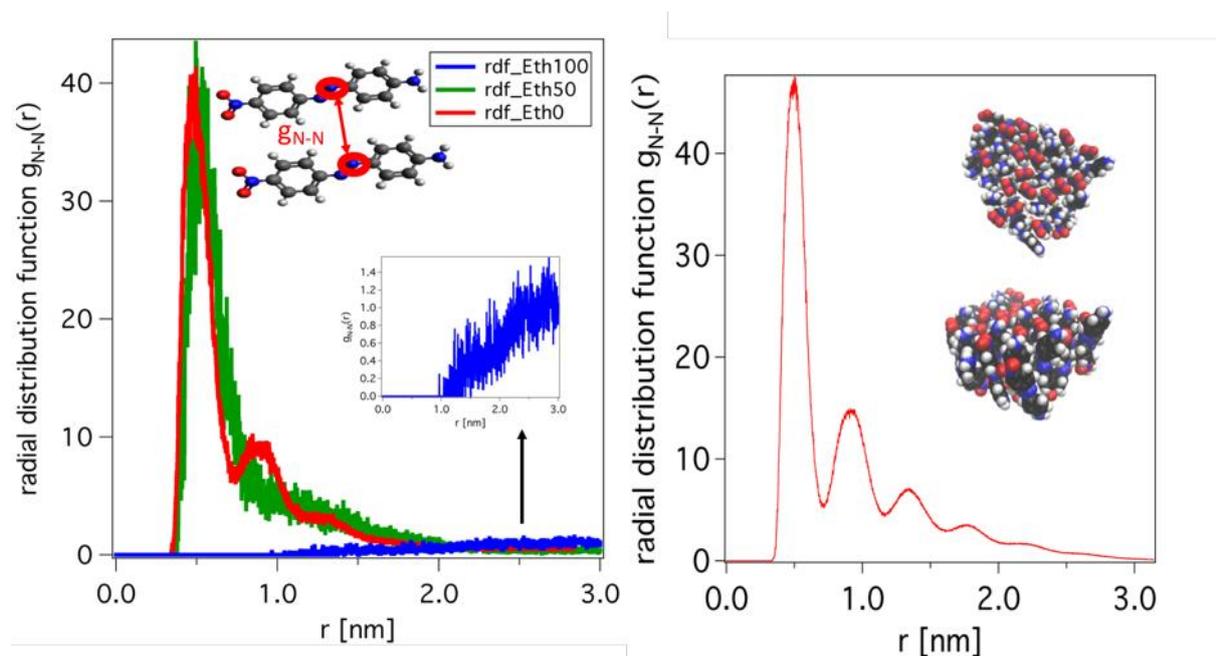


Figure 2. Left, the N-N radial distribution function as depicted in the scheme for different ethanol-water ratio with 6 DO3 molecules in the box. Right, the same with 40 DO3 molecules in the box. Insert is snapshots of the structure obtained.

The Figure 3 right presents the input structure used as a model for the calculation. The orientation of the dye is randomly distributed between “up” and “down” with a ratio equal to 0.51. This ratio is defined by the number of “up” dye divided by the total number of the dye in the aggregate. This ratio has been chosen close to 0.5 because of the results given by the molecular dynamics calculation but is not strictly equal to 0.5. Indeed, in this case, for centrosymmetric reason, the SHS intensity would be close to zero. Additional model presented in the SI, shows that the value of this ratio doesn’t change significantly the computed values of the coefficients $I_{2,SHS}^I$ and $I_{4,SHS}^I$ presented in Table 2. As it is expected, it only changes the absolute SHS Intensity. The simulated polarization plots presented in Figure 3 left reproduced very nicely the experimental data presented in Figure 1 both in

absolute intensity and in the global shape. The table 2 gives the different coefficients obtained by the model and they can be directly compared to the experimentally coefficients presented in table 1. It shows that the model of supramolecular nanosheet seems to describe correctly the spatial arrangement of the dyes in the aggregate. The evolution of those plots with the size of the aggregate also shows that increasing the water ratio in the mixture close to 100% leads to the formation of very large aggregates in the range of around 200 nm.

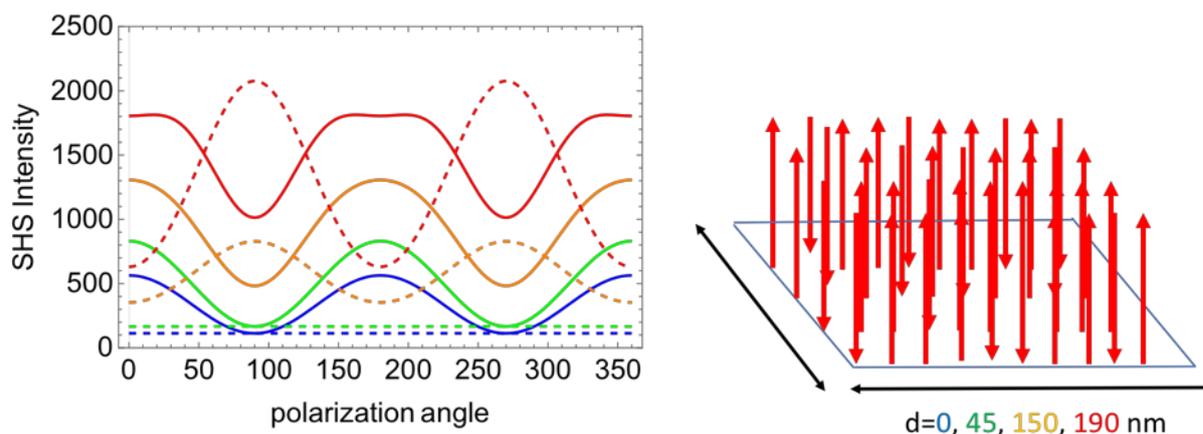


Figure 3. Simulation of the polarization resolved SHS for various 2D nanosheets with regularly spaced molecule as depicted by the scheme on the right. Blue curve is for $d=0$ nm, green curve for $d=45$ nm, orange for $d=150$ nm and red curve for $d=190$ nm. The solid lines are for the $I(\alpha, V_{out})$ polar plots and dashed lines are $I(\alpha, H_{out})$ polar plots.

Table 2. The coefficients obtained by the calculation performed by PySHS software on the 2D nanosheet organization

Size (nm)	0	45	150	190
I_2^V	0.67	0.66	0.49	0.26
I_4^V	0.00	-0.00	-0.04	-0.07
I_2^H	0.00	0.00	-0.26	-0.54
I_4^H	0.00	0.00	0.01	0.02

To summarize this first part, the experimental

SHS results show that this technique is sensitive to the growth of aggregates which occurs upon increasing the water/ethanol ratio. We now examine the effect of IMO-CH₃ addition in the aqueous dye solution.

3.2 DO3 in water with hybrid imogolite nanotube

The dye behavior in pure water after being in contact with an IMO-CH₃ nanotubes suspension is considered. Figure 4 shows the experimental polarization plot after 24h equilibration.

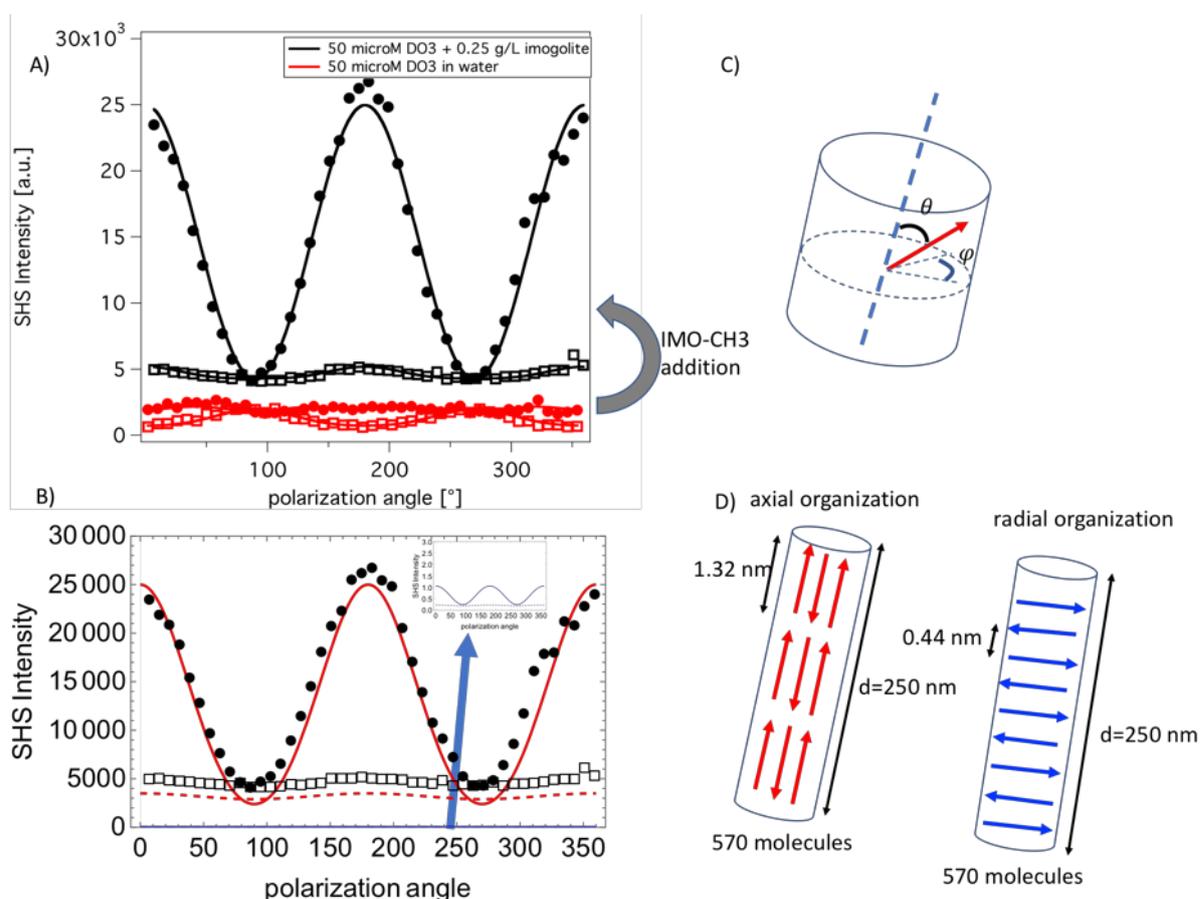


Figure 4. A) Black curves are the experimental polarization plot of 0.050 mM DO3 in water with 0.25 g/L of imogolite nanotube (full point are Vout and empty square are Hout plot). The red curves are obtained in the same condition without imogolite nanotube. The solid lines are fit according to equation 2. B) The red curves are the simulated polar plots obtained with an axial organization of the dye in the nanotube as depicted in the scheme D). The blue curves are the simulated polar plot obtained with a radial organization of the dye in the nanotube as depicted in the scheme. The black points and squares are the same data as A).

C) The definition of the angle used to describe the different organizations. D) Description of the two models used in the computation.

In presence of IMO-CH₃ the polarization plots drastically change compared to the case without IMO-CH₃. Both a very high SHS intensity and a modified pattern of the polar plots are observed with IMO-CH₃. It was known that IMO-CH₃ itself does not have a significative contribution in the SHS signal, see additional data given, therefore the change in the SHS intensity can be attributed to a confinement effect of the dye aggregate within the nanotubes. To explore this hypothesis, axial and radial dye organizations inside the nanotube are considered. In all computation, the nanotube suspension is modeled by a 250 nm long nanotube in agreement with the typically observed average length observed by Atomic Force Microscopy³⁵, additional details are given in the Supplementary Information. The axial model assume that the dyes fill the nanotube with an angle theta, equal to 0° (“up”) or to 180° (“down”). More precisely, 570 dyes are regularly positioned along the nanotube, as depicted in Figure 4, with triplets of tail-head DO3 molecules forming a chain spanning the whole nanotube. The second model assume a radial arrangement of the dipole in the nanotube as depicted in Figure 4. In this case, the 570 DO3 molecules are positioned with an angle theta equals to 90° and an angle phi fixed to the same value to all molecules. The dye are spaced by 0.44 nm, an average distance in agreement with the RDF results obtained by molecular dynamics. The polar plots obtained with the two models are presented in Figure 4 B.

The comparison between the experimental and simulated polarization plots unambiguously evidences that the axial organization is more probable. Indeed, this model reproduces much better the absolute intensity of the experimental data while the radial organization would produce a nearly zero SHS signal, as expected for this centrosymmetric arrangement. The numerical value of the $I_{i,SHS}^T$ coefficients obtained from the simulation are given in table 3. In particular, the I_2^H coefficient is nicely reproduced both in sign and in absolute value by the parallel model simulation.

Table 3. The coefficients obtained experimentally and by the calculation performed by PySHS software on the different filling organization

	Experiment DO3 in pure water with IMO-CH3	axial organization model	radial organization model
I_2^V	0.72	0.87	0.61
I_4^V	0.01	0.06	-0.02
I_2^H	0.10	0.10	-0.03
I_4^H	0.01	0.00	0.00

4 Conclusion and perspectives

In pure ethanol or at low water/ethanol ratio, SHS measurements demonstrate that there is no correlation between the dyes. This means that the dyes are well dispersed and randomly distributed in solution with no assembly. In pure water or at high water/ethanol ratio, the dye is poorly soluble and SHS measurements coupled to MD simulations enable to demonstrate that DO3 form aggregates having a 2D nanosheet shape. In pure water those aggregate reach very large size about 200 nm. When IMO-CH₃ are added in the dye aqueous solution, the dye molecules fill the nanotube. The SHS signal is then very different from the pure aqueous DO3 solution case, thus revealing a strong confining effect in the dye organization. The comparison of the SHS data with a radial and axial model enables to unambiguously evidence that the dyes are organized with their axis parallel to the nanotube walls. To our knowledge, it is the first time that polarization resolved SHS technique demonstrates its ability to probe molecular organization inside a nanotube. The different models and organizations proposed in this work reproduce nicely, without any adjustable parameters, the experimental data in all the different conditions: dye alone well solubilized in ethanol, dye aggregate in water-ethanol mixture and dye filling the nanotube. The models and the related numerical code developed here are quite general and open new perspectives to probe confined organization inside IMO-CH₃ nanotubes. The extension to different tube loading and the correlation with the confined chemical reactions would be very interesting and will be the subject of future experiments.

Conflicts of interest

There are no conflicts to declare.

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1. N. T. Southall, K. A. Dill and A. D. J. Haymet, *The Journal of Physical Chemistry B*, 2002, **106**, 521-533.
2. C. Tanford, *The hydrophobic effect: formation of micelles and biological membranes 2d ed*, J. Wiley., 1980.
3. A. Y. Ben-Naim, *Hydrophobic interactions*, Springer Science & Business Media, 2012.
4. K. Patil, R. Pawar and P. Talap, *Physical Chemistry Chemical Physics*, 2000, **2**, 4313-4317.
5. P. Bauduin, A. Renoncourt, A. Kopf, D. Touraud and W. Kunz, *Langmuir*, 2005, **21**, 6769-6775.
6. N. F. A. van der Vegt and D. Nayar, *The Journal of Physical Chemistry B*, 2017, **121**, 9986-9998.
7. J. Gregory, *Polymer International*, 1994, **35**, 105-106.
8. I. Bottero, B. Bonelli, S. E. Ashbrook, P. A. Wright, W. Zhou, M. Tagliabue, M. Armandi and E. Garrone, *Physical Chemistry Chemical Physics*, 2011, **13**, 744-750.
9. M. S. Amara, E. Paineau, S. Rouzière, B. Guiose, M.-E. M. Krapf, O. Taché, P. Launois and A. Thill, *Chemistry of Materials*, 2015, **27**, 1488-1494.
10. P. Picot, F. Gobeaux, T. Coradin and A. Thill, *Applied Clay Science*, 2019, **178**, 105133.
11. S. Patra, D. Schaming, P. Picot, M.-C. Pignié, J.-B. Brubach, L. Sicard, S. Le Caër and A. Thill, *Environmental Science: Nano*, 2021, **8**, 2523-2541.
12. G. Martin-Gassin, E. Paineau, P. Launois and P.-M. Gassin, *The Journal of Physical Chemistry Letters*, 2022, **13**, 6883-6888.
13. Y. Chen, H. I. Okur, N. Gomopoulos, C. Macias-Romero, P. S. Cremer, P. B. Petersen, G. Tocci, D. M. Wilkins, C. Liang, M. Ceriotti and S. Roke, *Science Advances*, 2016, **2**, e1501891.
14. G. Revillod, J. Duboisset, I. Russier-Antoine, E. Benichou, G. Bachelier, C. Jonin and P.-F. Brevet, *The Journal of Physical Chemistry C*, 2008, **112**, 2716-2723.
15. M. Moris, M.-P. Van Den Eede, G. Koeckelberghs, O. Deschaume, C. Bartic, S. Van Cleuvenbergen, K. Clays and T. Verbiest, *Communications Chemistry*, 2019, **2**, 130.
16. S. Van Cleuvenbergen, Z. J. Smith, O. Deschaume, C. Bartic, S. Wachsmann-Hogiu, T. Verbiest and M. A. van der Veen, *Nature Communications*, 2018, **9**, 3418.
17. G. Gonella and H.-L. Dai, *Langmuir*, 2014, **30**, 2588-2599.
18. M. Bischoff, N. Y. Kim, J. B. Joo and A. Marchioro, *The Journal of Physical Chemistry Letters*, 2022, **13**, 8677-8683.
19. W. M. K. P. Wijekoon, Y. Zhang, S. P. Karna, P. N. Prasad, A. C. Griffin and A. M. Bhatti, *J. Opt. Soc. Am. B*, 1992, **9**, 1832-1842.
20. B. Derkowska-Zielinska, K. Fedus, H. Wang, C. Cassagne and G. Boudebs, *Optical Materials*, 2017, **72**, 545-548.
21. M. Boyer, E. Paineau, M. Bacia-Verloop and A. Thill, *Applied Clay Science*, 2014, **96**, 45-49.

22. P. Picot, Y. Liao, E. Barruet, F. Gobeaux, T. Coradin and A. Thill, *Langmuir*, 2018, **34**, 13225-13234.
23. P.-M. Gassin, S. Bellini, J. Zajac and G. Martin-Gassin, *The Journal of Physical Chemistry C*, 2017, **121**, 14566-14571.
24. J. Duboisset and P.-F. Brevet, *Physical Review Letters*, 2018, **120**, 263001.
25. N. Schmid, A. P. Eichenberger, A. Choutko, S. Riniker, M. Winger, A. E. Mark and W. F. van Gunsteren, *European Biophysics Journal*, 2011, **40**, 843-856.
26. A. K. Malde, L. Zuo, M. Breeze, M. Stroet, D. Poger, P. C. Nair, C. Oostenbrink and A. E. Mark, *Journal of Chemical Theory and Computation*, 2011, **7**, 4026-4037.
27. H. J. C. Berendsen, D. van der Spoel and R. van Drunen, *Computer Physics Communications*, 1995, **91**, 43-56.
28. B. Hess, H. Bekker, H. J. C. Berendsen and J. G. E. M. Fraaije, *Journal of Computational Chemistry*, 1997, **18**, 1463-1472.
29. L. Boudjema, H. Aarrass, M. Assaf, M. Morille, G. Martin-Gassin and P.-M. Gassin, *Journal of Chemical Information and Modeling*, 2020, **60**, 5912-5917.
30. L. Boudjema, H. Aarrass, M. Assaf, M. Morille, G. Martin-Gassin and P.-M. Gassin, *Journal of Chemical Information and Modeling*, 2021, **61**, 5719-5719.
31. P. Kaatz and D. P. Shelton, *The Journal of Chemical Physics*, 1996, **105**, 3918-3929.
32. R. Bersohn, Y. H. Pao and H. L. Frisch, *The Journal of Chemical Physics*, 1966, **45**, 3184-3198.
33. J. Duboisset, F. Rondepierre and P.-F. Brevet, *The Journal of Physical Chemistry Letters*, 2020, **11**, 9869-9875.
34. P.-M. Gassin, B. Prelot, B. Grégoire and G. Martin-Gassin, *Langmuir*, 2018, **34**, 12206-12213.
35. P. Picot, O. Taché, F. Malloggi, T. Coradin and A. Thill, *Faraday Discussions*, 2016, **191**, 391-406.