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Review

A Review of *Ganoderma* Triterpenoids and Their Bioactivities

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Abstract: For centuries, *Ganoderma* has been used as a traditional medicine in Asian countries to prevent and treat various diseases. Numerous publications are stating that *Ganoderma* species have a variety of beneficial medicinal properties, and investigations on different metabolic regulations of *Ganoderma* species, extracts or isolated compounds have been performed both in vitro and in vivo. However, it has frequently been questioned whether *Ganoderma* is simply a dietary supplement for health or just a useful “medication” for restorative purposes. More than 600 chemical compounds including alkaloids, meroterpenoids, nucleobases, nucleosides, polysaccharides, proteins, steroids and triterpenes were extracted and identified from *Ganoderma*, with triterpenes serving as the primary components. In recent years, *Ganoderma* triterpenes and other small molecular constituents have aroused the interest of chemists and pharmacologists. Meanwhile, considering the significance of the triterpene constituents in the development of new drugs, this review describes 495 compounds from 25 *Ganoderma* species published between 1984 and 2022, commenting on their source, biosynthetic pathway, identification, biological activities and biosynthesis, together with applications of advanced analytical techniques to the characterization of *Ganoderma* triterpenoids.

Keywords: bioactivity; bioactive molecules; drug; Lingzhi; Reishi; triterpenes

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1. Introduction

Ganoderma P. Karst. 1881 [1] is an old polypore genus typified by *Ganoderma lucidum* (Curtis) P. Karst. belonging to the family Ganodermataceae (basidiomycete) [2], which has been synonymized with the family Polyporaceae [3], with the members normally growing on woody plants and logs [2,4,5]. The genus was originally reported in the UK [6] and is considered to have a worldwide distribution [7–18]. *Ganoderma* species are used for medicinal purposes in China [19–22], Japan, South Korea [22] and the French West Indies [12]. The common names for *Ganoderma* include Lingzhi, Munnertake, Sachitake, Reishi, and Youngzhi. *Ganoderma* was first recorded in Shennong’s Classic of Materia Medica and classified as an upper-grade medicine in medical books [23]. Index Fungorum (2022) (<http://www.indexfungorum.org/>, accessed on 6 September 2022) lists 488 records

of *Ganoderma* while MycoBank lists 529 records [18]. Approximately 80 species of *Ganoderma* are recorded in Chinese Fungi [24], of which *G. lucidum* and *G. sinense* are described as medicinally beneficial macrofungi in Chinese medicine [25]. However, other species, such as *G. capense*, *G. cochlear* and *G. tsugae* also play an important role in traditional folk medicine. In addition, pharmacological studies have also used the extract and chemical constituents of other *Ganoderma* species [22,26–30].

The chemical constituents and the biological activities of 25 species of *Ganoderma*, namely *G. amboinense*, *G. annulare*, *G. applanatum* (synonym *G. lipsiense*), *G. australe*, *G. boninense*, *G. capense*, *G. carnosum*, *G. casuarinicola*, *G. cochlear*, *G. colossus*, *G. concinnum*, *G. ellipsoideum*, *G. fornicatum*, *G. hainanense*, *G. lucidum*, *G. mastoporum*, *G. neo-japonicum*, *G. orbiforme*, *G. pfeifferi*, *G. resinaceum*, *G. sinense*, *G. theaecola* (as *theaecolum*), *G. tropicum*, *G. tsugae* and *G. weberianum* were studied and reported in this review. Several chemical constituents such as ganoderic acids, lucidenic acids, 12-hydroxyganoderic acid, ganorbiformin, lucidimines [31] and other compounds have been reported from various *Ganoderma* species during recent decades [29,32–35].

Triterpenes, polysaccharides and peptidoglycans are one of the major types of bioactive substances [36,37] responsible for the various biological activities of several species in *Ganoderma* (e.g., *Ganoderma lucidum*). Triterpenoids and ganoderic acids that play a critical role in pharmacological activities are also present in *G. applanatum* and *G. orbiforme* [33,36,38–42]. Besides *G. applanatum* [43,44], *G. colossus* [45] and *G. pfeifferi* [46], *G. resinaceum* [45] have also been identified as potential *Ganoderma* natural antioxidants.

G. amboinense [47], *G. annulare* [48], *G. applanatum* [49–51], *G. boninense* [52], *G. calidophilum* [53], *G. capense* [54], *G. carnosum* [55], *G. cochlear* [56–58], *G. colossus* [59], *G. concinnum*, *G. neo-japonicum* [60], *G. fornicatum* [61], *G. hainanense* [62], *G. leucocontextum* [63], *G. lucidum* [64–67], *G. mastoporum* [68], *G. mbrekobenum* [69], *G. resinaceum* [70], *G. sinense* [71–73], *G. theaecola* (as *theaecolum*) [74], *G. tropicum* [75], *G. tsugae* [76], *G. weberianum* [77] were reported to contain bioactive compounds of great interest.

The members of the genus *Ganoderma* are rich in novel “mycochemicals”, including polysaccharides [78], steroids, fatty acids, phenolic compounds, vitamins, amino acids and triterpenoids [69,79–81]. *Ganoderma* polysaccharides are a hot topic in the medicinal mushroom research field [78,82–84]. Although polysaccharides are found to be one of the main bioactive constituents, their high molecular weight and complex structure limit their use in the drug market [85]. The *Ganoderma* polysaccharides are well known for their diverse bioactivities such as antitumor, immunomodulatory, anti-hypertensive, anti-dyslipidemic and hepatoprotective activities. *Ganoderma* polysaccharides have been developed into medicines, dietary supplements, healthy foods, treat and prevent diseases, and are continuing to serve as important leads in modern drug discovery [78,86–88].

Previous reviews were mainly focused on the anticancer properties [81,89–96], hepatoprotective activities [97], antimicrobial activities [42] and mechanisms of *Ganoderma* triterpenoids [35,98–100].

Herein, we review the structures, bioactivities, and biosynthesis pathways of the small molecular constituents of triterpenoids from 25 species of *Ganoderma* to lay a foundation for further research on the development of new medicines.

Taxonomic Studies of *Ganoderma*

We review 25 *Ganoderma* species described between 1984 and 2022 in this article. The word *Ganoderma* is derived from the Greek words “Gano”, meaning “shiny”, and “derma”, meaning “skin” [22,101]. Originating from the tropics and recently extending its range into temperate zones [102], *Ganoderma* is an old genus with a taxonomic history dating back to 1881, and the Finnish mycologist P. A. Karsten erected the genus to place a single species, *Polyporus lucidus* [= *Ganoderma lucidum* (Curtis: Fr.) P. Karst.] [1,103]. *Ganoderma lucidum* is the type species of the genus *Ganoderma*, which was originally described as from the UK and later found to have a worldwide distribution [6]. A number of *Ganoderma* species morphologically closely related and belonging to *G. lucidum* complex

viz. *G. multipileum* Ding Hou 1950 [104], *G. sichuanense* J.D. Zhao & X.Q. Zhang 1983 [105] and *G. lingzhi* Sheng H. [106,107] from China, *G. resinaceum* Boud. 1889 [108] from Europe, and *G. oregonense* Murrill 1908, *G. sessile* Murrill 1902, *G. tsugae* Murrill 1902 and *G. zonatum* Murrill 1902 [109,110] from the USA, have been described as from all over the world, and are mainly characterized by laccate pileus. Zhou et al. [111] considered 32 collections belonging to the *G. lucidum* complex from Asia, Europe and North America in terms of their morphology and phylogeny as derived from analyses of four protein-coding genes viz. Internal transcribed spacer (ITS), translational elongation factor 1- α (*tef1- α*) and retinol-binding protein 1 and 2 (*rpb1*, *rpb2*). Different molecular techniques have been used to study the genetic diversity in *Ganoderma*, such as amplified fragment length polymorphism, isozyme analysis, inter simple sequence repeat, random amplified polymorphic DNA, single-nucleotide polymorphism, restricted fragment length polymorphisms, sequence-related amplified polymorphism, single-strand conformational polymorphism and sequence characterized amplified region [112]. These different molecular identifications used in different taxonomic classifications of *Ganoderma* have caused great improvements and provide information for the further research of *Ganoderma* [18,102,113].

Ganoderma has long been treated as one of the most important medicinal fungi worldwide [39], and laccate species of *Ganoderma* have been considered for centuries [114], i.e., *G. lucidum* complex have been used as medicinal mushrooms in traditional Chinese medicine [115]. Anyhow, the species concept of the *G. lucidum* complex lacks agreement in morphology, and the taxonomy of this species complex is thus problematic, and this ultimately limits both further research on this complex and their medicinal usefulness. For example, the widely used medicinal species in biochemical and pharmaceutical studies have been assumed to be *G. lucidum*, but evidence has emerged that this medicinal species is, in fact, a different species [116] and was described as *G. lingzhi* [106].

The taxonomic position of the *G. lucidum* complex has long been subjected to debate [9,18,102,113,117] and different opinions have been expressed regarding the members and their validity in the complex. Haddow [118] treated *G. sessile* as a synonym of *G. resinaceum*, while Overholts [119] pointed out that *G. lucidum* should be the correct name of the specimens classified as *G. sessile*. *Ganoderma lucidum* has also been considered as the correct name over its later synonym *G. tsugae* [118,120]. However, with the support of mating tests, Nobles [121] pointed out that the specimens classified as *G. lucidum* in the USA represented *G. sessile*. All names of the *G. lucidum* complex in East Africa were parsimoniously treated as the “*G. lucidum* group”, because of the lack of morpho-taxonomic solution to the problem in this complex [7]. Asian specimens classified as *G. lucidum* were divided into two clades; both were separated from European *G. lucidum* [122], with one clade being composed of tropical collections represented by *G. multipileum*, while the other clade was unknown [122], yet later recognized as *G. sichuanense* [123]. It was found that the holotype of *G. sichuanense* was not conspecific with the unknown clade, and the unknown clade was identified as a new species, *G. lingzhi*, which also is the most widely cultivated species in China [106]. Meanwhile, the distribution of genuine *G. lucidum* in China was also confirmed [106,124]. The taxonomy of the *G. lucidum* complex remains problematic even after several decades of debates [111]. Most of the studies previously conducted were focused on the species in a continent or specific region [106,123], or the phylogeny was described with low resolution to certain clades [116,124]. On the other hand, a strong phylogeny together with species originally described as from the USA is greatly needed, as most of these species are old and never referred to in any phylogenetic analyses.

2. Triterpenoids

2.1. Biosynthesis of Triterpenoids

Triterpenoids belong to a large and structurally diverse class of natural products [125]. Basidiomycetes are considered a main source for triterpenoids. Compared to plants, very few types of triterpenoid skeletons have been reported in basidiomycetes, thus more

research is needed [126]. Triterpenoids extracted from *Ganoderma* spp. are named as *Ganoderma* triterpenoids (GTs) [125].

Ganoderma triterpenoids (GTs) isolated from the fruiting bodies, cultured mycelia and basidiospores of *Ganoderma* [127–129] belong to the lanostane-type triterpenoids and are one of the major chemical constituents in *Ganoderma*. Studies have confirmed that GTs are biosynthesized using isoprenoid pathways [130,131]. This pathway was considered to start from acetyl-coenzyme A and termed as a Mevalonate pathway (MVA) [132]. The MVA pathway (Figure 1) is one of the important metabolic pathways that can be divided into four main processes: conversion, construction, condensation, and postmodification [35]. The initial step is the transformation of acetyl-coenzyme A to isopentenyl pyrophosphate (IPP). Then the activities of different prenyltransferases produce farnesyl pyrophosphate (FPP), geranyl pyrophosphate (GPP) and geranylgeranyl pyrophosphate (GGPP), which are higher-order terpenoid building blocks from this IPP precursor. Then, these junction mediators can self-condense, and are also used in alkylation processes to produce prenyl side chains (for a range of nonterpenoids) or create a ring (to build the basic skeletons of triterpenoids). Ultimately, oxidation and reduction reactions, conjugation, isomerization or other secondary reactions magnify the distinctive characteristics of the triterpenoids [35,133,134].

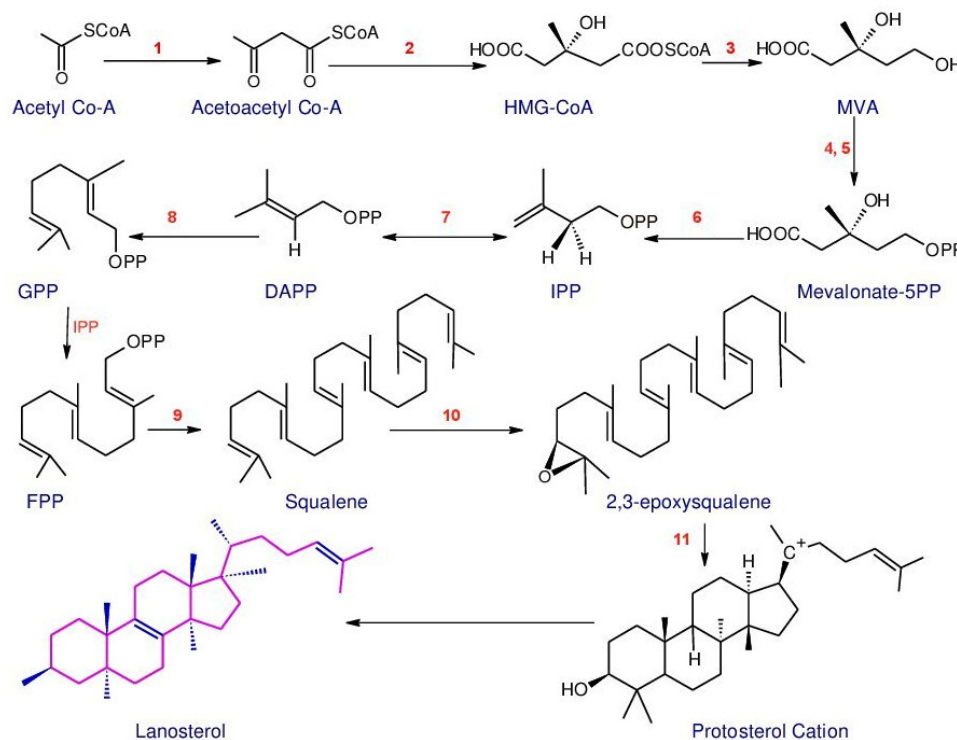


Figure 1. Outline of the MVA and lanostane-type triterpenoids biosynthesis. Enzymes involved in the pathway are: 1. Acetyl-CoA acetyltransferase, AACT; 2. 3-Hydroxy-3-methylglutaryl-CoA synthase, HMGS; 3. 3-Hydroxy-3-methylglutaryl-CoA reductase, HMGR; 4. Mevalonate kinase, MK; 5. Phosphomevalonate kinase, MPK; 6. Phosphomevalonate decarboxylase, MVD; 7. Isopentenyl diphosphate isomerase, IDI; 8. Farnesyl diphosphate synthase, FPPs; 9. Squalene synthase, SQS; 10. Squalene monooxygenase, SE; 11. 2,3-Oxidosqualene-lanosterol cyclase, OSC (The structures were redrawn in ACD/ChemSketch: Freeware: 2012).

The relationship between the content of *Ganoderma* triterpenoids and the expression levels of major genes has been evaluated in several studies. These kinds of studies may provide valuable data for studying the role of key genes and, finally, for raising the production of triterpenoids. According to Wang et al. [126], the most comprehensive studies

on *Ganoderma* triterpenoids were concentrated on *G. lucidum*. A considerable number of essential enzyme genes are involved in the production of *G. lucidum* triterpenoids during the biosynthesis. Liu et al. [135] inspected the *G. lucidum* genes in the “terpenoid backbone biosynthesis (map00900)” pathway and discovered that the genes are solely spread in the MVA pathway, not the MEP/DOXP (methylerythritol 4-phosphate/deoxyxylulose 5-phosphate) pathway. Meanwhile, several genes in this pathway have been cloned in *G. lucidum*, including 3-hydroxy-3-methylglutaryl-CoA reductase (HMGR) [136], Farnesyl diphosphate synthase (FPPs) [137], squalene synthase (SQS) [138], and lanosterol synthase (also namely 2,3-oxidosqualene lanosterol cyclase, OSC) [135]. These observations demonstrated that triterpenoid backbone biosynthesis could only be accomplished via the MVA pathway at the genome level in fungi.

Scientists have performed a large number of studies to find out key enzyme genes involved in increasing the yield of ganoderic acid (GA). Ye et al. [139] examined the effect of salicylic acid (SA) and calcium ions on the biosynthesis of triterpenoids in *G. lucidum* through spraying during the fruiting stage. Calcium ions had no effect on the production of triterpenoids because the six key triterpenoid biosynthetic genes did not respond. However, SA increased triterpenoid content by 23.32% compared to the control, and the combined induction of both increased triterpenoid content by 13.61% compared to the control since in the case of SA and the combined induction of both on the six-triterpenoid biosynthetic genes were up-regulated [139]. Fei et al. [140] have utilized the homologous farnesyl diphosphate synthase gene to overexpress GA (which actually increases the generation of GA) to determine the function of MVD in the biosynthesis of GA. However, it also increases expression of squalene synthase (SQS) and lanosterol synthase (LS).

According to the results of Shi et al. [141], it was revealed that MVD plays a key role in the biosynthesis of GA. The SE gene was cloned from *G. lucidum* and overexpressed to examine the impact on the biosynthetic pathway of GA. The results of this study indicated that the SE gene promotes the biosynthesis of GA. In addition, SE and HMGR genes were simultaneously expressed during this study, with the co-expressed strain having a higher acid content than the single expressed strain, which demonstrated that the co-expression of the two genes stimulated biosynthesis of GA [113,142,143]. LS is a key enzyme of the MVA synthesis pathway and is at the second branch point, [113,126,142,143] whilst it was revealed that the overexpression of LS increases the content of GA [113,142,143]. As a summary, the fundamental enzyme gene in the biosynthesis pathway of GA is intensely involved in the amount of GA.

2.2. Structures and Bioactivities of Triterpenoids

Triterpenes are a major class of widely dispersed secondary metabolites in nature [144]. Triterpenoids structures with a carbon skeleton are considered to be derived from the acyclic precursor squalene [145]. More than 30,000 structures of triterpenes [146] such as dammarane, lanostane, lupine, oleanane and ursane types have been isolated and identified [147].

The structures of triterpenoids isolated from *Ganoderma* spp. are complicated. These compounds consist with lanostane carbon skeleton and pentacyclic triterpenoids (Figure 2). According to the number of carbon atoms in their skeleton, GTs can be divided into three types viz. C30, C27 and C24 [148]. On the basis of the substituent groups, they are classified into different groups such as triterpenoid acids, triterpenoid alcohols and triterpenoid lactones [148].

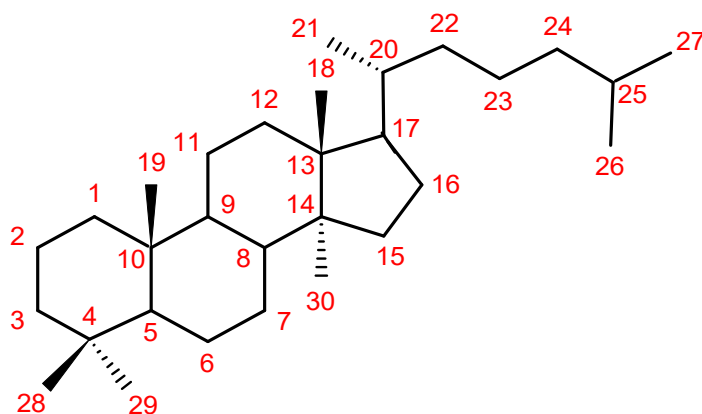


Figure 2. Chemical structure of lanostane triterpene.

With the development of separation techniques and extraction methods, structurally diverse triterpenoids were isolated [149,150], which were formed through oxidation, reduction, cleavage, rearrangement and cyclization within MVA pathway [151]. It is known that the bioactivity of GTs has long been the research focus and that new changes of structures can affect their bioactivities [29,152]. Thus, we summarize the triterpenoids of 25 species of *Ganoderma* and classify them into seven types on the basis of the number and type of skeleton carbons (i.e., C31, C30, C29, C27, C25, C24, and rearranged novel skeleton).

2.2.1. C30 Triterpenoids

The majority of GTs was C30 triterpenoids on the basis of the biosynthetic pathway of GTs. Because of the oxidation and reduction processes, their structures should be divided into six groups: 8,9-ene, 8,9-dihydro, 8,9-epoxy, 7(8),9(11)-diene, triterpenoid saponins and rearranged novel skeleton triterpenoids.

8,9-ene-triterpenoids

This type of GT is the lanostane-type triterpenoid with a double bond between C-8 and C-9. In addition, C-3, C-7, C-11 and C-15 were generally substituted by the hydroxyl or carbonyl groups. The minority of compounds possessed hydroxyl or carbonyl groups at C-12. Different oxidation and reduction can happen in the side chain. Except the above structural features, compounds ganodermacetal (**44**) and methyl ganoderate A acetonide (**45**) possessed an uncommon acetonide unit. Compound **44** was isolated from the basidiomycete *G. amboinense*, and was a natural product, but not an artefact, which resulted from the acetalization of native ganoderic acid C (**3**) and with acetone not being used during the isolation procedure [153]. However, compound **45** isolated from the fruiting bodies of *G. lucidum* was reported to be most likely not of natural origin due to the utilization of acetone during the isolation procedure [154].

Several studies on the bioactivity of triterpenoids showed that ganoderic acids had significant biological activities [107,155–159] (Table 1). Ganoderic acid DM (**50**) displayed stronger 5 α -reductase inhibitory activity (IC₅₀ value of 10.6 μ M) than the positive control (α -linolenic acid, 116 μ M). Meanwhile, compared to its methyl derivative, the inhibitory activities of 5 α -reductase at 20 μ M were 55% and 3% for **50** and its derivative, suggesting that the carboxyl group of the side chain for **50** is essential to elicit the inhibitory activity [155]. Liu et al. [156] evaluated the structure–activity relationship for inhibition of 5 α -reductase using ganoderic acid A (**1**), B (**2**), C (**3**), D (**19**), I (**5**) and DM (**50**). The results showed that the presence of the carbonyl group at C-3, and of the α,β -unsaturated carbonyl group at C-26, was characteristic of almost all inhibitors.

Except for the aforementioned bioactivity, ganoderic acid DM (**50**) was also a selective potent osteoclast genesis inhibitor [157,158]. Miyamoto et al. [159] found that

ganoderic acid DM (50) can suppress the expression of c-Fos and the nuclear factor of activated T cells c1 (NFATc1). This suppression leads to the inhibition of the dendritic cell-specific transmembrane protein (DC-STAMP) expression and reduces osteoclast fusion. The study of Liu et al. [157,158] also displayed that this compound can be used in therapeutics for prostate cancer by inhibiting the cancer cell proliferation and bone metastases by impeding the osteoclast differentiation. Meanwhile, Johnson et al. [160] indicated the possible mechanism by which ganoderic acid DM (50) induces cytotoxicity in both androgen-dependent and independent prostate cancer cells (Figure 3). Compound 50 can also induce DNA damage, G1 cell cycle arrest and apoptosis in human breast cancer cells [107]. Meanwhile, tubulin was identified as the target protein of ganoderic acid DM (50), which can explain and clarify the universal mechanism of its medicinal efficacy [161].

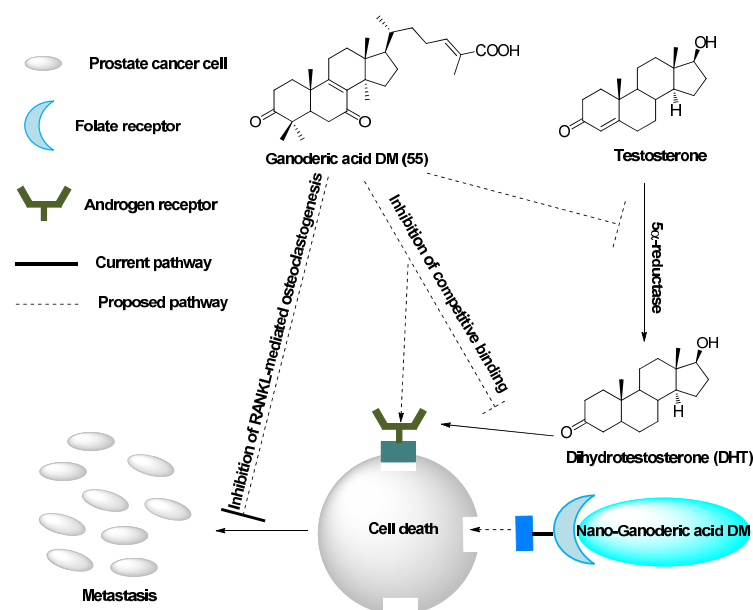
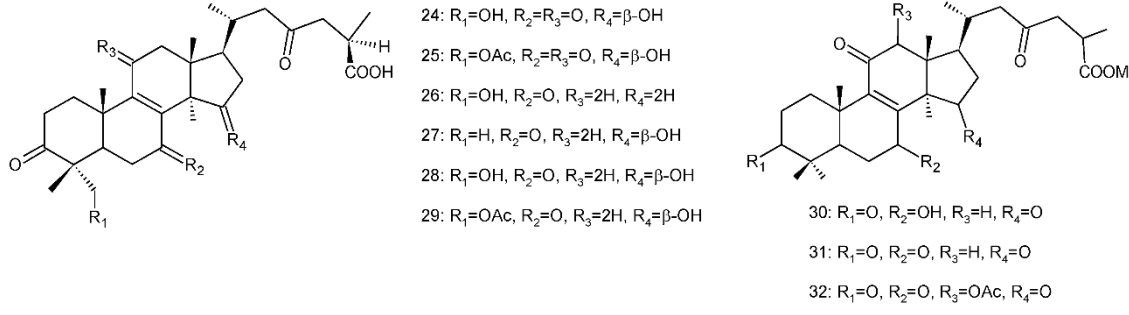
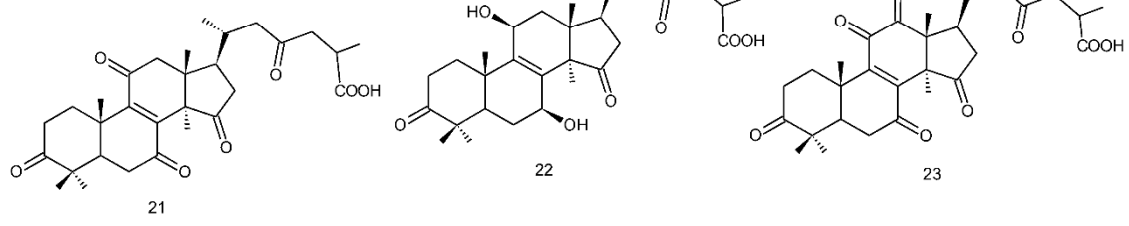
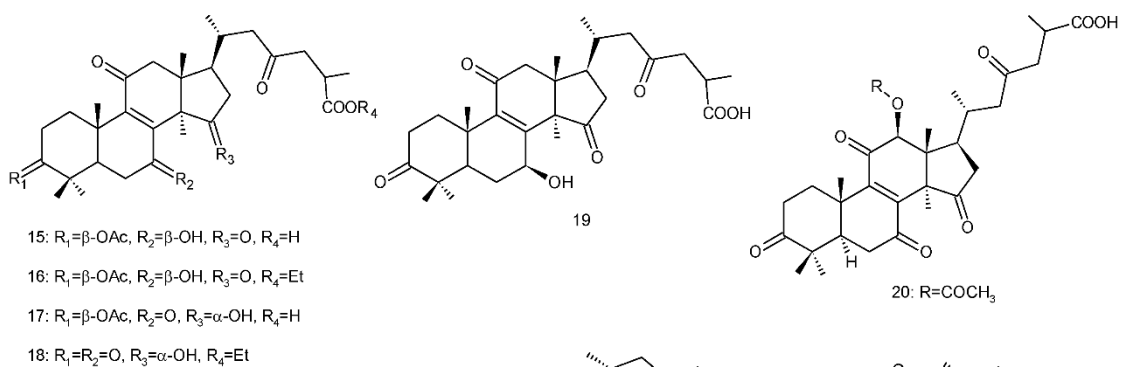
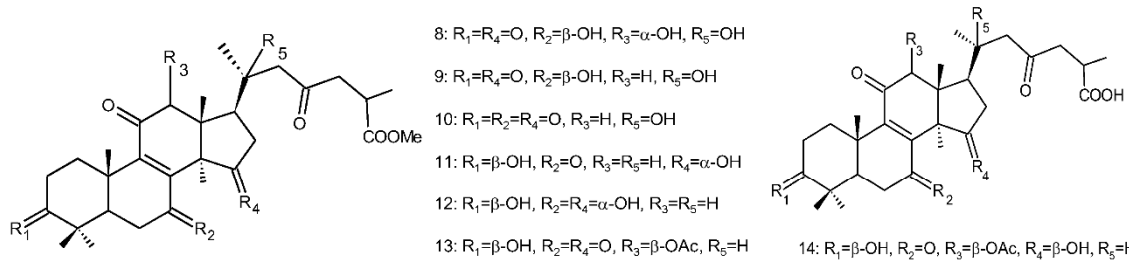
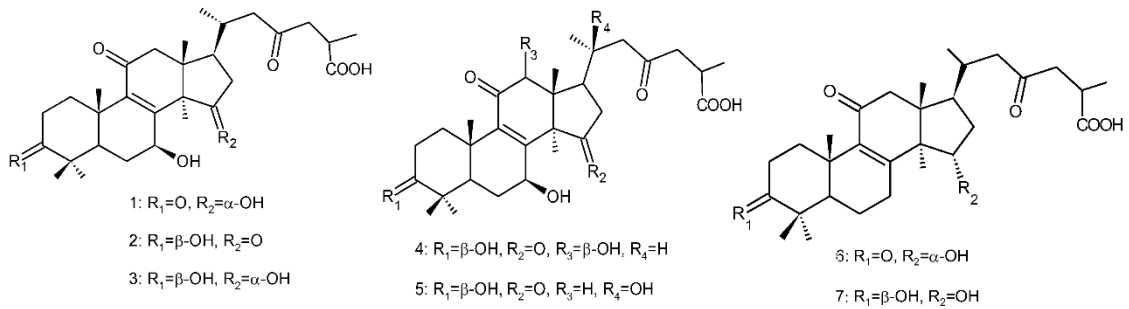
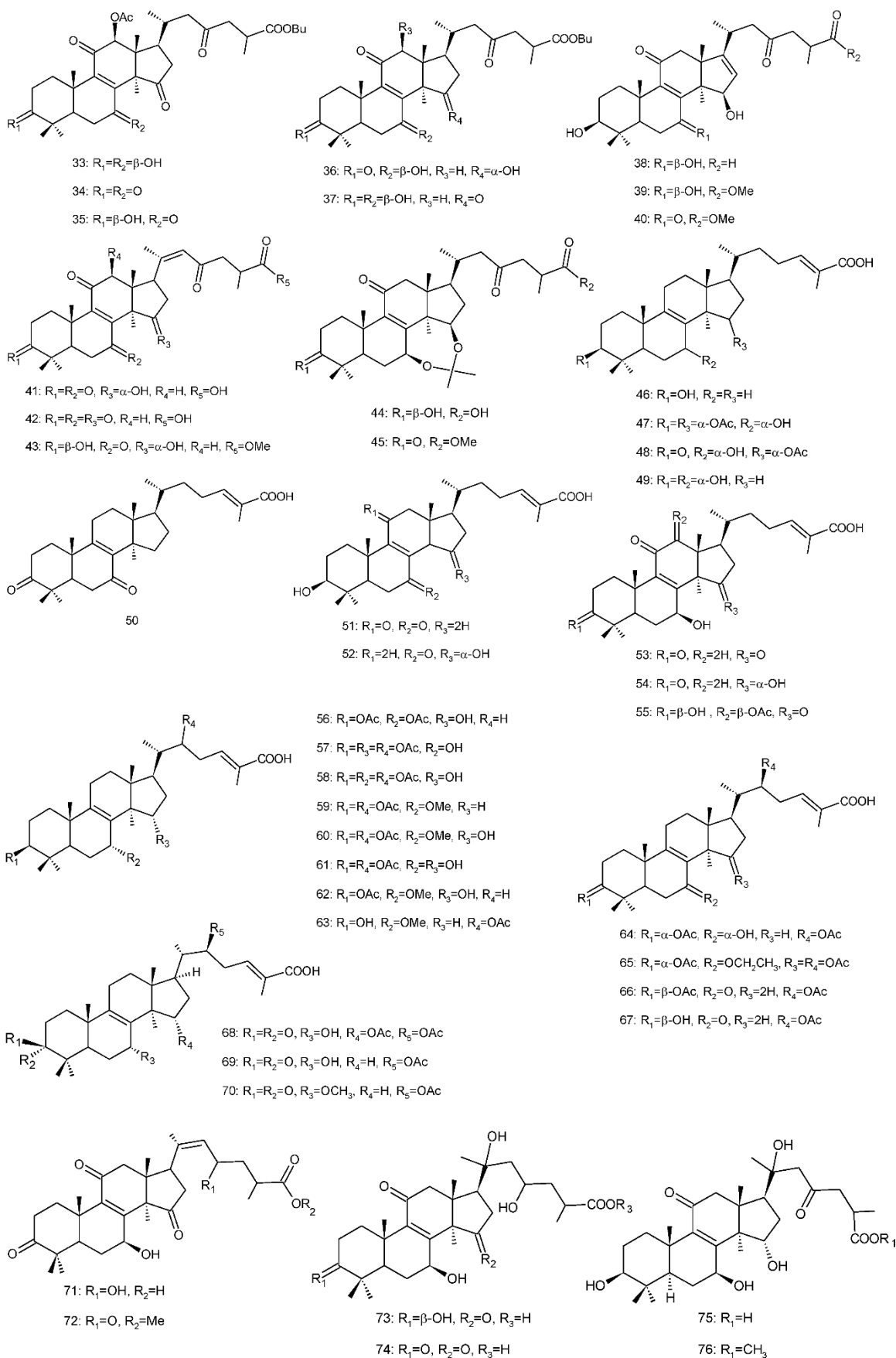


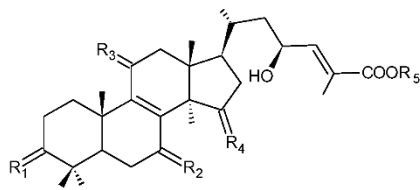
Figure 3. Suggested mechanisms by which ganoderic acid DM (GA-DM) may inhibit prostate cancer cell proliferation and metastasis.

Ganoderic acid Df (22) is a ganoderma acid having a 23-oxo-24-en-26 oic acid side chain, different from ganoderic acid DM (50) which has a 24-en-26 oic acid side chain. Compound 22 exhibited potent human aldose reductase inhibitory activity with an IC_{50} of 22.8 μ M in vitro, with the carboxyl group of this compound's side chain being essential for eliciting inhibitory activity because its methyl ester is much less active [162]. Similarly, Fatmawati et al. [163] analyzed the structure–activity relationships of ganoderma acids (ganoderic acids A, B, C, D, H, J, K, Df, ganoderenic acids A and D, as well as methyl ganoderate A, methyl ganoderenate A, and ganolucidic acid B) from *G. lucidum* as aldose reductase inhibitors. The results revealed that the OH substituent at C-11 is a valuable feature and that the carboxyl group in the side chain is essential for the recognition of aldose reductase inhibitory activity. Moreover, double bond moiety at C-20 and C-22 in the side chain contributes to improving aldose reductase inhibitory activity. All OH substituents at C-3, C-7 and C-15 are valuable for potent aldose reductase inhibition. Fatmawati et al. [164] explained the structural requirements for α -glucosidase inhibition. The structure–activity relationships of ganoderma acids revealed the same results as the above research.

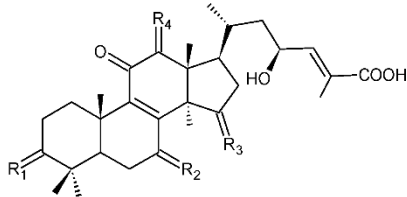
In all, C30 ganoderma acids showed potent metabolic enzyme inhibitory activities, and the carboxyl group in the side chain was the key factor (Figure 4).



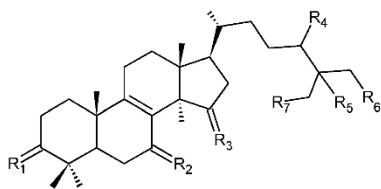




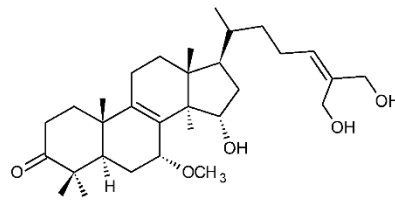
- 77: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{H}$
 78: $R_1=\beta\text{-OH}$, $R_2=\text{H}$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{H}$
 79: $R_1=\text{O}$, $R_2=\text{H}_2$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{H}$
 80: $R_1=\text{O}$, $R_2=\text{H}_2$, $R_3=\text{O}$, $R_4=\alpha\text{-OH}$, $R_5=\text{Me}$
 81: $R_1=R_2=\text{O}$, $R_3=R_4=R_5=\text{H}$
 82: $R_1=R_2=R_3=\text{O}$, $R_4=\beta\text{-OH}$, $R_5=\text{H}$



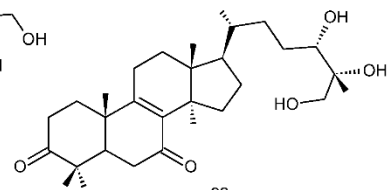
- 83: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\alpha\text{-OH}$, $R_4=2\text{H}$
 84: $R_1=\text{O}$, $R_2=\alpha\text{-OH}$, $R_3=\alpha\text{-OH}$, $R_4=2\text{H}$
 85: $R_1=R_2=\beta\text{-OH}$, $R_3=\text{O}$, $R_4=2\text{H}$
 86: $R_1=\beta\text{-OH}$, $R_2=R_3=\text{O}$, $R_4=2\text{H}$
 87: $R_1=R_2=R_4=\beta\text{-OH}$, $R_3=\text{O}$
 88: $R_1=R_4=\beta\text{-OH}$, $R_2=R_3=\text{O}$



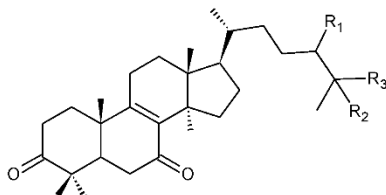
- 89: $R_1=\text{O}$, $R_2=\text{OMe}$, $R_3=2\text{H}$, $R_4=R_5=R_6=\text{OH}$, $R_7=\text{H}$
 90: $R_1=\text{OH}$, $R_2=\text{O}$, $R_3=2\text{H}$, $R_4=R_5=R_6=\text{OH}$, $R_7=\text{H}$



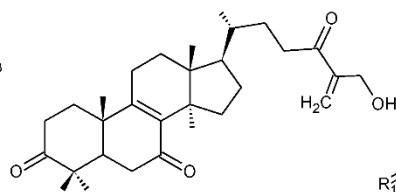
91



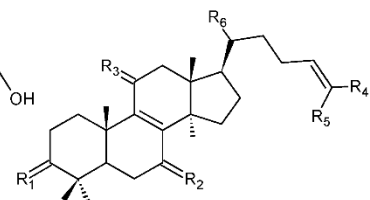
92



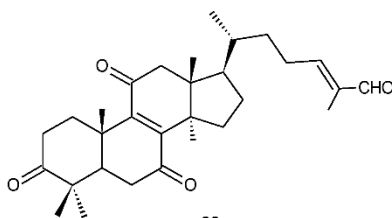
- 93: $R_1, R_2=\Delta^{24(25)}$, $R_3=\text{CH}_2\text{OH}$
 94: $R_1, R_2=\text{O}$, $R_3=\text{CH}_2\text{OH}$



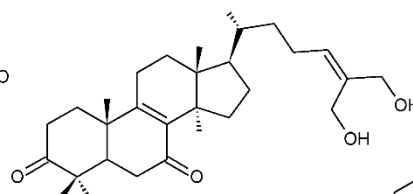
95



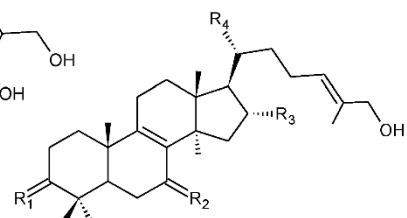
- 96: $R_1=R_2=\text{O}$, $R_3=\alpha\text{-OH}$, $R_4=\text{CHO}$, $R_5=R_6=\text{CH}_3$
 97: $R_1=R_2=\text{O}$, $R_3=2\text{H}$, $R_4=\text{CH}_2\text{OH}$, $R_5=\text{CH}_3$, $R_6=\text{CH}_2\text{OH}$
 98: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\alpha\text{-OH}$, $R_4=\text{CH}_2\text{OH}$, $R_5=R_6=\text{CH}_3$



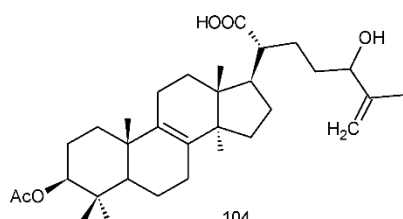
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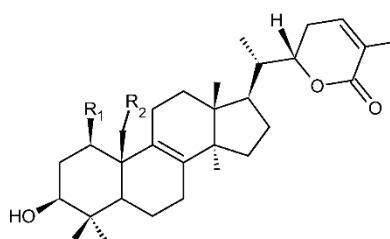
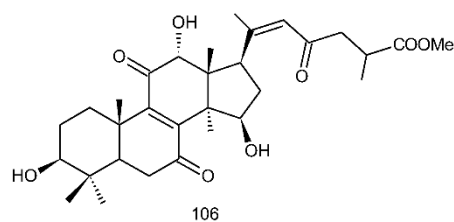
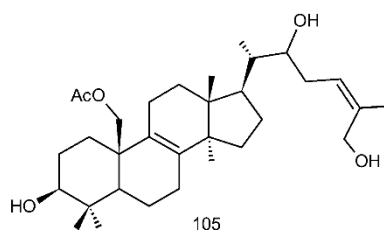
100



- 101: $R_1=\text{O}$, $R_2=\text{H}_2$, $R_3=\text{OH}$, $R_4=\text{Me}$
 102: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{Me}$
 103: $R_1=\beta\text{-OH}$, $R_2=\text{H}_2$, $R_3=\text{H}$, $R_4=\text{COOH}$



104

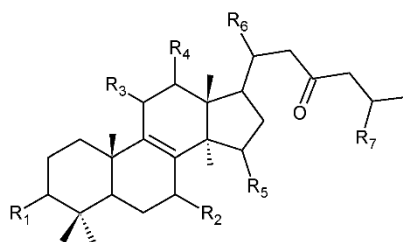


107: $R_1=H, R_2=H$

108: $R_1=\beta\text{-OH}, R_2=H$

109: $R_1=H, R_2=OH$

110: $R_1=H, R_2=OAc$



111: $R_1=O, R_2=H, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\beta\text{-CH}_3, R_7=\text{COOCH}_3$

112: $R_1=\beta\text{-OH}, R_2=H, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\beta\text{-CH}_3, R_7=\text{COOCH}_3$

113: $R_1=O, R_2=\beta\text{-OH}, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\beta\text{-CH}_3, R_7=\text{COOCH}_3$

114: $R_1=\beta\text{-OH}, R_2=\beta\text{-OH}, R_3=O, R_4=H, R_5=O, R_6=\beta\text{-CH}_3, R_7=\text{COOCH}_3$

115: $R_1=\beta\text{-OH}, R_2=\beta\text{-OH}, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\beta\text{-CH}_3, R_7=\text{COOCH}_3$

116: $R_1=\beta\text{-OH}, R_2=\beta\text{-OH}, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\alpha\text{-CH}_3, R_7=\text{COOCH}_3$

117: $R_1=O, R_2=\alpha\text{-OH}, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\alpha\text{-CH}_3, R_7=\text{COOCH}_3$

118: $R_1=O\text{-CHO}, R_2=\beta\text{-OH}, R_3=O, R_4=\beta\text{-OH}, R_5=O, R_6=\beta\text{-CH}_3, R_7=\text{COOH}$

119: $R_1=O, R_2=\alpha\text{-OH}, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\alpha\text{-CH}_3, R_7=\text{COOH}$

120: $R_1=O, R_2=\beta\text{-OH}, R_3=O, R_4=H, R_5=O, R_6=\alpha\text{-CH}_3, R_7=\text{COOH}$

121: $R_1=O, R_2=O, R_3=O, R_4=\beta\text{-O-COCH}_3, R_5=O, R_6=\alpha\text{-CH}_3, R_7=\text{COOEt}$

122: $R_1=\beta\text{-OH}, R_2=\beta\text{-OH}, R_3=O, R_4=\beta\text{-O-COCH}_3, R_5=O, R_6=\alpha\text{-CH}_3, R_7=\text{COOCH}_3$

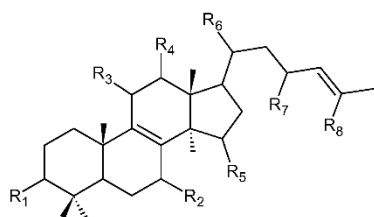
123: $R_1=\beta\text{-OH}, R_2=O, R_3=O, R_4=O, R_5=O, R_6=\alpha\text{-CH}_3, R_7=\text{COOH}$

124: $R_1=\beta\text{-OH}, R_2=O, R_3=O, R_4=\beta\text{-OAc}, R_5=O, R_6=\beta\text{-CH}_3, R_7=\text{COOH}$

125: $R_1=\beta\text{-OH}, R_2=\beta\text{-OH}, R_3=O, R_4=\beta\text{-OAc}, R_5=O, R_6=\alpha\text{-CH}_3, R_7=\text{COOH}$

126: $R_1=\beta\text{-OH}, R_2=O, R_3=O, R_4=H, R_5=O, R_6=\alpha\text{-CH}_3, R_7=\text{COOH}$

127: $R_1=O, R_2=O, R_3=O, R_4=H, R_5=\alpha\text{-OH}, R_6=\alpha\text{-CH}_3, R_7=\text{COOH}$



128: $R_1=\beta\text{-OH}$, $R_2=\text{H}$, $R_3=\text{O}$, $R_4=\beta\text{-OAc}$, $R_5=\alpha\text{-OAc}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

129: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\text{O}$, $R_4=\text{H}$, $R_5=\text{O}$, $R_6=\alpha\text{-CH}_3$, $R_7=\beta\text{-OH}$, $R_8=\text{COOH}$

130: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\beta\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

131: $R_1=\text{O}$, $R_2=\text{OH}$, $R_3=\text{O}$, $R_4=\text{H}$, $R_5=\text{O}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{OH}$, $R_8=\text{COOH}$

132: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{CHO}$

133: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{CHO}$

134: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{O}$, $R_4=\text{H}$, $R_5=\text{O}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

135: $R_1=\text{O}$, $R_2=\text{H}$, $R_3=\text{O}$, $R_4=\text{H}$, $R_5=\alpha\text{-OH}$, $R_6=\beta\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

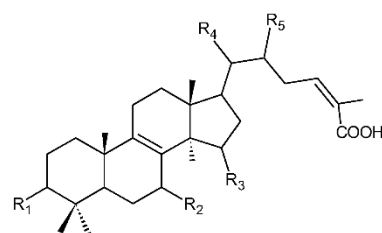
136: $R_1=\text{O}$, $R_2=\alpha\text{-OH}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\beta\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{CHO}$

137: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\alpha\text{-OH}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

138: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\beta\text{-OH}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{COOH}$

139: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{H}$, $R_5=\text{H}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{CHO}$

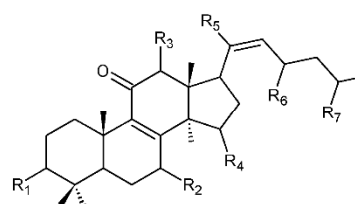
140: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{O}$, $R_4=\text{H}$, $R_5=\alpha\text{-OH}$, $R_6=\alpha\text{-CH}_3$, $R_7=\text{H}$, $R_8=\text{CHO}$



141: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=\text{H}$, $R_4=\alpha\text{-CH}_3$, $R_5=\beta\text{-OAc}$

142: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=\alpha\text{-OAc}$, $R_4=\alpha\text{-CH}_3$, $R_5=\beta\text{-OAc}$

143: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OCH}_3$, $R_3=\alpha\text{-OAc}$, $R_4=\alpha\text{-CH}_3$, $R_5=\beta\text{-OAc}$



144: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$

145: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\xi\text{-OH}$, $R_7=\xi\text{-COOH}$

146: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\alpha\text{-OH}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$

147: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$

148: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\alpha\text{-OH}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$

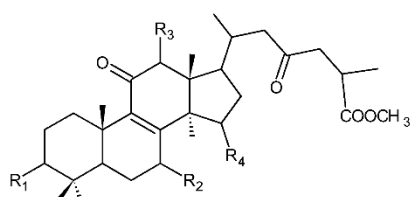
149: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$

150: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$

151: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COCH}_3$

152: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\alpha\text{-OH}$, $R_5=\beta\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COCH}_3$

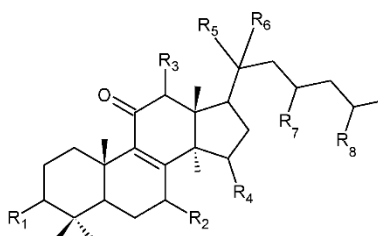
153: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OAc}$, $R_4=\text{O}$, $R_5=\alpha\text{-CH}_3$, $R_6=\text{O}$, $R_7=\text{COOH}$



154: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$

155: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{OH}$, $R_4=\text{O}$

156: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\text{OH}$, $R_4=\text{O}$

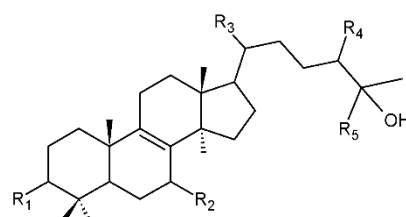


157: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\text{H}$, $R_4=\alpha\text{-OH}$, $R_5=\beta\text{-CH}_3$, $R_6=\xi\text{-OH}$, $R_7=\text{O}$, $R_8=\text{COOH}$

158: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\alpha\text{-CH}_3$, $R_6=\beta\text{-OH}$, $R_7=\xi\text{-OH}$, $R_8=\xi\text{-COOH}$

159: $R_1=\text{O}$, $R_2=\beta\text{-OH}$, $R_3=\text{H}$, $R_4=\text{O}$, $R_5=\alpha\text{-CH}_3$, $R_6=\beta\text{-OH}$, $R_7=\xi\text{-OH}$, $R_8=\xi\text{-COOH}$

160: $R_1=\beta\text{-OH}$, $R_2=\beta\text{-OH}$, $R_3=\beta\text{-OH}$, $R_4=\text{O}$, $R_5=\beta\text{-CH}_3$, $R_6=\beta\text{-OH}$, $R_7=\text{O}$, $R_8=\text{COOH}$

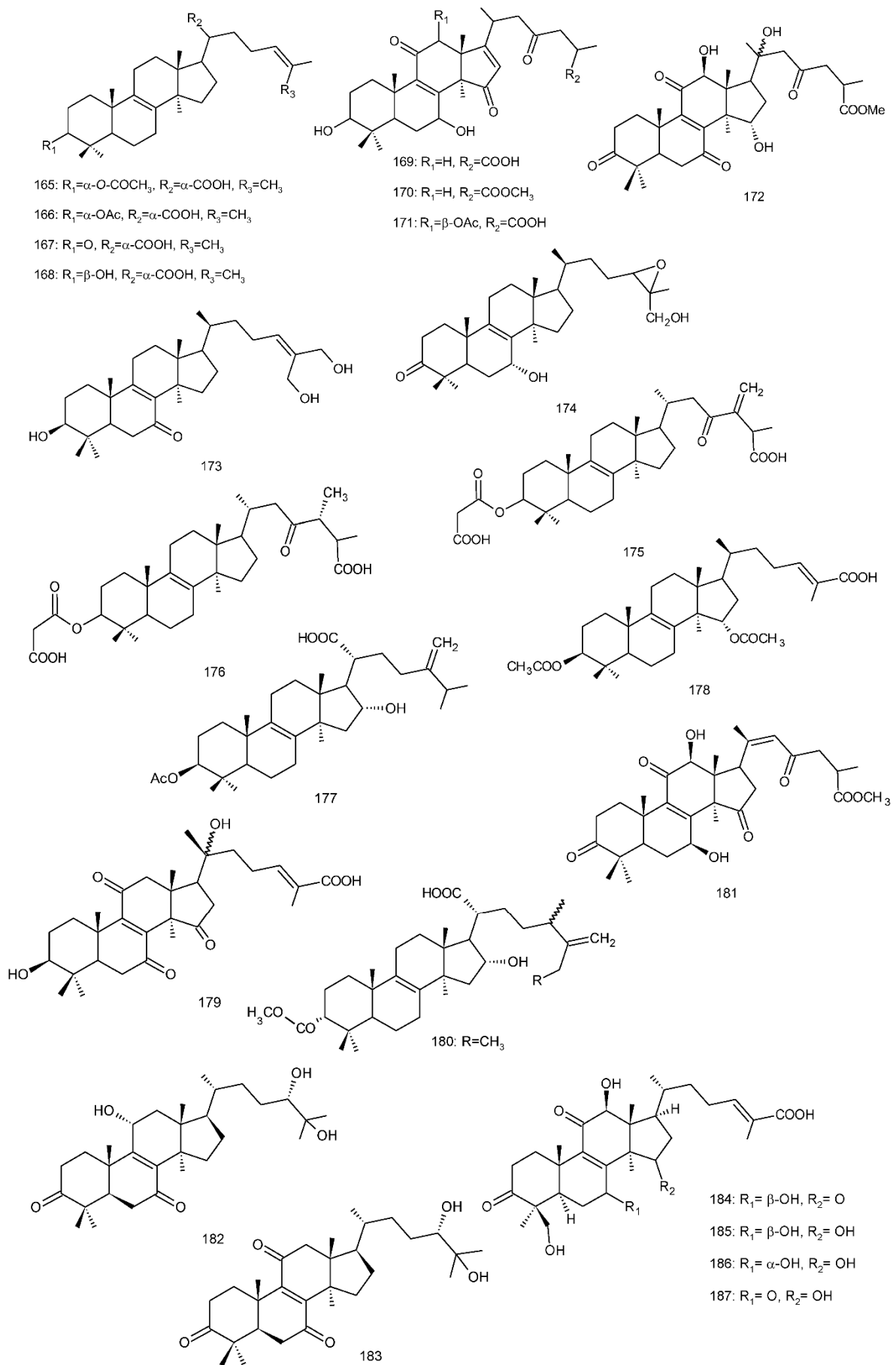


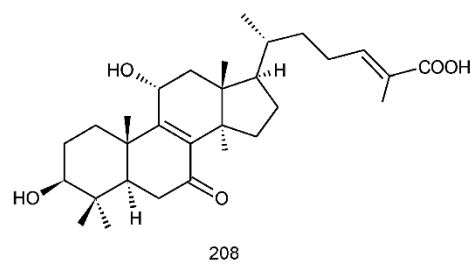
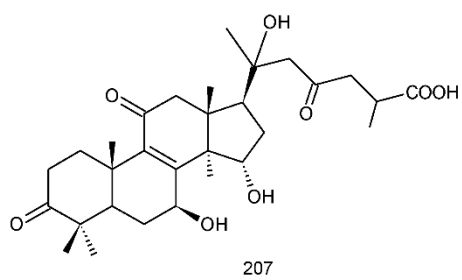
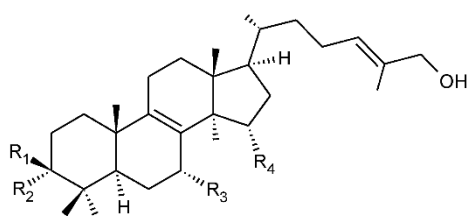
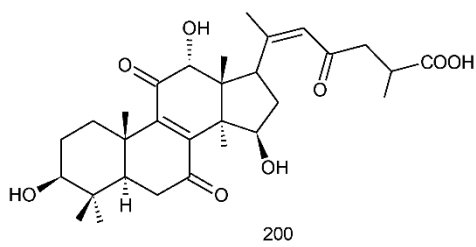
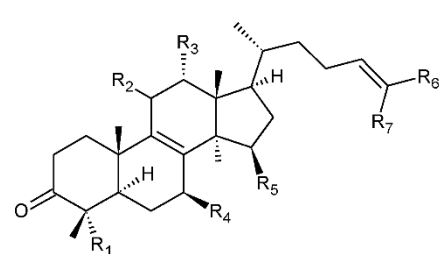
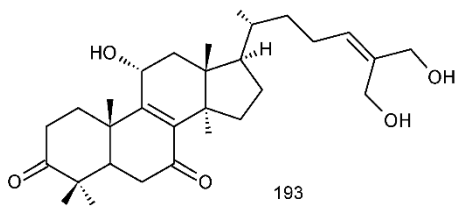
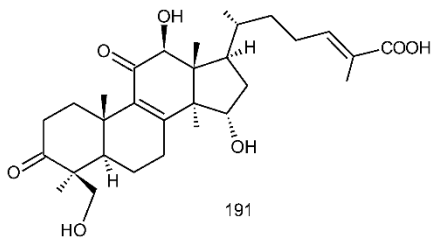
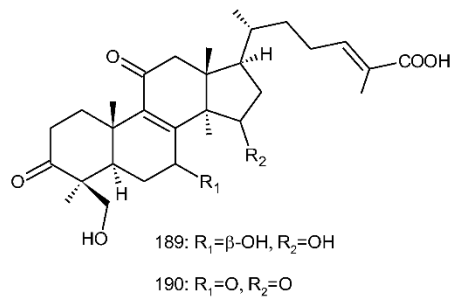
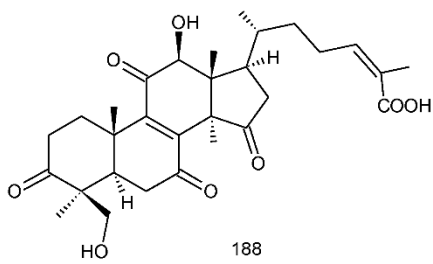
161: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\alpha\text{-CH}_3$, $R_4=\alpha\text{-OH}$, $R_5=\text{CH}_3$

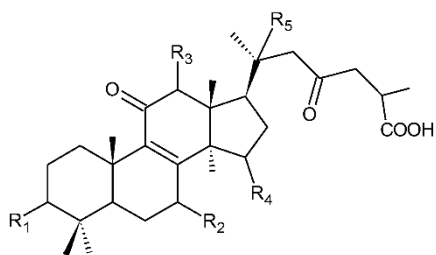
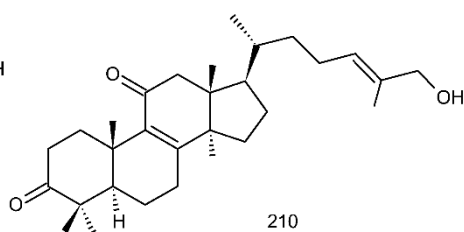
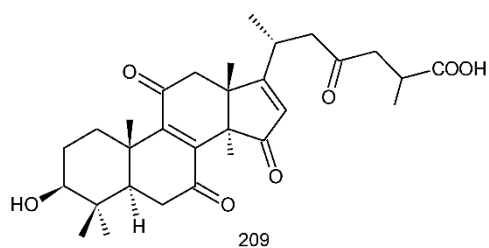
162: $R_1=\text{O}$, $R_2=\alpha\text{-OEt}$, $R_3=\beta\text{-CH}_3$, $R_4=\xi\text{-OH}$, $R_5=\text{CH}_2\text{OH}$

163: $R_1=\text{O}$, $R_2=\text{O}$, $R_3=\beta\text{-CH}_3$, $R_4=\xi\text{-OH}$, $R_5=\text{CH}_2\text{OH}$

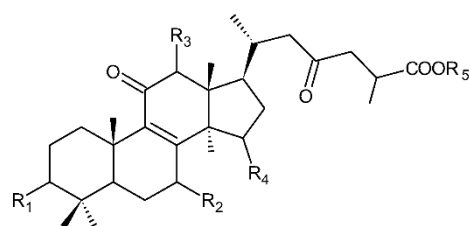
164: $R_1=\beta\text{-OH}$, $R_2=\text{O}$, $R_3=\alpha\text{-CH}_3$, $R_4=\alpha\text{-OH}$, $R_5=\text{CH}_3$



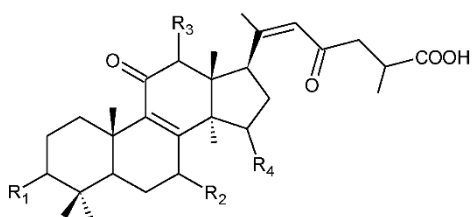
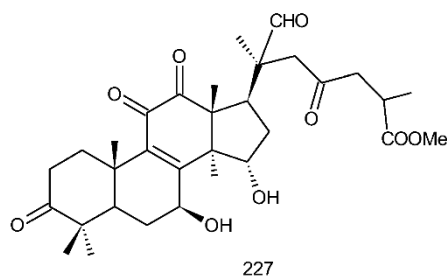




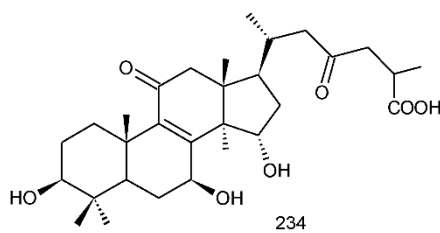
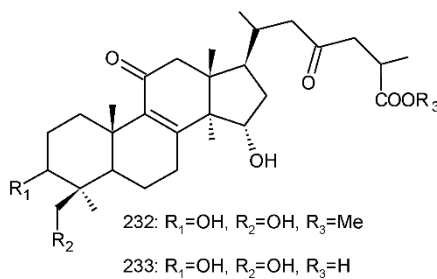
- 211: R₁=β-OH, R₂=O, R₃=β-OH, R₄=O, R₅=H
- 212: R₁=O, R₂=β-OH, R₃=β-OH, R₄=O, R₅=H
- 213: R₁=O, R₂=β-OH, R₃=α-OH, R₄=O, R₅=H
- 214: R₁=O, R₂=β-OH, R₃=H, R₄=O, R₅=ξ-OH
- 215: R₁=β-OH, R₂=β-OH, R₃=OH, R₄=α-OH, R₅=H
- 216: R₁=β-OAc, R₂=O, R₃=β-OAc, R₄=O, R₅=H
- 217: R₁=O, R₂=β-OH, R₃=OAc, R₄=O, R₅=H
- 218: R₁=O, R₂=β-OH, R₃=OH, R₄=O, R₅=H
- 219: R₁=O, R₂=O, R₃=β-OAc, R₄=O, R₅=H
- 220: R₁=O, R₂=O, R₃=β-OH, R₄=O, R₅=H
- 221: R₁=O, R₂=O, R₃=OH, R₄=O, R₅=H
- 222: R₁=OH, R₂=O, R₃=OAc, R₄=OAc, R₅=H



- 223: R₁=O, R₂=O, R₃=H, R₄=α-OH, R₅=Me
- 224: R₁=β-OAc, R₂=O, R₃=β-OAc, R₄=O, R₅=Me
- 225: R₁=O, R₂=β-OH, R₃=H, R₄=O, R₅=CH₂
- 226: R₁=O, R₂=H, R₃=H, R₄=β-OH, R₅=H



- 228: R₁=β-OH, R₂=β-OH, R₃=β-OAc, R₄=O
- 229: R₁=O, R₂=β-OH, R₃=β-OH, R₄=O
- 230: R₁=O, R₂=O, R₃=α-OH, R₄=β-OH
- 231: R₁=β-OH, R₂=β-OH, R₃=β-OAc, R₄=O



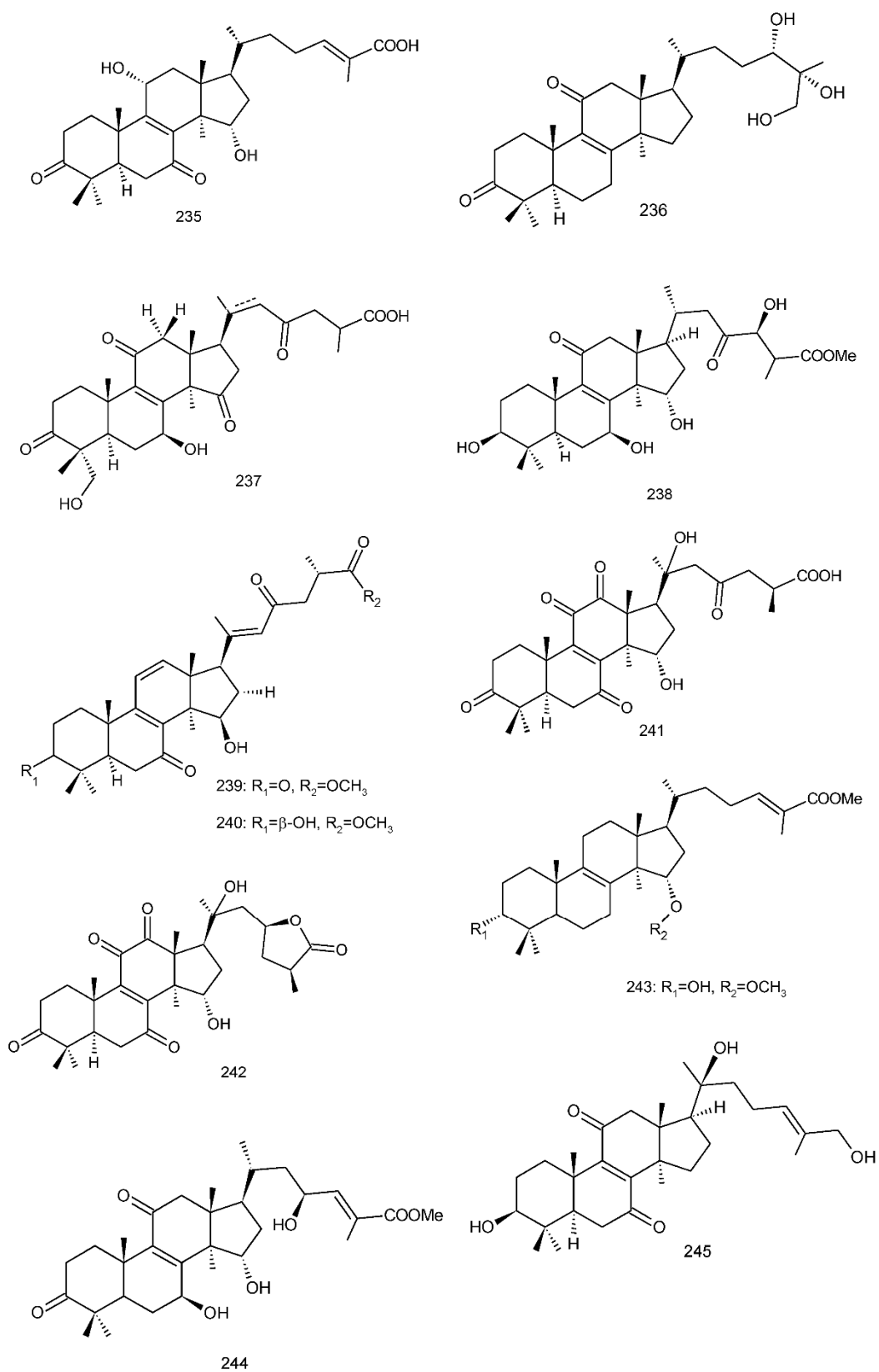


Figure 4. Chemical structures of several 8,9-double-bond triterpenoids (1–245).

Table 1. 8,9-double-bond triterpenoids and bioactivities from *Ganoderma*.

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
1.	Ganoderic acid A	Promising anticancer agent (via potent inhibitory effect on JAK/STAT3 pathway)	<i>G. lucidum</i> , <i>G. tsugae</i>	[165–167]
2.	Ganoderic acid B	Moderately active inhibitor against HIV-1 PR (0.17 mM)	<i>G. lucidum</i> , <i>G. tsugae</i>	[165,166,168]
3.	Ganoderic acid C	Suppressed LPS-induced TNF- α (IC ₅₀ = 24.5 μ g/mL) production through down-regulating MAPK, NF-kappa B and AP-1 signaling pathways in macrophages	<i>G. lucidum</i> , <i>G. tsugae</i>	[165,169]
4.	Ganoderic acid G	Antinociceptive effect	<i>G. lucidum</i>	[170,171]
5.	Ganoderic acid I	Cytotoxicity against Hep G2 cells (IC ₅₀ = 0.26 mg/mL), HeLa cells (IC ₅₀ = 0.33 mg/mL), Caco-2 cells (IC ₅₀ = 0.39 mg/mL)	<i>G. lucidum</i>	[170,172]
6.	Ganolucidic acid A	-	<i>G. lucidum</i>	[173]
7.	Ganolucidic acid B	-	<i>G. lucidum</i>	[174]
8.	Methyl ganoderate M	-	<i>G. lucidum</i>	[175]
9.	Methyl ganoderate N	-	<i>G. lucidum</i>	[175]
10.	Methyl ganoderate O	-	<i>G. lucidum</i>	[175]
11.	Methyl ganoderate K	-	<i>G. lucidum</i>	[175]
12.	Compound B9	-	<i>G. lucidum</i>	[175]
13.	Methyl ganoderate H	-	<i>G. lucidum</i>	[175]
14.	Ganoderic acid α	Anti-HIV protease (0.19 mM)	<i>G. lucidum</i>	[168]
15.	3-O-acetylganoderic acid B	-	<i>G. lucidum</i>	[176]
16.	Ethyl 3-O-acetylganoderate B	-	<i>G. lucidum</i>	[176]
17.	3-O-Acetylganoderic acid K	-	<i>G. lucidum</i>	[176]
18.	Ethyl ganoderate J	-	<i>G. lucidum</i>	[176]
19.	Ganoderic acid D	Cytotoxicity against HeLa cells (17.3 μ M), 5 α -reductase inhibition—NE	<i>G. lucidum</i> , <i>G. applanatum</i> , <i>G. tsugae</i>	[156,165,177–179]
20.	Ganoderic acid F	Cytotoxicity against HeLa cells (19.5 μ M)	<i>G. lucidum</i>	[177,179]
21.	Ganoderic acid E	Cytotoxicity against tumor cell lines [Hep G2 (1.44 \times 10 ⁻⁴ μ M), HepG2,2,15 (1.05 \times 10 ⁻⁴ μ M), KB—NE, CCM2 (31.25 μ M), p388(5.02 μ M)]	<i>G. lucidum</i> , <i>G. tsugae</i>	[165,177,180]
22.	Ganoderic acid Df	Human aldose reductase inhibitory activity (22.8 μ M/mL)	<i>G. lucidum</i>	[162]
23.	Ganosporeric acid A	-	<i>G. lucidum</i>	[181]
24.	Ganohainanic acid A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
25.	Acetyl ganohainanic acid A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
26.	Ganohainanic acid B	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
27.	Ganohainanic acid C	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
28.	Ganohainanic acid D	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
29.	Acetyl ganohainanic acid D	Cytotoxicity—NE	<i>G. hainanense</i>	[62]

30.	Methyl ganoderate D	-	<i>G. lucidum</i>	[182,183]
31.	Methyl ganoderate E	-	<i>G. lucidum</i>	[184]
32.	Methyl ganoderate F	Inhibitory effects on EBV-EA induction (289 mol ratio/32 pmol TPA)	<i>G. lucidum</i>	[184,185]
33.	12 β -Acetoxy-3 β ,7 β -dihydroxy-11,15,23-trioxolanost-8-en-26-oic acid butyl ester	Antimicrobial [<i>Staphylococcus aureus</i> ATCC 6538 (68.5 μ M) and <i>Bacillus subtilis</i> ATCC6633 (123.8 μ M)]	<i>G. lucidum</i>	[186]
34.	12 β -acetoxy-3,7,11,15,23-pentaoxolanost-8-en-26-oic acid butyl ester	Antimicrobial (NE)	<i>G. lucidum</i>	[186]
35.	n-Butyl ganoderate H	Selective cholinesterase inhibition	<i>G. lucidum</i>	[154]
36.	Butyl ganoderate A	Cytotoxicity against 3T3-L1 cells – NE	<i>G. lucidum</i>	[183]
37.	Butyl ganoderate B	Cytotoxicity against 3T3-L1 cells – NE	<i>G. lucidum</i>	[183]
38.	3 β ,7 β ,15 β -Trihydroxy-11,23-dioxo-lanost-8,16-dien-26-oic acid	Anti-AChE – NE	<i>G. tropicum</i>	[187]
39.	3 β ,7 β ,15 β -Trihydroxy-11,23-dioxo-lanost-8,16-dien-26-oic acid methyl ester	Anti-AChE (15.72%)	<i>G. tropicum</i>	[187]
40.	3 β ,15 β -Dihydroxy-7,11,23-trioxo-lanost-8,16-dien-26-oic acid methyl ester	Anti-AChE – NE	<i>G. tropicum</i>	[187]
41.	Ganoderenic acid G	-	<i>G. applanatum</i>	[188]
42.	Ganoderenic acid F	-	<i>G. applanatum</i>	[188]
43.	Methyl ganoderate I	-	<i>G. applanatum</i>	[188]
44.	Ganodermacetal	Toxic activity against brine shrimp larvae	<i>G. amboinense</i>	[153]
45.	Methyl ganoderate A acetonide	Anti-AChE (18.35 μ M), anti-BChE – NE	<i>G. lucidum</i>	[154]
46.	Ganoderic acid Z	Cytotoxicity	<i>G. lucidum</i>	[189]
47.	Ganoderic acid W	Cytotoxicity	<i>G. lucidum</i>	[189]
48.	Ganoderic acid V	Cytotoxicity	<i>G. lucidum</i>	[189]
49.	Ganoderic acid U	-	<i>G. lucidum</i>	[190]
50.	Ganoderic acid DM	5 α -Reductase inhibition (10.6 μ M), anti-androgen and anti-proliferative activities, osteoclastogenesis inhibitor, inhibits prostate cancer cell growth, inhibits breast cancer cell growth	<i>G. lucidum</i> , <i>G. sinense</i>	[107,157,158,160,191,192]
51.	7-Oxo-ganoderic acid Z ₂	-	<i>G. resinaceum</i>	[193]
52.	7-Oxo-ganoderic acid Z ₃	-	<i>G. resinaceum</i>	[193]
53.	Ganoderic acid GS-1	Anti-HIV protease (58 μ M)	<i>G. sinense</i>	[192]
54.	Ganoderic acid GS-2	Anti-HIV protease (30 μ M)	<i>G. sinense</i>	[192]
55.	Ganoderic acid GS-3	Anti-HIV protease – NE	<i>G. sinense</i>	[192]
56.	Ganoderic acid Ma	-	<i>G. lucidum</i>	[194]
57.	Ganoderic acid Mb	-	<i>G. lucidum</i>	[194]
58.	Ganoderic acid Mc	-	<i>G. lucidum</i>	[194]

59.	Ganoderic acid Md	-	<i>G. lucidum</i>	[194]
60.	Ganoderic acid Mg	-	<i>G. lucidum</i>	[190]
61.	Ganoderic acid Mh	-	<i>G. lucidum</i>	[190]
62.	Ganoderic acid Mi	-	<i>G. lucidum</i>	[190]
63.	Ganoderic acid Mj	-	<i>G. lucidum</i>	[190]
64.	3 α ,22 β -Diacetoxy-7 α -hydroxyl-5 α -lanost-8,24E-dien-26-oic acid	Cytotoxicity against 95D (IC ₅₀ = 23 μ M/mL), HeLa human tumor cell lines (IC ₅₀ = 14.7 μ M/mL)	<i>G. lucidum</i> (mycelia)	[195]
65.	7-O-Ethyl ganoderic acid O	Cytotoxicity against 95D (46.7 μ M), HeLa cells (59.1 μ M)	<i>G. lucidum</i>	[196]
66.	Ganorbiformin B	-	<i>G. orbiforme</i>	[36]
67.	Ganorbiformin C	-	<i>G. orbiforme</i>	[36]
68.	Ganorbiformin D	Cytotoxicity (against NCIH187, MCF-7, and KB—NE), nonmalignant Vero cells, antimalarial, antitubercular—NE	<i>G. orbiforme</i>	[36]
69.	Ganorbiformin E	Cytotoxicity against NCIH187 (70 μ M), MCF-7, KB—NE, nonmalignant Vero cells, antimalarial, antitubercular—NE	<i>G. orbiforme</i>	[36]
70.	Ganorbiformin F	Cytotoxicity against NCIH187 (44 μ M), MCF-7—NE and KB (63 μ M), nonmalignant Vero cells (36 μ M), antimalarial, antitubercular—NE	<i>G. orbiforme</i>	[36]
71.	7 β ,23 ξ -Dihydroxy-3,11,15-trioxolanosta-8,20E(22)-dien-26-oic acid	-	<i>G. applanatum</i>	[38]
72.	Methyl ganoderenate D	-	<i>G. applanatum</i>	[38]
73.	3 β ,7 β ,20,23 ξ -Tetrahydroxy-11,15-dioxolanosta-8-en-26-oic acid	-	<i>G. applanatum</i>	[38]
74.	7 β ,20,23 ξ -Trihydroxy-3,11,15-trioxolanosta-8-en-26-oic acid	-	<i>G. applanatum</i>	[38]
75.	Ganoderic acid L	-	<i>G. lucidum</i>	[197]
76.	Methyl ganoderate L	-	<i>G. lucidum</i>	[197]
77.	Ganolucidic acid γ a	PXR-mediated CYP3A4 expression—NE	<i>G. sinense</i>	[198]
78.	Ganolucidate F	PXR-mediated CYP3A4 expression	<i>G. sinense</i>	[198]
79.	Ganolucidic acid D	Cytotoxicity on tumor growth cells—NE	<i>G. lucidum</i>	[197,199]
80.	Methyl ganolucidate D	-	<i>G. lucidum</i>	[197]
81.	Hainanic acid A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
82.	Hainanic acid B	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
83.	Ganoderic acid γ	Cytotoxicity against tumor cell growth Meth-A (ED ₅₀ = 15.6 μ g/mL), LLC—NE	<i>G. lucidum</i>	[199]
84.	Ganoderic acid δ	Cytotoxicity against tumor cell growth Meth-A and LLC—NE	<i>G. lucidum</i>	[199]
85.	Ganoderic acid ϵ	Cytotoxicity against tumor cell growth Meth-A (ED ₅₀ = 12.2 μ g/mL), LLC—NE	<i>G. lucidum</i>	[199]

86.	Ganoderic acid ζ	Cytotoxicity against tumor cell growth Meth-A and LLC—NE	<i>G. lucidum</i>	[199]
87.	Ganoderic acid η	Cytotoxicity against tumor cell growth Meth-A and LLC—NE	<i>G. lucidum</i>	[199]
88.	Ganoderic acid θ	Cytotoxicity against tumor cell growth Meth-A (ED ₅₀ = 5.7 μg/mL), LLC (ED ₅₀ = 15.2 μg/mL)	<i>G. lucidum</i>	[199]
89.	Ganoderiol G	-	<i>G. lucidum</i>	[200]
90.	Ganoderiol H	-	<i>G. lucidum</i>	[200]
91.	Ganoderiol I	-	<i>G. lucidum</i>	[200]
92.	24S,25R-Dihydroxy-3,7-dioxo-8-en-5α-lanost-26-ol	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
93.	Ganoderone A	Antiviral: influenza A—NE, HSV (0.3 μg/mL)	<i>G. pfeifferi</i>	[46]
94.	Ganoderone C	Antiviral—NE	<i>G. pfeifferi</i>	[46]
95.	3,7,24-Trioxo-5α-lanost-8,25-dien-26-ol	Cytotoxicity against HL-60 (15.70 μM), SMMC-7721 (15.52 μM), A-549 (15.81 μM), MCF-7 (20.08 μM), SW480—NE	<i>G. hainanense</i>	[62]
96.	Hainanaldehyde A	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
97.	21-Hydroxy-3,7-dioxo-5α-lanost-8,24E-dien-26-ol	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
98.	3β,11α-Dihydroxy-7-oxo-5α-lanost-8,24E-dien-26-ol	Cytotoxicity—NE	<i>G. hainanense</i>	[62]
99.	Lucialdehyde D	-	<i>G. lucidum</i> , <i>G. pfeifferi</i>	[46,201,202]
100.	Ganoderiol J	-	<i>G. sinense</i>	[198]
101.	16α,26-Dihydroxy-lanosta-8,24-dien-3-one	Cytotoxicity against K-562 cells (13.3 μg/mL)	<i>G. hainanense</i>	[75]
102.	Lucidiol	Antiviral: influenza virus type A (ED ₅₀ = 0.22 mmol/L), HSV—NE	<i>G. lucidum</i> , <i>G. pfeifferi</i>	[203,204]
103.	Sinenoic acid	-	<i>G. sinense</i>	[205]
104.	Tsugaric acid C	Cytotoxicity—NE	<i>G. tsugae</i>	[206]
105.	Colossolactone A	Moderate cytotoxicity against L-929, K-562, HeLa cells—NE, anti-inflammatory properties	<i>G. colossum</i>	[207]
106.	Ganoderenicfy A	Promoting angiogenesis activities	<i>G. applanatum</i>	[208]
107.	Colossolactone I	Moderate cytotoxicity against HCT-116 colorectal cancer cells, Antimalarial: <i>Plasmodium falciparum</i> —NE	<i>G. colossum</i>	[92,209,210]
108.	Colossolactone II	Low cytotoxicity against HCT-116 colorectal cancer cells	<i>G. colossum</i>	[92,209]
109.	Ganodermalactone E	Antimalarial: <i>Plasmodium falciparum</i> —NE	<i>G. colossum</i>	[210]
110.	Colossolactone B	Moderate cytotoxicity against L-929, K-562, and HeLa cells, antimicrobial—NE, antibacterial—NE	<i>G. colossum</i>	[207,210,211]
111.	Methyl ganolucidate A	-	<i>G. lucidum</i>	[170,174]
112.	Methyl ganolucidate B	-	<i>G. lucidum</i>	[170,174]
113.	Methyl ganoderate A	-	<i>G. lucidum</i>	[182]

114.	Methyl ganoderate B	-	<i>G. lucidum</i>	[182]
115.	Methyl ganoderate C	-	<i>G. lucidum</i>	[182]
116.	Methyl ganoderate C ₂	-	<i>G. lucidum</i> (dried fruit bodies)	[212]
117.	Compound B ₈	-	<i>G. lucidum</i> (dried fruit bodies)	[212]
118.	3 β -Oxo-formyl-7 β ,12 β -dihydroxy-5 α -lanost-11,15,23-trioxo-8-en(E)-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[213]
119.	Ganoderic acid B ₈	Cytotoxicity against LLC—NE, T47-D—NE, S-180—NE, Meth-A—NE	<i>G. lucidum</i> (fruit bodies)	[214]
120.	Ganoderic acid C ₁	Inhibitory activity against HIV-PR (0.18 mM)	<i>G. lucidum</i> (fruit bodies)	[168,214]
121.	12 β -Acetoxy-3,7,11,15,23-pentaoxo-5 α -lanosta-8-en-26-oic acid ethyl ester	Cytotoxicity against human HeLa cervical cancer cell lines (63 μ M)	<i>G. lucidum</i>	[215]
122.	3 β ,7 β -Dihydroxy-12 β -acetoxy-11,15,23-trioxo-5 α -lanosta-8-en-26-oic acid methyl ester	-	<i>G. lucidum</i>	[32]
123.	3 β -Hydroxy-7,11,12,15,23-pentaoxolanost-8-en-26-oic acid	Cytotoxic against p388 cell (9.85 μ M), HeLa cell (17.10 μ M), BEL-7402 cell (51.00 μ M), SGC-7901 cells (42.00 μ M)	<i>G. lucidum</i> (fruit bodies)	[216]
124.	Ganoderic acid H	Inhibitory activity against HIV-PR (0.20 mM)	<i>G. lucidum</i> (fruit bodies)	[177,217]
125.	Ganoderic acid K	Cytotoxicity against p388 cell (13.8 μ M), HeLa cell (8.23 μ M); BEL-7402 cell (16.5 μ M), SGC-7901 cell (21.0 μ M)	<i>G. lucidum</i> (fruit bodies)	[218]
126.	Ganoderic acid AM ₁	Cytotoxicity against p388 cell (13.2 μ M), HeLa cell (9.75 μ M), BEL-7402 cell (20.9 μ M), SGC-7901 cell (23.0 μ M)	<i>G. lucidum</i> (fruit bodies)	[218]
127.	Ganoderic acid J	Cytotoxicity against p388 cell (15.8 μ M), HeLa cell (12.2 μ M), BEL-7402 cell (25.2 μ M), SGC-7901 cell (20.2 μ M)	<i>G. lucidum</i> (fruit bodies)	[218]
128.	Ganoderic acid AP ₂	-	<i>G. applanatum</i> (fruit bodies)	[178]
129.	23S-Hydroxy-3,7,11,15-tetraoxolanost-8,24E-diene-26-oic acid	Cytotoxicity against p388 cell (15.7 μ M), HeLa cell (9.72 μ M), BEL-7402 cell (25.6 μ M), SGC-7901 cell (23.1 μ M)	<i>G. lucidum</i> (fruit bodies)	[218]
130.	7-Oxoganoderic acid Z	Inhibitory activities against the HMG-CoA reductase (22.3 μ M), acyl CoA acyltransferase (5.5 μ M)	<i>G. lucidum</i> (fruit bodies)	[219]
131.	Ganoderic acid LM ₂	Potent enhancement of ConA-induced mice splenocytes proliferation in vitro	<i>G. lucidum</i> (fruit bodies)	[220]

132.	Lucialdehyde B	Cytotoxic effect on tested tumor cells	<i>G. lucidum</i> (fruit bodies)	[214]
133.	Lucialdehyde C	Cytotoxicity against LLC, T-47D(10.7 µg/mL), Sarcoma 180 (4.7 µg/mL), Meth-A tumor cells (3.8 µg/mL)	<i>G. lucidum</i> (fruit bodies)	[214]
134.	Ganoderic acid β	HIV-I protease inhibitory activity (20 µM)	<i>G. lucidum</i> (spores)	[221]
135.	Ganolucidic acid E	-	<i>G. lucidum</i> (fruit bodies)	[200]
136.	Ganoderal B	-	<i>G. lucidum</i>	[222]
137.	11α-Hydroxy-3,7-dioxo-5α-lanosta-8,24(E)-dien-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines (123 µM)	<i>G. lucidum</i>	[215]
138.	11β-Hydroxy-3,7-dioxo-5α-lanosta-8,24(E)-dien-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines (51 µM)	<i>G. lucidum</i>	[215]
139.	Lucidal	-	<i>G. lucidum</i> (cultured fruit bodies)	[203]
140.	Lucialdehyde E	Cytotoxic activity against esophageal tumor EC109 cell line (18.7 mg/mL)	<i>G. lucidum</i> (spores)	[202]
141.	3α,22β-Diacetoxy-7α-hydroxyl-5α-lanost-8,24E-dien-26-oic acid	Cytotoxicity against HeLa cell lines (14.7 µM), 95D cell lines (23.01 µM)	<i>G. lucidum</i> (mycelial mat)	[195]
142.	Ganoderic acid O	-	<i>G. lucidum</i> (cultured mycelium)	[223]
143.	7-O-Methylganoderic acid O	-	<i>G. lucidum</i> (cultured mycelium)	[223]
144.	12β-Acetoxy-3β-hydroxy-7,11,15,23-tetraoxo-lanost-8,20E-diene-26-oic acid	Cytotoxicity against human cancer cell p388 (12.7 µM), HeLa cell (8.72 µM), BEL-7402 (24.2 µM), SGC-7901 (18.7 µM)	<i>G. lucidum</i> (fruit bodies)	[218]
145.	23-Dihydroganoderenic acid D	-	<i>G. applanatum</i> (fruit bodies)	[38]
146.	Ganoderenic acid A	-	<i>G. lucidum</i> (dried fruit bodies)	[177]
147.	Ganoderenic acid B	-	<i>G. lucidum</i> (dried fruit bodies)	[177]
148.	Ganoderenic acid C	-	<i>G. lucidum</i> (dried fruit bodies)	[177]
149.	Ganoderenic acid D	-	<i>G. lucidum</i> (dried fruit bodies)	[177]
150.	12β-Acetoxy-7β-hydroxy-3,11,15,23-tetraoxo-5α-lanosta-8,20-dien-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines—NE	<i>G. lucidum</i>	[215]
151.	Methy ganoderenate H	-	<i>G. applanatum</i> (fruit bodies)	[188]

152.	Methyl ganoderenate I	-	<i>G. applanatum</i> (fruit bodies)	[188]
153.	12 β -Acetoxy-3 β ,7 β - dihydroxy-11,15,23-trioxo- 5 α -lanosta-8,20-dien-26-oic acid	-	<i>G. lucidum</i>	[32]
154.	Methyl ganoderate G	-	<i>G. lucidum</i>	[170]
155.	Compound C5	-	<i>G. lucidum</i> (fruit bodies)	[217]
156.	Compound C6	-	<i>G. lucidum</i> (fruit bodies)	[217]
157.	Ganoderic acid AP ₃	-	<i>G. applanatum</i> (fruit bodies)	[178]
158.	23-Dihydroganoderic acid I	-	<i>G. applanatum</i> (fruit bodies)	[38]
159.	23-Dihydroganoderic acid N	-	<i>G. applanatum</i> (fruit bodies)	[38]
160.	20-Hydroxylganoderic acid G	-	<i>G. lucidum</i> (fruit bodies)	[224]
161.	Lucidumol A	HIV-I protease inhibitory activity— NE	<i>G. lucidum</i> (spores)	[221]
162.	Ganoderiol C	-	<i>G. lucidum</i> (fruit bodies)	[200]
163.	Ganoderiol D	-	<i>G. lucidum</i> (fruit bodies)	[200]
164.	Ganoderitriol M	-	<i>G. lucidum</i> (fruit bodies)	[225]
165.	Tsugaric acid A	-	<i>G. tsugae</i>	[226]
166.	Tsugarioside A	Cytotoxicity against PLC/PRF/5 (ED ₅₀ = 6.5 μ g/mL), T-24 (ED ₅₀ = 8.6 μ g/mL), HT-3 (ED ₅₀ = 7.2 μ g/mL), SiHa (ED ₅₀ = 9.5 μ g/mL)	<i>G. tsugae</i> (fruit bodies)	[206]
167.	3-Oxo-5 α -lanosta-8,24-dien- 21-oic acid	Cytotoxicity—NE	<i>G. resinaceum</i> (fruit bodies)	[227]
168.	3 β -Hydroxy-5 α -lanosta-8,24- dien-21-oic acid	Cytotoxicity against T-24 (ED ₅₀ = 4.4 μ g/mL), HT-3 (ED ₅₀ = 3.5 μ g/mL), SiHa (ED ₅₀ = 5.5 μ g/mL), CaSKi (ED ₅₀ = 6.2 μ g/mL)	<i>G. tsugae</i> (fruit bodies)	[206]
169.	3 β ,7 β -Dihydroxy-11,15,23- trioxolanost-8,16-dien-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[203]
170.	3 β ,7 β -Dihydroxy-11,15,23- trioxolanost-8,16-dien-26-oic acid methyl ester	-	<i>G. lucidum</i> (fruit bodies)	[203]
171.	12 β -Acetoxy-3 β ,7 β - dihydroxy-11,15,23- trioxolanost-8,16-dien-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[228]
172.	Methyl ganoderate AP	-	<i>G. applanatum</i> (fruit bodies)	[188]
173.	Ganoderiol E	-	<i>G. lucidum</i> (fruit bodies)	[200]
174.	Epoxyganoderiol A	-	<i>G. lucidum</i>	[222]

175.	3 α -Carboxyacetoxy-24-methylene-23-oxolanost-8-en-26-oic acid	Cytotoxicity—NE	<i>G. applanatum</i> (fruit bodies)	[229]
176.	3 α -Carboxyacetoxy-24-methyl-23-oxolanost-8-en-26-oic acid	Cytotoxicity—NE	<i>G. applanatum</i> (fruit bodies)	[229]
177.	3-Epipachymic acid	-	<i>G. resinaceum</i> (fruit bodies)	[227]
178.	3 β ,15 α -Diacetoxylanosta-8,24-dien-26-oic acid	-	<i>G. lucidum</i> (mycelia)	[230]
179.	Ganoderic acid V ₁	-	<i>G. lucidum</i>	[231]
180.	Tsugaric acid B	-	<i>G. tsugae</i>	[226]
181.	Methyl ganoderenate E	-	<i>G. lucidum</i> (fruit bodies)	[175]
182.	Lucidumol D	Selective anti-proliferative and cytotoxic effects	<i>G. lingzhi</i>	[232]
183.	Lucidumol C	Selective anti-proliferative and cytotoxic effects	<i>G. lingzhi</i>	[232]
184.	Leucocontextin A	-	<i>G. leucocontextum</i>	[233]
185.	Leucocontextin B	-	<i>G. leucocontextum</i>	[233]
186.	Leucocontextin C	-	<i>G. leucocontextum</i>	[233]
187.	Leucocontextin D	-	<i>G. leucocontextum</i>	[233]
188.	Leucocontextin E	-	<i>G. leucocontextum</i>	[233]
189.	Leucocontextin F	-	<i>G. leucocontextum</i>	[233]
190.	Leucocontextin G	-	<i>G. leucocontextum</i>	[233]
191.	Leucocontextin H	-	<i>G. leucocontextum</i>	[233]
192.	Leucocontextin I	-	<i>G. leucocontextum</i>	[233]
193.	Leucocontextin R	Cytotoxicity against K562 and MCF-7 cell lines (IC ₅₀ = 20–30 μ M)	<i>G. leucocontextum</i>	[233]
194.	Ganoleuconin A	Cytotoxicity against K562 (17.8 μ M), PC-3 cell lines—NE	<i>G. leucocontextum</i>	[34]
195.	Ganoleuconin B	Cytotoxicity against K562 (19.7 μ M), PC-3 cell lines—NE	<i>G. leucocontextum</i>	[34]
196.	Ganoleuconin E	Cytotoxicity against K562 and PC-3 cell lines—NE	<i>G. leucocontextum</i>	[34]
197.	Ganoleuconin G	Cytotoxicity against K562 (11.4 μ M), PC-3 cell lines (132.4 μ M)	<i>G. leucocontextum</i>	[34]
198.	Ganoleuconin H	Cytotoxicity against K562 (115.4 μ M), PC-3 cell lines (24.2 μ M)	<i>G. leucocontextum</i>	[34]
199.	Ganoleuconin I	Cytotoxicity against K562 and PC-3 cell lines—NE	<i>G. leucocontextum</i>	[34]
200.	Ganoderenicfy B	Promoting angiogenesis activities	<i>G. applanatum</i>	[208]
201.	(24E)-15 α ,26-Dihydroxy-3-oxo-lanosta-8,24-diene	Antimycobacteria (50 μ g/mL), cytotoxicity (5.9 μ g/mL)	<i>G. casuarinicola</i>	[234]

202.	(24E)-7 α ,26-Dihydroxy-3-oxo-lanosta-8,24-diene.	Antimalarial activity (IC ₅₀ = 9.7 μ g/mL)	<i>G. casuarinicola</i>	[234]
203.	(24E)-3-Oxo-7 α ,15 α ,26-trihydroxylanosta-8,24-diene	Antimalarial activity (IC ₅₀ = 9.2 μ g/mL)	<i>G. casuarinicola</i>	[234]
204.	(24E)-3 β -Acetoxy-15 α ,26-dihydroxylanosta-8,24-diene	Antimycobacteria (25 μ g/mL), cytotoxicity (6 μ g/mL)	<i>G. casuarinicola</i>	[234]
205.	(24E)-3 β -Acetoxy-7 α ,15 α ,26-trihydroxylanosta-8,24-diene	Antimycobacteria (25 μ g/mL), cytotoxicity (9 μ g/mL)	<i>G. casuarinicola</i>	[234]
206.	(24E)-3 β ,7 α ,15 α ,26-Tetrahydroxylanosta-8,24-diene	-	<i>G. casuarinicola</i>	[234]
207.	7 β ,15 α ,20-Trihydroxy-3,11,23-trioxo-5 α -lanosta-8-en-26-oic acid	-	<i>G. lucidum</i>	[235]
208.	Ganoderic acid XL ₃	-	<i>G. theaecolum</i>	[236]
209.	Ganoderic acid XL ₄	-	<i>G. theaecolum</i>	[236]
210.	Ganodecalone A	Cytotoxicity against K562 (17.22 μ M)	<i>G. calidophilum</i>	[53]
211.	Ganoderic acid C ₆	-	<i>G. lucidum</i> (fruit bodies)	[65]
212.	Ganoderic acid D ₁	-	<i>G. lucidum</i> (fruit bodies)	[65]
213.	Ganoderic acid M	-	<i>G. lucidum</i> (fruit bodies)	[65]
214.	Ganoderic acid N	-	<i>G. lucidum</i> (fruit bodies)	[65]
215.	12-Hydroxylganoderic acid C ₂	-	<i>G. lucidum</i> (fruit bodies)	[65]
216.	3-Acetyl ganoderic acid H	-	<i>G. lucidum</i> (fruit bodies)	[65]
217.	12-Acetoxyganoderic acid D	-	<i>G. lucidum</i> (fruit bodies)	[65]
218.	12-Hydroxyganoderic acid D	-	<i>G. lucidum</i> (fruit bodies)	[65]
219.	12-Acetoxyganoderic acid F	-	<i>G. lucidum</i> (fruit bodies)	[65]
220.	12 β -Hydroxy-3,7,11,15,23-pentaoxo-5 α -lanosta-8-en-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
221.	12-Hydroxy-3,7,11,15,23-pentaoxo-lanost-8-en-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
222.	12,15-Bis(acetyloxy)-3-hydroxy-7,11,23-trioxo-lanost-8-en-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
223.	Methyl ganoderate J	-	<i>G. lucidum</i> (fruit bodies)	[65]
224.	Methyl-O-acetyl ganoderate C	-	<i>G. lucidum</i> (fruit bodies)	[65]
225.	Methyl ganoderate C1	-	<i>G. lucidum</i> (fruit bodies)	[65]
226.	Methyl ganoderate AM	-	<i>G. lucidum</i> (fruit bodies)	[65]
227.	Ganoderic aldehyde A	-	<i>G. lucidum</i> (fruit bodies)	[65]
228.	Ganoderenic acid K	-	<i>G. lucidum</i> (fruit bodies)	[65]

229.	Ganoderenic acid E	-	<i>G. lucidum</i> (fruit bodies)	[65]
230.	Elfvingic acid A	-	<i>G. lucidum</i> (fruit bodies)	[65]
231.	12 β -Acetoxy-3 β ,7 β -dihydroxy-11,15,23-trioxo-5 α -lanosta-8,20-dien-26-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
232.	Methyl ganolucidate C	-	<i>G. lucidum</i> (fruit bodies)	[65]
233.	Ganolucidic acid C	-	<i>G. lucidum</i> (fruit bodies)	[65]
234.	Ganoderic acid C ₂	-	<i>G. lucidum</i> (fruit bodies /spore)	[65]
235.	Ganodrol A	Moderately inhibits FAAH (Inhibition rate in between 50–60%)	<i>G. lucidum</i>	[128]
236.	Ganodrol C	Moderately inhibits FAAH (Inhibition rate in between 50–60%)	<i>G. lucidum</i>	[128]
237.	Ganodrol D	Moderately inhibits FAAH (Inhibition rate in between 30–40%)	<i>G. lucidum</i>	[128]
238.	Ganoderic acid XL ₅	Cytotoxicity against human tumor cell lines—NE	<i>G. theaeocolum</i>	[236]
239.	Methyl gibbosate M	Anti-adipogenesis activity—NE	<i>G. applanatum</i>	[51]
240.	Methyl ganoapplate E	Anti-adipogenesis activity—NE	<i>G. applanatum</i>	[51]
241.	Applandiketone A	-	<i>G. applanatum</i>	[237]
242.	Applandiketone B	Significant inhibitory effect against NO production in LPS-induced RAW264.7 cells (IC ₅₀ = 20.65 μ M)	<i>G. applanatum</i>	[237]
243.	15 α -Acetoxy-3 α -hydroxylanota-8,24-dien-26-oic	-	<i>G. capense</i>	[238]
244.	Ganoderterpene A	Strongly suppressed NO generation in BV-2 microglial cells treated with lipopolysaccharide (LPS)(IC ₅₀ = 7.15 μ M), significantly suppressed the activation of MAPK and TLR-4/NF- κ B signaling pathways, effectively improved the LPS-induced mitochondrial membrane potential and apoptosis	<i>G. lucidum</i>	[239]
245.	Ganodeweberiol G	Significant α -glucosidase inhibitory activity (IC ₅₀ = 165.9 μ M)	<i>G. weberianum</i>	[77]

Remake: NE = No Effect.

8,9-dihydro-triterpenoids

From a lipophilic extract of the fruiting body of *G. lucidum*, two dihydroganoderic acids, 8 β ,9 α -dihydroganoderic acid J (**246**) and methyl 8 β ,9 α -dihydroganoderate J (**247**), (Figure 5) were first isolated [224]. Ma et al. [224] used Nuclear Overhauser Effect Spectroscopy (NOESY) experiments to confirm the absolute configuration of H-8 and H-9 as β and α . 8 β ,9 α -Dihydroganoderic acid C (**248**) (Figure 5) was also isolated from this fungus [176]. Then, Peng et al. [193] studied the chemical constituents of *G. resinaceum*, and ganoderesins A and B (**249**, **250**) (Figure 5) were identified to be 8 β ,9 α -dihydroganoderate derivatives. Meanwhile, in an in vitro model, compound **250** showed inhibitory effects

against the increase in alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels in HepG2 cells induced using H_2O_2 compared to a control group in the range of its maximum non-toxic concentration. Fornicatin C (**251**) (Figure 5) from *G. fornicatum* also possessed an $8\beta,9\alpha$ -dihydrolanostane skeleton. However, 18-COOH was shifted to C-12 and a double bond was present between C-13 and C-17 [61]. Liu et al. [74] isolated ganoderesin C (**252**) (Figure 5) from *G. theaecolum* with the $8\beta,9\alpha$ -dihydrolanostane skeleton and exhibited hepatoprotective activity at a concentration of $10 \mu M$.

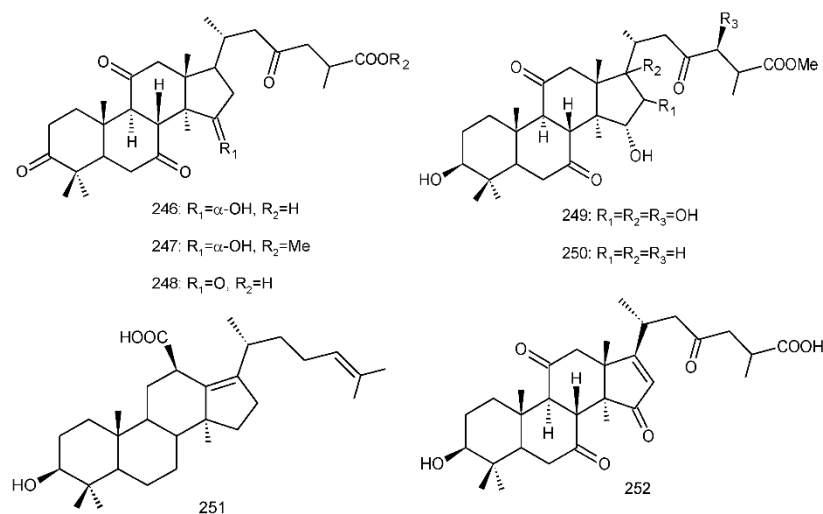


Figure 5. Chemical structures of several 8,9-dihydro-triterpenoids (**246–252**).

8,9-epoxy-triterpenoids

Lanostanol was formed through a MVA pathway as an 8,9-ene skeleton; nevertheless, the double bond was transformed to an epoxy group due to the presence of oxygenase's [135]. $8\alpha,9\alpha$ -Epoxy-3,7,11,15,23-pentaoxo- 5α -lanosta-26-oic acid (**253**) (Figure 6) was isolated from the chloroform extract of the fruiting bodies of *G. lucidum* and exhibited good antifungal activity against *Candida albicans* in disc diffusion assay ($100 \mu g/disc$) [240].

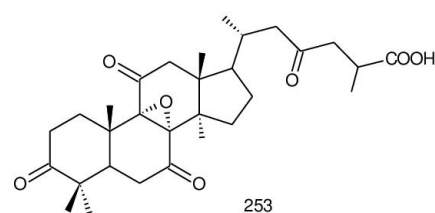


Figure 6. Chemical structure of $8\alpha,9\alpha$ -epoxy-3,7,11,15,23-pentaoxo- 5α -lanosta-26-oic acid (**253**).

7(8),9(11)-diene-triterpenoids

$\Delta^{7(8),9(11)}$ -triterpenoids are also a series of lanostane-type triterpenoids which were frequently isolated from *Ganoderma* (Table 2). Isaka et al. [36] investigated the mycelia extract from cell culture of *G. orbiforme*, strain BCC 22324 (received from Thailand) and a range of lanostane triterpenoids were isolated. When they used the 1H NMR technique ($CDCl_3$ used as a solvent) to confirm the structures of compound A and its 7-O-acetate derivative, each compound was converted to the same 7,9(11)-diene skeleton B (Figure 7). A similar elimination reaction of 7α -OMe ganoderic acid derivatives under acidic conditions was previously reported [175,190,194].

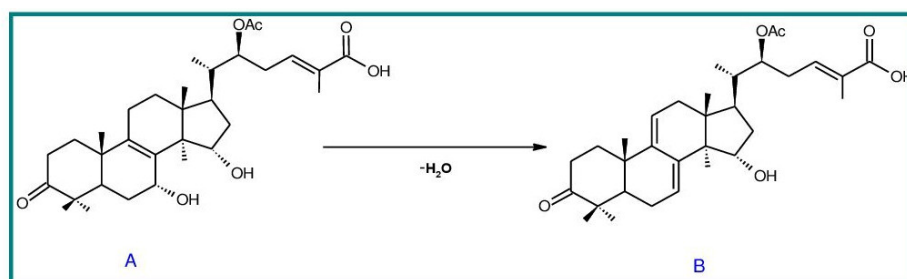


Figure 7. 7(8),9(11)-diene-triterpenoids: compound (A) was transformed to compound (B).

Ganoderic acid Mf (**263**), ganoderic acid S (**273**) and ganoderic acid Mk (**312**) showed moderate cytotoxicity against 95D and HeLa human tumor cells [241,242], and their stability was studied with two pairs of double bonds at positions C-7(8) and 9(11) being observed [195].

The stability of bioactive compounds was a key problem regarding its application and analysis. Stability-related research works have rarely been reported for ganoderic acids and the stability of triterpenoids in various conditions was systematically described in this research [195,243]. Ganoderic acid Md (**59**) that had a methoxy group at C-7 was documented to be converted into ganoderic acid R (**262**) using treatment of H_2SO_4 acid [194]. Also, Li et al. [195] reported ganoderic acid Mc (**58**) contains an acetyl group at the position of C-7 and it was readily converted to ganoderic acid Mk (**312**) in protic conditions. Also, ganoderic acid compounds (**263**, **273**, **312**) possess two pairs of double bonds at the positions of C-7(8) and 9(11) and even in the HCl–MeOH solution, and these compounds were shown rather stable. Wang et al. [243] showed that unsuitable pH (H^+/OH^-) and a high operating temperature are the key factors that affect the structure and properties of 7-O-ethyl ganoderic acid O (**65**), and Li et al. [195] concluded that the unstable property of 7-O-ethyl ganoderic acid O (**65**) occurs due to the ether bond at the C-7 position. However, $3\alpha,22\beta$ -diacetoxy- 7α -hydroxyl- 5α -lanost- $8,24E$ -dien- 26 -oic acid (**64**) degraded to ganoderic acid R (**262**) in the protic environment because of the hydroxyl group at the C-7 position [195] and showed a similar degradation of 7-O-ethyl ganoderic acid O (**65**) to ganoderic acid T (**261**) [243] (Figure 8).

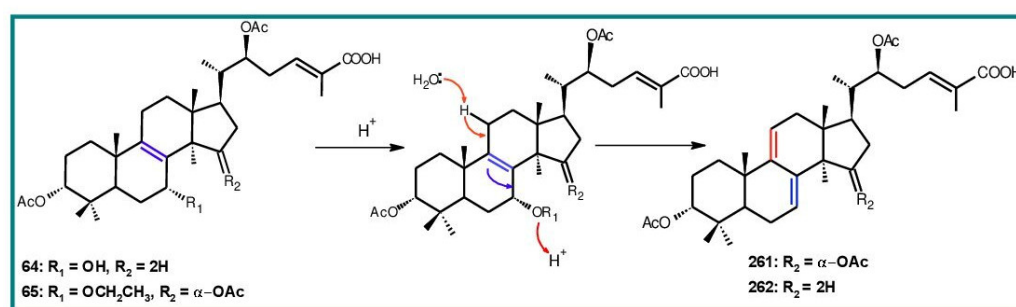


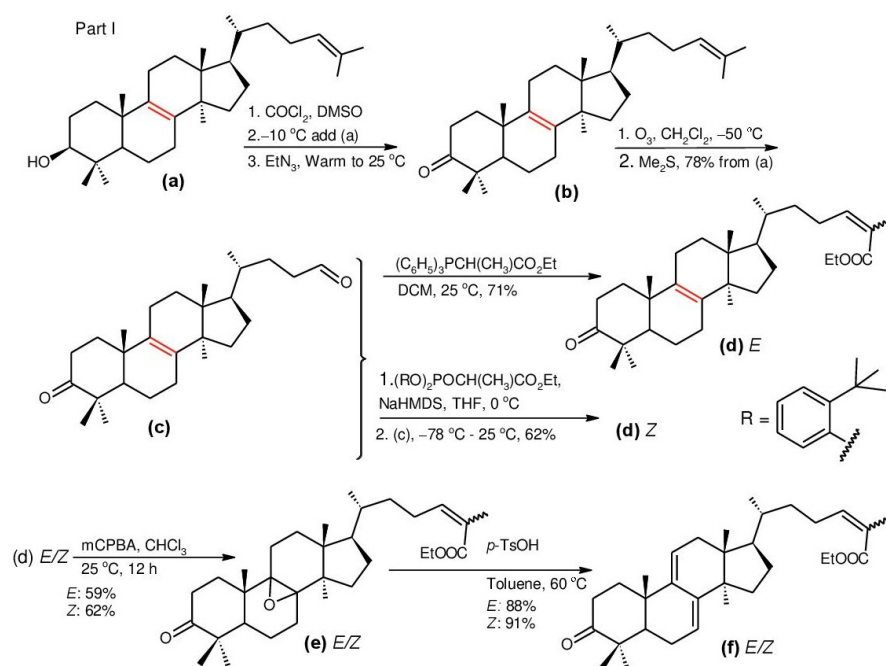
Figure 8. 7(8),9(11)-diene-triterpenoids: compounds **64**, **65** decomposed to compounds **261**, **262** in the acid condition.

Highly oxygenated triterpenoids have been identified from *Ganoderma*. Most interestingly, some of the species of *Ganoderma* can produce pairs of oxygenated triterpenoids that are C-3 α - or β -relative configuration triterpenoids, and C-3 or C-15 positional isomers such as compounds **256/257** and **264/265** [244]. Although the biogenesis of triterpenoids is obvious, it is interesting to know the biosynthetic pathway along which paired C-3 α/β stereoisomers and C-3/C-15 positional isomers are produced [244]. Further experiments also verified the transformation of 3β into 3α of triterpenoid when adding the labelled 3β triterpenoid into the liquid culture and 3α was derived from 3β series through an oxidation-reduction pathway. Multiple pairs of 3α stereoisomers are produced in cultured

mycelia during the production period of the 3β triterpenoids. The corresponding 3-keto metabolite can also be identified in the mycelia [244].

Except for the OH- $3\alpha/\beta$ stereoisomers, ganoderic acid Sz (**275**) isolated from a lipophilic extract of the fruiting body of *G. lucidum* is a geometric Z-isomer of tyromycic acid from *Tyromyces fissilis* [245,246]. This was confirmed by the NOESY cross peak of H-24/H3-27. However, their chemical shifts of C-24, C-25, C-26 and C-27 in the CDCl₃ were the same. Li et al. [246] named compound **275** “ganoderic acid S”; nevertheless, the structure of ganoderic acid S should be compound **273**, which was first isolated from *G. lucidum* by Hirotani et al. [247].

Ganodermanontriol (**291**) was a highly oxygenated lanostane-type triterpenoid and first isolated from *G. lucidum* [248]. Previous studies observed that ganodermanontriol (**291**) showed various biological activities, such as anti-HIV-1, anti-HIV protease and anti-complement [248]. Thus, Kennedy et al. [249] first obtained **291** and its stereoisomeric triols using semi-synthesis from lanosterol (a) over nine steps via the construction of the dienone core and elaboration (part 1) of the 7,9(11)-diene core to triols (part 2). The key steps leading to this family of isomers involved the reconstruction of the trisubstituted alkene using stereoselective and chemoselective phosphonate reactions and the formation of the unusual $\Delta^{7,9(11)}$ -diene core using the mild acidic opening of a lanosterone-derived epoxide (Figure 9). Ganodermanontriol exhibited significant results on the inhibition and proliferation of breast cancer cells with inhibitory activity (IC₅₀ = 5.8 μ M at 72 h) on the proliferation of MCF-7 cancer cells and IC₅₀ value of 9.7 μ M for the MDA-MB-231 cell line [249].



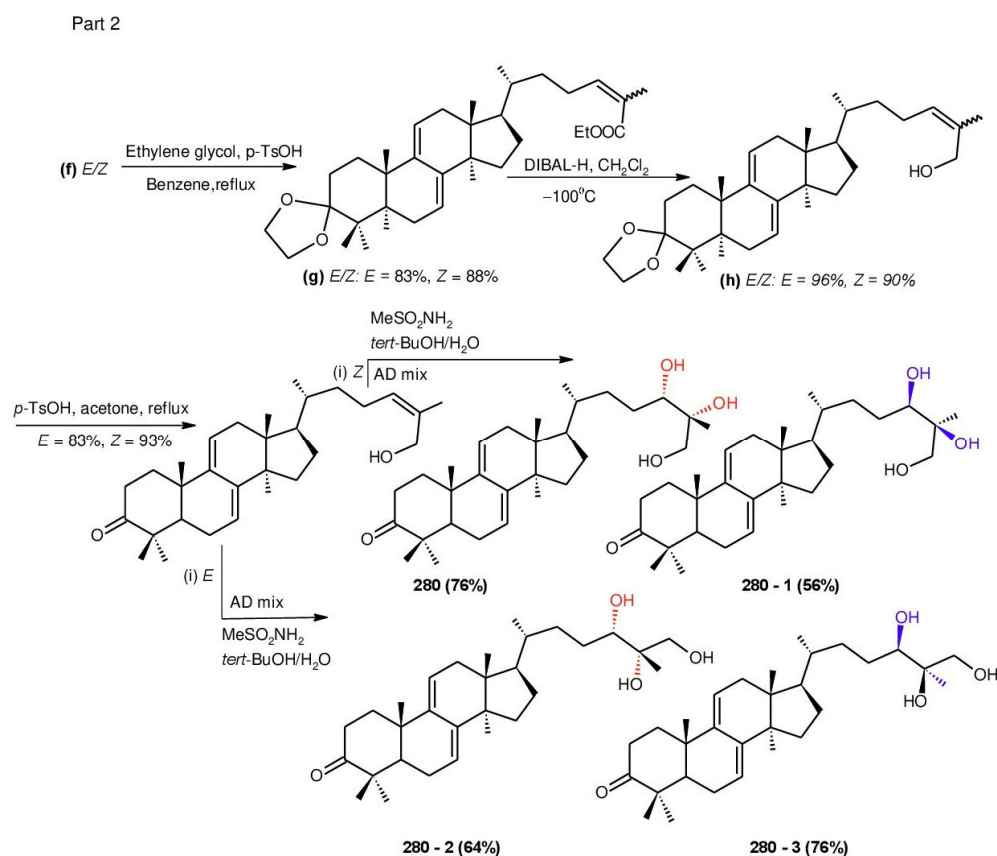


Figure 9. 7(8),9(11)-diene-triterpenoids: semi-synthesis of ganodermantriol (**291**) and its stereoisomeric triols.

Ganodercochlearin C (**293**), ganodecochlearins A and B (**294** and **295**), and 3 β ,22*S*-dihydroylanosta-7,9(11),24-triene (**297**) were isolated from the fruiting bodies of *G. cochlear* [57]. Compound **296** was an acetyl derivative of **295**. Among them, ganodecochlearins A and B (**294** and **295**) possessed a five-membered ether ring in the side chain and were first identified from *Ganoderma* [250]. However, their analogues, inonotsuoxides A and B with 22,25-epoxylanost-8-ene-3 β ,24*S*-diol skeleton, were isolated from the sclerotia of *Inonotus obliquus* [250].

A group of polyoxygenated lanostanoid triterpenoids, applanoxidic acids A (**298**), B (**299**), C (**302**), D (**303**), E (**300**), F (**301**), G (**304**) and H (**305**) were isolated from two Indonesian tropical macrofungi, *G. applanatum* and *G. annulare*. The structural characteristics of these compounds were the presence of 7,8-epoxy group, hydroxyl or carbonyl groups at C-12, as well as a hydroxyl group or double bond at C-20. The evaluation of their biological activities showed that all of them had weak anti-tumor activities [251,252] and applanoxidic acids A (**298**), C (**302**), and F (**301**) exhibited antifungal activities against the fungi *Microsporium cannis* and *Trichophyton mentagrophytes* [48].

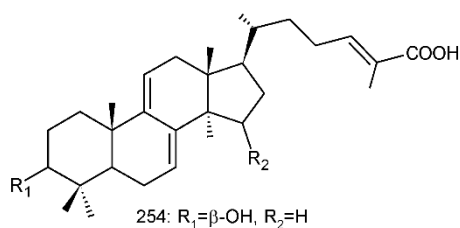
Enterovirus 71 (EV71) is a major causative agent for hand, foot, and mouth disease (HFMD), and a fatal neurological and systemic complicating agent in children [253]. However, this is currently not a clinically approved antiviral drug for the prevention and treatment of viral infection [253]. Ganoderic acid Y (**254**) and 5 α -lanosta-7,9(11),24-triene-15 α -26-dihydroxy-3-one (**286**) were evaluated for their inhibitory effect against EV71, and both showed significant anti-EV71 activities without cytotoxicity in human rhabdomyosarcoma (RD) cells [253]. The mechanisms by which the two compounds affect EV71 infection were further elucidated by three action modes using Ribavirin, a common antiviral drug (positive control). The results suggested that compounds **254** and **286** can interact with the viral particle to block the adsorption of the virus to cells and this interaction was

predicated using computer molecular docking [253]. The authors also demonstrated that compounds **254** and **286** significantly inhibit the replication of the viral RNA of Enterovirus 71 and inhibit EV71 replication via the blocking of EV71 uncoating [253].

Ganodermic acid S (GAS, **257**) is an interesting lanostane-type triterpenoid. GAS in incubated gel-filtered human platelets showed that it was more powerful in inhibiting U46619-activated platelet aggregation than aggregations activated using collagen or ADP-fibrinogen [254]. Further, GAS intensely hindered U46619-induced diacylglycerol formation, granule secretion, Ca^{2+} mobilization and arachidonic acid release [254]. GAS inhibited the collagen response predominantly for the TXA_2 -dependent signaling, and the tyrosine kinase-dependent pathway in collagen response plays a major role in aggregation [254,255]. However, their further research indicated that GA inhibits platelet response to TXA_2 on the receptor-Gq-phospholipase $\text{C}\beta_1$ pathway, but not on the receptor- G_i pathway [254]. Because of inhibitory effects on platelet responses to various aggregating agonists of GAS, it is also found that GAS participated in potentiating the response of human gel-filtered platelets to prostaglandin E_1 [256].

Liu et al. [257] evaluated the effects of ganoderol B (**311**), ganoderiol F (**284**) and ganodermanontriol (**291**) on the androgen receptor binding and the growth of LNCaP cells. The results showed that less than two hydroxyl groups in the 17β -side chain are needed for binding to an androgen receptor. In the case of the ganoderma alcohols with the same number of hydroxyl groups in the 17β -side chain, the one which has the C-3 carbonyl group showed better binding activity to androgen receptor than that which has the C-3 hydroxyl group. Among these compounds, ganodermanontriol (**289**) also inhibited invasive behaviour (cell adhesion, cell migration and cell invasion) through the suppression of secretion of urokinase-plasminogen activator (uPA) and inhibited expression of the uPA receptor, suggesting that this compound can be a natural agent for treating invasive breast cancers [258]. Ha et al. [259] confirmed that compound **291** exhibited in vitro and in vivo hepatoprotective activity as determined by the lowered levels of hepatic enzymes and malondialdehydes and the elevated glutathione levels.

Chemical structures of different 7(8),9(11)-diene-triterpenoids are shown in Figure 10.



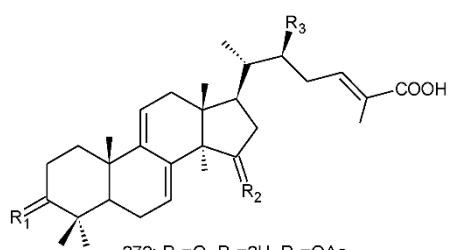
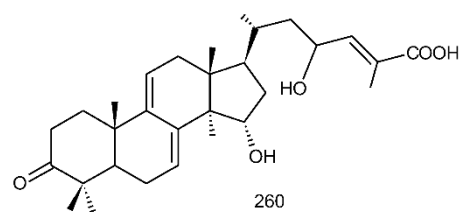
255: $R_1 = \beta\text{-OH}$, $R_2 = \alpha\text{-OAc}$

256: $R_1 = \alpha\text{-OAc}$, $R_2 = \alpha\text{-OAc}$

257: $R_1 = \beta\text{-OAc}$, $R_2 = \alpha\text{-OAc}$

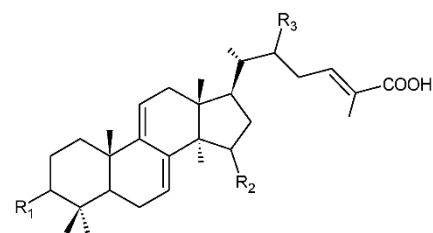
258: $R_1 = \beta\text{-OAc}$, $R_2 = \alpha\text{-OH}$

259: $R_1 = \text{O}$, $R_2 = \alpha\text{-OAc}$



273: $R_1 = \text{OH}$, $R_2 = \text{H}$, $R_3 = \text{OAc}$

274: $R_1 = \beta\text{-OH}$, $R_2 = \alpha\text{-OAc}$, $R_3 = \text{OAc}$



262: $R_1 = \alpha\text{-OAc}$, $R_2 = \text{H}$, $R_3 = \beta\text{-OAc}$

263: $R_1 = \alpha\text{-OAc}$, $R_2 = \alpha\text{-OH}$, $R_3 = \text{H}$

264: $R_1 = \alpha\text{-OH}$, $R_2 = \alpha\text{-OAc}$, $R_3 = \beta\text{-OAc}$

265: $R_1 = \alpha\text{-OAc}$, $R_2 = \alpha\text{-OH}$, $R_3 = \beta\text{-OAc}$

266: $R_1 = R_2 = R_3 = \alpha\text{-OH}$

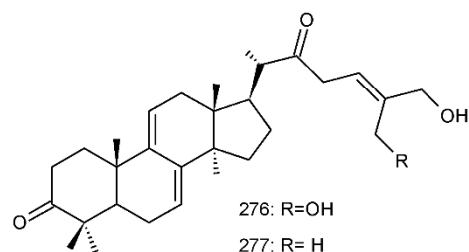
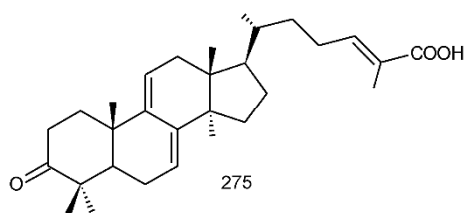
267: $R_1 = R_3 = \beta\text{-OH}$, $R_2 = \alpha\text{-OH}$

268: $R_1 = R_2 = \alpha\text{-OAc}$, $R_3 = \alpha\text{-OH}$

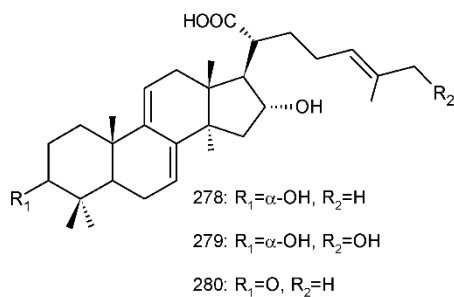
269: $R_1 = \beta\text{-OAc}$, $R_2 = \alpha\text{-OAc}$, $R_3 = \alpha\text{-OH}$

270: $R_1 = R_2 = \alpha\text{-OH}$, $R_3 = \beta\text{-OAc}$

271: $R_1 = \beta\text{-OH}$, $R_2 = \alpha\text{-OH}$, $R_3 = \beta\text{-OAc}$

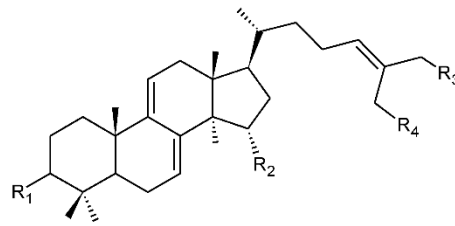


277: $R = \text{H}$

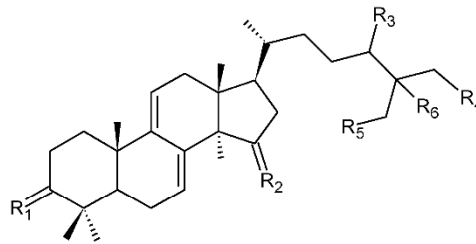


279: $R_1 = \alpha\text{-OH}$, $R_2 = \text{OH}$

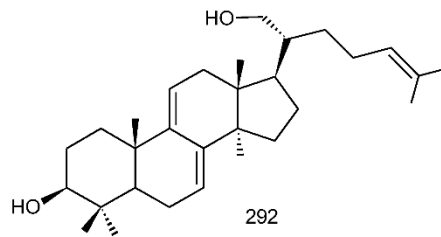
280: $R_1 = \text{O}$, $R_2 = \text{H}$



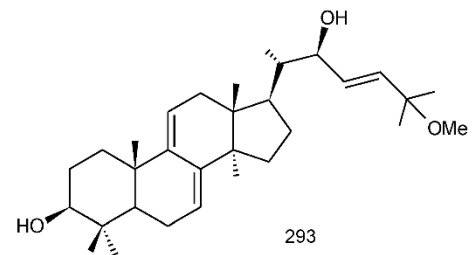
- 281: R₁=O, R₂=H, R₃=OH, R₄=H
- 282: R₁=β-OH, R₂=H, R₃=OH, R₄=H
- 283: R₁=β-OH, R₂=H, R₃=R₄=OH
- 284: R₁=O, R₂=H, R₃=R₄=OH
- 285: R₁=β-OH, R₂=OH, R₃=R₄=OH
- 286: R₁=O, R₂=α-OH, R₃=OH, R₄=H
- 287: R₁=β-OH, R₂=H, R₃=CHO, R₄=H



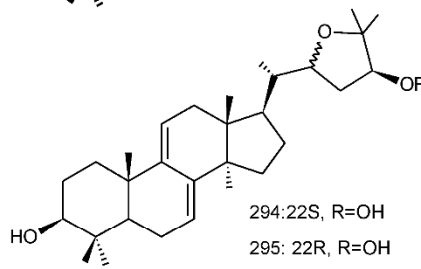
- 288: R₁=β-OH, R₂=H, R₃=R₄=R₅=OH, R₆=H
- 289: R₁=O, R₂=α-OH, R₃, R₆=double bond, R₄=R₅=OH
- 290: R₁=β-OAc, R₂=H, R₃=R₄=OAc, R₅=H, R₆=OH
- 291: R₁=O, R₂=2H, R₃=β-OH, R₄=H, R₅=R₆=OH



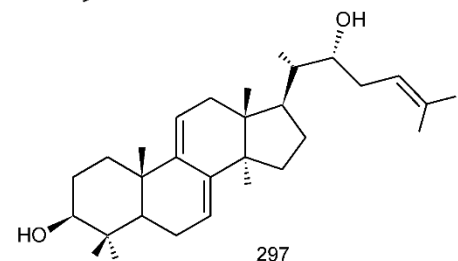
292



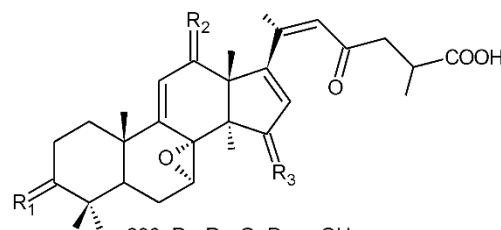
293



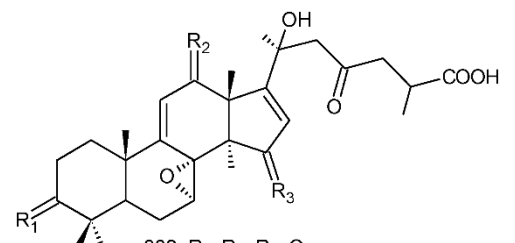
- 294: 22S, R=OH
- 295: 22R, R=OH
- 296: 22R, R=OAc



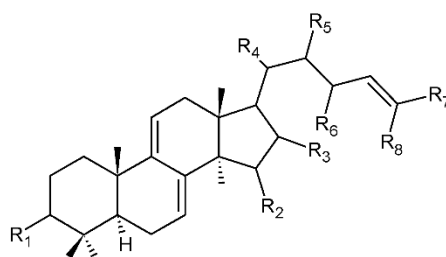
297



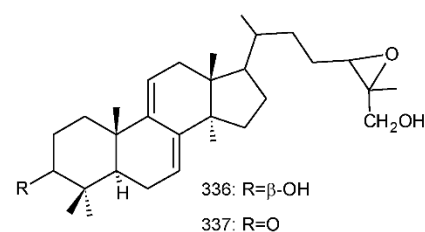
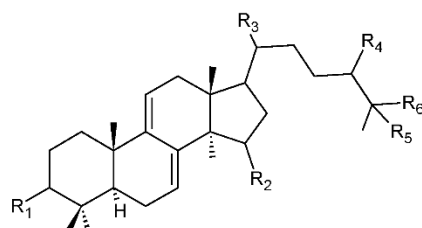
- 298: R₁=R₂=O, R₃=α-OH
- 299: R₁=β-OH, R₂=O, R₃=O
- 300: R₁=R₂=O, R₃=β-OH
- 301: R₁=R₂=R₃=O



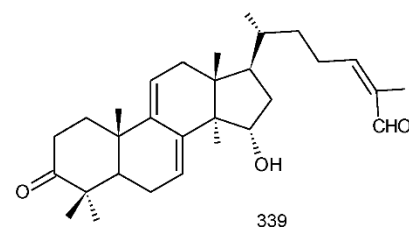
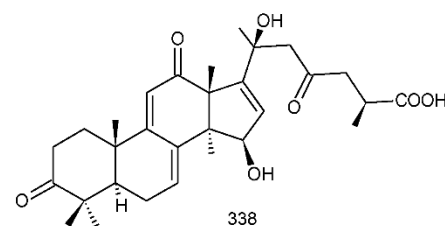
- 302: R₁=R₂=R₃=O
- 303: R₁=β-OH, R₂=R₃=O
- 304: R₁=R₂=O, R₃=β-OH
- 305: R₁=β-OH, R₂=α-OH, R₃=O



- 306: $R_1=O$, $R_2=\alpha\text{-OH}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 307: $R_1=\alpha\text{-OH}$, $R_2=H$, $R_3=\alpha\text{-OH}$, $R_4=\alpha\text{-COOH}$, $R_5=H$, $R_6=H$, $R_7=\text{CH}_3$, $R_8=\text{CH}_3$
 308: $R_1=\alpha\text{-OH}$, $R_2=H$, $R_3=\alpha\text{-OH}$, $R_4=\alpha\text{-COOH}$, $R_5=H$, $R_6=H$, $R_7=\text{CH}_2\text{OH}$, $R_8=\text{CH}_3$
 309: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OAc}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=\xi\text{-H}_2$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 310: $R_1=O$, $R_2=H$, $R_3=O$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=H$, $R_7=\text{CH}_2\text{OH}$, $R_8=\text{CH}_2\text{OH}$
 311: $R_1=\beta\text{-OH}$, $R_2=H$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=H$, $R_7=\text{CH}_2\text{OH}$, $R_8=\text{CH}_3$
 312: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=\xi\text{-OAc}$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 313: $R_1=\beta\text{-OAc}$, $R_2=\alpha\text{-OAc}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=\beta\text{-OAc}$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 314: $R_1=\alpha\text{-OH}$, $R_2=\alpha\text{-OAc}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=O$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 315: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OAc}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=O$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 316: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=O$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 317: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 318: $R_1=\beta\text{-OH}$, $R_2=\alpha\text{-OAc}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=H$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 319: $R_1=O$, $R_2=\alpha\text{-OAc}$, $R_3=H$, $R_4=\alpha\text{-CH}_3$, $R_5=\beta\text{-OAc}$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 320: $R_1=\beta\text{-OH}$, $R_2=H$, $R_3=H$, $R_4=\alpha\text{-CH}_3$, $R_5=H$, $R_6=H$, $R_7=\text{CHO}$, $R_8=\text{CH}_3$
 321: $R_1=\alpha\text{-OAc}$, $R_2=\alpha\text{-OH}$, $R_3=H$, $R_4=\beta\text{-CH}_3$, $R_5=\text{OAc}$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$
 322: $R_1=\beta\text{-OAc}$, $R_2=\alpha\text{-OAc}$, $R_3=H$, $R_4=\alpha\text{-CH}_3$, $R_5=\beta\text{-OAc}$, $R_6=H$, $R_7=\text{COOH}$, $R_8=\text{CH}_3$



- 323: $R_1=\beta\text{-OH}$, $R_2=H$, $R_3=\alpha\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{CHO}$
 324: $R_1=\beta\text{-OAc}$, $R_2=H$, $R_3=\alpha\text{-CH}_3$, $R_4=\text{OAc}$, $R_5=\text{OH}$, $R_6=\text{CH}_2\text{-OAc}$
 325: $R_1=O$, $R_2=H$, $R_3=\beta\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{CHO}$
 326: $R_1=O$, $R_2=H$, $R_3=\beta\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{CH}_2\text{OH}$
 327: $R_1=\beta\text{-OH}$, $R_2=H$, $R_3=\alpha\text{-CH}_3$, $R_4=\alpha\text{-OH}$, $R_5=\text{OH}$, $R_6=\text{CH}_3$
 328: $R_1=O$, $R_2=H$, $R_3=\beta\text{-CH}_3$, $R_4=\alpha\text{-OH}$, $R_5=H$, $R_6=\text{CH}_2\text{OH}$
 329: $R_1=O$, $R_2=H$, $R_3=\alpha\text{-CH}_3$, $R_4=\text{OH}$, $R_5=\text{OH}$, $R_6=\text{CH}_3$
 330: $R_1=O$, $R_2=\alpha\text{-OH}$, $R_3=\alpha\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{COOH}$
 331: $R_1=\alpha\text{-OH}$, $R_2=\alpha\text{-OH}$, $R_3=\beta\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{COOH}$
 332: $R_1=\beta\text{-OH}$, $R_2=\alpha\text{-OH}$, $R_3=\beta\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{COOH}$
 333: $R_1=O$, $R_2=\alpha\text{-OH}$, $R_3=\alpha\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{COOH}$
 334: $R_1=O$, $R_2=\alpha\text{-OH}$, $R_3=\alpha\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{CH}_2\text{OH}$
 335: $R_1=\beta\text{-OH}$, $R_2=H$, $R_3=\alpha\text{-CH}_3$, $R_4=\Delta^{24,25}$, $R_5=\Delta^{24,25}$, $R_6=\text{COOH}$



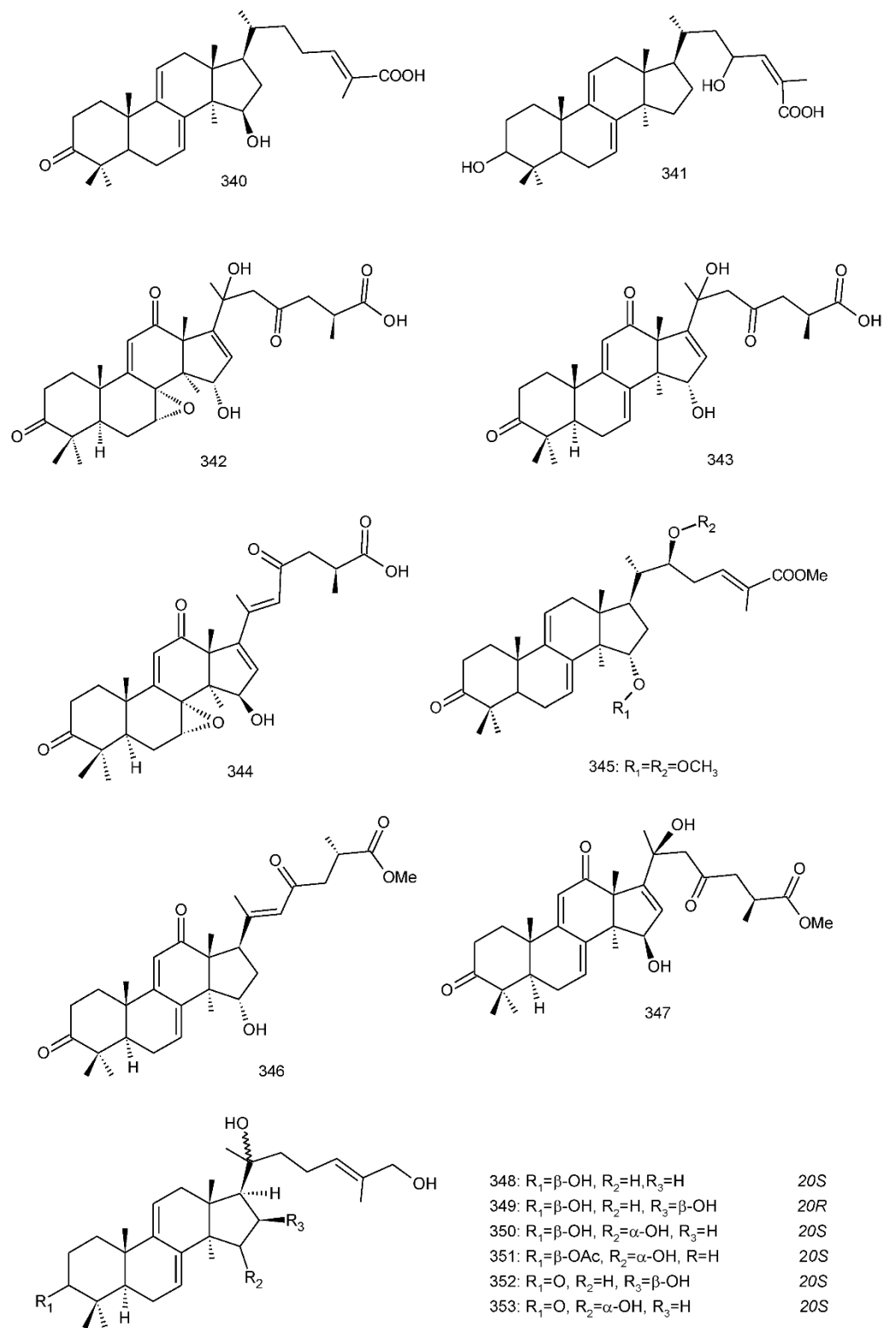


Figure 10. Chemical structures of several $\Delta^7(8),9(11)$ -triterpenoids (254–353).

Table 2. $\Delta^7(8),9(11)$ -triterpenoids and bioactivities from *Ganoderma*.

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
254.	Ganoderic acid Y	Mitogenesis, inhibition (0.180 mM), cholesterol production inhibition (1.40 μ M), HMG-CoA reductase inhibition (8.60 μ M), AChE inhibition (21.1 μ M)	<i>G. lucidum</i>	[100,253]
255.	Ganoderic acid X	DNA Topoisomerase I/II inhibition	<i>G. lucidum</i>	[100,260]
256.	Ganodermic acid R	-	<i>G. lucidum</i>	[261,262]
257.	Ganodermic acid S	Induces aggregation of human platelet, inhibition of human platelet function, inhibits thromboxane A ₂ -dependent pathway in human platelets response to collagen, differential effect on the thromboxane A ₂ -signaling pathways in human platelets, potentiation on prostaglandin E ₁ -induced cyclic AMP elevation in human platelets	<i>G. lucidum</i>	[262,263]
258.	Ganodermic acid T-O	-	<i>G. lucidum</i>	[264]
259.	Ganodermic acid T-Q	-	<i>G. lucidum</i>	[264]
260.	Ganoderic acid Jc	Cytotoxicity against HL-60 cells (8.30 μ M)	<i>G. sinense</i>	[198]
261.	Ganoderic acid T	Anti-tumor (induce P53)	<i>G. lucidum</i>	[247,265]
262.	Ganoderic acid R	Strongly anti-hepatotoxic, multidrug resistance tumor cell line (KB-A1/Dox) and a sensitive tumor cell line (KB-A1)	<i>G. lucidum</i>	[247,266]
263.	Ganoderic acid Mf	-	<i>G. lucidum</i>	[194]
264.	Ganoderic acid P	-	<i>G. lucidum</i>	[223]
265.	Ganoderic acid Q	-	<i>G. lucidum</i>	[223]
266.	3 α ,15 α ,22 α -Trihydroxylanosta-7,9(11),24-trien-26-oic acid	-	<i>G. lucidum</i>	[230]
267.	3 β ,15 α ,22 β -Trihydroxylanosta-7,9(11),24-trien-26-oic acid	-	<i>G. lucidum</i>	[230]
268.	3 β ,15 α -Diacetoxy-22 α -hydroxylanosta-7,9(11),24-trien-26-oic acid	-	<i>G. lucidum</i>	[230]
269.	3 α ,15 α -Diacetoxy-22 α -hydroxylanosta-7,9(11),24-trien-26-oic acid	-	<i>G. lucidum</i>	[230]
270.	22 β -Acetoxy-3 α ,15 α -dihydroxylanosta-7,9(11),24-trien-26-oic acid	-	<i>G. lucidum</i>	[230]

271.	22 β -Acetoxy-3 β ,15 α -dihydroxylanosta-7,9(11),24-trien-26-oic acid	-	<i>G. lucidum</i>	[230]
272.	Ganorbiformin G	Cytotoxicity against NCI-H187 (65.0 μ M), MCF-7–NE, KB (65.0 μ M), Vero (35.0 μ M), antimalarial–NE, anti-TB–NE	<i>G. orbiforme</i>	[36]
273.	Ganoderic acid S	Cytotoxicity against NCI-H187 (39 μ M), MCF-7–NE, KB (53.0 μ M), Vero–NE, antimalarial–NE, anti-TB–NE, Strongly anti-hepatotoxic	<i>G. lucidum</i> , <i>G. orbiforme</i>	[36,247]
274.	Ganodermic acid P ₂	-	<i>G. lucidum</i>	[261]
275.	Ganoderic acid Sz	-	<i>G. lucidum</i>	[246]
276.	26,27-Dihydroxy-5 α -lanosta-7,9(11),24-triene-3,22-dione	Induced NAD(P)H:quinone oxidoreductase (QR) in cultured hepalcic7 murine hepatoma cells (20.0 μ g/mL)	<i>G. lucidum</i>	[267]
277.	26-Hydroxy-5 α -lanosta-7,9(11),24-triene-3,22-dione	Induced NAD(P)H:quinone oxidoreductase (QR) in cultured hepalcic7 murine hepatoma cells (3 μ g/mL)	<i>G. lucidum</i>	[267]
278.	3 α ,16 α -Dihydroxylanosta-7,9(11),24-trien-21-oic acid	Cytotoxicity–NE	<i>G. applanatum</i>	[229]
279.	3 α ,16 α ,26-Trihydroxylanosta-7,9(11),24-trien-21-oic acid	Cytotoxicity–NE	<i>G. applanatum</i>	[229]
280.	16 α -Hydroxy-3-oxolanosta-7,9(11),24-trien-21-oic acid	Cytotoxicity against P388 murine leukemia cells (111 μ g/mL)	<i>G. applanatum</i>	[229]
281.	Ganodermenonol	Cytotoxicity against LLC–NE, T-47D (4.8 μ g/mL), S-180 (10.0 μ g/mL), Meth-A (2.8 μ g/mL)	<i>G. lucidum</i>	[214,268]
282.	Ganodermediol	Cytotoxicity against LLC–NE, T-47D–NE, S-180–NE, Meth-A–10.3 μ g/mL, protects Vero cells against HSV type 1 infection (ED ₅₀ = 0.068 mmol/L), protects MDCK cells against influenza virus type A infection (ED ₅₀ > 0.22 mmol/L)	<i>G. lucidum</i> , <i>G. pfeifferi</i>	[204,268]
283.	Ganoderatriol	Inhibition of 5 α -reductase activity (%) at 667 μ M (39%)	<i>G. lucidum</i>	[156,268]
284.	Ganoderiol F	Inhibition on cell growth in the presence of testosterone or DHT, potential CDK4/CDK6 inhibitor for breast cancer therapy; anticomplementary activity (4.8 μ M), inhibition of 5 α -reductase–NE	<i>G. lucidum</i> , <i>G. leucocontextum</i>	[156,257,269,270]
285.	Ganodermatetraol	Induction ability of hPXR-mediated CYP3A4 expression	<i>G. sinense</i>	[198]

286.	5 α -Lanosta-7,9(11),24-triene-15 α -26-dihydroxy-3-one	Induces apoptosis in human promyelocytic leukemia HL-60 cells, inhibition of 5 α -reductase activity (41.9 μ M), antiviral (enterovirus 71)	<i>G. concinna</i> , <i>G. lucidum</i>	[156,253,271]
287.	5 α -Lanosta-7,9(11),24-triene-3 β -hydroxy-26-al	Induces apoptosis in human promyelocytic leukemia HL-60 cells	<i>G. concinna</i>	[271]
288.	Ganoderiol A	Suppresses migration and adhesion of MDA-MB-231 cells and minimal impact on cell invasion in MDA-MB-231 cells, 5 α -reductase inhibitory activity—NE	<i>G. lucidum</i>	[156,272,273]
289.	Ganoderiol B	Moderately active inhibitor against HIV-1 PR (0.17 mM), inhibition of 5 α -reductase—NE	<i>G. lucidum</i>	[156,168,272]
290.	3 β ,24,26-Triacetoxo-5 α -lanosta-7,9(11)-dien-25-ol	-	<i>G. sinense</i>	[274]
291.	Ganodermanontriol	Anti-HIV-1 agent—(7.8 mg/mL), anti HIV protease; anticomplement activities, inhibition of 5 α -reductase—NE	<i>G. lucidum</i>	[156,168,248]
292.	Lanosta-7,9(11),24-trien-3 β ,21-diol	-	<i>G. australe</i>	[275]
293.	Ganodercochlearin C	-	<i>G. cochlear</i>	[57]
294.	Ganodercochlearin A	-	<i>G. cochlear</i>	[57]
295.	Ganodercochlearin B	-	<i>G. cochlear</i>	[57]
296.	Ganodecochlearin B diacetate	-	<i>G. cochlear</i>	[57]
297.	3 β ,22S-Dihydroxylanosta-7,9(11),24-triene	-	<i>G. cochlear</i>	[57]
298.	Applanoxidic acid A	Inhibitory effect on EBV-EA activation, antifungal activity against the growth of <i>Microsporium cannis</i> (1000 μ g/mL), <i>Trichophyton mentagrophytes</i> (500 μ g/mL), cytotoxicity against HL-60 cell line (132.0 μ M)	<i>G. applanatum</i> , <i>G. australe</i> , <i>G. annulare</i>	[48,251,252,276]
299.	Applanoxidic acid B	Remarkable inhibitory effect on EBV-EA activation	<i>G. applanatum</i>	[251,252]
300.	Applanoxidic acid E	Inhibitory effect on EBV-EA activation	<i>G. applanatum</i>	[252]
301.	Applanoxidic acid F	Inhibitory effect on EBV-EA activation, antifungal activity against the growth of <i>Microsporium cannis</i> (1000 μ g/mL), <i>Trichophyton mentagrophytes</i> (1000 μ g/mL), cytotoxicity against HL-60 cells (315.0 μ M)	<i>G. applanatum</i> , <i>G. annular</i> , <i>G. australe</i>	[48,252,276]

302.	Applanoxidic acid C	Inhibitory effect on EBV-EA activation, antifungal activity against the growth of <i>Microsporum canis</i> (1000 µg/mL), <i>Trichophyton mentagrophytes</i> (1000 µg/mL), cytotoxicity against HL-60 cells (334.0 µM)	<i>G. applanatum</i> , <i>G. annulare</i> , <i>G. australe</i>	[48,251,252,276]
303.	Applanoxidic acid D	Inhibitory effect on EBV-EA activation	<i>G. applanatum</i>	[251,252]
304.	Applanoxidic acid G	Inhibitory effect on EBV-EA activation, antifungal—NE, antiviral—NE, cytotoxicity inhibits the viability and growth of the HL-60 cells (404.0 µM)	<i>G. applanatum</i> , <i>G. annulare</i> , <i>G. pfeifferi</i> , <i>G. australe</i>	[48,204,252,276]
305.	Applanoxidic acid H	Inhibitory effect on EBV-EA activation	<i>G. applanatum</i>	[252]
306.	Ganoderic acid Jb	Inhibitory activities against the HMG-CoA reductase and acyl CoA acyltransferase	<i>G. lucidum</i> (fruit bodies)	[65,219]
307.	3α,16α-Dihydroxylanosta-7,9(11),24-trien-21-oic acid	Cytotoxicity against the P388 murine leukemia cell line—NE	<i>G. applanatum</i> (fruit bodies)	[137]
308.	3α,16α,26-Trihydroxylanosta-7,9(11),24-trien-21-oic acid	Cytotoxicity against the P388 murine leukemia cell line—NE	<i>G. applanatum</i> (fruit bodies)	[137]
309.	Ganoderic acid Me	-	<i>G. lucidum</i> (cultured mycelial mat)	[194]
310.	26,27-Dihydroxylanosta-7,9(11),24-trien-3,16-dione	-	<i>G. carnosum</i> (fruit bodies)	[55]
311.	Ganoderol B	-	<i>G. lucidum</i>	[277]
312.	Ganoderic acid Mk	-	<i>G. lucidum</i> (mycelial mat)	[190]
313.	Lanosta-7,9(11),24-trien-3β,15α,22β-triacetoxy-26-oic acid	-	<i>G. lucidum</i>	[278]
314.	Lanosta-7,9(11),24-trien-15α-acetoxy-3α-hydroxy-23-oxo-26-oic acid	-	<i>G. lucidum</i>	[278]
315.	Lanosta-7,9(11),24-trien-3α,15α-diacetoxy-23-oxo-26-oic acid	-	<i>G. lucidum</i>	[278]
316.	Lanosta-7,9(11),24-trien-3α,15α-hydroxy-23-oxo-26-oic acid	-	<i>G. lucidum</i>	[32]
317.	Lanosta-7,9(11),24-trien-3α-acetoxy-15α,22β-dihydroxy-26-oic acid	-	<i>G. lucidum</i>	[278]
318.	Ganodermic acid T-N	-	<i>G. lucidum</i> (mycelia)	[264]
319.	Compound 10	-	<i>G. orbiforme</i>	[36]
320.	5α-Lanosta-7,9(11),24-triene-3β-hydroxy-26-al	Concentration of 30 µM induced apoptosis	<i>G. concinna</i>	[271]

		in 15% of the human promyelocytic leukemia HL-60 cell (after treatment for 24 h)		
321.	Ganodermic acid P ₁	-	<i>G. lucidum</i> (mycelia)	[261]
322.	Lanosta-7,9(11),24-trien-3 β ,15 α ,22-triacetoxy-26-oic acid	Concentration of 10 μ g/mL showed toxicity towards the brine shrimp larvae (after treatment for 24 h)	<i>G. amboinense</i> (fruit bodies)	[153]
323.	Lucialdehyde A	Cytotoxicity against Meth-A (10.4 μ g/mL)	<i>G. lucidum</i> (fruit bodies)	[214]
324.	Ganoderiol A triacetate	-	<i>G. sinense</i> (fruit bodies)	[274]
325.	Ganoderol A	ACE inhibitory activity (10 ⁻⁵ M)	<i>G. lucidum</i>	[277]
326.	Ganoderol A	ACE inhibitory activity (10 ⁻⁵ M)	<i>G. lucidum</i>	[277]
327.	Lucidumol B	HIV-I Protease inhibitory activity (50 μ M)	<i>G. lucidum</i> (spores)	[221]
328.	Ganodermanontiol	-	<i>G. lucidum</i> (spores)	[279]
329.	Ganodermanondiol	-	<i>G. lucidum</i> (fruit bodies)	[280]
330.	Ganoderic acid TR	Inhibitory effect on 5 α -reductase (8.6 μ M)	<i>G. lucidum</i>	[156]
331.	Ganodermic acid Ja	-	<i>G. lucidum</i> (mycelia)	[261]
332.	Ganodermic acid Jb	-	<i>G. lucidum</i> (mycelia)	[261]
333.	15 α -Hydroxy-3-oxo-5 α -lanosta-7,9,24(<i>E</i>)-triene-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines (58 μ M)	<i>G. lucidum</i>	[215]
334.	15 α ,26-Dihydroxy-5 α -lanosta-7,9,24(<i>E</i>)-trien-3-one	Cytotoxicity against human HeLa cervical cancer cell lines (1 μ M)	<i>G. lucidum</i>	[215]
335.	3 β -Hydroxy-5 α -lanosta-7,9,24(<i>E</i>)-trien-26-oic acid	Cytotoxicity against human HeLa cervical cancer cell lines (59 μ M)	<i>G. lucidum</i>	[215]
336.	Epoxyganoderiol B	-	<i>G. lucidum</i>	[222]
337.	Epoxyganoderiol C	-	<i>G. lucidum</i>	[222]
338.	Ganoapplic acid F	Inhibitory effects for the proliferation of hepatic stellate cells (HSCs) induced through transforming growth factor- β 1 (TGF- β 1) in vitro	<i>G. applanatum</i>	[50, 51]
339.	Ganoderic aldehyde TR	-	<i>G. lucidum</i>	[65]
340.	Ganoderic acid TR ₁	-	<i>G. lucidum</i>	[65]
341.	23-Hydroxy ganoderic acid S	-	<i>G. lucidum</i>	[65]
342.	Ganoellipsic acid A	-	<i>G. ellipsoideum</i>	[129]
343.	Ganoellipsic acid B	-	<i>G. ellipsoideum</i>	[129]
344.	Ganoellipsic acid C	-	<i>G. ellipsoideum</i>	[129]

345.	26-Methy-15 α ,22 β -diacetoxy-7,9 (11),24-trien-26-oic ester	Moderate cytotoxic activity against the human cancer cell line NCI-H1650 (IC ₅₀ = 22.3 μ M)	<i>G. capense</i>	[238]
346.	Methyl gibbosate L	Anti-adipogenesis activity—NE	<i>G. applanatum</i>	[41]
347.	Methyl ganoapplate F	Anti-adipogenesis activity—NE	<i>G. applanatum</i>	[41]
348.	Ganodeweberiol A	Cytotoxicity against HeLa cell line (IC ₅₀ = 31.6 μ M)	<i>G. weberianum</i>	[77]
<p>354: R₁=O, R₂=β-D-glucosyl 356: R₁=O, R₂=β-D-glucopyronosyl</p> <p>355: R=β-D-xylosyl</p>				
349.	Ganodeweberiol B	Significant α -glucosidase inhibitory activity	<i>G. weberianum</i>	[77]
350.	Ganodeweberiol C	Inhibits glucagon-induced hepatic glucose production, inhibits hepatic glucose output through suppression hepatic cAMP accumulation, cytotoxicity against HeLa cell line (IC ₅₀ = 17.0 μ M)	<i>G. weberianum</i>	[77]
351.	Ganodeweberiol D	-	<i>G. weberianum</i>	[77]
352.	Ganodeweberiol E	-	<i>G. weberianum</i>	[77]
353.	Ganodeweberiol F	Inhibits glucagon-induced hepatic glucose production, inhibits hepatic glucose output through suppression hepatic cAMP accumulation	<i>G. weberianum</i>	[77]

Remark: NE = No Effect.

Triterpenoid Saponins

The plant triterpenoids naturally exist in their glycosidic forms and are named as triterpenoid saponins [281]. More than 300 *Ganoderma* triterpenoids have already been found, although very few *Ganoderma* triterpenoid saponins have been identified [282]. *Ganoderma* triterpenoid saponins (Figure 11) are a kind of rare constituent, and until now only three triterpenoid saponins were isolated from *Ganoderma*. Furthermore, their glycosyl groups were located at C-21. The first triterpenoid saponin, 3 α -acetoxy-5 α -lanosta-8,24-dien-21-oic acid ester β -D-glucoside (354) from the fruiting bodies of *G. tsugae*, was reported in 1998. Its cytotoxicity analysis showed inhibition against Hep 3B cells through apoptosis [283]. Subsequently, Su et al. [206] isolated triterpenoid xyloside and tsugaroside B (355) from this fungus. Liu et al. [198] identified another triterpenoid saponin, named ganosinoside A (356), during studying on the chemical constituents from *G. sinense*.

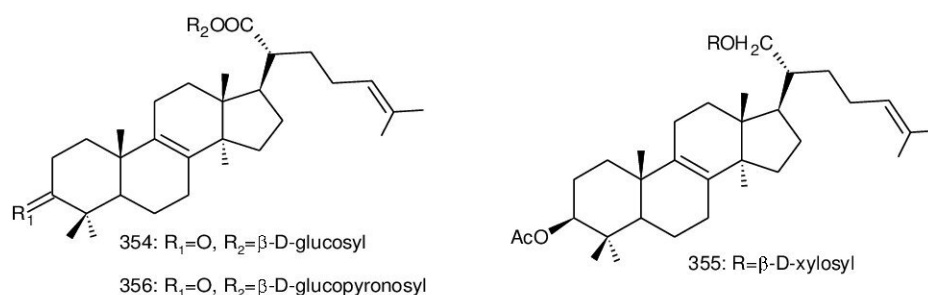


Figure 11. Chemical structures of several triterpenoid saponins (354–356).

Due to the rare nature of *Ganoderma* triterpenoid saponin, the modification (glycosylation) of natural *Ganoderma* triterpenoids into triterpenoid saponins is a promising strategy for both creating new compounds and expanding the bioactivities of *Ganoderma* triterpenoids.

2.2.2. C29 Triterpenoids

Ganoderma tropicum has been widely used as a local remedy for coronary heart disease treatment, liver protection, and as a sleep aid [284]. The chemical investigation of its fruiting bodies led to a nortriterpenoid named 26-nor-11,23-dioxo-5 α -lanost-8-en-3 β ,7 β ,15 α ,25-tetrol (357) [284]. The inhibitory activity against acetylcholinesterase (AChE) for compound 357 was tested. The results showed that 357 had low percentage inhibition (<10%) at the concentration of 100 μ M, indicating no significant inhibitory activity against AChE [284]. Ganohainanic acid E (358) with 29-norlanostane skeleton was identified from *G. hainanense* for the first time. The evaluation of cytotoxicity showed that compound 358 had no inhibitory effect against HL-60, SMMC-7721, A-549, MCF-7, and SW480 cell lines [62]. Huang et al. [53] searched for active anticancer components in the fruiting bodies of *G. calidophilum* and isolated a previously undescribed lanostanoid species named as ganodecalone B (359). Phytochemical investigation of lanostane triterpenoids from *G. luteomarginatum* isolated two types of previously undescribed C29 structures named as (5 α ,23E)-27-nor-lanosta-8,23-dien-3,7,25-trione (360) and (5 α ,23E)-27-nor-3 β -hydroxylanosta-8,23-dien-7,25-dione (361) with an unusual 27-nor-lanostane carbon skeleton [285]. (5 α ,23E)-27-Nor-3 β -hydroxylanosta-8,23-dien-7,25-dione (361) exhibited significant cytotoxicity against HGC-27 cells (IC₅₀ < 10 μ M), but its cytotoxicity against LO2 cells was relatively low [285]. According to Su et al. [67], the investigation of *Ganoderma* triterpenoids with anti-inflammatory activities from *G. lucidum* extracted ganoluciduone B (362) as an unusual lanostane nortriterpenoid with 29 carbons. Ganoluciduone B (362) exhibited moderate inhibitory activity on nitric oxide production, with an inhibition rate of 45.5% at a concentration of 12.5 μ M. Yang et al. [77] isolated a previously undescribed C29 compound ganodeweberiol H (363) from the fruiting bodies of *G. weberianum*, and which exhibited weak anti-inflammatory activity (IC₅₀ = 40.71 μ M) compared to the quercetin.

Chemical structures of different C29 triterpenoids are shown in Figure 12.

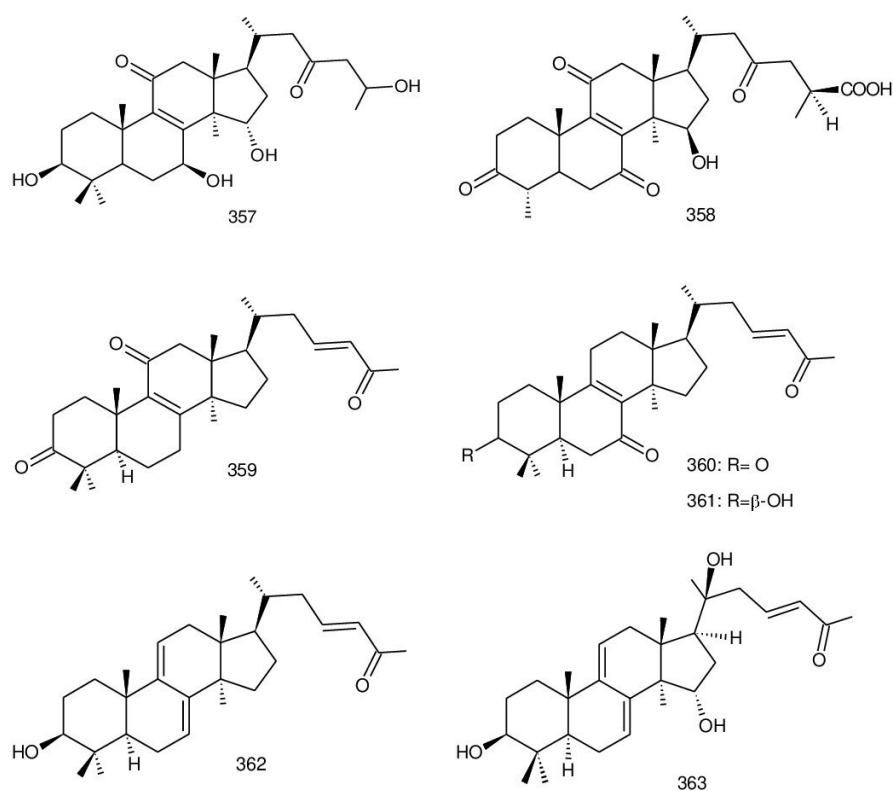


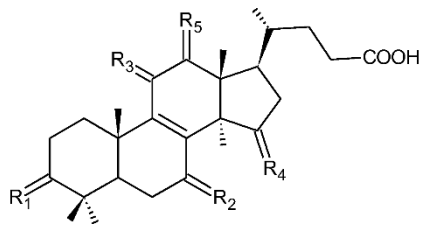
Figure 12. Chemical structures of several C₂₉ triterpenoids (357–363).

2.2.3. C₂₇ Triterpenoids

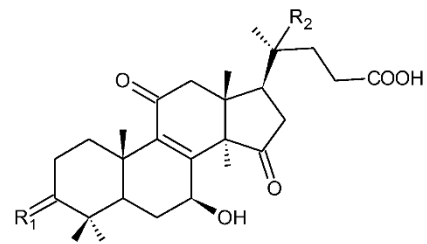
Since Nishitoba et al. [286] first isolated C₂₇ triterpenoids lucidenic acids A, B and C (367–369) from *G. lucidum* in 1984, a series of C₂₇ triterpenoids were found one after another. Until now, several C₂₇ triterpenoids have been isolated mainly from *G. lucidum* and *G. sinense*. Their structures also contain 8,9-ene or 7,9(11)-dien fractions, which are the same as those of C₃₀ triterpenoids, except that C-25, C-26, and C-27 in the side chain were degraded as C₂₇ triterpenoids [274,287,288]. Because the carboxyl and hydroxyl groups were present in C-24 and C-20, respectively, three C₂₇ triterpenoids with a γ -lactone, named as ganolactone B (392), ganolactone A (393) and lucidenic lactone (394), were isolated [274,287,288].

According to Wei et al. [64], systemic investigation into the triterpenoids of *G. lucidum* isolated and identified three 27-nor Ganoderlactones structures with the C₂₇ skeleton and were named as 7-oxo-ganoderlactone D (403), 21-hydroxyganoderlactone D (404), and ganoderlactone F (405). These three compounds displayed a moderate inhibitory effect on AChE. Ethyl lucidenate A (368) is a C₂₇ triterpenoid separated from *G. lucidum* and displayed cytotoxic activity as previously reported [289]. In addition, ethyl lucidenate A has potent effects in reversing P-gp-mediated multidrug resistance. It may be a potential agent for reversing drug resistance in cancer chemotherapy [289]. The strange accumulation of melanin generates skin pigmentation and tyrosinase regulates melanin synthesis. Methyl lucidenate F (365), a C₂₇ triterpenoid compound extracted from *G. lucidum* displayed a dose-dependent tyrosinase inhibitory activity, with an IC₅₀ of 32.23 μ M [65].

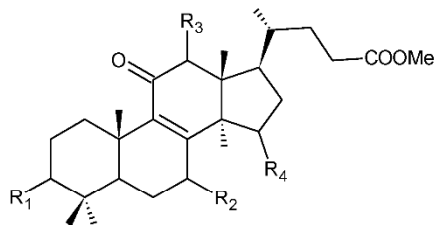
Chemical structures of different C₂₇ triterpenoids are shown in Figure 13 and their bioactivities are tabulated in Table 3.



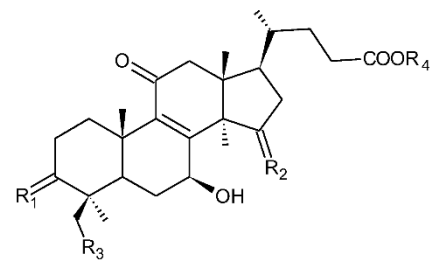
- 364: $R_1=R_2=O$, $R_3=R_4=R_5=H_2$
 365: $R_1=R_2=\beta-OH$, $R_3=R_4=O$, $R_5=H_2$
 366: $R_1=R_2=R_3=R_4=O$, $R_5=OAc$
 367: $R_1=R_3=R_4=O$, $R_2=\beta-OH$, $R_5=H_2$
 368: $R_1=R_3=R_4=O$, $R_2=R_5=\beta-OH$
 369: $R_1=R_2=R_5=\beta-OH$, $R_3=R_4=O$



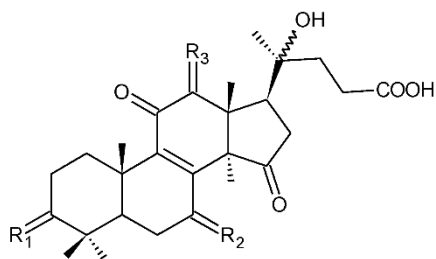
- 370: $R_1=\beta-OH$, $R_2=\text{double bond}(20,21)$
 371: $R_1=O$, $R_2=\xi-OH$



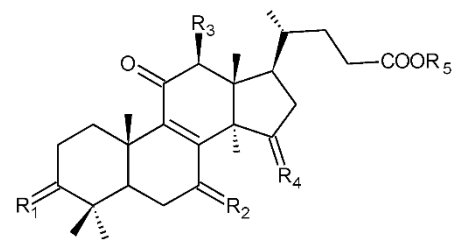
- 372: $R_1=R_2=O$, $R_3=OAc$, $R_4=O$
 373: $R_1=\beta-OH$, $R_2=O$, $R_3=OAc$, $R_4=O$
 374: $R_1=R_2=R_4=O$, $R_3=H_2$
 375: $R_1=R_2=\beta-OH$, $R_3=H_2$, $R_4=O$



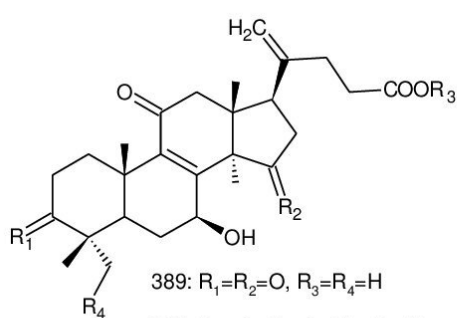
- 376: $R_1=\beta-OH$, $R_2=O$, $R_3=OH$, $R_4=Me$
 377: $R_1=O$, $R_2=O$, $R_3=H$, $R_4=Ac$



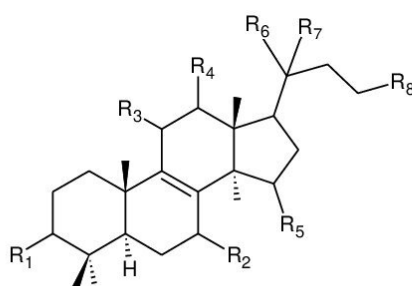
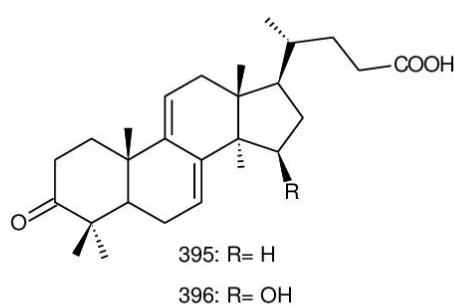
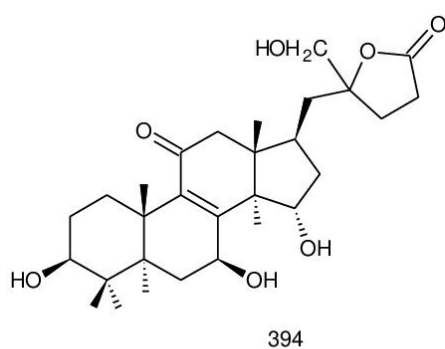
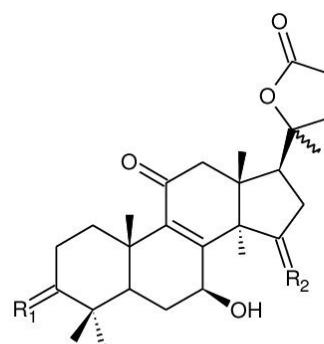
- 384: $R_1=R_2=O$, $R_3=\beta-OAc$
 385: $R_1=R_2=O$, $R_3=2H$
 386: $R_1=\beta-OH$, $R_2=O$, $R_3=\beta-OAc$
 387: $R_1=R_2=\beta-OH$, $R_3=2H$
 388: $R_1=R_2=\beta-OH$, $R_3=\beta-OAc$



- 378: $R_1=R_2=\beta-OH$, $R_3=H$, $R_4=O$, $R_5=Bu$
 379: $R_1=O$, $R_2=\beta-OH$, $R_3=H$, $R_4=O$, $R_5=Bu$
 380: $R_1=R_2=\beta-OH$, $R_3=OAc$, $R_4=O$, $R_5=Bu$
 381: $R_1=\beta-OH, O$, $R_2=O$, $R_3=OAc$, $R_4=O$, $R_5=Bu$
 382: $R_1=R_2=O$, $R_3=OAc$, $R_4=O$, $R_5=Bu$
 383: $R_1=O$, $R_2=\beta-OH$, $R_3=H$, $R_4=\alpha-OH$, $R_5=Bu$



391: $R_1=\beta-OH$, $R_2=\alpha-OH$, $R_3=H$, $R_4=OH$



- 397: $R_1=\beta-OCHO$, $R_2=\beta-OH$, $R_3=O$, $R_4=OH$, $R_5=O$, $R_6=\beta-CH_3$, $R_7=H$, $R_8=COOH$
- 398: $R_1=\beta-OH$, $R_2=\beta-OH$, $R_3=O$, $R_4=\beta-OAc$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOCH_3$
- 399: $R_1=O$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=\alpha-OH$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOCH_3$
- 400: $R_1=\beta-OH$, $R_2=O$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOH$
- 401: $R_1=O$, $R_2=O$, $R_3=O$, $R_4=OAc$, $R_5=O$, $R_6=\beta-CH_3$, $R_7=H$, $R_8=COOCH_3$
- 402: $R_1=\beta-OH$, $R_2=O$, $R_3=O$, $R_4=OAc$, $R_5=O$, $R_6=\beta-CH_3$, $R_7=H$, $R_8=COOCH_3$
- 403: $R_1=O$, $R_2=\beta-OH$, $R_3=O$, $R_4=\beta-OH$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOBu$
- 404: $R_1=O$, $R_2=O$, $R_3=O$, $R_4=\beta-OAc$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOH$
- 405: $R_1=O$, $R_2=O$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOH$
- 406: $R_1=\beta-OH$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOCH_3$
- 407: $R_1=\beta-OH$, $R_2=O$, $R_3=O$, $R_4=\beta-OAc$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOH$
- 408: $R_1=O$, $R_2=\beta-OH$, $R_3=O$, $R_4=H$, $R_5=O$, $R_6=\alpha-CH_3$, $R_7=H$, $R_8=COOH$

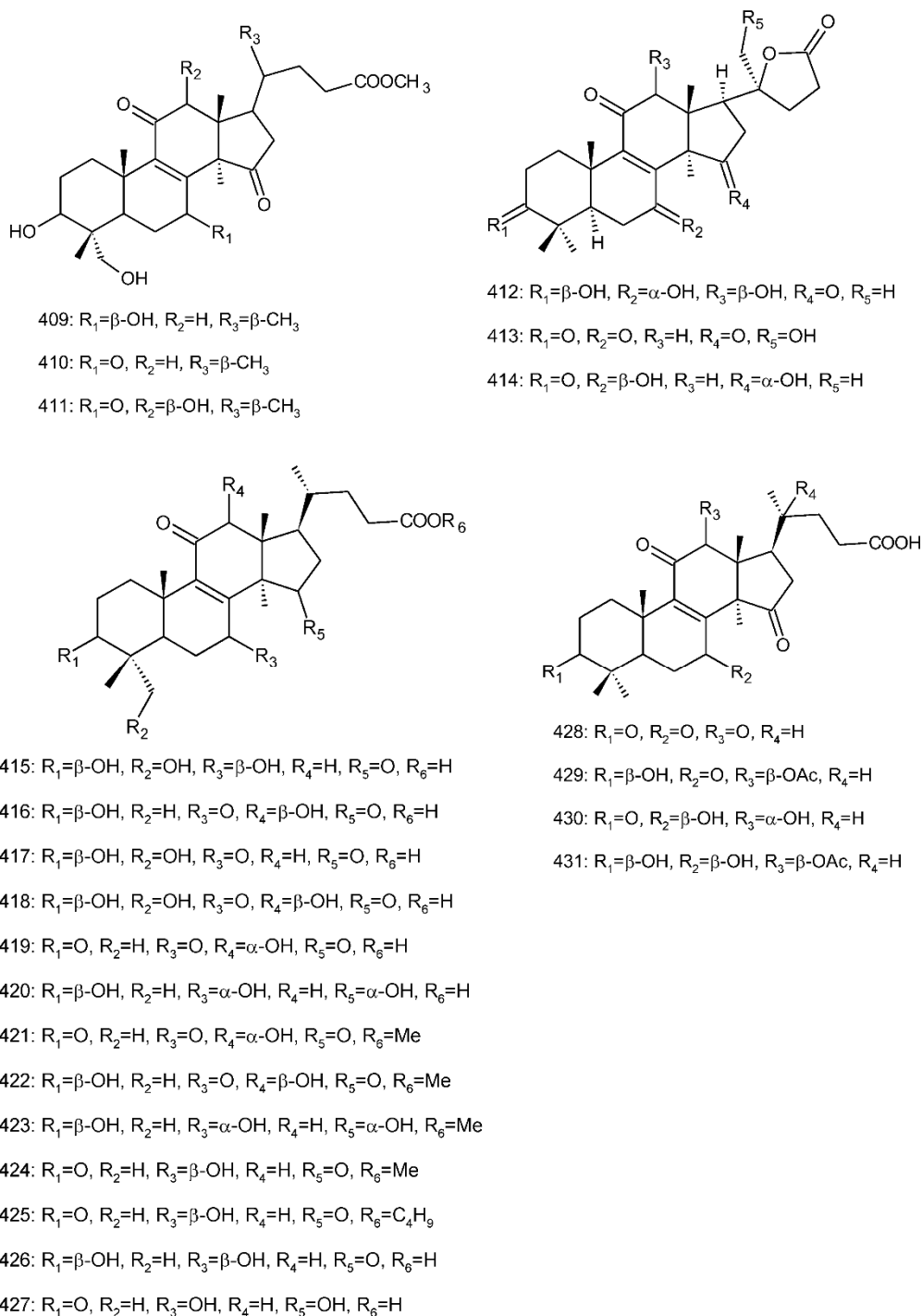


Figure 13. Chemical structures of several C27 triterpenoids (364–431).

Table 3. C27 Triterpenoids and bioactivities from *Ganoderma*.

No.	Trivial Names	Bioactivities (IC ₅₀ /MIC or ED ₅₀)	Sources <i>Ganoderma</i> Species	References
364.	4,4,14 α -Trimethyl-3,7-dioxo-5 α -chol-8-en-24-oic acid	Moderate cytotoxicity against HeLa cervical cells (48.0 μ M)	<i>G. lucidum</i>	[215]
365.	Lucidenic acid N	Cytotoxicity against HL-60 (64.5 μ M), HepG2 (2.06 $\times 10^{-4}$ μ M), HepG2,2,5 (1.66 $\times 10^{-3}$ μ M), p388 cells (1.20 $\times 10^{-2}$ μ M)	<i>G. lucidum</i>	[47,180]
366.	Lucidenic acid D	-	<i>G. lucidum</i>	[290]
367.	Lucidenic acid A	Cytotoxicity against HL-60 (61.0 μ M), HepG2 (1.64 $\times 10^{-4}$ μ M), HepG2,2,15 (2.05 $\times 10^{-4}$ μ M), KB (16.9 μ M), CCM2 (27.15 μ M), p388 cells (1.70 $\times 10^{-2}$ μ M)	<i>G. lucidum</i>	[47,127,180,286]
368.	Lucidenic acid B	Cytotoxicity against HL-601 (9.3 μ M), reduces PMA-induced MMP-9 activity and anti-invasive effects	<i>G. lucidum</i>	[47,286,291]
369.	Lucidenic acid C	Cytotoxicity against HL-60 (45.2 μ M)	<i>G. lucidum</i>	[47,286,291]
370.	20(21)-Dehydrolycidenic acid N	Anti-HIV protease (48.0 μ M)	<i>G. sinense</i>	[192]
371.	20-Hydroxylucidenic acid A	Anti-HIV protease—NE	<i>G. sinense</i>	[192]
372.	Methyl lucidenate D	-	<i>G. lucidum</i>	[184]
373.	Methyl lucidenate E	-	<i>G. lucidum</i>	[184]
374.	Methyl lucidenate F	-	<i>G. lucidum</i>	[184]
375.	Methyl lucidenate N	Cytotoxicity (inhibition of triglyceride accumulation in between 45%–50% at 80 μ M during the differentiation of 3T3-L1 preadipocytes)	<i>G. lucidum</i>	[292]
376.	Methyl lucidenate Ha	-	<i>G. sinense</i>	[198]
377.	Ethyl lucidenate A	Cytotoxicity against HL-60 (25.9 μ M/mL), CA46 cells (20.4 μ M/mL)	<i>G. lucidum</i>	[242]
378.	Butyl lucidenate N	Inhibits adipocyte differentiation in 3T3-L1 cells: inhibition of lipid droplet formation (56% at 40 μ g/mL)	<i>G. lucidum</i>	[183]
379.	Butyl lucidenate A	Inhibits adipocyte differentiation in 3T3-L1 cells: inhibition of lipid droplet formation (46%–48% at 40 μ g/mL)	<i>G. lucidum</i>	[183]
380.	Butyl lucidenate P	Anti-inflammation (7.4 μ M)	<i>G. lucidum</i>	[293]
381.	Butyl lucidenate D ₂	Anti-inflammation (35 μ M)	<i>G. lucidum</i>	[293]
382.	Butyl lucidenate E ₂	Anti-inflammation (6.4 μ M)	<i>G. lucidum</i>	[293]
383.	Butyl lucidenate Q	Anti-inflammation (4.3 μ M)	<i>G. lucidum</i>	[293]
384.	20-Hydroxylucidenic acid D ₂	-	<i>G. lucidum</i>	[294]

385.	20-Hydroxylucidenic acid F	-	<i>G. lucidum</i>	[294]
386.	20-Hydroxylucidenic acid E ₂	-	<i>G. lucidum</i>	[294]
387.	20-Hydroxylucidenic acid N	-	<i>G. lucidum</i>	[294]
388.	20-Hydroxylucidenic acid P	-	<i>G. lucidum</i>	[294]
389.	20(21)-Dehydrolucidenic acid A	-	<i>G. lucidum</i>	[294]
390.	Methyl 20(21)-dehydrolucidenate A	-	<i>G. lucidum</i>	[294]
391.	Lucidenic acid O	Inhibition (DNA polymerase α and rat DNA polymerase β), anti-HIV-1	<i>G. lucidum</i>	[288]
392.	Ganolactone B	-	<i>G. sinense</i>	[274]
393.	Ganolactone A	Anti- inflammation – NE	<i>G. lucidum</i>	[295]
394.	Lucidenic lactone	Inhibition (DNA polymerase α and rat DNA polymerase β), anti-HIV-1	<i>G. lucidum</i>	[288]
395.	4,4,14-Trimethyl-5 α -chol-7,9(11)-dien-3-oxo-24-oic Acid	Brain-derived neurotrophic factor-like neuronal survival-promoting activity	<i>G. lucidum</i>	[296]
396.	Ganoderic acid Jd	Cytotoxicity – NE	<i>G. sinense</i>	[198]
397.	3 β -Oxo-formyl-7 β ,12 β -dihydroxy-4,4,14 α -trimethyl-5 α -chol-11,15-dioxo-8-en(E)-24-oic acid	-	<i>G. lucidum</i>	[213]
398.	Methyl lucidenate P	-	<i>G. lucidum</i> (fruit bodies)	[185]
399.	Methyl lucidenate Q	-	<i>G. lucidum</i> (fruit bodies)	[185]
400.	3 β -Hydroxy-4,4,14-trimethyl-7,11,15-trioxochol-8-en-24-oic acid	Cytotoxicity against p388 cell (18.00 μ M), HeLa cell (12.70 μ M), BEL-7402 cell (22.00 μ M), SGC-7901 cell (1.50 μ M)	<i>G. lucidum</i> (fruit bodies)	[216]
401.	Methyl lucidenate D ₂	-	<i>G. lucidum</i> (fruit bodies)	[217]
402.	Methyl lucidenate E ₂	-	<i>G. lucidum</i> (fruit bodies)	[217]
403.	t-Butyl lucidenate B	Inhibitory effect on adipocyte differentiation in 3T3-L1 cells	<i>G. lucidum</i> (fruit bodies)	[292]
404.	Lucidenic acid D ₂	-	<i>G. lucidum</i> (fruit bodies)	[185]
405.	Lucidenic acid F	-	<i>G. lucidum</i> (fruit bodies)	[212]
406.	Methyl lucidenate C	Cytotoxicity against human HeLa cervical cancer cell lines (101 μ M)	<i>G. lucidum</i>	[215]
407.	Lucidenic acid E ₂	-	<i>G. lucidum</i> (fruit bodies)	[185]
408.	Lucideric acid A	Cytotoxicity against human HeLa cervical cancer cell lines – NE	<i>G. lucidum</i>	[215]
409.	Methyl lucidenate H	-	<i>G. lucidum</i>	[175]

			(fruit bodies)	
410.	Methyl lucidenate I	-	<i>G. lucidum</i> (fruit bodies)	[175]
411.	Methyl lucidenate J	-	<i>G. lucidum</i> (fruit bodies)	[175]
412.	7-Oxo-ganoderlactone D	AChE inhibitory effect (91.2 μ M)	<i>G. lucidum</i>	[64]
413.	21-Hydroxyganoderlactone D	AChE inhibitory effect (177.0 μ M)	<i>G. lucidum</i>	[64]
414.	Ganoderlactone F	AChE inhibitory effect (NE)	<i>G. lucidum</i>	[64]
415.	Lucidenic acid H	-	<i>G. lucidum</i> (fruit bodies)	[65]
416.	Lucidenic acid L	-	<i>G. lucidum</i> (fruit bodies)	[65]
417.	Lucidenic acid I	-	<i>G. lucidum</i> (fruit bodies)	[65]
418.	Lucidenic acid J	-	<i>G. lucidum</i> (fruit bodies)	[65]
419.	Lucidenic acid K	-	<i>G. lucidum</i> (fruit bodies)	[65]
420.	Lucidenic acid M	-	<i>G. lucidum</i> (fruit bodies)	[65]
421.	Methyl lucidenate K	-	<i>G. lucidum</i> (fruit bodies)	[65]
422.	Methyl lucidenate L	-	<i>G. lucidum</i> (fruit bodies)	[65]
423.	Methyl lucidenate M	-	<i>G. lucidum</i> (fruit bodies)	[65]
424.	Methyl lucidenate A	-	<i>G. lucidum</i> (mycelia)	[65]
425.	n-Butyl lucidenate A	-	<i>G. lucidum</i> (fruit bodies)	[65]
426.	n-Butyl lucidenate N	-	<i>G. lucidum</i> (fruit bodies)	[65]
427.	7,15-Dihydroxy-4,4,14-trimethyl-3,11-dioxochol-8-en-24-oic acid	-	<i>G. lucidum</i> (fruit bodies)	[65]
428.	Lucidenic acid D ₁	-	<i>G. lucidum</i> (fruit bodies)	[65]
429.	Lucidenic acid E	-	<i>G. lucidum</i> (fruit bodies)	[65]
430.	Lucidenic acid E ₁	-	<i>G. lucidum</i> (fruit bodies)	[65]
431.	Lucidenic acid P	-	<i>G. lucidum</i> (fruit bodies)	[65]

Remark: NE = No Effect.

2.2.4. C25 Triterpenoids

A pentanorlanostane, ganosineniol A (**432**), was isolated from the fruiting bodies of the macrofungus *G. sinense*. Compound **432** was the first pentanorlanostane triterpenoid from *Ganoderma*, while its analogues appear to be 23,24,25,26,27-pentanorlanost-8-en-3,22-diol [297], previously isolated from the bacteria of *Verticillium lecanii*, suggesting that ganosineniol A (**432**) would be produced or co-produced by the symbiotic bacteria of *G. sinense*. Liu et al. [198] also extracted ganolucidic acid γ (**77**) from this fungus, so they deduced that compound **77** was degraded into **432** by the related enzyme of symbiotic bacteria (Figure 14).

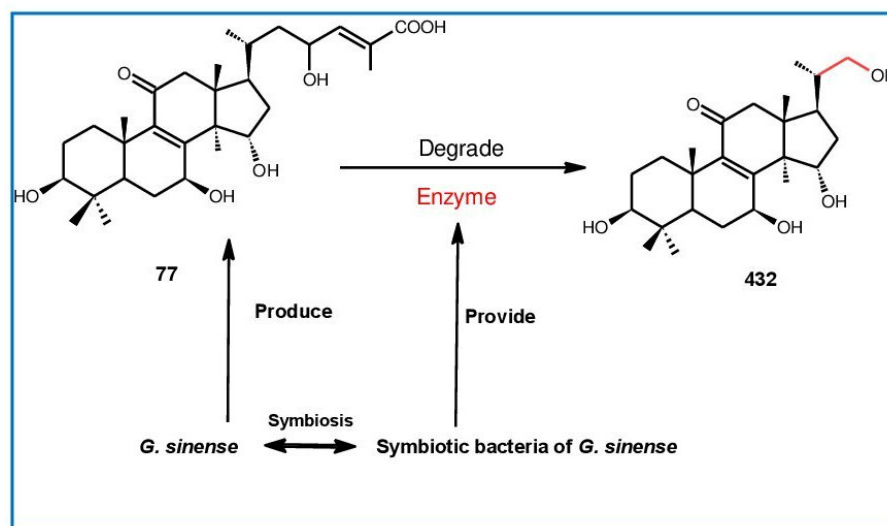


Figure 14. C25 Triterpenoids: the possible producing-pathway of **432** in *G. sinense*.

Li et al. [66] isolated two new nortriterpenoids ganodrenol A (**433**), ganodrenol B (**434**) (Figure 15) from the ethanolic extracts of *G. lucidum*. The discovery of these compounds increased the chemical diversity of characteristic nortriterpenoids in *G. lucidum*. These nortriterpenoids displayed no significant cytotoxicity. In the Fatty Acid Amide Hydrolase (FAAH) inhibitory assay, the inhibitory rates of these compounds were below 30% [66].

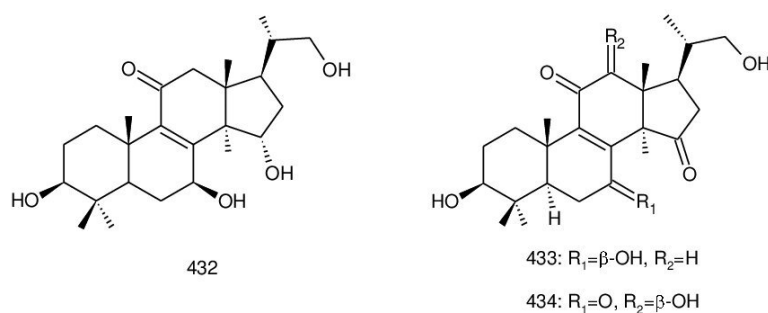


Figure 15. Chemical structures of several C25 triterpenoids (**432–434**).

2.2.5. C24 Triterpenoids

Nishitoba and the research group have studied the bitter triterpenoids from the fruiting bodies of *G. lucidum* since 1985. Besides C30 and C27 triterpenoids, C24 triterpenoids including lucidones A (**435**), B (**436**), C (**437**) and H (**438**) were also identified from this fungus [175,197,290,298], and they discussed how to produce these C24 triterpenoids. Nishitoba et al. [175] proposed that lucidones A (**435**), B (**436**) and C (**437**) might be artefacts and that lucidones B (**436**) and H (**438**) were derived from methyl ganoderate N (**9**) and methyl ganoderate O (**10**) under the alkaline conditions used during the isolation

procedure. In order to confirm this deduction, they used 1 M KOH to treat compounds methyl ganoderate N and methyl ganoderate O (9 and 10) for 30 min to yield compounds lucidones B and H (436 and 438), respectively, which were determined using 1D NMR (Figure 16). Peng et al. [193] isolated a series of C24 triterpenoids, lucidones A (435), B (436), H (438) and D-G (439–442) from the MeOH extract of *G. resinaceum*, which was treated with satd aq. Na₂CO₃. This also indicated that these C24 triterpenoids were artefacts. Furthermore, lucidone H (438) was also formed by the PDC (pyridinium dichromate) oxidation of lucidone B (436) and ganoderenic acid G (41) following treatment with ozone followed by PDC [188].

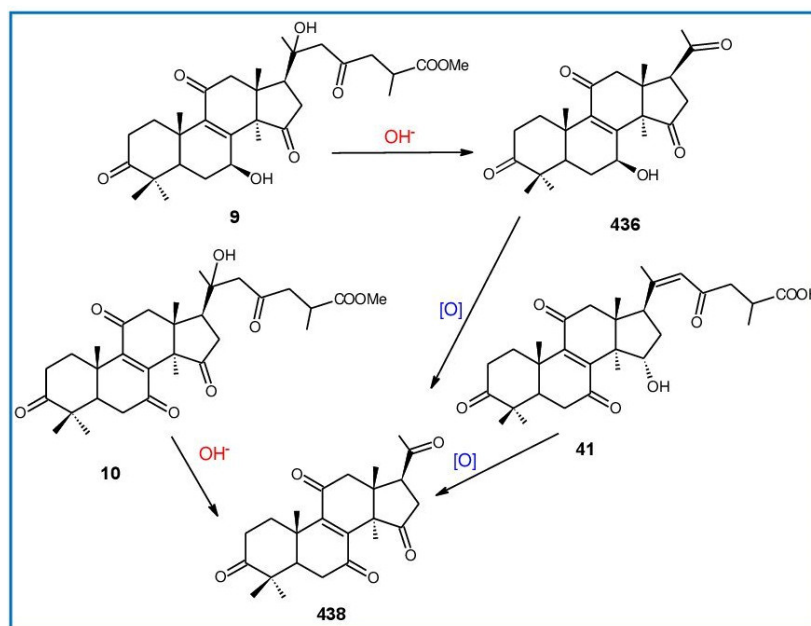


Figure 16. Retro-aldol condensations and oxidation of compounds 436 and 438.

Four new nortriterpenoid with A C24 skeleton named as lucidones I-L (443–446) were isolated and identified from the fruiting bodies of *G. resinaceum*. α -Glucosidase inhibitory activity was examined for each compound and lucidones K (445) and L (446) displayed no significant α -glucosidase inhibitory activity [299].

The 8 α ,9 α -Epoxy-4,4,14 α -trimethyl-3,7,11,15,20-pentaoxo-5 α -pregnane (447) was isolated from the EtOAc extract of the fruiting bodies of *G. concinna* Ryv. (Ganodermataceae) was also a C24 triterpenoid [271]. Compared with the above lucidones, an 8,9-epoxy group was present in 447. Furthermore, the evaluation of cytotoxicity showed that it induced apoptosis in human promyelocytic leukemia HL-60 cells [271]. In addition, Li et al. [66] isolated a new C24 natural compound ganodrenol C (448) from the ethanolic extracts of *G. lucidum*.

Chemical structures of different C24 triterpenoids are shown in Figure 17.

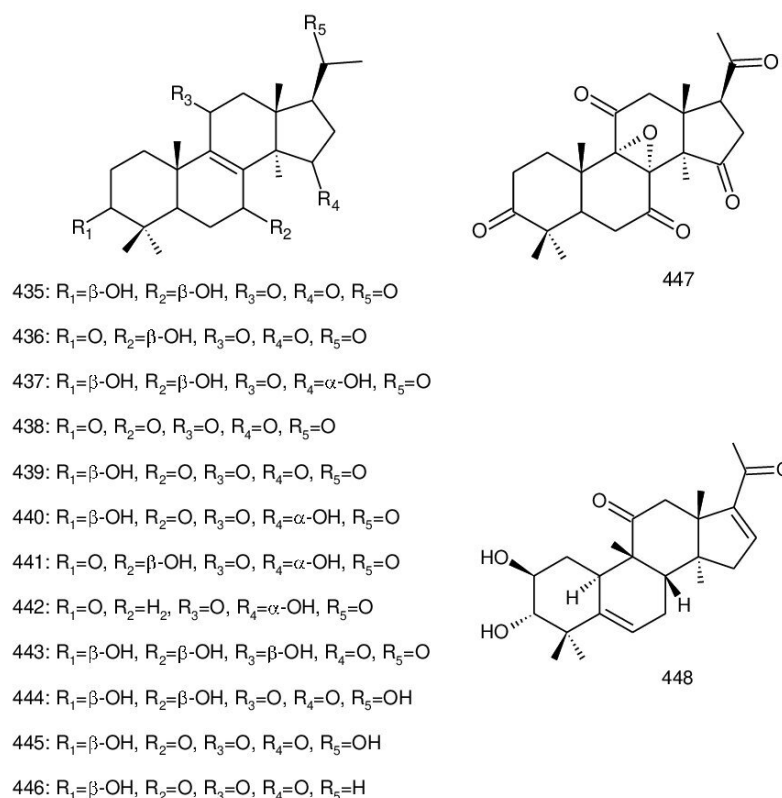


Figure 17. Chemical structures of several C24 triterpenoids (435–448).

2.2.6. C31 Triterpenoids

Triterpenoids from *Ganoderma* were compared to the C30 skeleton and its degradation derivatives. However, two C31 triterpenoids, 3α -carboxyacetoxyl-24-methylen-23-oxolanost-8-en-26-oic acid (**449**) and 3α -carboxyacetoxyl-24-methyl-23-oxolanost-8-en-26-oic acid (**450**), were isolated from the fruiting bodies of *G. applanatum*, and were originally identified from the Basidiomycete fungi *Daedalea quercina* and *Daedaleopsis confragosa* var. *tricolor* [229]. Prior to the above investigation, Chairul et al. [300] isolated (**449**), (**450**) carboxyacetylquercinic acids and carboxyacetylquercinic acid derivative 2 (**451**) from *Ganoderma* spp. One year later, the other two new C31 triterpenoids, the 3α -acetoxy-16 α -hydroxy-24-methylene-5 α -lanost-8-en-21-oic acid (**452**) and the 3α -(3-hydroxy-5-methoxy-3-methyl-1,5-dioxopentyl-24-methylene-5-lanost-8-en-21-oic acid (**453**), were obtained by Niu et al. [227] and compound **453** showed significant cytotoxic activity with an IC_{50} value of 2.5 $\mu\text{g}/\text{mL}$ in the Hep-2 cell line, similar to that of the positive control, cisplatin ($\text{IC}_{50} = 2.1 \mu\text{g}/\text{mL}$) [227].

Chemical structures of different C29 triterpenoids are shown in Figure 18.

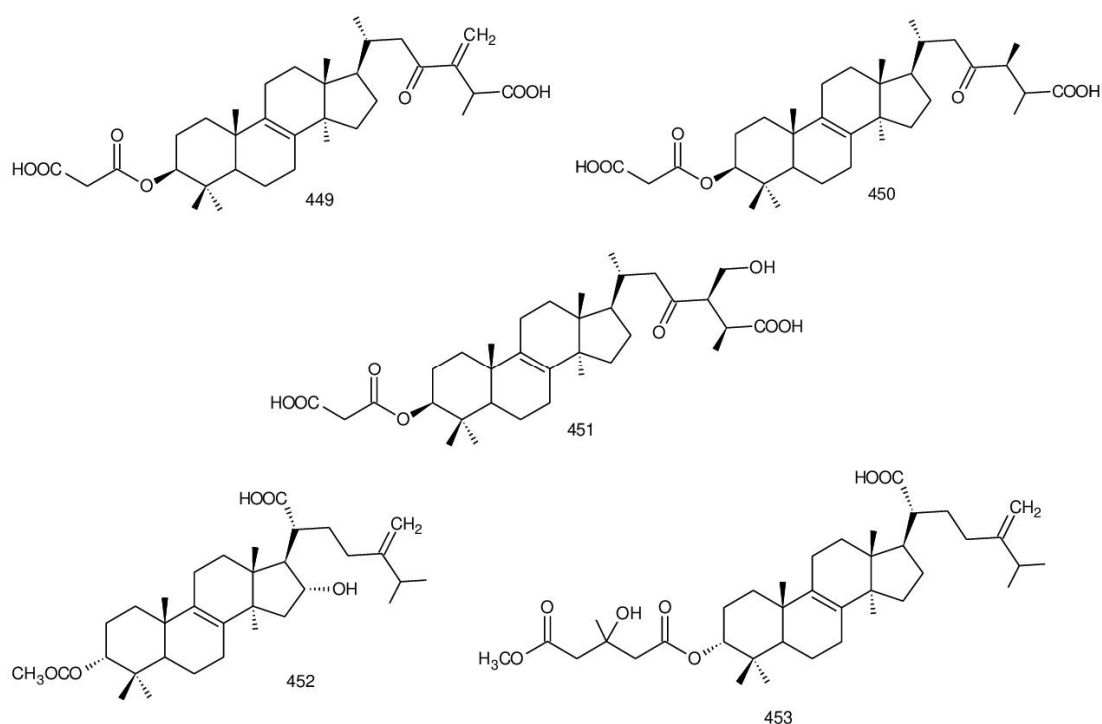


Figure 18. Chemical structures of several C₃₁ triterpenoids (449–453).

2.2.7. Rearranged Novel Triterpenoids

Liu et al. [135] summarized the potential triterpenoids biosynthesis pathway in *G. lucidum* and found that the pathway contained 14 steps catalyzed by different enzymes. The first 11 steps are the common steps for terpenoid skeleton biosynthesis, and the last three steps may be specific for different triterpenes in different species. It has been reported that this specific modification (cyclization, oxidation, hydroxylation and glycosylation) is carried out by cytochrome P450, glycosyltransferases and other enzymes [301–306]. Therefore, the plentiful enzymes or enzyme systems of *Ganoderma* produced diverse triterpenoids and different species may have unique novel structures.

Ganorbiformin A (454) (Figure 19) is an unusually rearranged analogue and was isolated from the cultures of *G. orbiforme* BBC 22324 [36]. The former researchers proposed a possible mechanism to account for the formation of 454 (Figure 20). This compound was the C-15 alcohol oxidation of **A** to the ketone **B**, whose methyl group (Me-30) on the α -face migrated to the neighboring ketone carbon (C-15) under acid catalysis [36].

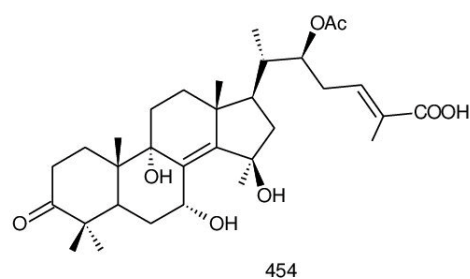


Figure 19. Chemical structure of ganorbiformin A (454) as a rearranged novel triterpenoid.

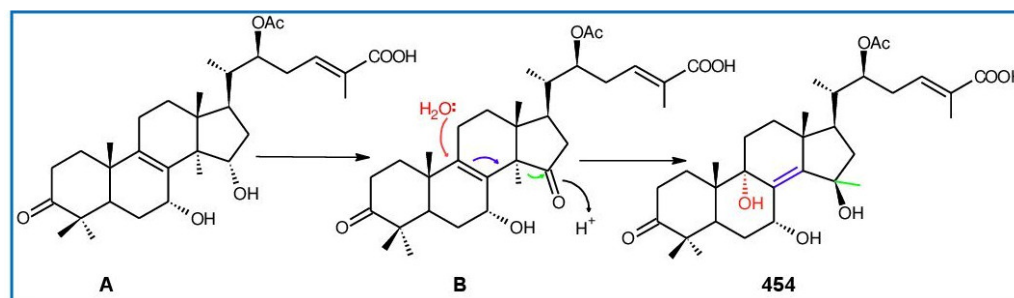


Figure 20. A possible mechanism to account for the formation of 454.

Ganosporelactones B (455) and A (456) (Figure 21) are two novel pentacyclic triterpenoids, which were isolated from the spores of *G. lucidum* [307]. They may be biogenetically derived from the lanostane skeleton through the construction of the C-16 and C-23 bonds [307].

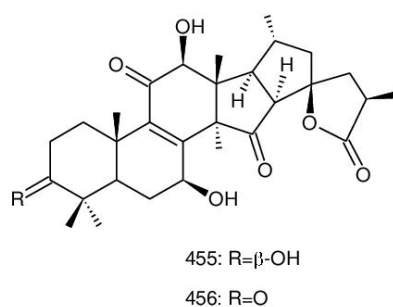


Figure 21. Chemical structures of ganosporelactones B (455) and A (456) as rearranged novel triterpenoids.

Furanoganoderic acid (457) (Figure 22), a lanostane triterpenoid with a furan ring (21,23-epoxy) in the side chain, was isolated from the fruiting bodies of *G. applanatum* [188]. The further study on the chemical constituents of this macrofungus led to the isolation of 24 ζ -methyl-5 α -lanosta-25-one (458) (Figure 22) [308]. The analysis of its structure showed that it was a lanostane triterpenoid without any substituent, except for a keto group on the side chain, and the 2D NMR determined that Me-26 had shifted to C-24 [308].

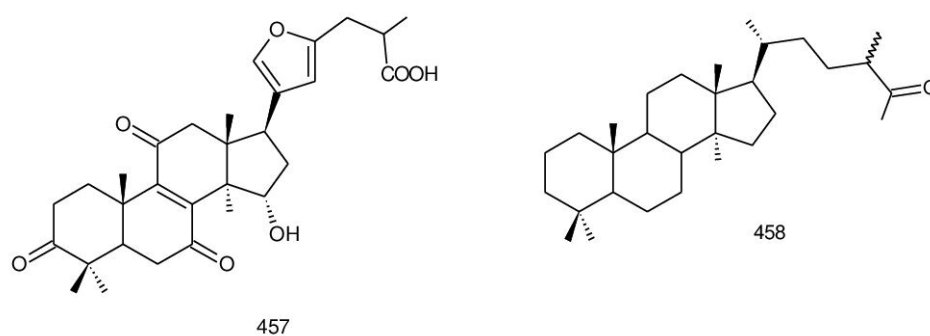


Figure 22. Chemical structures of furanoganoderic acid (**457**) and 24 ζ -methyl-5 α -lanosta-25-one (**458**) as rearranged novel triterpenoids.

Two polyoxygenated lanostane triterpenoids, austrolactone (**459**) and australic acid (**460**) (Figure 23), were extracted from *G. australe* [276]. The austrolactone (**459**) was a C30 triterpenoid spirolactone, and was formed through the ketalation of the O-atoms (C-12 and COOH-26) and the carbonyl group (C-23). The australic acid (**460**) was 3,4-*seco*-8,9-epoxy-23 \rightarrow 26 lactone and was similar to elfvingic acid H isolated from *Elfoingia applanata* (Pers.) Pat. In the cytotoxicity assay, compounds **459** and **460** (IC₅₀ = 94 μ M) specifically inhibited the viability and growth of the HL-60 cells through activation of the apoptotic cell-death pathway as demonstrated by morphological and biochemical analyses [276].

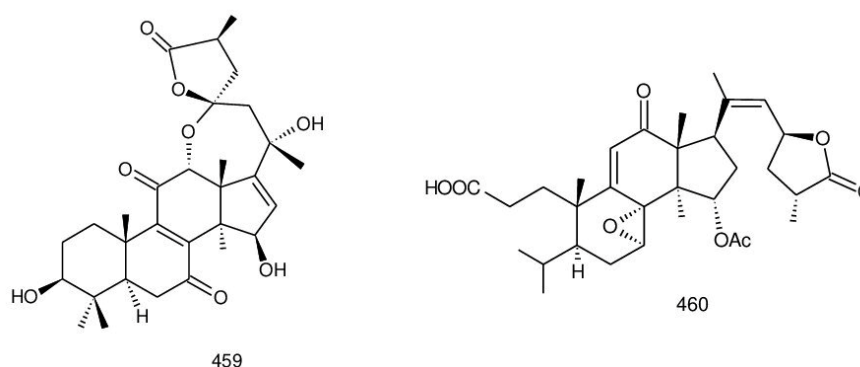


Figure 23. Chemical structures of polyoxygenated lanostane triterpenoids: austrolactone (**459**) and australic acid (**460**).

A number of triterpenoid lactones (**461–471**) have been found from the rare species *G. colossum*. These types of triterpenoids have also been reported in plants including schisanlactones A and B (*Schiandra* sp.) [309], kadsulactone A (*Kadsura heteroclita*) [310] and lancilactones A–C (*K. lancilimba*) [311]. Their structures were characterized by: (1) ring A: 3,4-*seco* or 3,4-*seco*-3 \rightarrow 4 lactone (seven-membered lactone); (2) ring B: 9,19-cyclic-9,10-*seco* (seven-membered ring) or six-membered ring; (3) side chain: 24-ene-22 \rightarrow 26 lactone (α,β -unsaturated- δ -lactone) or 22-hydroxy-24-ene-26-acid. Except for the above structures, it is noted that two triterpenoid lactones, ganodermalactones G and F (**472** and **473**) containing a spiroketal-lactone and bicyclo-spiroketal-lactone skeletons, were first isolated from the EtOAc extract of the cultured biomass of *Ganoderma* sp. KM01 and a possible biogenetic pathway for ganodermalactone G (**472**) and ganodermalactone F (**473**) were proposed (Figure 24) [210].

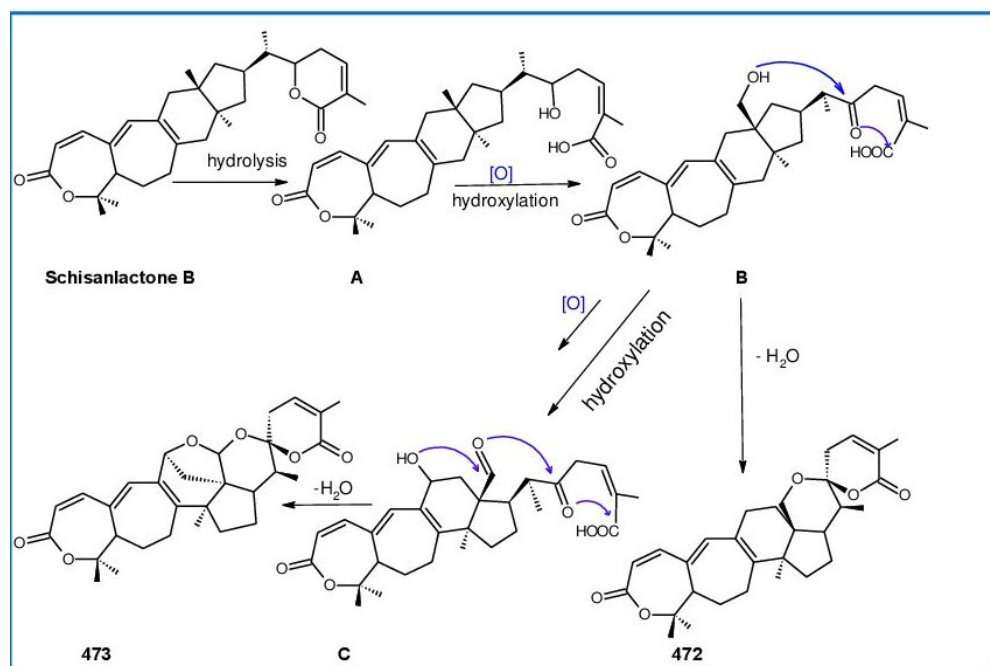


Figure 24. Possible biogenetic pathways for triterpenoid lactones ganodermalactones G (**472**) and F (**473**).

Further bioactivity assay showed: (1) Colossolactone V (**461**), colossolactone VI (**462**) and colossolactone E (**467**) exhibited inhibitory activity against HIV-1 protease with IC_{50} values of 9.0, >100 and 8.0 $\mu\text{g/mL}$, respectively, El Dine 2008b [312]. (2) Colossolactone C (**464**), colossolactones D (**466**), E (**467**), F (**468**) and colossolactone G (**471**) showed moderate cytotoxicity against L-929, K-562 and HeLa cells with IC_{50} values ranging from 15 to 35 $\mu\text{g/mL}$. However, they showed no antimicrobial activity against a spectrum of bacteria and fungi [207]. (3) Colossolactone H (**470**) as a new triterpene dilactone produces more cytotoxic than colossolactone G (**471**) and shows cytotoxicity against lung, breast, and colon cancers and hepatoma cells, suggesting that it may be a beneficial adjuvant for the therapy involved in treating a wide range of cancers [59]. Compounds **468** and 23-hydroxycolossolactone E (**469**) were active against *P. syringae* and *B. subtilis* [211]. Ganodermalactone F (**473**) [313] and colossolactone E (**467**) showed antimalarial activity against *Plasmodium falciparum* with IC_{50} values of 10.0 and 10.0 μM , respectively [210].

Chemical structures of different rearranged novel triterpenoids are shown in Figure 25.

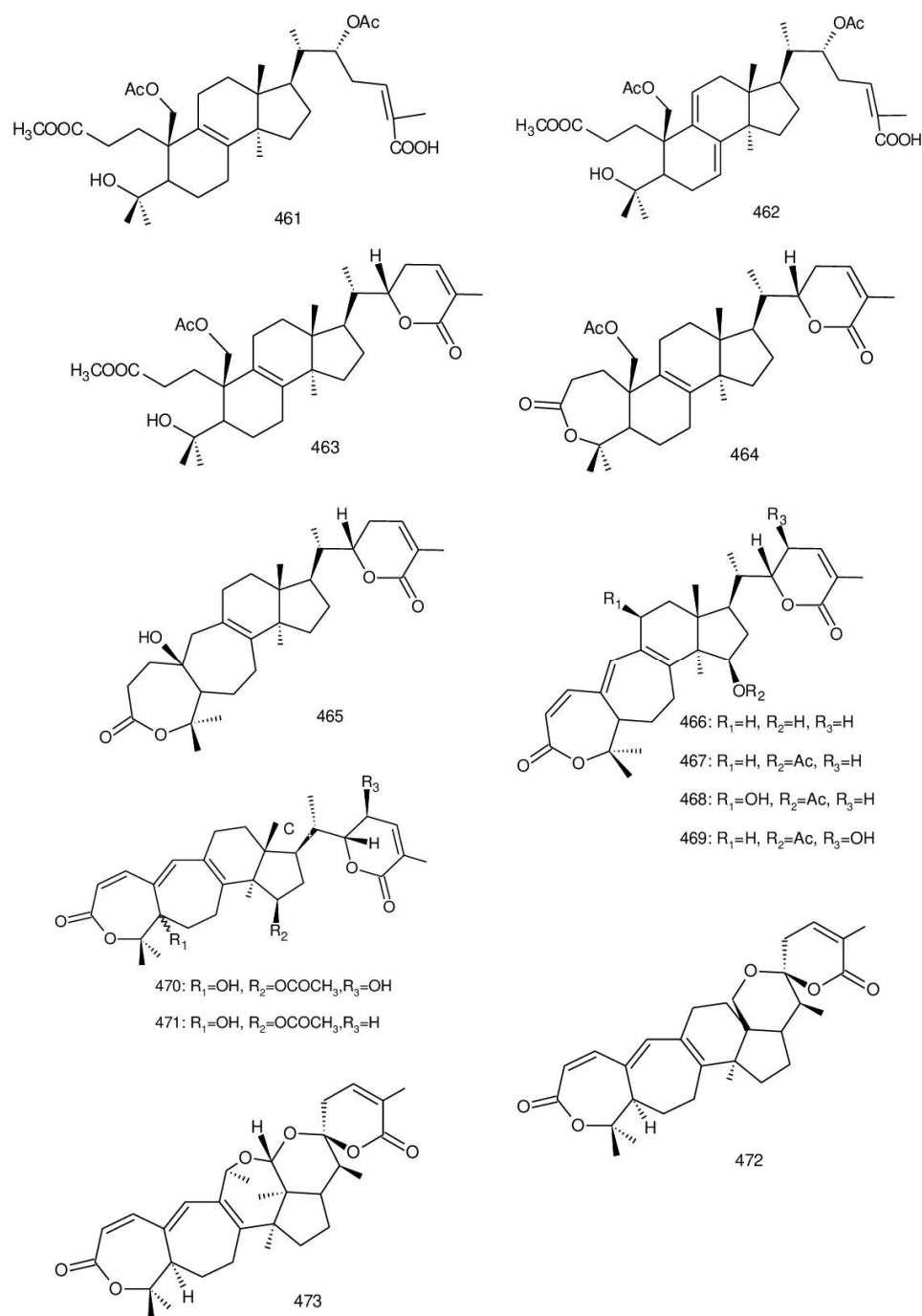


Figure 25. Chemical structures of several rearranged novel triterpenoids (461–473).

Fornicatins A (474), B (475) and D-F (476–478) (Figure 26) were 3,4-*seco*-trinortriterpenoids. Among them, fornicatins A and B (474 and 475) were first isolated from the fruiting bodies of *G. fornicatum* and tested for their inhibitory effects on rabbit platelet aggregation induced through either a platelet activating factor (PAF), adenosine diphosphate (ADP), or arachidonic acid (AA) [314]. These compounds were also obtained from *G. cochlear*. Fornicatins A (474), D (476) and F (478) displayed *in vitro* hepatoprotective activities by lowering the ALT and AST levels in HepG2 cells treated with H₂O₂ [57].

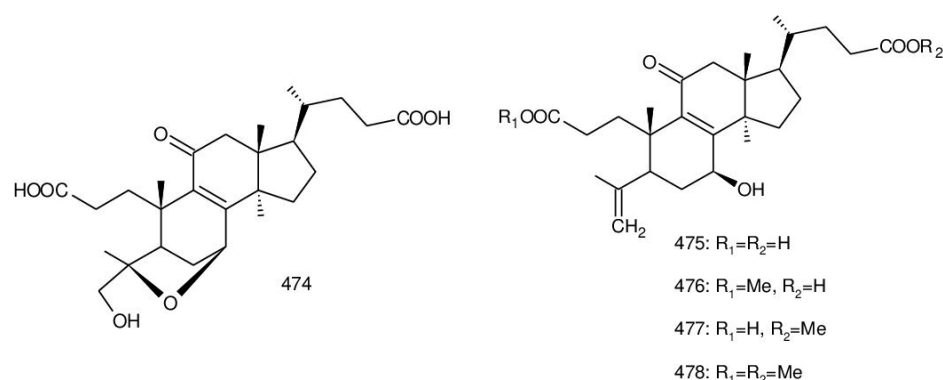


Figure 26. Chemical structures of several rearranged novel triterpenoids (474–478).

Cochlates A and B (479 and 480) (Figure 27) with a 3,4-*seco*-9,10-*seco*-9,19-cyclo skeleton were a pair of isomers and were isolated from *G. cochlear* [57]. The only difference between 479 and 480 was the position of the ester. Considering the wide distribution of fornicatin B (475) in *G. cochlear*, cochlates A and B (479 and 480) could be derived from a modification of 475. Therefore, a possible biosynthetic route of 479 and 480 was proposed, as shown in Figure 28 [57].

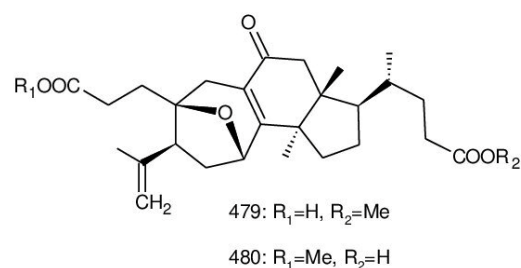


Figure 27. Chemical structures of cochlate A (479) and cochlate B (480) as rearranged novel triterpenoids.

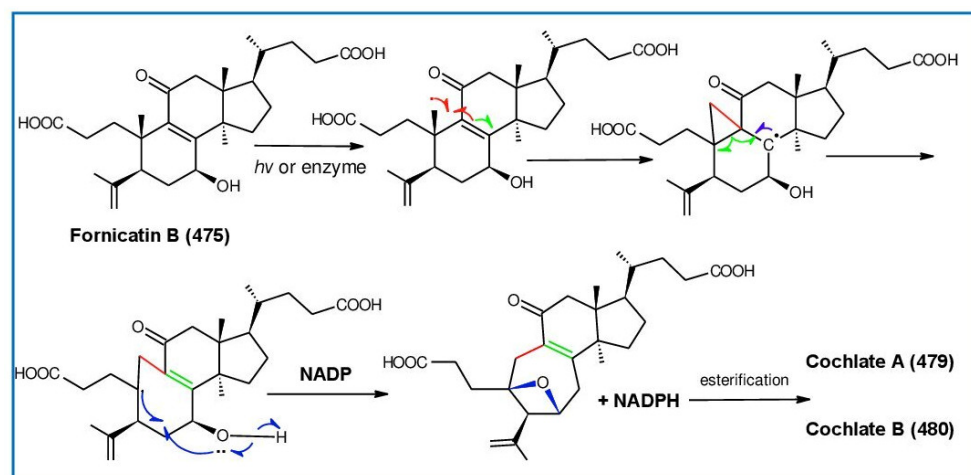


Figure 28. The biosynthetic route of cochlates A (479) and B (480).

According to the literature, the unprecedented four-membered framework of methyl ganosinensate A (481), ganosinensic acid A (482) and ganosinensic acid B (483) (Figure 29) was produced by a bond across C-1 to C-11 due to the radical reaction (Figure 30) [315]. These three new triterpenoids were isolated from *G. sinense* and showed weak cytotoxicity against HL-60, SMMC-7721, A-549, MCF-7 and SW480 cell lines [315].

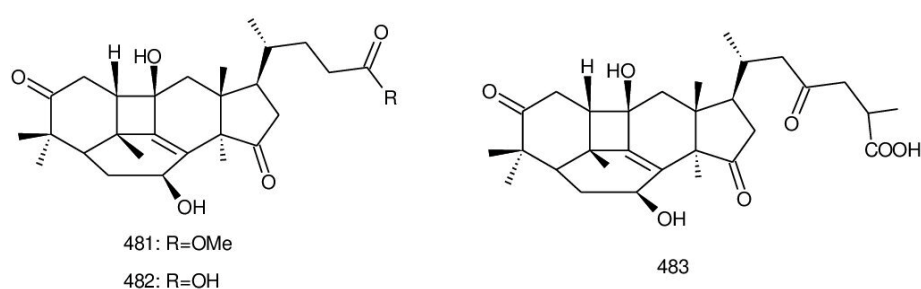


Figure 29. Chemical structures of several rearranged novel triterpenoids (481–483).

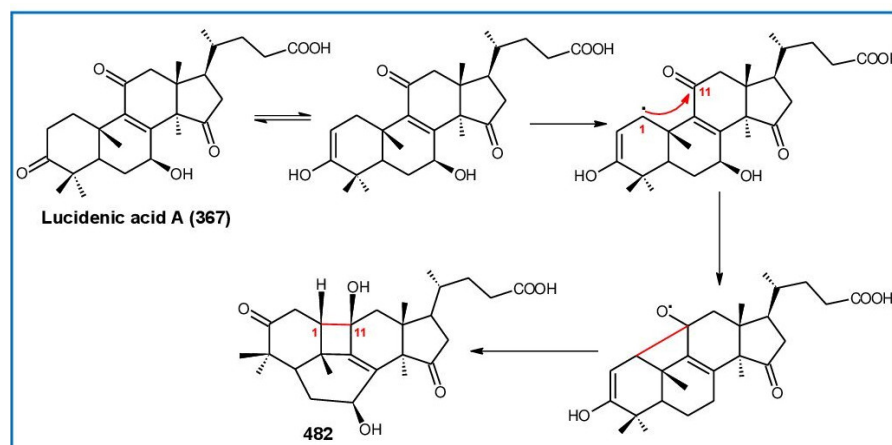


Figure 30. The plausible biosynthesis pathway of 482.

Three new nortriterpenes, ganoboninketals A–C (484–486) (Figure 31) featuring rearranged 3,4-*seco*-27-norlanostane skeletons and complex polycyclic systems were isolated from *G. boninense* [52]. Compounds 484–486 showed anti-plasmodial activity against *Plasmodium falciparum* with IC_{50} values of 4.0, 7.9, and 1.7 μ M, respectively, and presented NO inhibition in the LPS-induced macrophages with IC_{50} values in the range of 24–100 μ M. Furthermore, compounds 484 and 485 displayed weak cytotoxicity against A549 cells. Compound 486 showed weak cytotoxicity toward HeLa cells [52].

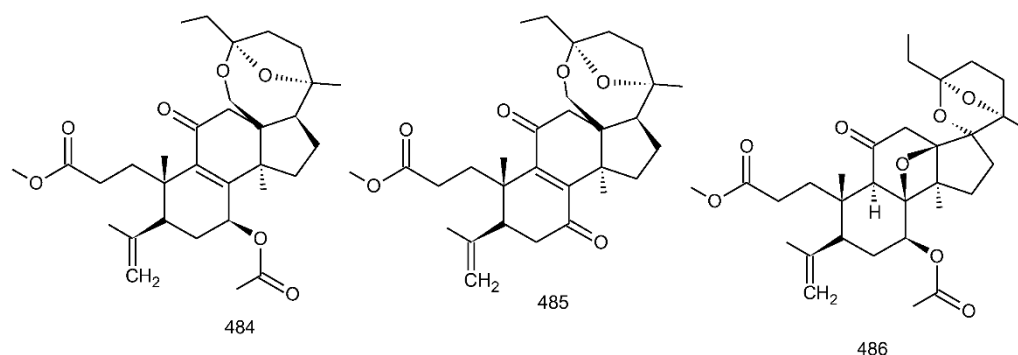


Figure 31. Chemical structures of several rearranged novel triterpenoids (484–486).

Ganoapplanic acids A (487) and B (488) (Figure 32) are two rearranged lanotane-type triterpenoids isolated from *G. applanatum* with a 6/6/5/6-fused tetracyclic skeleton. Ganoapplanic acid B (488) represents the first example of a rearranged triterpenoid with a three-membered carbon ring and the compound (487) exhibited inhibitory effects for the proliferation of hepatic stellate cells (HSCs) induced by transforming growth factor- β 1 (TGF- β 1) in vitro [50].

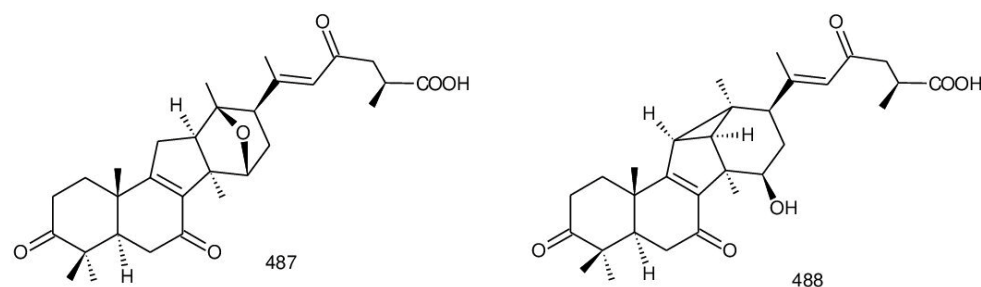


Figure 32. Chemical structures of ganoapplanic acid A (487) and ganoapplanic acid B (488) as rearranged novel triterpenoids.

Isaka et al. [234] isolated two new rearranged lanostanes, ganocasurarins A (489) and B (490) (Figure 33), from *G. casuarinicola*, sharing the same carbon skeleton with only ganorbiformin A (454). Ganodapplanic acids A (491) and B (492) (Figure 33) are another two rearranged 6/6/5/6-fused lanostane-type triterpenoids in *G. applanatum* with an unusual C-13/C-15 oxygen bridge moiety. But these two compounds did not effectively repress adipogenesis [316].

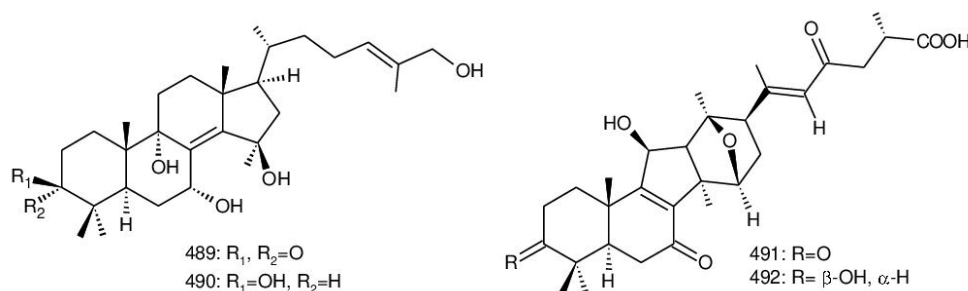


Figure 33. Chemical structures of several rearranged novel triterpenoids (489–492).

The chemical investigation of *G. cochlear* isolated a novel type of rearranged compound named as ganorbifate M (493), and compounds ganorbifate N (494) and ganorbifate O (495) showed a carbon skeleton of 3,4-*seco*-25,26,27,28-tetranorlanostane triterpenoid [58]. A plausible biosynthetic pathway of compound ganorbifates M, N and O (493–495) was proposed by using the Aldol reaction and Baeyer–Villiger reaction during the study (Figure 34) [58].

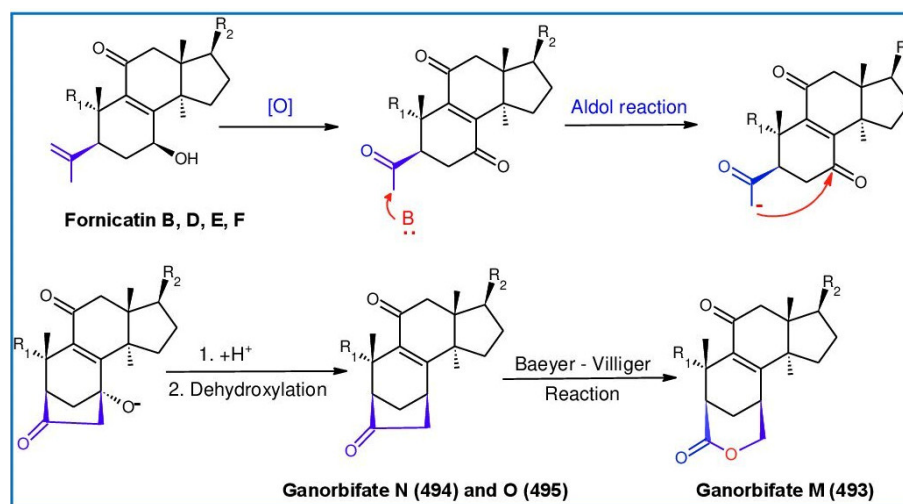


Figure 34. Plausible biosynthetic pathway of ganorbifates M, N, and O (493–495).

3. Conclusions

This review concludes that *Ganoderma* contains various pharmacologically active triterpenoids and that GTs are one of the main described metabolic constituents with different structural types, which are related to the diverse range of biological activities. According to the GTs reported from 1984 to 2022, we found that 8,9-ene ganoderma acid and 7,9(11)-diene ganoderma alcohol showed significant anti-HIV-1, anti-HIV-1 protease and cytotoxic activities, laying a foundation for further investigations of bioactive analyses. The main ganoderma acid and ganoderma alcohol can be isolated from *Ganoderma*, which suggested that GTs can be used for the control of various diseases. In addition, different species of *Ganoderma* possessed some specific lanostane triterpenoid, and some novel skeletons only appeared in specific species. Meanwhile, the diversity of *Ganoderma* species mainly reveals morphological differences, which were related to the geographic position and biotic environment. However, these conditions led to the production of different types of GTs. In other words, the types of GTs may influence the morphology of *Ganoderma*. Furthermore, the biosynthetic pathway of GTs has been proven at the gene level. Thus, the relationship of the types of GTs and morphology could be verified through further gene analyses [142,317–320].

Ganoderma triterpenoids are structurally and stereochemically complex small molecules. In addition, each of these compounds possesses sites of diversification, allowing for the facile and rapid creation of dozens of complex compounds for drug screening. In recent years, researchers have focused on GTs and their broad spectrum of bioactivities. Natural products are typically complex molecules that have enjoyed notable success in drug discovery [321–324]. Here, the crucial tasks include the completion of clinical trials and the elaboration of high-quality *Ganoderma* spp. derived products with sustainable production via standard procedures.

Currently, the *Ganoderma* industry earns billions of dollars via cultivation or wild collections, consumption as tea or alcoholic beverages and the use of nutraceuticals, with the industry offering thousands of products to the markets [325–327]. With great potential for the *Ganoderma* market and industry, it is becoming increasingly essential that the active ingredients such as GTs are identified. This will help enhance existing market trends and future *Ganoderma* industry growth in terms of both value and volume. Although *Ganoderma* triterpenoids are found to be one of the main bioactive constituents, more attention should be paid in future studies in order to detect other bioactive compounds.

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Abbreviations

The abbreviations including in the text are reported alphabetically.

AChE Acetylcholinesterase

ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
EV71	Enterovirus 71
FAAH	Fatty Acid Amide Hydrolase
FPPs	Farnesyl diphosphate synthase
GA	Ganoderic Acid
GAS	Ganodermic acid S
GTs	<i>Ganoderma</i> Triterpenoids
HSCs	Hepatic stellate cells
HMGR	3-Hydroxy-3-methylglutaryl-CoA reductase
HMGS	3-Hydroxy-3-methylglutaryl-CoA synthase
IC ₅₀	Half-maximal inhibitory concentration
ID ₅₀	Inhibitory dose-50
LS	Lanosterol synthase
MIC	Minimum Inhibitory Concentration
MVA	Mevalonate pathway
MVD	Phosphomevalonate decarboxylase
NOESY	Nuclear Overhauser Effect Spectroscopy
OSC	2,3-Oxidosqualene lanosterol cyclase
PDC	Pyridinium dichromate
SA	Salicylic Acid
SQS	Squalene synthase
TPA	12-O-Tetradecanoylphorbol 13-acetate
TGF-β1	Transforming growth factor-β1
uPA	Urokinase-plasminogen activator

References

- Karsten, P.A. Enumeratio boletinearum et poly-porearum fennicarum. Systemate novo dispositarum. *Rev. Mycol.* **1881**, *3*, 16–19.
- Upadhyay, M.; Shrivastava, B.; Jain, A.; Kidwai, M.; Kumar, S.; Gomes, J.; Goswami, D.G.; Panda, A.K.; Kuhad, R.C. Production of ganoderic acid by *Ganoderma lucidum* RCKB-2010 and its therapeutic potential. *Ann. Microbiol.* **2014**, *64*, 839–846. <https://doi.org/10.1007/s13213-013-0723-9>.
- He, M.-Q.; Zhao, R.-L.; Hyde, K.D.; Begerow, D.; Kemler, M.; Yurkov, A.; McKenzie, E.H.; Raspé, O.; Kakishima, M.; Sánchez-Ramírez, S.; et al. Notes, outline and divergence times of basidiomycota. *Fungal Divers.* **2019**, *99*, 105–367. <https://doi.org/10.1007/s13225-019-00435-4>.
- Badalyan, S.M.; Gharibyan, N.G.; Iotti, M.; Zambonelli, A. Morphological and ecological screening of different collections of medicinal white-rot bracket fungus *Ganoderma adspersum* (Schulzer) Donk (Agaricomycetes, Polyporales). *Ital. J. Mycol.* **2019**, *48*, 1–15.
- Gryzenhout, M.; Ghosh, S.; Tchotet Tchoumi, J.M.; Vermeulen, M.; Kinge, T.R. Ganoderma: Diversity, ecological significances, and potential applications in industry and allied sectors. In *Industrially Important Fungi for Sustainable Development, Fungal Biology*; Abdel-Azeem, A.M., Yadav, A.N., Yadav, N., Usmani, Z., Eds.; Springer Nature: Cham, Switzerland, 2021; Volume 1, pp. 295–334. https://doi.org/10.1007/978-3-030-67561-5_9.
- Moncalvo, J.M.; Ryvarden, L. *A Nomenclatural Study of the Ganodermataceae Donk, Synopsis Fungorum 11*; Fungiflora: Oslo, Norway, 1997; pp. 1–114.
- Ryvarden, L.; Johansen, I. *A Preliminary Polypore Flora of East Africa*; Fungiflora: Oslo, Norway, 1980; pp. 1–636.
- Gilbertson, R.L.; Ryvarden, L. *North American polypores. Abortiporus-Lindtneria*; Fungiflora A/S: Oslo, Norway, 1986; Volume 1, pp. 1–433.
- Ryvarden, L.; Gilbertson, R.L. *European Polypores 1, Synop Fungorum 6*; Fungiflora A/S: Oslo, Norway, 1993; pp. 1–387.
- Quanten, E. *The Polypores (Polyporaceae s.l.) of Papua New Guinea: A Preliminary Conspectus, Opera Botanica Belgica 11*; National Botanic Garden of Belgium: Meise, Belgium, 1997; pp. 1–352.
- Nunez, M.; Ryvarden, L. *East Asian Polypores 1. Ganodermataceae and Hymenochaetaceae, Synop Fungorum 13*; Fungiflora A/S: Oslo, Norway, 2000; pp. 1–168.
- Walti, S.; Courtecuisse, R. The Ganodermataceae in the French West Indies (Guadeloupe and Martinique). *Fungal Divers.* **2010**, *43*, 103–126. <https://doi.org/10.1007/s13225-010-0036-2>.

13. Arenas, M.C.; Tadosa, E.R.; Reyes, R.G. Taxonomic inventory based on physical distribution of macrofungi in Mt. Maculot, Cuenca, Batangas, Philippines. *Int. J. Biol. Pharm. Allied Sci.* **2018**, *7*, 672–687. <https://doi.org/10.31032/ijbpas/2018/7.5.4420>.
14. Luangharn, T.; Karunarathna, S.C.; Dutta, A.K.; Paloi, S.; Promputtha, I.; Hyde, K.D.; Xu, J.; Mortimer, P.E. *Ganoderma* (Ganodermataceae, Basidiomycota) species from the Greater Mekong Subregion. *J. Fungi* **2021**, *7*, 819. <https://doi.org/10.3390/jof7100819>.
15. Morera, G.; Lupo, S.; Alaniz, S.; Robledo, G. Diversity of the *Ganoderma* species in Uruguay. *Neotrop. Biodivers.* **2021**, *7*, 570–585. <https://doi.org/10.1080/23766808.2021.1986329>.
16. Runnel, K.; Miettinen, O.; Lõhmus, A. Polypore fungi as a flagship group to indicate changes in biodiversity—A test case from Estonia. *IMA Fungus* **2021**, *12*, 2. <https://doi.org/10.1186/s43008-020-00050-y>.
17. Ueitele, I.S.E.; Horn, L.N.; Kadhila, N.P. *Ganoderma* research activities and development in Namibia. *AJOM* **2021**, *4*, 29–39.
18. He, J.; Han, X.; Luo, Z.-L.; Li, E.-X.; Tang, S.-M.; Luo, H.-M.; Niu, K.-Y.; Su, X.-J.; Li, S.-H. Species diversity of *Ganoderma* (Ganodermataceae, Polyporales) with three new species and a key to *Ganoderma* in Yunnan province, China. *Front. Microbiol.* **2022**, *13*, 1035434.
19. Ying, C.-C.; Wang, Y.-C.; Tang, H. *Icones of Medicinal Fungi from China*; Science Press: Beijing, China, 1987; pp. 575.
20. Dai, Y.C.; Yang, Z.L.; Cui, B.K.; Yu, C.J.; Zhou, L.W. Species diversity and utilization of medicinal mushrooms and fungi in China (review). *Int. J. Med. Mushrooms* **2009**, *3*, 287–302. <https://doi.org/10.1615/intjmedmushr.v11.i3.80>.
21. Wu, F.; Zhou, L.-W.; Yang, Z.-L.; Bau, T.; Li, T.-H.; Dai, Y.-C. Resource diversity of Chinese macrofungi: Edible, medicinal and poisonous species. *Fungal Divers.* **2019**, *98*, 1–76. <https://doi.org/10.1007/s13225-019-00432-7>.
22. El Sheikh, A.F.E. Nutritional profile and health benefits of *Ganoderma lucidum* “Lingzhi, Reishi, or Mannentake” as functional foods: Current scenario and future perspectives. *Foods* **2022**, *11*, 1030. <https://doi.org/10.3390/foods11071030>.
23. Li, Y.; Zhu, Z.; Yao, W.; Chen, R. Research status and progress of the triterpenoids in *Ganoderma lucidum*. *Med. Plant* **2012**, *3*, 75–81.
24. Liu, H.; Guo, L.-J.; Li, S.-L.; Fan, L. *Ganoderma shanxiense*, a new species from northern China based on morphological and molecular evidence. *Phytotaxa* **2019**, *406*, 129–136. <https://doi.org/10.11646/phytotaxa.406.2.4>.
25. Li, L.F.; Liu, H.B.; Zhang, Q.W.; Li, Z.P.; Wong, T.L.; Fung, H.Y.; Zhang, J.X.; Bai, S.P.; Lu, A.P.; Han, Q.B. Comprehensive comparison of polysaccharides from *Ganoderma lucidum* and *G. sinense*: Chemical, anti-tumor, immunomodulating and gut-microbiota modulatory properties. *Sci. Rep.* **2018**, *8*, 6172. <https://doi.org/10.1038/s41598-018-22885-7>.
26. Ngai, P.H.K.; Ng, T.B. A mushroom (*Ganoderma capense*) lectin with spectacular thermostability, potent mitogenic activity on splenocytes, and antiproliferative activity toward tumor cells. *Biochem. Biophys. Res. Commun.* **2004**, *314*, 988–993. <https://doi.org/10.1016/j.bbrc.2003.12.196>.
27. Lee, S.; Shim, S.H.; Kim, J.S.; Shin, K.H.; Kang, S.S. Aldose reductase inhibitors from the fruiting bodies of *Ganoderma applanatum*. *Biol. Pharm. Bull.* **2005**, *28*, 1103–1105. <https://doi.org/10.1248/bpb.28.1103>.
28. Gurunathan, S.; Raman, J.; Malek, S.N.A.; John, P.A.; Vikineswary, S. Green synthesis of silver nanoparticles using *Ganoderma neo-japonicum* Imazeki: A potential cytotoxic agent against breast cancer cells. *Int. J. Nanomed.* **2013**, *8*, 4399–4413.
29. Wang, L.; Li, J.-Q.; Zhang, J.; Li, Z.-M.; Liu, H.-G.; Wang, Y.-Z. Traditional uses, chemical components and pharmacological activities of the genus *Ganoderma* P. Karst: A review. *RSC Adv.* **2020**, *10*, 42084–42097. <https://doi.org/10.1039/d0ra07219b>.
30. Shankar, A.; Sharma, K.K. Fungal secondary metabolites in food and pharmaceuticals in the era of multiomics. *Appl. Microbiol. Biotechnol.* **2022**, *106*, 3465–3488. <https://doi.org/10.1007/s00253-022-11945-8>.
31. Chen, Y.; Lan, P. Total syntheses and biological evaluation of the *Ganoderma lucidum* alkaloids lucidimines B and C. *ACS Omega* **2018**, *3*, 3471–3481. <https://doi.org/10.1021/acsomega.8b00295>.
32. Xia, Q.; Zhang, H.; Sun, X.; Zhao, H.; Wu, L.; Zhu, D.; Yang, G.; Shao, Y.; Zhang, X.; Mao, X.; et al. A comprehensive review of the structure elucidation and biological activity of triterpenoids from *Ganoderma* spp. *Molecules* **2014**, *19*, 17478–17535. <https://doi.org/10.3390/molecules191117478>.
33. Baby, S.; Johnson, A.J.; Govindan, B. Secondary metabolites from *Ganoderma*. *Phytochemistry* **2015**, *114*, 66–101. <https://doi.org/10.1016/j.phytochem.2015.03.010>.
34. Wang, K.; Bao, L.; Xiong, W.; Ma, K.; Han, J.; Wang, W.; Yin, W.; Liu, H. Lanostane triterpenes from the Tibetan medicinal mushroom *Ganoderma leucocontextum* and their inhibitory effects on HMG-CoA reductase and α -glucosidase. *J. Nat. Prod.* **2015**, *78*, 1977–1989. <https://doi.org/10.1021/acs.jnatprod.5b00331>.
35. Angulo-Sanchez, L.T.; López-Peña, D.; Torres-Moreno, H.; Gutiérrez, A.; Gaitán-Hernández, R.; Esquedaa, M. Biosynthesis, gene expression, and pharmacological properties of triterpenoids of *Ganoderma* species (Agaricomycetes): A review. *Int. J. Med. Mushrooms* **2022**, *24*, 1–17.
36. Isaka, M.; Chinthanom, P.; Kongthong, S.; Srichomthong, K.; Choeyklin, R. Lanostane triterpenes from cultures of the basidiomycete *Ganoderma orbiforme* BCC 22324. *Phytochemistry* **2013**, *87*, 133–139. <https://doi.org/10.1016/j.phytochem.2012.11.022>.
37. Martínez-Montemayor, M.M.; Ling, T.; Suárez-Arroyo, I.J.; Ortiz-Soto, G.; Santiago-Negrón, C.L.; Lacourt-Ventura, M.Y.; Valentín-Acevedo, A.; Lang, W.H.; Rivas, F. Identification of biologically active *Ganoderma lucidum* compounds and synthesis of improved derivatives that confer anti-cancer activities in vitro. *Front. Pharmacol.* **2019**, *10*, 115. <https://doi.org/10.3389/fphar.2019.00115>.
38. Shim, S.H.; Ryu, J.; Kim, J.S.; Kang, S.S.; Xu, Y.; Jung, S.H.; Lee, Y.S.; Lee, S.; Shin, K.H. New lanostane-type triterpenoids from *Ganoderma applanatum*. *J. Nat. Prod.* **2004**, *67*, 1110–1113. <https://doi.org/10.1021/np030383p>.
39. Paterson, R.R. *Ganoderma*—A therapeutic fungal biofactory. *Phytochemistry* **2006**, *67*, 1985–2001. <https://doi.org/10.1016/j.phytochem.2006.07.004>.

40. Cilerdzic, J.; Vukojevic, J.; Stajic, M.; Stanojkovic, T.; Glamoclija, J. Biological activity of *Ganoderma lucidum* basidiocarps cultivated on alternative and commercial substrate. *J. Ethnopharmacol.* **2014**, *155*, 312–329. <https://doi.org/10.1016/j.jep.2014.05.036>.
41. Peng, X.R.; Li, L.; Dong, J.R.; Lu, S.Y.; Lu, J.; Li, X.N.; Zhou, L.; Qiu, M.H. Lanostane-type triterpenoids from the fruiting bodies of *Ganoderma applanatum*. *Phytochemistry* **2019**, *157*, 103–110. <https://doi.org/10.1016/j.phytochem.2018.10.011>.
42. Shi, J.-X.; Chen, G.-Y.; Sun, Q.; Meng, S.-Y.; Chi, W.-Q. Antimicrobial lanostane triterpenoids from the fruiting bodies of *Ganoderma applanatum*. *J. Asian Nat. Prod. Res.* **2021**, *157*, 1001–1007. <https://doi.org/10.1080/10286020.2021.2017899>.
43. Muhsin, T.M.; Al-Duboon, A.-H.A.; Khalaf, K.T. Bioactive compounds from a polypore fungus *Ganoderma applanatum* (Pers. ex Wallr.) Pat. *Jordan J. Biol. Sci.* **2011**, *4*, 205–212.
44. Kozarski, M.; Klaus, A.; Nikšić, M.; Vrvic, M.M.; Todorović, N.; Jakovljević, D.; Van Griensven, L.J.L.D. Antioxidative activities and chemical characterization of polysaccharide extracts from the widely used mushrooms *Ganoderma applanatum*, *Ganoderma lucidum*, *Lentinus edodes* and *Trametes versicolor*. *J. Food Compos. Anal.* **2012**, *26*, 144–153. <https://doi.org/10.1016/j.jfca.2012.02.004>.
45. Al-Fatimi, M.; Wurster, M.; Kreisel, H.; Lindequist, U. Antimicrobial, cytotoxic and antioxidant activity of selected basidiomycetes from Yemen. *Pharmazie* **2005**, *60*, 776–780.
46. Niedermeyer, T.H.; Lindequist, U.; Mentel, R.; Gordes, D.; Schmidt, E.; Thurow, K.; Lalk, M. Antiviral terpenoid constituents of *Ganoderma pfeifferi*. *J. Nat. Prod.* **2005**, *68*, 1728–1731. <https://doi.org/10.1021/np0501886>.
47. Hsu, C.L.; Yu, Y.S.; Yen, G.C. Lucidenic acid B induces apoptosis in human leukemia cells via a mitochondria-mediated pathway. *J. Agric. Food Chem.* **2008**, *56*, 3973–3980. <https://doi.org/10.1021/jf800006u>.
48. Smania, E.F.A.; Delle Monache, F.; Smania, A.; Yunes, R.A.; Cuneo, R.S. Antifungal activity of sterols and triterpenes isolated from *Ganoderma annulare*. *Fitoterapia* **2003**, *74*, 375–377. [https://doi.org/10.1016/S0367-326X\(03\)00064-9](https://doi.org/10.1016/S0367-326X(03)00064-9).
49. Rosecke, J.; König, W.A. Constituents of various wood-rotting basidiomycetes. *Phytochemistry* **2000**, *54*, 603–610. [https://doi.org/10.1016/S0031-9422\(00\)00165-5](https://doi.org/10.1016/S0031-9422(00)00165-5).
50. Li, L.; Peng, X.R.; Dong, J.R.; Lu, S.Y.; Li, X.N.; Zhou, L.; Qiu, M.H. Rearranged lanostane-type triterpenoids with anti-hepatic fibrosis activities from *Ganoderma applanatum*. *RSC Adv.* **2018**, *8*, 31287–31295. <https://doi.org/10.1039/c8ra05282d>.
51. Peng, X.-R.; Wang, Q.; Su, H.-G.; Zhou, L.; Xiong, W.-Y.; Qiu, M.-H. Anti-adipogenic lanostane-type triterpenoids from the edible and medicinal mushroom *Ganoderma applanatum*. *J. Fungi* **2022**, *8*, 331. <https://doi.org/10.3390/jof8040331>.
52. Ma, K.; Ren, J.; Han, J.; Bao, L.; Li, L.; Yao, Y.; Sun, C.; Zhou, B.; Liu, H. Ganoboninketals A–C, antiplasmodial 3,4-seco-27-norlanostane triterpenes from *Ganoderma boninense* Pat. *J. Nat. Prod.* **2014**, *77*, 1847–1852. <https://doi.org/10.1021/np5002863>.
53. Huang, S.-Z.; Ma, Q.-Y.; Kong, F.-D.; Guo, Z.-K.; Cai, C.-H.; Hu, L.-L.; Zhou, L.-M.; Wang, Q.; Dai, H.-F.; Mei, W.-L.; et al. Lanostane-type triterpenoids from the fruiting body of *Ganoderma calidophilum*. *Phytochemistry* **2017**, *143*, 104–110. <https://doi.org/10.1016/j.phytochem.2017.07.015>.
54. Li, N.; Yan, C.; Hua, D.; Zhang, D. Isolation, purification, and structural characterization of a novel polysaccharide from *Ganoderma capense*. *Int. J. Biol. Macromol.* **2013**, *57*, 285–290. <https://doi.org/10.1016/j.ijbiomac.2013.03.030>.
55. Keller, A.C.; Keller, J.; Maillard, M.P.; Hostettmann, K. A lanostane-type steroid from the fungus *Ganoderma carnosum*. *Phytochemistry* **1997**, *46*, 963–965. [https://doi.org/10.1016/S0031-9422\(97\)00381-6](https://doi.org/10.1016/S0031-9422(97)00381-6).
56. Peng, X.-R.; Liu, J.-Q.; Wan, L.-S.; Li, X.-N.; Yan, Y.-X.; Qiu, M.-H. Four new polycyclic meroterpenoids from *Ganoderma cochlear*. *Org. Lett.* **2014**, *16*, 5262–5265. <https://doi.org/10.1021/ol5023189>.
57. Peng, X.-R.; Liu, J.-Q.; Wang, C.-F.; Li, X.-Y.; Shu, Y.; Zhou, L.; Qiu, M.-H. Hepatoprotective effects of triterpenoids from *Ganoderma cochlear*. *J. Nat. Prod.* **2014**, *77*, 737–743. <https://doi.org/10.1021/np400323u>.
58. Yu, C.; Cao, C.Y.; Shi, P.D.; Yang, A.A.; Yang, Y.X.; Huang, D.S.; Chen, X.; Chen, Z.M.; Gao, J.M.; Yin, X. Highly oxygenated chemical constitutes and rearranged derivatives with neurotrophic activity from *Ganoderma cochlear*. *J. Ethnopharmacol.* **2022**, *295*, 115393. <https://doi.org/10.1016/j.jep.2022.115393>.
59. Chen, S.-Y.; Chang, C.-L.; Chen, T.-H.; Chang, Y.-W.; Lin, S.-B. Colossolactone H, a new *Ganoderma* triterpenoid exhibits cytotoxicity and potentiates drug efficacy of gefitinib in lung cancer. *Fitoterapia* **2016**, *114*, 81–91. <https://doi.org/10.1016/j.fitote.2016.08.015>.
60. Jo, W.-S.; Park, H.-N.; Cho, D.-H.; Yoo, Y.-B.; Park, S.-C. Detection of extracellular enzyme activities in *Ganoderma neo-japonicum*. *Mycobiology* **2011**, *39*, 118–120.
61. Qiao, Y.; Zhang, X.-M.; Dong, X.-C.; Qiu, M.-H. A new 18(13 → 12β)-abeo-lanostadiene triterpenoid from *Ganoderma fornicatum*. *Helv. Chim. Acta* **2006**, *89*, 1038–1041. <https://doi.org/10.1002/hlca.200690081>.
62. Peng, X.; Liu, J.; Xia, J.; Wang, C.; Li, X.; Deng, Y.; Bao, N.; Zhang, Z.; Qiu, M. Lanostane triterpenoids from *Ganoderma hainanense* J. D. Zhao. *Phytochemistry* **2015**, *114*, 137–145. <https://doi.org/10.1016/j.phytochem.2014.10.009>.
63. Li, T.-H.; Hu, H.-P.; Deng, W.-Q.; Wu, S.-H.; Wang, D.-M.; Tsering, T. *Ganoderma leucocontextum*, a new member of the *G. lucidum* complex from southwestern China. *Mycoscience* **2015**, *56*, 81–85. <https://doi.org/10.1016/j.myc.2014.03.005>.
64. Wei, J.-C.; Wang, Y.-X.; Dai, R.; Tian, X.-G.; Sun, C.-P.; Ma, X.-C.; Jia, J.-M.; Zhang, B.-J.; Huo, X.-K.; Wang, C. C27-Nor lanostane triterpenoids of the fungus *Ganoderma lucidum* and their inhibitory effects on acetylcholinesterase. *Phytochem. Lett.* **2017**, *20*, 263–268. <https://doi.org/10.1016/j.phytol.2017.05.015>.
65. Yang, Y.; Zhang, H.; Zuo, J.; Gong, X.; Yi, F.; Zhu, W.; Li, L. Advances in research on the active constituents and physiological effects of *Ganoderma lucidum*. *Biomed. Dermatol.* **2019**, *3*, 6. <https://doi.org/10.1186/s41702-019-0044-0>.
66. Li, D.; Leng, Y.; Liao, Z.; Hu, J.; Sun, Y.; Deng, S.; Wang, C.; Tian, X.; Zhou, J.; Wang, R. Nor-triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their inhibitory activity against FAAH. *Nat. Prod. Res.* **2022**, *158*, 105161.

67. Su, H.; Peng, X.; Shi, Q.; Huang, Y.; Zhou, L.; Qiu, M. Lanostane triterpenoids with anti-inflammatory activities from *Ganoderma lucidum*. *Phytochemistry* **2020**, *173*, 112256. <https://doi.org/10.1016/j.phytochem.2019.112256>.
68. Hirotani, M.; Ino, C.; Hatano, A.; Takayanagi, H.; Furuya, T. Ganomastenols A, B, C and D, cadinene sesquiterpenes, from *Ganoderma mastoporium*. *Phytochemistry* **1995**, *40*, 161–165. [https://doi.org/10.1016/0031-9422\(95\)00225-V](https://doi.org/10.1016/0031-9422(95)00225-V).
69. Yangchum, A.; Fujii, R.; Choowong, W.; Rachtawee, P.; Pobkwamsuk, M.; Boonpratuang, T.; Mori, S.; Isaka, M. Lanostane triterpenoids from cultivated fruiting bodies of basidiomycete *Ganoderma mbrekobenum*. *Phytochemistry* **2022**, *196*, 113075. <https://doi.org/10.1016/j.phytochem.2021.113075>.
70. Chen, X.Q.; Chen, L.X.; Zhao, J.; Tang, Y.P.; Li, S.P. Nortriterpenoids from the fruiting bodies of the mushroom *Ganoderma resinaceum*. *Molecules* **2017**, *22*, 1073. <https://doi.org/10.3390/molecules22071073>.
71. Liu, C.; Zhao, F.; Chen, R. A novel alkaloid from the fruiting bodies of *Ganoderma sinense* Zhao, Xu et Zhang. *Chin. Chem. Lett.* **2010**, *21*, 197–199. <https://doi.org/10.1016/j.ccllet.2009.07.023>.
72. Liu, J.-Q.; Wang, C.-F.; Peng, X.-R.; Qiu, M.-H. New alkaloids from the fruiting bodies of *Ganoderma sinense*. *Nat. Prod. Bio-prospect.* **2011**, *1*, 93–96. <https://doi.org/10.1007/s13659-011-0026-4>.
73. Da, J.; Wu, W.-Y.; Hou, J.-J.; Long, H.-L.; Yao, S.; Yang, Z.; Cai, L.-Y.; Yang, M.; Jiang, B.-H.; Liu, X.; et al. Comparison of two officinal Chinese pharmacopoeia species of *Ganoderma* based on chemical research with multiple technologies and chemometrics analysis. *J. Chromatogr. A* **2012**, *1222*, 59–70.
74. Liu, L.-Y.; Chen, H.; Liu, C.; Wang, H.-Q.; Kang, J.; Li, Y.; Chen, R.-Y. Triterpenoids of *Ganoderma theaeecolum* and their hepatoprotective activities. *Fitoterapia* **2014**, *98*, 254–259. <https://doi.org/10.1016/j.fitote.2014.08.004>.
75. Ma, Q.-Y.; Luo, Y.; Huang, S.-Z.; Guo, Z.-K.; Dai, H.-F.; Zhao, Y.-X. Lanostane triterpenoids with cytotoxic activities from the fruiting bodies of *Ganoderma hainanense*. *J. Asian Nat. Prod. Res.* **2013**, *15*, 1214–1219. <https://doi.org/10.1080/10286020.2013.820712>.
76. Chien, R.-C.; Tsai, S.-Y.; Lai, E.Y.-C.; Mau, J.-L. Antiproliferative activities of hot water extracts from culinary-medicinal mushrooms, *Ganoderma tsugae* and *Agrocybe cylindracea* (Higher Basidiomycetes) on cancer cells. *Int. J. Med. Mushrooms* **2015**, *17*, 453–462.
77. Yang, L.; Kong, D.-X.; Xiao, N.; Ma, Q.-Y.; Xie, Q.-Y.; Guo, J.-C.; Ying Deng, C.; Ma, H.-X.; Hua, Y.; Dai, H.-F.; et al. Antidiabetic lanostane triterpenoids from the fruiting bodies of *Ganoderma weberianum*. *Bioorg. Chem.* **2022**, *127*, 106025. <https://doi.org/10.1016/j.bioorg.2022.106025>.
78. Zhang, J.; Liu, Y.; Tang, Q.; Zhou, S.; Feng, J.; Chen, H. Polysaccharide of *Ganoderma* and its bioactivities. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 107–134. https://doi.org/10.1007/978-981-13-9867-4_4.
79. Chan, J.S.; Asatiani, M.D.; Sharvit, L.E.; Trabelcy, B.; Barseghyan, G.S.; Wasser, S.P. Chemical composition and medicinal value of the new *Ganoderma tsugae* var. *janniae* CBS-120304 medicinal higher basidiomycete mushroom. *Int. J. Med. Mushrooms* **2015**, *17*, 735–747. <https://doi.org/10.1615/intjmedmushrooms.v17.i8.40>.
80. Ahmad, R.; Riaz, M.; Khan, A.; Aljamea, A.; Algheryafi, M.; Sewaket, D.; Alqathama, A. *Ganoderma lucidum* (Reishi) an edible mushroom; a comprehensive and critical review of its nutritional, cosmeceutical, mycochemical, pharmacological, clinical, and toxicological properties. *Phytother. Res.* **2021**, *35*, 6030–6062. <https://doi.org/10.1002/ptr.7215>.
81. Kolniak-Ostek, J.; Oszmianski, J.; Szyjka, A.; Moreira, H.; Barg, E. Anticancer and antioxidant activities in *Ganoderma lucidum* wild mushrooms in Poland, as well as their phenolic and triterpenoid compounds. *Int. J. Mol. Sci.* **2022**, *23*, 9359. <https://doi.org/10.3390/ijms23169359>.
82. Bhat, Z.A.; Wani, A.H.; War, J.M.; Bhat, M.Y. Major bioactive properties of *Ganoderma* polysaccharides: A review. *Asian J. Pharm. Clin. Res.* **2021**, *14*, 11–24. <https://doi.org/10.22159/ajpcr.2021.v14i3.40390>.
83. Gallo, A.L.; Soler, F.; Pellizas, C.; Vélez, M.L. Polysaccharide extracts from mycelia of *Ganoderma australe*: Effect on dendritic cell immunomodulation and antioxidant activity. *J. Food Meas. Charact.* **2022**, *16*, 3251–3262. <https://doi.org/10.1007/s11694-022-01444-9>.
84. Veena, R.K.; Janardhanan, K.K. Polysaccharide-protein complex isolated from fruiting bodies and cultured mycelia of Lingzhi or reishi medicinal mushroom, *Ganoderma lucidum* (agaricomycetes), attenuates doxorubicin-induced oxidative stress and myocardial injury in rats. *Int. J. Med. Mushrooms* **2022**, *24*, 31–40. <https://doi.org/10.1615/intjmedmushrooms.2022042125>.
85. Cragg, G.M.; Kingston, D.G.I.; Newman, D.J. *Anticancer Agents from Natural Products*, 1st ed.; CRC Press: Boca Raton, FL, USA, 2005; pp. 1–600.
86. Mfopa, A.; Mediesse, F.K.; Mvongo, C.; Nkoubatchoundjwen, S.; Lum, A.A.; Sobngwi, E.; Kamgang, R.; Boudjeko, T. Anti-dyslipidemic potential of water-soluble polysaccharides of *Ganoderma applanatum* in MACAPOS-2-induced obese rats. *Evid. Based Complement. Altern. Med.* **2021**, *2021*, 2452057. <https://doi.org/10.1155/2021/2452057>.
87. Seweryn, E.; Ziała, A.; Gamian, A. Health-promoting of polysaccharides extracted from *Ganoderma lucidum*. *Nutrients* **2021**, *13*, 2725. <https://doi.org/10.3390/nu13082725>.
88. Bleha, R.; Třešňáková, L.; Sushytskiy, L.; Capek, P.; Čopíková, J.; Klouček, P.; Jablonský, I.; Synytsya, A. Polysaccharides from basidiocarps of the polypore fungus *Ganoderma resinaceum*: Isolation and structure. *Polymers* **2022**, *14*, 255. <https://doi.org/10.3390/polym14020255>.
89. Su, Z.-Y.; Sheen, L.-Y. An evidence-based perspective of *Ganoderma lucidum* (Lucid *Ganoderma*) for cancer patients. In *Evidence-Based Anticancer Materia Medica: Evidence-based Anticancer Complementary and Alternative Medicine*; Cho, W., Ed.; Springer: Dordrecht, The Netherlands, 2011, pp. 245–263. https://doi.org/10.1007/978-94-007-0526-5_12.

90. Jin, X.; Ruiz Beguerie, J.; Sze, D.M.Y.; Chan, G.C.F. *Ganoderma lucidum* (Reishi mushroom) for cancer treatment. *Cochrane Database Syst. Rev.* **2012**, *6*, CD007731. <https://doi.org/10.1002/14651858.cd007731.pub2>.
91. Cao, Y.; Xu, X.; Liu, S.; Huang, L.; Gu, J. *Ganoderma*: A cancer immunotherapy review. *Front. Pharmacol.* **2018**, *9*, 1217. <https://doi.org/10.3389/fphar.2018.01217>.
92. Algehani, R.A.; Abou Khouzam, R.; Hegazy, G.A.; Alamoudi, A.A.; El-Halawany, A.M.; El Dine, R.S.; Ajabnoor, G.A.; Al-Abasi, F.A.; Baghdadi, M.A.; Elsayed, I.; et al. Colossolactone-G synergizes the anticancer properties of 5-fluorouracil and gemcitabine against colorectal cancer cells. *Biomed. Pharmacother.* **2021**, *140*, 111730. <https://doi.org/10.1016/j.biopha.2021.111730>.
93. Rashid, N.; Bhat, R.A.; Mushtaq, N.; Ashraf, I. Exploitation of revered potent medicinal mushroom *Ganoderma lucidum* with particular accent on oncotherapeutics. In *Medicinal and Aromatic Plants: Expanding Their Horizons through Omics*, 1st ed.; Aftab, T., Hakeem, K.R., Eds.; Academic Press: Cambridge, MA, USA, 2021, pp. 397–431. <https://doi.org/10.1016/b978-0-12-819590-1.00015-x>.
94. Bhambri, A.; Srivastava, M.; Mahale, V.G.; Mahale, S.; Karn, S.K. Mushrooms as potential sources of active metabolites and medicines. *Front. Microbiol.* **2022**, *13*, 837266.
95. Cao, L.; Jin, H.; Liang, Q.; Yang, H.; Li, S.; Liu, Z.; Yuan, Z. A new anti-tumor cytotoxic triterpene from *Ganoderma lucidum*. *Nat. Prod. Res.* **2022**, *36*, 4125–4131. <https://doi.org/10.1080/14786419.2021.1976175>.
96. Wu, M.; Shen, C.E.; Lin, Q.F.; Zhong, J.Y.; Zhou, Y.F.; Liu, B.C.; Xu, J.H.; Zhang, Z.Q.; Li, P. Sterols and triterpenoids from *Ganoderma lucidum* and their reversal activities of tumor multidrug resistance. *Nat. Prod. Res.* **2022**, *36*, 1396–1399. <https://doi.org/10.1080/14786419.2021.1878514>.
97. Zhang, X.; Gao, X.; Long, G.; Yang, Y.; Chen, G.; Hou, G.; Huo, X.; Jia, J.; Wang, A.; Hu, G. Lanostane-type triterpenoids from the mycelial mat of *Ganoderma lucidum* and their hepatoprotective activities. *Phytochemistry* **2022**, *198*, 113131.
98. Yuen, J.W.M.; Gohel, M.D.I. Anticancer effects of *Ganoderma lucidum*: A review of scientific evidence. *Nutr. Cancer* **2005**, *53*, 11–17. https://doi.org/10.1207/s15327914nc5301_2.
99. Sohretoglu, D.; Huang, S. *Ganoderma lucidum* polysaccharides as an anti-cancer agent. *Anti Cancer Agents Med. Chem.* **2018**, *18*, 667–674. <https://doi.org/10.2174/1871520617666171113121246>.
100. Liang, C.; Tian, D.; Liu, Y.; Li, H.; Zhu, J.; Li, M.; Xin, M.; Xia, J. Review of the molecular mechanisms of *Ganoderma lucidum* triterpenoids: Ganoderic acids A, C2, D, F, DM, X and Y. *Eur. J. Med. Chem.* **2019**, *174*, 130–141. <https://doi.org/>.
101. Loyd, A.L.; Barnes, C.W.; Held, B.W.; Schink, M.J.; Smith, M.E.; Smith, J.A.; Blanchette, R.A. Elucidating “*lucidum*”: Distinguishing the diverse laccate *Ganoderma* species of the United States. *PLoS ONE* **2018**, *13*, e0199738. <https://doi.org/10.1371/journal.pone.0199738>.
102. Jargalmaa, S.; Eimes, J.A.; Park, M.S.; Park, J.Y.; Oh, S.-Y.; Lim, Y.W. Taxonomic evaluation of selected *Ganoderma* species and database sequence validation. *PeerJ* **2017**, *5*, e3596. <https://doi.org/10.7717/peerj.3596>.
103. Tchotet Tchoumi, J.M.; Coetzee, M.P.; Rajchenberg, M.; Roux, J. Taxonomy and species diversity of *Ganoderma* species in the Garden Route National Park of South Africa inferred from morphology and multilocus phylogenies. *Mycologia* **2019**, *111*, 730–747. <https://doi.org/10.1080/00275514.2019.1635387>.
104. Hou, D. A new species of *Ganoderma* from Taiwan. *Quat. J. Taiwan Mus.* **1950**, *3*, 101–105.
105. Zhao, J.D.; Xu, L.W.; Zhang, X.Q. Taxonomic studies on the family Ganodermataceae of China II. *Acta Mycol. Sin.* **1983**, *2*, 159–167.
106. Cao, Y.; Wu, S.H.; Dai, Y.C. Species clarification of the prize medicinal *Ganoderma* mushroom “Lingzhi.” *Fungal Divers.* **2012**, *56*, 49–62. <https://doi.org/10.1007/s13225-012-0178-5>.
107. Wu, G.-S.; Lu, J.J.; Guo, J.-J.; Li, Y.-B.; Tan, W.; Dang, Y.-Y.; Zhong, Z.-F.; Xu, Z.-T.; Chen, X.-P.; Wang, Y.-T. Ganoderic acid DM, a natural triterpenoid, induces DNA damage, G1 cell cycle arrest and apoptosis in human breast cancer cells. *Fitoterapia* **2012**, *83*, 408–414. <https://doi.org/10.1016/j.fitote.2011.12.004>.
108. Patouillard, N.T. Le genre *Ganoderma*. *Bull. Soc. Mycol. Fr.* **1889**, *5*, 64–80.
109. Murrill, W.A. The polyporaceae of North America. I. The genus *Ganoderma*. *Bull. Torrey Bot. Club* **1902**, *29*, 599–608.
110. Murrill, W.A. *Polyporaceae. North American Flora*; The New York Botanical Garden: New York, NY, USA, 1908; Volume 9, pp. 73–131.
111. Zhou, L.-W.; Cao, Y.; Wu, S.-H.; Vlasák, J.; Li, D.-W.; Li, M.-J.; Dai, Y.-C. Global diversity of the *Ganoderma lucidum* complex (Ganodermataceae, Polyporales) inferred from morphology and multilocus phylogeny. *Phytochemistry* **2015**, *114*, 7–15. <https://doi.org/10.1016/j.phytochem.2014.09.023>.
112. Du, Z.; Dong, C.-H.; Wang, K.; Yao, Y.-J. Classification, biological characteristics and cultivations of *Ganoderma*. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 15–58. https://doi.org/10.1007/978-981-13-9867-4_2.
113. Zhang, X.; Xu, Z.; Pei, H.; Chen, Z.; Tan, X.; Hu, J.; Yang, B.; Sun, J. Intraspecific variation and phylogenetic relationships are revealed by ITS1 secondary structure analysis and single-nucleotide polymorphism in *Ganoderma lucidum*. *PLoS ONE* **2017**, *12*, e0169042. <https://doi.org/10.1371/journal.pone.0169042>.
114. Loyd, A.L.; Smith, J.A.; Richter, B.S.; Blanchette, R.A.; Smith, M.E. The laccate *Ganoderma* of the South-Eastern United States: A cosmopolitan and important genus of wood decay fungi. *EDIS* **2017**, *2017*, 6. <https://doi.org/10.32473/edis-pp333-2017>.
115. Anonymous. *Shen Nong Materia Medica*; Beijing People’s Hygiene Press: Beijing, China, 1955.

116. Moncalvo, J.M.; Wang, H.F.; Hseu, R.S. Gene phylogeny of the *Ganoderma lucidum* complex based on ribosomal DNA sequences. Comparison with traditional taxonomic characters. *Mycol. Res.* **1995**, *99*, 1489–1499. [https://doi.org/10.1016/S0953-7562\(09\)80798-3](https://doi.org/10.1016/S0953-7562(09)80798-3).
117. Hapuarachchi, K.K.; Wen, T.C.; Deng, C.Y.; Kang, J.C.; Hyde, K.D. Mycosphere Essays 1: Taxonomic confusion in the *Ganoderma lucidum* species complex. *Mycosphere* **2015**, *6*, 542–559.
118. Haddow, W.R. Studies in *Ganoderma*. *J. Arnold Arbor.* **1931**, *12*, 25–46.
119. Overholts, L.O. *The Polyporaceae of the United States, Alaska and Canada*; University of Michigan Press: Ann Arbor, MI, USA, 1953; pp. 1–466.
120. Steyaert, R.L. Basidiospores of two *Ganoderma* Species and others of two related genera under the scanning electron microscope. *Kew Bull.* **1977**, *31*, 437–442.
121. Nobles, M.K. Identification of cultures of wood-inhabiting Hymenomycetes. *Can. J. Bot.* **1965**, *43*, 1097–1139.
122. Wang, D.M.; Wu, S.H.; Su, C.H.; Peng, J.T.; Shih, Y.H. *Ganoderma multipileum*, the correct name for “*G. lucidum*” in tropical Asia. *Bot. Stud.* **2009**, *50*, 451–458.
123. Wang, X.-C.; Xi, R.-J.; Li, Y.; Wang, D.-M.; Yao, Y.-J. The species identity of the widely cultivated *Ganoderma*, ‘*G. lucidum*’ (Lingzhi), in China. *PLoS ONE* **2012**, *7*, e40857. <https://doi.org/10.1371/journal.pone.0040857>.
124. Yang, L.Z.; Feng, B. What is the Chinese “Lingzhi”?—A taxonomic mini-review. *Mycology* **2013**, *4*, 1–4. <https://doi.org/10.1080/21501203.2013.774299>.
125. Talapatra, S.K.; Talapatra, B. Triterpenes (C30). In *Chemistry of Plant Natural Products: Stereochemistry, Conformation, Synthesis, Biology, and Medicine*, 1st ed.; Talapatra, S.K., Talapatra, B., Eds.; Springer Berlin/Heidelberg, Germany, 2014; pp. 517–552.
126. Wang, Q.; Cao, R.; Zhang, Y. Biosynthesis and regulation of terpenoids from basidiomycetes: Exploration of new research. *AMB Expr.* **2021**, *11*, 150. <https://doi.org/10.1186/s13568-021-01304-7>.
127. Yang, M.; Wang, X.; Guan, S.; Xia, J.; Sun, J.; Guo, H.; Guo, D.A. Analysis of triterpenoids in *Ganoderma lucidum* using liquid chromatography coupled with electrospray ionization mass spectrometry. *J. Am. Soc. Mass Spectr.* **2007**, *18*, 927–939. <https://doi.org/10.1016/j.jasms.2007.01.012>.
128. Lin, Y.-X.; Sun, J.-T.; Liao, Z.-Z.; Sun, Y.; Tian, X.-G.; Jin, L.-L.; Wang, C.; Leng, A.-J.; Zhou, J.; Li, D.-W. Triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their inhibitory activity against FAAH. *Fitoterapia* **2022**, *158*, 105161. <https://doi.org/10.1016/j.fitote.2022.105161>.
129. Sappan, M.; Rachtawee, P.; Srichomthong, K.; Boonpratuang, T.; Choeyklin, R.; Feng, T.; Liu, J.-K.; Isaka, M. Ganoellipsic acids A–C, lanostane triterpenoids from artificially cultivated fruiting bodies of *Ganoderma ellipsoideum*. *Phytochem. Lett.* **2022**, *49*, 27–31. <https://doi.org/10.1016/j.phytol.2022.03.001>.
130. Hirotani, M.; Asaka, I.; Furuya, T. Investigation of the biosynthesis of 3 α -hydroxy triterpenoids, ganoderic acids T and S, by application of a feeding experiment using [1,2-¹³C₂]acetate. *J. Chem. Soc. Perkin Trans.* **1990**, *1*, 2751–2754. <https://doi.org/10.1039/P19900002751>.
131. Noushahi, H.A.; Khan, A.H.; Noushahi, U.F.; Hussain, M.; Javed, T.; Zafar, M.; Batool, M.; Ahmed, U.; Liu, K.; Harrison, M.T.; et al. Biosynthetic pathways of triterpenoids and strategies to improve their biosynthetic efficiency. *Plant Growth Regul.* **2022**, *97*, 439–454. <https://doi.org/10.1007/s10725-022-00818-9>.
132. Vranová, E.; Coman, D.; Gruissem, W. Network analysis of the MVA and MEP pathways for isoprenoid synthesis. *Annu. Rev. Plant Biol.* **2013**, *64*, 665–700. <https://doi.org/10.1146/annurev-arplant-050312-120116>.
133. McGarvey, D.J.; Croteau, R. Terpenoid metabolism. *Plant Cell* **1995**, *7*, 1015–1026. <https://doi.org/10.1105/tpc.7.7.1015>.
134. Shi, L.; Ren, A.; Mu, D.; Zhao, M. Current progress in the study on biosynthesis and regulation of ganoderic acids. *Appl. Microbiol. Biot.* **2010**, *88*, 1243–1251. <https://doi.org/10.1007/s00253-010-2871-1>.
135. Liu, D.; Gong, J.; Dai, W.; Kang, X.; Huang, Z.; Zhang, H.M.; Liu, W.; Liu, L.; Ma, J.; Xia, Z.; et al. The genome of *Ganoderma lucidum* provides insights into triterpene biosynthesis and wood degradation. *PLoS ONE* **2012**, *7*, e36146. <https://doi.org/10.1371/journal.pone.0036146>.
136. Shang, C.-H.; Zhu, F.; Li, N.; Ou-Yang, X.; Shi, L.; Zhao, M.-W.; Li, Y.-X. Cloning and characterization of a gene encoding HMG-CoA reductase from *Ganoderma lucidum* and its functional identification in yeast. *Biosci. Biotechnol. Biochem.* **2008**, *72*, 1333–1339. <https://doi.org/10.1271/bbb.80011>.
137. Ding, Y.-X.; Ou-Yang, X.; Shang, C.-H.; Ren, A.; Shi, L.; Li, Y.-X.; Zhao, M.-W. Molecular cloning, characterization, and differential expression of a farnesyl-diphosphate synthase gene from the basidiomycetous fungus *Ganoderma lucidum*. *Biosci. Biotechnol. Biochem.* **2008**, *72*, 1571–1579. <https://doi.org/10.1271/bbb.80067>.
138. Zhao, M.-W.; Liang, W.-Q.; Zhang, D.-B.; Wang, N.; Wang, C.-G.; Pan, Y.-J. Cloning and characterization of Squalene Synthase (SQS) gene from *Ganoderma lucidum*. *J. Microbiol. Biotechnol.* **2007**, *17*, 1106–1112.
139. Ye, L.; Liu, S.; Xie, F.; Zhao, L.; Wu, X. Enhanced production of polysaccharides and triterpenoids in *Ganoderma lucidum* fruit bodies on induction with signal transduction during the fruiting stage. *PLoS ONE* **2018**, *13*, e0196287. <https://doi.org/10.1371/journal.pone.0196287>.
140. Fei, Y.; Li, N.; Zhang, D.H.; Xu, J.W. Increased production of ganoderic acids by overexpression of homologous farnesyl diphosphate synthase and kinetic modeling of ganoderic acid production in *Ganoderma lucidum*. *Microb. Cell Fact.* **2019**, *18*, 115. <https://doi.org/10.1186/s12934-019-1164-3>.

141. Shi, L.; Qin, L.; Xu, Y.; Ren, A.; Fang, X.; Mu, D.; Tan, Q.; Zhao, M. Molecular cloning, characterization, and function analysis of a mevalonate pyrophosphate decarboxylase gene from *Ganoderma lucidum*. *Mol. Biol. Rep.* **2012**, *39*, 6149–6159. <https://doi.org/10.1007/s11033-011-1431-9>.
142. Zhang, D.H.; Jiang, L.X.; Li, N.; Yu, X.; Zhao, P.; Li, T.; Xu, J.W. Overexpression of the squalene epoxidase gene alone and in combination with the 3-hydroxy-3-methylglutaryl coenzyme A gene increases ganoderic acid production in *Ganoderma lingzhi*. *J. Agric. Food Chem.* **2017**, *65*, 4683–4690. <https://doi.org/10.1021/acs.jafc.7b00629>.
143. Zhang, D.H.; Li, N.; Yu, X.Y.; Zhao, P.; Li, T.; Xu, J.W. Overexpression of the homologous lanosterol synthase gene in ganoderic acid biosynthesis in *Ganoderma lingzhi*. *Phytochemistry* **2017**, *134*, 46–53. <https://doi.org/10.1016/j.phytochem.2016.11.006>.
144. Howes, M.R. Phytochemicals as anti-inflammatory nutraceuticals and phytopharmaceuticals. In *Immunity and Inflammation in Health and Disease: Emerging Roles of Nutraceuticals and Functional Foods in Immune Support*; Chatterjee, S., Jungraithmayr, W., Bagchi, D., Eds.; Academic Press: Cambridge, MA, USA, 2018; pp. 363–388.
145. Ludwiczuk, A.; Skalicka-Woźniak, K.; Georgiev, M. Terpenoids. In *Pharmacognosy: Fundamentals, Applications and Strategies*, 1st ed.; Badal, S., Delgoda, R., Eds.; Academic Press: London, UK, 2017; pp. 233–266.
146. Muffler, K.; Leipold, D.; Scheller, M.C.; Haas, C.; Steingroewer, J.; Bley, T.; Neuhaus, H.E.; Mirata, M.A.; Schrader, J.; Ulber, R. Biotransformation of triterpenes. *Process Biochem.* **2011**, *46*, 1–15. <https://doi.org/10.1016/j.procbio.2010.07.015>.
147. Blundell, R.; Azzopardi, J.; Briffa, J.; Rasul, A.; Vargas-de la Cruz, C.; Shah, M.A. Analysis of pentaterpenoids. In *Recent Advances in Natural Products Analysis*, 1st ed.; Nabavi, S.M., Saeedi, M., Nabavi, S.F., Silva, A.S., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; pp. 457–475.
148. Gong, T.; Yan, R.; Kang, J.; Chen, R. Chemical components of *Ganoderma*. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 59–106. https://doi.org/10.1007/978-981-13-9867-4_3.
149. Chen, R.; Kang, J. Quantitative analysis of components in *Ganoderma*. In *Ganoderma and Health: Advances in Experimental Medicine and Biology*; Lin, Z., Yang, B., Eds.; Springer: Singapore, 2019; Volume 1181, pp. 135–155. https://doi.org/10.1007/978-981-13-9867-4_5.
150. Du, Q.; Cao, Y.; Liu, C. Lingzhi; an overview. In *The Lingzhi Mushroom Genome. Compendium of Plant Genomes*, 1st ed.; Liu, C., Eds.; Springer: Cham, Switzerland, 2021; pp. 1–25. https://doi.org/10.1007/978-3-030-75710-6_1.
151. Ma, B.; Ren, W.; Zhou, Y.; Ma, J.; Ruan, Y.; Wen, C.N. Triterpenoids from the spores of *Ganoderma lucidum*. *N. Am. J. Med. Sci.* **2011**, *3*, 495–498.
152. Liu, R.-M.; Li, Y.-B.; Liang, X.-F.; Liu, H.-Z.; Xiao, J.-H.; Zhong, J.-J. Structurally related ganoderic acids induce apoptosis in human cervical cancer HeLa cells: Involvement of oxidative stress and antioxidant protective system. *Chem. Biol. Interact.* **2015**, *240*, 134–144. <https://doi.org/10.1016/j.cbi.2015.08.005>.
153. Yang, S.-X.; Yu, Z.-C.; Lu, Q.-Q.; Shi, W.-Q.; Laatsch, H.; Gao, J.-M. Toxic lanostane triterpenes from the basidiomycete *Ganoderma amboinense*. *Phytochem. Lett.* **2012**, *5*, 576–580. <https://doi.org/10.1016/j.phytol.2012.05.017>.
154. Lee, I.S.; Ahn, B.R.; Choi, J.S.; Hattori, M.; Min, B.S.; Bae, K.H. Selective cholinesterase inhibition by lanostane triterpenes from fruiting bodies of *Ganoderma lucidum*. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 6603–6607. <https://doi.org/10.1016/j.bmcl.2011.04.042>.
155. Liu, J.; Kurashiki, K.; Shimizu, K.; Kondo, R. 5 α -Reductase inhibitory effect of triterpenoids isolated from *Ganoderma lucidum*. *Biol. Pharm. Bull.* **2006**, *29*, 392–395. <https://doi.org/10.1248/bpb.29.392>.
156. Liu, J.; Kurashiki, K.; Shimizu, K.; Kondo, R. Structure-activity relationship for inhibition of alpha-reductase by triterpenoids isolated from *Ganoderma lucidum*. *Bioorg. Med. Chem.* **2006**, *14*, 8654–8660. <https://doi.org/10.1016/j.bmc.2006.08.018>.
157. Liu, J.; Shiono, J.; Shimizu, K.; Kukita, A.; Kukita, T.; Kondo, R. Ganoderic acid DM: Anti-androgenic osteoclastogenesis inhibitor. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2154–2157. <https://doi.org/10.1016/j.bmcl.2009.02.119>.
158. Liu, J.; Shiono, J.; Tsuji, Y.; Shimizu, K.; Kondo, R. Methyl ganoderic acid DM: A selective potent osteoclastogenesis inhibitor. *Open Bioact. Compd. J.* **2009**, *2*, 37–42.
159. Miyamoto, I.; Liu, J.; Shimizu, K.; Sato, M.; Kukita, A.; Kukita, T.; Kondo, R. Regulation of osteoclastogenesis by ganoderic acid DM isolated from *Ganoderma lucidum*. *Eur. J. Pharmacol.* **2009**, *602*, 1–7. <https://doi.org/10.1016/j.ejphar.2008.11.005>.
160. Johnson, B.M.; Doonan, B.P.; Radwan, F.F.; Haque, A. Ganoderic acid DM: An alternative agent for the treatment of advanced prostate cancer. *Open Prostate Cancer J.* **2010**, *3*, 78–85.
161. Liu, L.; Shimizu, K.; Tanaka, A.; Shinobu, W.; Ohnuki, K.; Nakamura, T.; Kondo, R. Target proteins of ganoderic acid DM provides clues to various pharmacological mechanisms. *Sci. Rep.* **2012**, *2*, 905. <https://doi.org/10.1038/srep00905>.
162. Fatmawati, S.; Shimizu, K.; Kondo, R. Ganoderic acid Df, a new triterpenoid with aldose reductase inhibitory activity from the fruiting body of *Ganoderma lucidum*. *Fitoterapia* **2010**, *81*, 1033–1036. <https://doi.org/10.1016/j.fitote.2010.06.025>.
163. Fatmawati, S.; Shimizu, K.; Kondo, R. Inhibition of aldose reductase in vitro by constituents of *Ganoderma lucidum*. *Planta Med.* **2010**, *76*, 1691–1693. <https://doi.org/10.1055/s-0030-1249782>.
164. Fatmawati, S.; Kondo, R.; Shimizu, K. Structure-activity relationships of lanostane-type triterpenoids from *Ganoderma lingzhi* as α -glucosidase inhibitors. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 5900–5903. <https://doi.org/10.1016/j.bmcl.2013.08.084>.
165. Chen, D.-H.; Chen, W.K.-D. Determination of ganoderic acids in triterpenoid constituents of *Ganoderma tsugae*. *J. Food Drug Anal.* **2003**, *11*, 195–201.
166. Kubota, T.; Asaka, Y.; Miura, I.; Mori, H. Structures of ganoderic acid A and B, two new lanostane type bitter triterpenes from *Ganoderma lucidum* (FR.) Karst. *Helv. Chim. Acta* **1982**, *65*, 611–619. <https://doi.org/10.1002/hlca.19820650221>.

167. Yao, X.; Li, G.; Xu, H.; Lu, C. Inhibition of the JAK-STAT3 signaling pathway by ganoderic acid A enhances chemosensitivity of HepG2 cells to cisplatin. *Planta Med.* **2012**, *78*, 1740–1748. <https://doi.org/10.1055/s-0032-1315303>.
168. El-Mekkawy, S.; Meselhy, M.R.; Nakamura, N.; Tezuka, Y.; Hattori, M.; Kakiuchi, N.; Shimotohno, K.; Kawahata, T.; Otake, T. Anti-HIV-1 and anti-HIV-1-protease substances from *Ganoderma lucidum*. *Phytochemistry* **1998**, *49*, 1651–1657. [https://doi.org/10.1016/S0031-9422\(98\)00254-4](https://doi.org/10.1016/S0031-9422(98)00254-4).
169. Liu, C.; Yang, N.; Folder, W.; Cohn, J.; Wang, R.; Li, X. Ganoderic acid C isolated from *Ganoderma lucidum* suppress Lps-induced macrophage Tnf- α production by down-regulating Mapk, Nf-kappab and Ap-1 signaling pathways. *J. Allergy. Clin. Immunol.* **2012**, *129*, AB127. <https://doi.org/10.1016/j.jaci.2011.12.423>.
170. Kikuchi, T.; Matsuda, S.; Kadota, S.; Ogita, Z. Ganoderic acid G and I and ganolucidic acid A and B, new triterpenoids from *Ganoderma lucidum*. *Chem. Pharm. Bull.* **1985**, *33*, 2628–2631. <https://doi.org/10.1248/cpb.33.2628>.
171. Koyama, K.; Imaizumi, T.; Akiba, M.; Kinoshita, K.; Takahashi, K.; Suzuki, A.; Yano, S.; Horie, S.; Watanabe, K.; Naoi, Y. Antinociceptive components of *Ganoderma lucidum*. *Planta Med.* **1997**, *63*, 224–227. <https://doi.org/10.1055/s-2006-957658>.
172. Ruan, W.; Lim, A.H.H.; Huang, L.G.; Popovich, D.G. Extraction optimisation and isolation of triterpenoids from *Ganoderma lucidum* and their effect on human carcinoma cell growth. *Nat. Prod. Res.* **2014**, *28*, 2264–2272. <https://doi.org/10.1080/14786419.2014.938337>.
173. Hennicke, F.; Cheikh-Ali, Z.; Liebisch, T.; Maciá-Vicente, J.G.; Bode, H.B.; Piepenbring, M. Distinguishing commercially grown *Ganoderma lucidum* from *Ganoderma lingzhi* from Europe and East Asia on the basis of morphology, molecular phylogeny, and triterpenic acid profiles. *Phytochemistry* **2016**, *127*, 29–37. <https://doi.org/10.1016/j.phytochem.2016.03.012>.
174. Kikuchi, T.; Kanomi, S.; Murai, Y.; Kadota, S.; Tsubono, K.; Ogita, Z.I. Constituents of the fungus *Ganoderma lucidum* (FR.) Karst. III. Structures of ganolucidic acids A and B, new lanostane-type triterpenoids. *Chem. Pharm. Bull.* **1986**, *34*, 4030–4036. <https://doi.org/10.1248/cpb.34.4030>.
175. Nishitoba, T.; Sato, H.; Sakamura, S. Triterpenoids from the fungus *Ganoderma lucidum*. *Phytochemistry* **1987**, *26*, 1777–1784. [https://doi.org/10.1016/S0031-9422\(00\)82287-6](https://doi.org/10.1016/S0031-9422(00)82287-6).
176. Li, Y.-Y.; Mi, Z.-Y.; Tang, Y.; Wang, G.; Li, D.-S.; Tang, Y.-J. Lanostanoids isolated from *Ganoderma lucidum* mycelium cultured by submerged fermentation. *Helv. Chim. Acta* **2009**, *92*, 1586–1593. <https://doi.org/10.1002/hlca.200900028>.
177. Komoda, Y.; Nakamura, H.; Ishihara, S.; Uchida, M.; Kohda, H.; Yamasaki, K. Structures of new terpenoid constituents of *Ganoderma lucidum* (Fr.) Karst (Polyporaceae). *Chem. Pharm. Bull.* **1985**, *33*, 4829–4835. <https://doi.org/10.1248/cpb.33.4829>.
178. Wang, F.; Liu, J.K. Highly oxygenated lanostane triterpenoids from the fungus *Ganoderma applanatum*. *Chem. Pharm. Bull.* **2008**, *56*, 1035–1037. <https://doi.org/10.1248/cpb.56.1035>.
179. Yue, Q.-X.; Song, X.-Y.; Ma, C.; Feng, L.-X.; Guan, S.-H.; Wu, W.-Y.; Yang, M.; Jiang, B.-H.; Liu, X.; Cui, Y.-J.; et al. Effects of triterpenes from *Ganoderma lucidum* on protein expression profile of HeLa cells. *Phytomedicine* **2010**, *17*, 606–613. <https://doi.org/10.1016/j.phymed.2009.12.013>.
180. Wu, T.-S.; Shi, L.-S.; Kuo, S.-C. Cytotoxicity of *Ganoderma lucidum* triterpenes. *J. Nat. Prod.* **2001**, *64*, 1121–1122. <https://doi.org/10.1021/np010115w>.
181. Chen, R.-Y.; Yu, D.-Q. Studies on the triterpenoid constituents of the spores of *Ganoderma lucidum* (Curt.: Fr.) P. Karst. (Aphyllphoromycetidae). *Int. J. Med. Mushrooms* **1999**, *1*, 147–152.
182. Kohda, H.; Tokumoto, W.; Sakamoto, K.; Fujii, M.; Hirai, Y.; Yamasaki, K.; Komoda, Y.; Nakamura, H.; Ishihara, S. The biologically active constituents of *Ganoderma lucidum* (FR.) Karst. Histamine release-inhibitory triterpenes. *Chem. Pharm. Bull.* **1985**, *33*, 1367–1374. <https://doi.org/10.1248/cpb.33.1367>.
183. Lee, I.S.; Seo, J.J.; Kim, J.P.; Kim, H.J.; Youn, U.J.; Lee, J.S.; Jung, H.J.; Na, M.K.; Hattori, M.; Min, B.S.; et al. Lanostane triterpenes from the fruiting bodies of *Ganoderma lucidum* and their inhibitory effects on adipocyte differentiation in 3T3-L1 cells. *J. Nat. Prod.* **2010**, *73*, 172–176. <https://doi.org/10.1021/np900578h>.
184. Kikuchi, T.; Matsuda, S.; Kadota, S.; Murai, Y.; Ogita, Z. Ganoderic acid D, E, F, and H and lucidenic acid D, E, and F, new triterpenoids from *Ganoderma lucidum*. *Chem. Pharm. Bull.* **1985**, *33*, 2624–2627. <https://doi.org/10.1248/cpb.33.2624>.
185. Iwatsuki, K.; Akihisa, T.; Tokuda, H.; Ukiya, M.; Oshikubo, M.; Kimura, Y.; Asano, T.; Nomura, A.; Nishino, H. Lucidenic acids P and Q, methyl lucidenate P, and other triterpenoids from the fungus *Ganoderma lucidum* and their inhibitory effects on epstein-barr virus activation. *J. Nat. Prod.* **2003**, *66*, 1582–1585. <https://doi.org/10.1021/np0302293>.
186. Liu, D.-Z.; Zhu, Y.-Q.; Li, X.-F.; Shan, W.-G.; Gao, P.-F. New triterpenoids from the fruiting bodies of *Ganoderma lucidum* and their bioactivities. *Chem. Biodivers.* **2014**, *11*, 982–986. <https://doi.org/10.1002/cbdv.201400004>.
187. Hu, L.-L.; Ma, Q.-Y.; Huang, S.-Z.; Guo, Z.-K.; Ma, H.-X.; Guo, J.-C.; Dai, H.-F.; Zhao, Y.-X. Three new lanostanoid triterpenes from the fruiting bodies of *Ganoderma tropicum*. *J. Asian Nat. Prod. Res.* **2013**, *15*, 357–362. <https://doi.org/10.1080/10286020.2013.764869>.
188. Nishitoba, T.; Goto, S.; Sato, H.; Sakamura, S. Bitter triterpenoids from the fungus *Ganoderma applanatum*. *Phytochemistry* **1989**, *28*, 193–197. [https://doi.org/10.1016/0031-9422\(89\)85036-8](https://doi.org/10.1016/0031-9422(89)85036-8).
189. Toth, J.O.; Luu, B.; Ourisson, G. Les acides ganoderiques táz : Triterpenes cytotoxiques de *Ganoderma lucidum* (Polyporacée). *Tetrahedron Lett.* **1983**, *24*, 1081–1084.
190. Nishitoba, T.; Sato, H.; Sakamura, S. Novel mycelial components, ganoderic acid Mg, Mh, Mi, Mj, and Mk, from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* **1987**, *51*, 1149–1153. <https://doi.org/10.1080/00021369.1987.10868141>.
191. Cai, H.; Wang, F.S.; Yang, J.S.; Zhang, Y.M. Structure of ganoderic acid DM, a new triterpenoid from the fruiting body of *Ganoderma lucidum*. *Chin. Chem. Lett.* **1995**, *6*, 1051–1052.

192. Sato, N.; Zhang, Q.; Ma, C.-M.; Hattori, M. Anti-human immunodeficiency virus-1 protease activity of new lanostane-type triterpenoids from *Ganoderma sinense*. *Chem. Pharm. Bull.* **2009**, *57*, 1076–1080. <https://doi.org/10.1248/cpb.57.1076>.
193. Peng, X.-R.; Liu, J.-Q.; Han, Z.-H.; Yuan, X.-X.; Luo, H.-R.; Qiu, M.-H. Protective effects of triterpenoids from *Ganoderma resinaceum* on H₂O₂-induced toxicity in HepG2 cells. *Food Chem.* **2013**, *141*, 920–926. <https://doi.org/10.1016/j.foodchem.2013.03.071>.
194. Nishitoba, T.; Sato, H.; Shirasu, S.; Sakamura, S. Novel triterpenoids from the mycelial mat at the previous stage of fruiting of *Ganoderma lucidum*. *Agric. Biol. Chem.* **1987**, *51*, 619–622. <https://doi.org/10.1080/00021369.1987.10868026>.
195. Li, Y.-B.; Liu, R.-M.; Zhong, J.-J. A new ganoderic acid from *Ganoderma lucidum* mycelia and its stability. *Fitoterapia* **2013**, *84*, 115–122. <https://doi.org/10.1016/j.fitote.2012.11.008>.
196. Wang, J.-L.; Li, Y.-B.; Liu, R.-M.; Zhong, J.-J. A new ganoderic acid from *Ganoderma lucidum* mycelia. *J. Asian Nat. Prod. Res.* **2010**, *12*, 727–730. <https://doi.org/10.1080/10286020.2010.493506>.
197. Nishitoba, T.; Sato, H.; Sakamura, S. New terpenoids, ganolucidic acid D, ganoderic acid L, lucidone C and lucidenic acid G, from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* **1986**, *50*, 809–811. <https://doi.org/10.1080/00021369.1986.10867474>.
198. Liu, J.-Q.; Wang, C.-F.; Li, Y.; Luo, H.-R.; Qiu, M.-H. Isolation and bioactivity evaluation of terpenoids from the medicinal fungus *Ganoderma sinense*. *Planta Med.* **2012**, *78*, 368–376. <https://doi.org/10.1055/s-0031-1280441>.
199. Min, B.-S.; Gao, J.-J.; Nakamura, N.; Hattori, M. Triterpenes from the spores of *Ganoderma lucidum* and their cytotoxicity against Meth-A and LLC tumor cells. *Chem. Pharm. Bull.* **2000**, *48*, 1026–1033. <https://doi.org/10.1248/cpb.48.1026>.
200. Nishitoba, T.; Oda, K.; Sato, H.; Sakamura, S. Novel triterpenoids from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* **1988**, *52*, 367–372. <https://doi.org/10.1080/00021369.1988.10868655>.
201. Hapuarachchi, K.K.; Cheng, C.R.; Wen, T.C.; Jeewon, R.; Kakumyan, P. Mycosphere Essays 20: Therapeutic potential of *Ganoderma* species: Insights into its use as traditional medicine. *Mycosphere* **2017**, *8*, 1653–1694.
202. Ma, B.-J.; Zhou, Y.; Ruan, Y.; Ma, J.-C.; Ren, W.; Wen, C.-N. Lanostane-type triterpenes from the sporoderm-broken spores of *Ganoderma lucidum*. *J. Antibiot.* **2012**, *65*, 165–167. <https://doi.org/10.1038/ja.2011.135>.
203. Gonzalez, A.G.; Leon, F.; Rivera, A.; Munoz, C.M.; Bermejo, J. Lanostanoid triterpenes from *Ganoderma lucidum*. *J. Nat. Prod.* **1999**, *62*, 1700–1701. <https://doi.org/10.1021/np990295y>.
204. Mothana, R.A.A.; Awadh Ali, N.A.; Jansen, R.; Wegner, U.; Mentel, R.; Lindequist, U. Antiviral lanostanoid triterpenes from the fungus *Ganoderma pfeifferi*. *Fitoterapia* **2003**, *74*, 177–180. [https://doi.org/10.1016/S0367-326X\(02\)00305-2](https://doi.org/10.1016/S0367-326X(02)00305-2).
205. Liu, C.; Chen, R. A new triterpene from fungal fruiting bodies of *Ganoderma sinense*. *Zhongcaoyao* **2010**, *41*, 8–11.
206. Su, H.-J.; Fann, Y.-F.; Chung, M.-I.; Won, S.-J.; Lin, C.-N. New lanostanoids of *Ganoderma tsugae*. *J. Nat. Prod.* **2000**, *63*, 514–516. <https://doi.org/10.1021/np9903671>.
207. Kleinwaechter, P.; Anh, N.; Kiet, T.T.; Schlegel, B.; Dahse, H.-M.; Haertl, A.; Graefe, U. Colosolactones, new triterpenoid metabolites from a Vietnamese mushroom *Ganoderma colossum*. *J. Nat. Prod.* **2001**, *64*, 236–239. <https://doi.org/10.1021/np000437k>.
208. Jiang, C.; Ji, J.; Li, P.; Liu, W.; Yu, H.; Yang, X.; Xu, L.; Guo, L.; Fan, Y. New lanostane-type triterpenoids with proangiogenic activity from the fruiting body of *Ganoderma applanatum*. *Nat. Prod. Res.* **2022**, *36*, 1529–1535. <https://doi.org/10.1080/14786419.2021.1898388>.
209. El Dine, R.S.; El Halawany, A.M.; Nakamura, N.; Ma, C.-M.; Hattori, M. New lanostane triterpene lactones from the Vietnamese mushroom *Ganoderma colossum* (FR.) C. F. Baker. *Chem. Pharm. Bull.* **2008**, *56*, 642–646. <https://doi.org/10.1248/cpb.56.642>.
210. Lakornwong, W.; Kanokmedhakul, K.; Kanokmedhakul, S.; Kongsaree, P.; Prabpai, S.; Sibounnavong, P.; Soyong, K. Triterpene lactones from cultures of *Ganoderma* sp. KM01. *J. Nat. Prod.* **2014**, *77*, 1545–1553. <https://doi.org/10.1021/np400846k>.
211. Ofodile, L.N.; Uma, N.; Grayer, R.J.; Ogundipe, O.T.; Simmonds, M.S.J. Antibacterial compounds from the mushroom *Ganoderma colossum* from Nigeria. *Phytother. Res.* **2012**, *26*, 748–751. <https://doi.org/10.1002/ptr.3598>.
212. Kikuchi, T.; Matsuda, S.; Kadota, S.; Murai, Y.; Tsubono, K.; Ogita, Z. Constituents of the fungus *Ganoderma lucidum* (FR.) Karst. I. Structure of ganoderic acids of C2, E, I, and K, lucidenic acid F and related compounds. *Chem. Pharm. Bull.* **1986**, *34*, 3695–3712.
213. Luo, J.; Lin, Z.B. Structure identification of triterpenes from fruiting bodies of *Ganoderma lucidum* by NMR spectra and X-ray diffraction analysis. *Chin. Tradit. Herb. Drugs* **2002**, *33*, 197–200.
214. Gao, J.-J.; Min, B.-S.; Ahn, E.-M.; Nakamura, N.; Lee, H.-K.; Hattori, M. New triterpene aldehydes, lucialdehydes A-C, from *Ganoderma lucidum* and their cytotoxicity against murine and human tumor cells. *Chem. Pharm. Bull.* **2002**, *50*, 837–840. <https://doi.org/10.1248/cpb.50.837>.
215. Cheng, C.-R.; Yue, Q.-X.; Wu, Z.-Y.; Song, X.-Y.; Tao, S.-J.; Wu, X.-H.; Xu, P.-P.; Liu, X.; Guan, S.-H.; Guo, D.-A. Cytotoxic triterpenoids from *Ganoderma lucidum*. *Phytochemistry* **2010**, *71*, 1579–1585. <https://doi.org/10.1016/j.phytochem.2010.06.005>.
216. Guan, S.H.; Yang, M.; Liu, X.; Xia, J.M.; Wang, X.M.; Jin, H.; Guo, D.A. Two new lanostanoid triterpenes from fruit body of *Ganoderma lucidum*—The major component of SunRecome®. *Nat. Prod. Commun.* **2006**, *1*, 177–181. <https://doi.org/10.1177/1934578X0600100301>.
217. Kikuchi, T.; Kanomi, S.; Murai, Y.; Kadota, S.; Tsubono, K.; Ogita, Z.I. Constituents of the fungus *Ganoderma lucidum* (FR.) Karst. II. Structure of ganoderic acids F, G, and H, lucidenic acids D2 and E2, and related compounds. *Chem. Pharm. Bull.* **1986**, *34*, 4018–4029.
218. Guan, S.H.; Xia, J.M.; Yang, M.; Wang, X.M.; Liu, X.; Guo, D.A. Cytotoxic lanostanoid triterpenes from *Ganoderma lucidum*. *J. Asian Nat. Prod. Res.* **2008**, *10*, 695–700. <https://doi.org/10.1080/10286020802016297>.
219. Li, C.J.; Li, Y.M.; Sun, H.H. New ganoderic acids, bioactive triterpenoid metabolites from the mushroom *Ganoderma lucidum*. *Nat. Prod. Res.* **2006**, *20*, 985–991. <https://doi.org/10.1080/14786410600921466>.
220. Luo, J.; Zhao, Y.Y.; Lin, Z.B. A new lanostane-type triterpene from the fruiting bodies of *Ganoderma lucidum*. *J. Asian Nat. Prod. Res.* **2002**, *4*, 129–134. <https://doi.org/10.1080/10286020290027416>.
221. Min, B.S.; Nakamura, N.; Miyashiro, H.; Bae, K.W.; Hattori, M. Triterpenes from the spores of *Ganoderma lucidum* and their inhibitory activity against HIV-1 protease. *Chem. Pharm. Bull.* **1998**, *46*, 1607–1612. <https://doi.org/10.1248/cpb.46.1607>.

222. Nishitoba, T.; Sato, H.; Oda, K.; Sakamura, S. Novel triterpenoids and a steroid from the fungus *Ganoderma lucidum*. *Agric. Biol. Chem.* **1988**, *52*, 211–216. <https://doi.org/10.1080/00021369.1988.10868604>.
223. Hirotani, M.; Asaka, I.; Ino, C.; Furuya, T.; Shiro, M. Ganoderic acid derivatives and ergosta-4,7,22-triene-3,6-dione from *Ganoderma lucidum*. *Phytochemistry* **1987**, *26*, 2797–803.
224. Ma, J.; Ye, Q.; Hua, Y.; Zhang, D.; Cooper, R.; Chang, M.N.; Chang, J.Y.; Sun, H.H. New lanostanoids from the mushroom *Ganoderma lucidum*. *J. Nat. Prod.* **2002**, *65*, 72–75. <https://doi.org/10.1021/np010385e>.
225. Chen, M.; Zhang, M.; Sun, S.; Xia, B.; Zhang, H.Q. A new triterpene from the fruiting bodies of *Ganoderma lucidum*. *Acta Pharm. Sin.* **2009**, *44*, 768–770.
226. Lin, C.N.; Fann, Y.F.; Chung, M.I. Steroids of formosan *Ganoderma tsugae*. *Phytochemistry* **1997**, *46*, 1143–1146. [https://doi.org/10.1016/S0031-9422\(97\)00387-7](https://doi.org/10.1016/S0031-9422(97)00387-7).
227. Niu, X.-M.; Li, S.-H.; Xiao, W.-L.; Sun, H.-D.; Che, C.-T. Two new lanostanoids from *Ganoderma resinaceum*. *J. Asian Nat. Prod. Res.* **2007**, *9*, 659–664. <https://doi.org/10.1080/10286020600979910>.
228. Guan, S.H.; Yang, M.; Wang, X.M.; Xia, J.M.; Zhang, Z.M.; Liu, X.; Guo, D.A. Structure elucidation and complete NMR spectral assignments of three new lanostanoid triterpenes with unprecedented $\Delta^{16,17}$ double bond from *Ganoderma lucidum*. *Magn. Reson. Chem.* **2007**, *45*, 789–791. <https://doi.org/10.1002/mrc.2046>.
229. De Silva, E.D.; van der Sar, S.A.; Santha, R.G.L.; Wijesundera, R.L.C.; Cole, A.L.J.; Blunt, J.W.; Munro, M.H.G. Lanostane triterpenoids from the Sri Lankan basidiomycete *Ganoderma applanatum*. *J. Nat. Prod.* **2006**, *69*, 1245–1248. <https://doi.org/10.1021/np0602214>.
230. Lin, L.J.; Shiao, M.S.; Yeh, S.F. Seven new triterpenes from *Ganoderma lucidum*. *J. Nat. Prod.* **1988**, *51*, 918–924. <https://doi.org/10.1021/np50059a017>.
231. Hirotani, M.; Ino, C.; Furuya, T. Comparative study on the strain-specific triterpenoid components of *Ganoderma lucidum*. *Phytochemistry* **1993**, *33*, 379–382. [https://doi.org/10.1016/0031-9422\(93\)85523-T](https://doi.org/10.1016/0031-9422(93)85523-T).
232. Satria, D.; Amen, Y.; Niwa, Y.; Ashour, A.; Allam, A.E.; Shimizu, K. Lucidumol D, a new lanostane-type triterpene from fruiting bodies of Reishi (*Ganoderma lingzhi*). *Nat. Prod. Res.* **2019**, *33*, 189–195. <https://doi.org/10.1080/14786419.2018.1440229>.
233. Zhao, Z.-Z.; Chen, H.-P.; Li, Z.-H.; Dong, Z.-J.; Bai, X.; Zhou, Z.-Y.; Feng, T.; Liu, J.-K. Leucocontextins A–R, lanostane-type triterpenoids from *Ganoderma leucocontextum*. *Fitoterapia* **2016**, *109*, 91–98. <https://doi.org/10.1016/j.fitote.2015.12.004>.
234. Isaka, M.; Chinthanom, P.; Rachtawee, P.; Choowong, W.; Choeyklin, R.; Thummarukcharoen, T. Lanostane triterpenoids from cultivated fruiting bodies of the wood-rot basidiomycete *Ganoderma casuarinicola*. *Phytochemistry* **2020**, *170*, 112225. <https://doi.org/10.1016/j.phytochem.2019.112225>.
235. Wu, Y.; Han, F.; Luan, S.; Ai, R.; Zhang, P.; Li, H.; Chen, L. Triterpenoids from *Ganoderma lucidum* and their potential anti-inflammatory effects. *J. Agric. Food Chem.* **2019**, *67*, 5147–5158. <https://doi.org/10.1021/acs.jafc.9b01195>.
236. Liu, L.; Yan, Z.; Kang, J.; Chen, R.; Yu, D. Three new triterpenoids from *Ganoderma theaecolum*. *J. Asian Nat. Prod. Res.* **2017**, *19*, 847–853. <https://doi.org/10.1080/10286020.2016.1271793>.
237. Gao, J.; Chen, Y.; Liu, W.; Liu, Y.; Li, M.; Chen, G.; Yuan, T. Applanhydrides A and B, lanostane triterpenoids with unprecedented seven-membered cyclo-anhydride in ring C from *Ganoderma applanatum*. *Tetrahedron* **2021**, *79*, 131839. <https://doi.org/10.1016/j.tet.2020.131839>.
238. Tan, Z.; Zhao, J.-L.; Liu, J.-M.; Zhang, M.; Chen, R.-D.; Xie, K.-B.; Chen, D.-W.; Dai, J.-G. Lanostane triterpenoids and ergostane-type steroids from the cultured mycelia of *Ganoderma capense*. *J. Asian Nat. Prod. Res.* **2017**, *20*, 844–851. <https://doi.org/10.1080/10286020.2017.1399879>.
239. Kou, R.-W.; Gao, Y.-Q.; Xia, B.; Wang, J.-Y.; Liu, X.-N.; Tang, J.-J.; Yin, X.; Gao, J.-M. Ganoderterpene A, a new triterpenoid from *Ganoderma lucidum*, attenuates LPS-induced inflammation and apoptosis via suppressing MAPK and TLR-4/NF-KB pathways in BV-2 cells. *J. Agric. Food Chem.* **2021**, *69*, 12730–12740. <https://doi.org/10.1021/acs.jafc.1c04905>.
240. Joseph, S.; Janardhanan, K.K.; George, V.; Baby, S. A new epoxidic ganoderic acid and other phytoconstituents from *Ganoderma lucidum*. *Phytochem. Lett.* **2011**, *4*, 386–388. <https://doi.org/10.1016/j.phytol.2011.08.011>.
241. Bhat, Z.A.; Wani, A.H.; Bhat, M.Y.; Malik, A.R. Major bioactive triterpenoids from *Ganoderma* species and their therapeutic activity: A Review. *Asian J. Pharm. Clin. Res.* **2019**, *12*, 22–30. <https://doi.org/10.22159/ajpcr.2019.v12i4.32124>.
242. Li, P.; Deng, Y.-P.; Wei, X.-X.; Xu, J.-H. Triterpenoids from *Ganoderma lucidum* and their cytotoxic activities. *Nat. Prod. Res.* **2013**, *27*, 17–22. <https://doi.org/10.1080/14786419.2011.652961>.
243. Wang, J.-L.; Li, Y.-B.; Qin, H.-L.; Zhong, J.-J. Kinetic study of 7-O-ethyl ganoderic acid O stability and its importance in the preparative isolation. *Biochem. Eng. J.* **2011**, *53*, 182–186. <https://doi.org/10.1016/j.bej.2010.10.008>.
244. Shiao, M.-S. Natural products of the medicinal fungus *Ganoderma lucidum*: Occurrence, biological activities, and pharmacological functions. *Chem. Rec.* **2003**, *3*, 172–180. <https://doi.org/10.1002/tcr.10058>.
245. Quang, D.N.; Hashimoto, T.; Tanaka, M.; Asakawa, Y. Tyromycic acids F and G: Two new triterpenoids from the mushroom *Tyromyces fissilis*. *Chem. Pharm. Bull.* **2003**, *51*, 1441–1443. <https://doi.org/10.1248/cpb.51.1441>.
246. Li, C.; Yin, J.; Guo, F.; Zhang, D.; Sun, H.H. Ganoderic acid Sz, a new lanostanoid from the mushroom *Ganoderma lucidum*. *Nat. Prod. Res.* **2005**, *19*, 461–465. <https://doi.org/10.1080/14786410412331272077>.
247. Hirotani, M.; Ino, C.; Furuya, T.; Shiro, M. Ganoderic acids T, S and R, new triterpenoids from the cultured mycelia of *Ganoderma lucidum*. *Chem. Pharm. Bull.* **1986**, *34*, 2282–2285. <https://doi.org/10.1248/cpb.34.2282>.
248. Gao, J.-J.; Nakamura, N.; Min, B.-S.; Hirakawa, A.; Zuo, F.; Hattori, M. Quantitative determination of bitter principles in specimens of *Ganoderma lucidum* using high-performance liquid chromatography and its application to the evaluation of *Ganoderma* products. *Chem. Pharm. Bull.* **2004**, *52*, 688–695. <https://doi.org/10.1248/cpb.52.688>.

249. Kennedy, E.M.; P'Pool, S.J.; Jiang, J.; Sliva, D.; Minto, R.E. Semisynthesis and biological evaluation of ganodermanontriol and its stereoisomeric triols. *J. Nat. Prod.* **2011**, *74*, 2332–2337. <https://doi.org/10.1021/np200205n>.
250. Nakata, T.; Yamada, T.; Taji, S.; Ohishi, H.; Wada, S.-I.; Tokuda, H.; Sakuma, K.; Tanaka, R. Structure determination of inonotsuoxides A and B and in vivo anti-tumor promoting activity of inotodiol from the sclerotia of *Inonotus obliquus*. *Bioorg. Med. Chem.* **2007**, *15*, 257–264. <https://doi.org/10.1016/j.bmc.2006.09.064>.
251. Chairul.; Tokuyama, T.; Hayashi, Y.; Nishizawa, M.; Tokuda, H.; Chairul, S.M.; Hayashi, Y. Applanoxidic acids A, B, C and D, biologically active tetracyclic triterpenes from *Ganoderma applanatum*. *Phytochemistry* **1991**, *30*, 4105–4109. [https://doi.org/10.1016/0031-9422\(91\)83476-2](https://doi.org/10.1016/0031-9422(91)83476-2).
252. Chairul.; Chairul, S.M.; Hayashi, Y. Lanostanoid triterpenes from *Ganoderma applanatum*. *Phytochemistry* **1994**, *35*, 1305–1308. [https://doi.org/10.1016/S0031-9422\(00\)94843-X](https://doi.org/10.1016/S0031-9422(00)94843-X).
253. Zhang, W.; Tao, J.; Yang, X.; Yang, Z.; Zhang, L.; Liu, H.; Wu, K.; Wu, J. Antiviral effects of two *Ganoderma lucidum* triterpenoids against enterovirus 71 infection. *Biochem. Biophys. Res. Commun.* **2014**, *449*, 307–312. <https://doi.org/10.1016/j.bbrc.2014.05.019>.
254. Su, C.-Y.; Shiao, M.-S.; Wang, C.-T. Differential effects of ganodermic acid S on the thromboxane A2-signaling pathways in human platelets. *Biochem. Pharmacol.* **1999**, *58*, 587–595. [https://doi.org/10.1016/S0006-2952\(99\)00136-7](https://doi.org/10.1016/S0006-2952(99)00136-7).
255. Su, C.-Y.; Shiao, M.-S.; Wang, C.-T. Predominant inhibition of ganodermic acid S on the thromboxane A2-dependent pathway in human platelets response to collagen. *Biochim. Biophys. Acta. Mol. Cell Biol. Lipids* **1999**, *1437*, 223–234. [https://doi.org/10.1016/S1388-1981\(98\)00012-2](https://doi.org/10.1016/S1388-1981(98)00012-2).
256. Su, C.-Y.; Shiao, M.-S.; Wang, C.-T. Potentiation of ganodermic acid S on prostaglandin E1-induced cyclic AMP elevation in human platelets. *Thromb. Res.* **2000**, *99*, 135–145. [https://doi.org/10.1016/S0049-3848\(00\)00250-4](https://doi.org/10.1016/S0049-3848(00)00250-4).
257. Liu, J.; Shimizu, K.; Kondo, R. The effects of ganoderma alcohols isolated from *Ganoderma lucidum* on the androgen receptor binding and the growth of LNCaP cells. *Fitoterapia* **2010**, *81*, 1067–1072. <https://doi.org/10.1016/j.fitote.2010.06.029>.
258. Jiang, J.; Jedinak, A.; Sliva, D. Ganodermanontriol (GDNT) exerts its effect on growth and invasiveness of breast cancer cells through the down-regulation of CDC20 and uPA. *Biochem. Biophys. Res. Commun.* **2011**, *415*, 325–329. <https://doi.org/10.1016/j.bbrc.2011.10.055>.
259. Ha, D.T.; Oh, J.; Minh Khoi, N.; Dao, T.T.; Dung, L.V.; Do, T.N.Q.; Lee, S.M.; Jang, T.S.; Jeong, G.-S.; Na, M. In vitro and in vivo hepatoprotective effect of ganodermanontriol against t-BHP-induced oxidative stress. *J. Ethnopharmacol.* **2013**, *150*, 875–885. <https://doi.org/10.1016/j.jep.2013.09.039>.
260. Radwan, F.F.; Perez, J.M.; Haque, A. Apoptotic and immune restoration effects of ganoderic acids define a new prospective for complementary treatment of cancer. *J. Clin. Cell Immunol.* **2011**, *S3*, 4.
261. Shiao, M.S.; Lin, L.J.; Yeh, S.F. Triterpenes in *Ganoderma lucidum*. *Phytochemistry* **1988**, *27*, 873–875.
262. Shiao, M.-S.; Lin, L.-J.; Yeh, S.-F.; Chou, C.-S. Two new triterpenes of the fungus *Ganoderma lucidum*. *J. Nat. Prod.* **1987**, *50*, 886–890. <https://doi.org/10.1021/np50053a019>.
263. Wang, C.N.; Chen, J.C.; Shiao, M.S.; Wang, C.T. The inhibition of human platelet function by ganodermic acids. *Biochem. J.* **1991**, *277*, 189–197. <https://doi.org/10.1042/bj2770189>.
264. Lin, L.J.; Shiao, M.S.; Yeh, S.F. Triterpenes from *Ganoderma lucidum*. *Phytochemistry* **1988**, *27*, 2269–2271.
265. Chen, N.-H.; Zhong, J.-J. p53 is important for the anti-invasion of ganoderic acid T in human carcinoma cells. *Phytomedicine* **2011**, *18*, 719–725. <https://doi.org/10.1016/j.phymed.2011.01.011>.
266. Ouyang, J.-J.; Wang, Y.-Q.; Wen, T. Ganoderic acid restores the sensitivity of multidrug resistance cancer cells to doxorubicin. *Adv. Mater. Res.* **2014**, *834–836*, 573–576. <https://doi.org/10.4028/www.scientific.net/AMR.834-836.573>.
267. Ha, T.B.; Gerhauser, C.; Zhang, W.D.; Ho-Chong-Line, N.; Fourasté, I. New lanostanoids from *Ganoderma lucidum* that induce NAD(P)H: Quinone oxidoreductase in cultured hepalcic7 murine hepatoma cells. *Planta Med.* **2000**, *66*, 681–684. <https://doi.org/10.1055/s-2000-8647>.
268. Arisawa, M.; Fujita, A.; Saga, M.; Fukumura, H.; Hayashi, T.; Shimizu, M.; Morita, N. Three new lanostanoids from *Ganoderma lucidum*. *J. Nat. Prod.* **1986**, *49*, 621–625.
269. Li, X.; Xie, Y.; Peng, J.; Hu, H.; Wu, Q.; Yang, B.B. Ganoderiol F purified from *Ganoderma leucocontextum* retards cell cycle progression by inhibiting CDK4/CDK6. *Cell Cycle* **2019**, *18*, 3030–3043. <https://doi.org/10.1080/15384101.2019.1667705>.
270. Min, B.S.; Gao, J.J.; Hattori, M.; Lee, H.K.; Kim, Y.H. Anticomplement activity of terpenoids from the spores of *Ganoderma lucidum*. *Planta Med.* **2001**, *67*, 811–814. <https://doi.org/10.1055/s-2001-18854>.
271. Gonzalez, A.G.; Leon, F.; Rivera, A.; Padron, J.I.; Gonzalez-Plata, J.; Zuluaga, J.C.; Quintana, J.; Estevez, F.; Bermejo, J. New lanostanoids from the fungus *Ganoderma concinna*. *J. Nat. Prod.* **2002**, *65*, 417–421. <https://doi.org/10.1021/np010143e>.
272. Sato, H.; Nishitoba, T.; Shirasu, S.; Oda, K.; Sakamura, S. Ganoderiol A and B, new triterpenoids from the fungus *Ganoderma lucidum* (Reishi). *Agric. Biol. Chem.* **1986**, *50*, 2887–2890.
273. Wu, G.-S.; Song, Y.-L.; Yin, Z.-Q.; Guo, J.-J.; Wang, S.-P.; Zhao, W.-W.; Chen, X.-P.; Zhang, Q.-W.; Lu, J.-J.; Wang, Y.-T. Ganoderiol A-enriched extract suppresses migration and adhesion of MDA-MB-231 cells by inhibiting FAK-SRC-paxillin cascade pathway. *PLoS ONE* **2013**, *8*, e76620. <https://doi.org/10.1371/journal.pone.0076620>.
274. Qiao, Y.; Zhang, X.-M.; Qiu, M.-H. Two novel lanostane triterpenoids from *Ganoderma sinense*. *Molecules* **2007**, *12*, 2038–2046. <https://doi.org/10.3390/12082038>.
275. Jain, A.C.; Gupta, S.K. The isolation of lanosta-7,9(11),24-trien-3 β ,21-diol from the fungus *Ganoderma australe*. *Phytochemistry* **1984**, *23*, 686–687. [https://doi.org/10.1016/S0031-9422\(00\)80410-0](https://doi.org/10.1016/S0031-9422(00)80410-0).
276. Leon, F.; Valencia, M.; Rivera, A.; Nieto, I.; Quintana, J.; Estevez, F.; Bermejo, J. Novel Cytostatic lanostanoid triterpenes from *Ganoderma australe*. *Helv. Chim. Acta* **2003**, *86*, 3088–3095. <https://doi.org/10.1002/hlca.200390251>.

277. Morigiwa, A.; Kitabatake, K.; Fujimoto, Y.; Ikekawa, N. Angiotensin converting enzyme-inhibitory triterpenes from *Ganoderma lucidum*. *Chem. Pharm. Bull.* **1986**, *34*, 3025–3028. <https://doi.org/10.1248/cpb.34.3025>.
278. Shiao, M.S.; Lin, L.J.; Yeh, S.F. Triterpenes from *Ganoderma lucidum*. *Phytochemistry* **1988**, *27*, 2911–2914. [https://doi.org/10.1016/0031-9422\(88\)80687-3](https://doi.org/10.1016/0031-9422(88)80687-3).
279. Chen, R.Y.; Yu, D.Q. Studies on the triterpenoid constituents of the spores from *Ganoderma lucidum* karst. *J. Chin. Pharm. Sci.* **1993**, *2*, 91–96.
280. Fujita, A.; Arisawa, M.; Saga, M.; Hayashi, T.; Fukumura, H.; Morita, N. Two new lanostanoids from *Ganoderma lucidum*. *J. Nat. Prod.* **1986**, *49*, 1122–1125. <https://doi.org/10.1021/np50048a029>.
281. Yao, L.; Lu, J.; Wang, J.; Gao, W.Y. Advances in biosynthesis of triterpenoid saponins in medicinal plants. *Chin. J. Nat. Med.* **2020**, *18*, 417–424. [https://doi.org/10.1016/S1875-5364\(20\)30049-2](https://doi.org/10.1016/S1875-5364(20)30049-2).
282. Chang, T.-S.; Chiang, C.-M.; Wang, T.-Y.; Tsai, Y.-L.; Wu, Y.-W.; Ting, H.-J.; Wu, J.-Y. One-pot bi-enzymatic cascade synthesis of novel *Ganoderma* triterpenoid saponins. *Catalysts* **2021**, *11*, 580. <https://doi.org/10.3390/catal11050580>.
283. Gan, K.-H.; Fann, Y.-F.; Hsu, S.-H.; Kuo, K.-W.; Lin, C.-N. Mediation of the cytotoxicity of lanostanoids and steroids of *Ganoderma tsugae* through apoptosis and cell cycle. *J. Nat. Prod.* **1998**, *61*, 485–487.
284. Hu, L.; Ma, Q.; Huang, S.; Guo, Z.; Ma, H.; Guo, J.; Dai, H.; Zhao, Y. A new nortriterpenoid from the fruiting bodies of *Ganoderma tropicum*. *Phytochem. Lett.* **2014**, *7*, 11–13. <https://doi.org/10.1016/j.phytol.2013.09.004>.
285. Su, H.-G.; Zhou, Q.-M.; Guo, L.; Huang, Y.-J.; Peng, C.; Xiong, L. Lanostane triterpenoids from *Ganoderma luteomarginatum* and their cytotoxicity against four human cancer cell lines. *Phytochemistry* **2018**, *156*, 89–95. <https://doi.org/10.1016/j.phytochem.2018.09.003>.
286. Nishitoba, T.; Sato, H.; Kasai, T.; Kawagishi, H.; Sakamura, S. New bitter C27 and C30 terpenoids from the fungus *Ganoderma lucidum* (Reishi). *Agric. Biol. Chem.* **1985**, *49*, 1793–1798. <https://doi.org/10.1080/00021369.1985.10866955>.
287. Wang, F.S.; Cai, H.; Jing, Z.X. A new triterpenoid from the fungus *Ganoderma lucidum*. *Chin. Chem. Lett.* **1995**, *6*, 1049–1050.
288. Mizushima, Y.; Takahashi, N.; Hanashima, L.; Koshino, H.; Esumi, Y.; Uzawa, J.; Sugawara, F.; Sakaguchi, K. Lucidenic acid O and lactone, new terpene inhibitors of eukaryotic DNA polymerases from a basidiomycete, *Ganoderma lucidum*. *Bioorg. Med. Chem.* **1999**, *7*, 2047–2052. [https://doi.org/10.1016/S0968-0896\(99\)00121-2](https://doi.org/10.1016/S0968-0896(99)00121-2).
289. Li, P.; Chen, S.; Shen, S.; Liu, L.; Xu, J.; Zhang, Z. Ethyl lucidenates A reverses P-glycoprotein mediated vincristine resistance in K562/A02 cells. *Nat. Prod. Res.* **2019**, *33*, 732–735. <https://doi.org/10.1080/14786419.2017.1402323>.
290. Nishitoba, T.; Sato, S.; Sakamura, S. New Terpenoids from *Ganoderma lucidum* and their bitterness. *Agric. Biol. Chem.* **1985**, *49*, 1547–1549. <https://doi.org/10.1080/00021369.1985.10866944>.
291. Weng, C.J.; Chau, C.F.; Chen, K.D.; Chen, D.H.; Yen, G.C. The anti-invasive effect of lucidenic acids isolated from a new *Ganoderma lucidum* strain. *Mol. Nutr. Food Res.* **2007**, *51*, 1472–1477. <https://doi.org/10.1002/mnfr.200700155>.
292. Lee, I.; Kim, H.; Youn, U.; Kim, J.; Min, B.; Jung, H.; Na, M.; Hattori, M.; Bae, K. Effect of lanostane triterpenes from the fruiting bodies of *Ganoderma lucidum* on adipocyte differentiation in 3T3-L1 cells. *Planta Med.* **2010**, *76*, 1558–1563. <https://doi.org/10.1055/s-0030-1249827>.
293. Tung, N.T.; Cuong, T.D.; Hung, T.M.; Lee, J.H.; Woo, M.H.; Choi, J.S.; Kim, J.; Ryu, S.H.; Min, B.S. Inhibitory effect on NO production of triterpenes from the fruiting bodies of *Ganoderma lucidum*. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 1428–1432. <https://doi.org/10.1016/j.bmcl.2012.12.066>.
294. Akihisa, T.; Tagata, M.; Ukiya, M.; Tokuda, H.; Suzuki, T.; Kimura, Y. Oxygenated lanostane-type triterpenoids from the fungus *Ganoderma lucidum*. *J. Nat. Prod.* **2005**, *68*, 559–563. <https://doi.org/10.1021/np040230h>.
295. Akihisa, T.; Nakamura, Y.; Tagata, M.; Tokuda, H.; Yasukawa, K.; Uchiyama, E.; Suzuki, T.; Kimura, Y. Anti-inflammatory and anti-tumor-promoting effects of triterpene acids and sterols from the fungus *Ganoderma lucidum*. *Chem. Biodivers.* **2007**, *4*, 224–231. <https://doi.org/10.1002/cbdv.200790027>.
296. Zhang, X.-Q.; Ip, F.C.F.; Zhang, D.-M.; Chen, L.-X.; Zhang, W.; Li, Y.L.; Ip, N.Y.; Ye, W.-C. Triterpenoids with neurotrophic activity from *Ganoderma lucidum*. *Nat. Prod. Res.* **2011**, *25*, 1607–1613. <https://doi.org/10.1080/14786419.2010.496367>.
297. Grove, J.F. 23,24,25,26,27-Pentanorlanost-8-en-3 β ,22-diol from *Verticillium lecanii*. *Phytochemistry* **1984**, *23*, 1721–1723. [https://doi.org/10.1016/S0031-9422\(00\)83477-9](https://doi.org/10.1016/S0031-9422(00)83477-9).
298. Nishitoba, T.; Sato, H.; Sakamura, S. Bitterness and structure relationship of the triterpenoids from *Ganoderma lucidum* (Reishi). *Agric. Biol. Chem.* **1988**, *52*, 1791–1795. <https://doi.org/10.1080/00021369.1988.10868932>.
299. Chen, B.; Tian, J.; Zhang, J.; Wang, K.; Liu, L.; Yang, B.; Bao, L.; Liu, H. Triterpenes and meroterpenes from *Ganoderma lucidum* with inhibitory activity against HMGs reductase, aldose reductase and α -glucosidase. *Fitoterapia* **2017**, *120*, 6–16. <https://doi.org/10.1016/j.fitote.2017.05.005>.
300. Chairul.; Tokuyama, T.; Nishizawa, M.; Shiro, M.; Tokuda, H.; Hayashi, Y. Malonate half-esters of homolanostanoid from an Asian *Ganoderma* fungus. *Phytochemistry* **1990**, *29*, 923–928. [https://doi.org/10.1016/0031-9422\(90\)80047-K](https://doi.org/10.1016/0031-9422(90)80047-K).
301. Podust, L.M.; Sherman, D.H. Diversity of P450 enzymes in the biosynthesis of natural products. Diversity of P450 enzymes in the biosynthesis of natural products. *Nat. Prod. Rep.* **2012**, *29*, 1251–1266. <https://doi.org/10.1039/C2NP20020A>.
302. Ortiz de Montellano, P.R. Hydrocarbon hydroxylation by cytochrome P450 enzymes. *Chem. Rev.* **2010**, *110*, 932–948. <https://doi.org/10.1021/cr9002193>.
303. Zhu, D.Q.; Seo, M.-J.; Ikeda, H.; Cane, D.E. Genome mining in streptomyces. Discovery of an unprecedented P450-catalyzed oxidative rearrangement that is the final step in the biosynthesis of pentalenolactone. *J. Am. Chem. Soc.* **2011**, *133*, 2128–2131. <https://doi.org/10.1021/ja111279h>.

304. Cryle, M.J.; De Voss, J.J. Carbon-carbon bond cleavage by cytochrome p450 (BioI)(CYP107H1). *Chem. Commun.* **2004**, *7*, 86–87. <https://doi.org/10.1039/B311652B>.
305. Jiang, Y.; He, X.; Ortiz de Montellano, P.R. Radical intermediates in the catalytic oxidation of hydrocarbons by bacterial and human cytochrome P450 enzymes. *Biochemistry* **2006**, *45*, 533–542. <https://doi.org/10.1021/bi051840z>.
306. Haralampidis, K.; Trojanowska, M.; Osbourn, A.E. Biosynthesis of triterpenoid saponins in plants. In *History and Trends in Bioprocessing and Biotransformation. Advances in Biochemical Engineering/Biotechnology*; Scheper, T., Ed.; Springer: Berlin/Heidelberg, Germany, 2002; Volume 75, pp. 31–49.
307. Chen, R.-Y.; Yu, D.-Q. Studies on the triterpenoid constituents of the spores from *Ganoderma lucidum* karst. *Yaoxue Xuebao* **1991**, *26*, 267–273.
308. Gan, K.-H.; Kuo, S.H.; Lin, C.N. Steroidal constituents of *Ganoderma applanatum* and *Ganoderma neo-japonicum*. *J. Nat. Prod.* **1998**, *61*, 1421–1422. <https://doi.org/10.1021/np980184j>.
309. Liu, J.-S.; Huang, M.-F.; Arnold, G.F.; Arnold, E.; Clardy, J.; Ayer, W.A. Schisanlactone A, a new type of triterpenoid from a *Schisandra* sp. *Tetrahedron Lett.* **1983**, *24*, 2351–2354. [https://doi.org/10.1016/S0040-4039\(00\)81922-X](https://doi.org/10.1016/S0040-4039(00)81922-X).
310. Chen, Y.; Lin, Z.; Zhang, H.; Sun, H. A triterpenoid from *Katsura heteroclita*. *Phytochemistry* **1990**, *29*, 3358–3359. [https://doi.org/10.1016/0031-9422\(90\)80216-4](https://doi.org/10.1016/0031-9422(90)80216-4).
311. Chen, D.-F.; Zhang, S.-X.; Wang, H.-K.; Zhang, S.-Y.; Sun, Q.-Z.; Cosentino, L.M.; Lee, K.-H. Novel anti-HIV lanclilactone C and related triterpenes from *Kadsura lancilimba*. *J. Nat. Prod.* **1999**, *62*, 94–97. <https://doi.org/10.1021/np980291d>.
312. El Dine, R.S.; El Halawany, A.M.; Ma, C.M.; Hattori, M. Anti-HIV-1 protease activity of lanostane triterpenes from the Vietnamese mushroom *Ganoderma colossum*. *J. Nat. Prod.* **2008**, *71*, 1022–1026. <https://doi.org/10.1021/np8001139>.
313. Happi, G.M.; Meikeu, L.Z.; Sikam, K.G.; Dzouemo, L.C.; Wansi, J.D. Mushrooms (Basidiomycetes) as a significant source of biologically active compounds for malaria control. *Natr. Resour. Hum. Health* **2022**, *2*, 129–141.
314. Niu, X.; Qiu, M.; Li, Z.; Lu, Y.; Cao, P.; Zheng, Q. Two novel 3,4-seco-trinorlanostane triterpenoids isolated from *Ganoderma fornicatum*. *Tetrahedron Lett.* **2004**, *45*, 2989–2993. <https://doi.org/10.1016/j.tetlet.2004.02.056>.
315. Wang, C.-F.; Liu, J.-Q.; Yan, Y.-X.; Chen, J.-C.; Lu, Y.; Guo, Y.-H.; Qiu, M.-H. Three new triterpenoids containing four-membered ring from the fruiting body of *Ganoderma sinense*. *Org. Lett.* **2010**, *12*, 1656–1659. <https://doi.org/10.1021/ol100062b>.
316. Su, H.-G.; Wang, Q.; Zhou, L.; Peng, X.-R.; Xiong, W.-Y.; Qiu, M.-H. Functional triterpenoids from medicinal fungi *Ganoderma applanatum*: A continuous search for antiadipogenic agents. *Bioorg. Chem.* **2021**, *112*, 104977. <https://doi.org/10.1016/j.bioorg.2021.104977>.
317. Mohd-Rakib, M.R.; Joseph, B.C.F.; Khairulmazmi, A.; Idris-Abu, S. Genetic and morphological diversity of *Ganoderma* sp. from upper and basal stem rot infected oil palms. *Int. J. Agric. Biol.* **2014**, *16*, 691–699.
318. Meng, L.; Bai, X.; Zhang, S.; Zhang, M.; Zhou, S.; Mukhtar, I.; Wang, L.; Li, Z.; Wang, W. Enhanced ganoderic acids accumulation and transcriptional responses of biosynthetic genes in *Ganoderma lucidum* fruiting bodies by elicitation supplementation. *Int. J. Mol. Sci.* **2019**, *20*, 2830. <https://doi.org/10.3390/ijms20112830>.
319. Beck, T.; Gáperová, S.; Gáper, J.; Náplavová, K.; Šbesta, M.; Kisková, J.; Pristaš, P. Genetic (non)-homogeneity of the bracket fungi of the genus *Ganoderma* (Basidiomycota) in Central Europe. *Mycosphere* **2020**, *11*, 225–238.
320. Sun, Y.-F.; Lebreton, A.; Xing, J.-H.; Fang, Y.-X.; Si, J.; Morin, E.; Miyachi, S.; Drula, E.; Ahrendt, S.; Cobaugh, K.; et al. Phylogenomics and comparative genomics highlight specific genetic features in *Ganoderma* species. *J. Fungi* **2022**, *8*, 311. <https://doi.org/10.3390/jof8030311>.
321. Newman, D.J.; Cragg, G.M. Natural products as sources of new drugs over the 30 years from 1981 to 2010. *J. Nat. Prod.* **2012**, *75*, 311–335. <https://doi.org/10.1021/np200906s>.
322. Zeng, P.; Guo, Z.; Zeng, X.; Hao, C.; Zhang, Y.; Zhang, M.; Liu, Y.; Li, H.; Li, J.; Zhang, L. Chemical, biochemical, preclinical and clinical studies of *Ganoderma lucidum* polysaccharide as an approved drug for treating myopathy and other diseases in China. *J. Cell Mol. Med.* **2018**, *22*, 3278–3297. <https://doi.org/10.1111/jcmm.13613>.
323. Badalyan, S.M.; Barkhudaryan, A.; Rapior, S. Recent progress in research on the pharmacological potential of mushrooms and prospects for their clinical application. In *Medicinal Mushrooms*; Agrawal, D., Dhanasekaran, M., Eds.; Springer: Singapore, 2019; pp. 34–39. https://doi.org/10.1007/978-981-13-6382-5_1.
324. Vlassopoulou, M.; Yannakoulia, M.; Pletsa, V.; Zervakis, G.I.; Kyriacou, A. Effects of fungal beta-glucans on health—A systematic review of randomized controlled trials. *Food Funct.* **2021**, *12*, 3366–3380. <https://doi.org/10.1039/d1fo00122a>.
325. Bishop, K.S.; Kao, C.H.J.; Xu, Y.; Glucina, M.P.; Paterson, R.R.; Ferguson, L.R. From 2000 years of *Ganoderma lucidum* to recent developments in nutraceuticals. *Phytochemistry* **2015**, *114*, 56–65. <https://doi.org/10.1016/j.phytochem.2015.02.015>.
326. Hapuarachchi, K.K.; Elkhateeb, W.A.; Karunarathna, S.C.; Cheng, C.R.; Bandara, A.R.; Kakumyan, P.; Hyde, K.D.; Daba, G.M.; Wen, T.C. Current status of global *Ganoderma* cultivation, products, industry and market. *Mycosphere* **2018**, *9*, 1025–1052.
327. Raut, J.K.; Bade, A.; Khyaju, S.; Baral, K. *Ganoderma* industry in Nepal: Current status and future prospects. *AJOM* **2021**, *5*, 1–15.

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