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Settleable atmospheric particulate matter induces stress and affects the oxygen-carrying capacity and innate immunity in Nile tilapia (*Oreochromis niloticus*)

Michelly Pereira Soares, Carolina Fernandes de Angelis, Israel Luz Cardoso, David J Mckenzie, Iara da Costa Souza, Daniel Wunderlin, Magdalena Monferrán, Marisa Narciso Fernandes, Cléo Alcantara Costa Leite

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1 **Sedimentable atmospheric particulate matter induces stress and affects the**
2 **oxygen-carrying capacity and innate immunity in Nile tilapia (*Oreochromis***
3 ***niloticus*)**

4

5 Michelly Pereira Soares¹, Carolina Fernandes De Angelis¹, Israel Luz Cardoso¹, David
6 J. McKenzie², Iara C Souza¹, Daniel A. Wunderlin³, Magdalena V. Monferrán³, Marisa
7 Narciso Fernandes¹, Cleo Alcantara Costa Leite^{1*}

8

9 ¹Department of Physiological Sciences, Federal University of São Carlos, Rod
10 Washington Luis km 235,13565-905 São Carlos, SP, Brazil.

11 ²MARBEC, Université de Montpellier, CNRS, Ifremer, IRD, Montpellier 34095, France.

12 ³ICYTAC: Instituto de Ciencia y Tecnología de Alimentos Córdoba, CONICET and
13 Departamento de Química Orgánica, Facultad de Ciencias Químicas, Universidad
14 Nacional de Córdoba, Bv. Medina Allende s/n, Ciudad Universitaria, 5000- Córdoba,
15 Argentina.

16

17 *Corresponding author: cleo.leite@ufscar.br

18 Email addresses of co-authors:

19 Michelly Pereira Soares: michelly_psoares@hotmail.com

20 Carolina De Angelis: caroldeangelis3@hotmail.com

21 Israel Luz Cardoso: israelcardoso2008@hotmail.com

22 David J McKenzie: david.mckenzie@cnr.fr

23 Daniel A. Wunderlin: danielwunderlin@gmail.com

24 Magdalena V. Monferrán: mmonferran@unc.edu.ar

25 Marisa Narciso Fernandes: dmnf@ufscar.br

Abbreviations: APM - atmospheric particulate matter, SeAPM - sedimentable atmospheric particulate matter

26 **Abstract**

27 Steel industry emissions of atmospheric particulate matter (APM) are responsible for
28 air to water cross contamination, which deposits metal/metalloid contaminants in
29 aquatic ecosystems. This source of contamination is not considered in most
30 environmental monitoring protocols being largely neglected. Using sedimentable
31 atmospheric particulate matter (SeAPM) collected in an area with steel industry
32 influence, we analyzed the sublethal effects on the hematological and innate
33 immunological variables in a teleost fish, Nile tilapia *Oreochromis niloticus* after short-
34 term exposure (96 h). After exposure to raw SeAPM ($1 \text{ g}\cdot\text{L}^{-1}$), blood samples were
35 analyzed to evaluate functional indices related to oxygen transport capacity, immune
36 activity and stress. SeAPM exposure reduced blood oxygen carrying capacity by
37 lessening hematocrit, hemoglobin, erythrocyte, and mean corpuscular hemoglobin
38 concentration. Compensatory increments in mean corpuscular volume and mean
39 corpuscular hemoglobin were observed. SeAPM exposure also impacted elements of
40 immune activity by reducing the number of leukocytes, thrombocytes and monocytes;
41 total plasma protein, and leukocyte respiratory activity, and by increasing lysozyme
42 concentrations. Furthermore, SeAPM elicited endocrine stress response, raising
43 plasma cortisol and glucose. The alterations caused by acute exposure to raw SeAPM
44 threatened the capacity to sustain aerobic metabolism and to respond to pathogens
45 which may reduce fitness of fish populations. These results highlight the need to
46 develop proper protocols for monitoring air-water cross-contamination and, also, for
47 further ecotoxicological research to evaluate the dangers of exposure to sublethal
48 SeAPM contamination by steel industries.

49

50 **Keywords:** APM; Steel industry; Metals/Metalloids; nanoparticles; environmental
51 risks; physiological responses.

52

53 **1. Introduction**

54 The steel industry is a significant source of atmospheric particulate matter, a
55 complex mixture of particles (Callén et al., 2009; Park and Kim, 2005; Tsai and Cheng,
56 2004) which may vary in composition, shape and size, including metallic nanoparticles
57 and emergent metallic compounds (Salgado, 2003). APM emission has been
58 associated with adverse health effects for humans, notably chronic respiratory
59 diseases, cardiac problems, lung cancer, and other serious health issues (Costa and
60 Dreher, 1997; Magas et al., 2007; Prieditis and Adamson, 2002; Wild et al., 2009).

61 In Brazil, sublethal APM contamination has been reported in continental and
62 estuarine waters (Souza et al., 2018b), which could result in important ecological
63 imbalances, reduced fish genetic diversity and even cause extirpation of specific fish
64 population (Bourret et al., 2008; Durrant et al., 2011). Bioaccumulation and
65 biomagnification of APM has been reported in aquatic food chains (Idrus et al., 2018;
66 Souza et al., 2021b and c). However, the worldwide regulatory agencies does not,
67 consider as a rule the air to water cross-contamination in existing environmental
68 monitoring protocols (CONAMA 491/2018 - BRASIL, 2018, Environmental Protection
69 Agency – EPA, European Union and Canada), being case to case studied separately.

70 Metallic compounds have a number of toxicological impacts on fishes, causing
71 oxidative stress (Cuny et al., 2004; Farombi et al., 2007; Valavanidis et al., 2006);
72 hematological and biochemical disturbances, and immunological depression (Bakshi
73 and Panigrahi, 2018; Hedayati et al., 2016; Hedayati and Darabitabar, 2017; Javed
74 and Usmani, 2015; Khabbazi et al., 2015; Lal Shah, 2010; Lavanya et al., 2011). The
75 susceptibility of fishes to metals and metaloids makes them especially vulnerable to
76 this component of anthropogenic contamination (Saurabh and Sahoo, 2008). Fishes
77 provide essential ecosystem services and are important resources, therefore they are

78 important indicator organisms to assess and monitor the presence of water
79 contaminants and their deleterious ecological consequences (Campos-Garcia et al.,
80 2015). It is especially valuable to investigate sublethal effects of contaminants, whose
81 presence and impact are still not well understood.

82 Hematological, biochemical and immunological biomarkers are used as
83 physiological early warning signals and are an essential tool to evaluate the general
84 health status of fish species. We used these markers to evaluate the effects of
85 exposure to a sublethal concentration of SeAPM collected from an area influenced
86 mainly by the steel industry, on a representative fish, the Nile tilapia, *Oreochromis*
87 *niloticus*. It's worldwide distribution and physiological knowledge base make them a
88 good experimental model for general ecotoxicological predictions (Solis et al., 2007).

89

90 **2. Material and Methods**

91

92 *2.1. Atmospheric particulate matter*

93 In order to fully represent the complexity of environmental APM contamination,
94 SeAPM was collected at Ilha do Boi (20°18'32"S, 40°16'33"O) in the city of Vitória,
95 state of Espírito Santo (ES), Brazil. The location has major influence of SeAPM emitted
96 by the steel industry from the Tubarão Complex, which is integrated by an iron pelleting
97 industry and an iron and steel exportation harbor (Santos et al, 2017). This SeAPM
98 has a complex composition, and most of it is composed by inorganic metallic
99 substances (Souza et al, 2021a, Santos et al, 2017, Machado et al, 2018 and Galvão
100 et al 2019). Studies have shown that there may be seasonal variation in Vitória
101 estuaries contamination (Souza et al., 2013). Therefore, for repeatability, SePM was
102 taken from a mixed sample of SeAPM derived from continuous collection over one

103 year, as described by Arrivabene and Souza (Arrivabene et al., 2015; Souza et al.,
104 2013). The exposure concentration of $1 \text{ g}\cdot\text{L}^{-1}$ refers to the SeAPM mass and it was
105 chosen because the environmental relevance, as it is based in the maximum SePM
106 recorded from the Institute of Environment and Hydric Resources (IEMA, 2021) and
107 according to previous pilot tests it also produces a level of aquatic metal contamination
108 similar to what has been reported for the estuarine area close to Vitoria city (Souza et
109 al., 2013). We have analyzed both nominal and real APM composition.

110

111 2.2. Fish

112 Nile tilapia, *Oreochromis niloticus*, were obtained from the Polettini fish farm in
113 Mogi Mirim, state of São Paulo, Brazil, and transported to the Department of
114 Physiological Sciences at the Federal University of São Carlos (UFSCar) in São Carlos
115 (SP). There, they were maintained in 1000L tanks under natural photoperiod (~12 h:12
116 h), supplied with biofiltered water at $25 \pm 1^\circ\text{C}$ and continuous aeration. They were
117 acclimated for 30 days prior to experimentation, and fed *ad libitum* with commercial
118 pellets (Supra® Acqua Nile tilapia feed: 5-8 mm; 12 % moisture; 32 % protein; 6 % fat).

119 The same water quality was provided for experimental exposures. The water
120 was continuously monitored (temperature, $25.3 \pm 0.91^\circ\text{C}$; dissolved oxygen, 7.1 ± 0.89
121 mg L^{-1} ; and pH, 6.3 ± 0.04), and stated suitable for experiment (Ibrahim and Nagggar,
122 2010). All procedures were approved by the UFSCar Committee of Ethics in Animal
123 Experimentation (CEUA-UFSCar protocol nº 8105110718).

124

125 2.3. SeAPM exposure and tissue sampling

126 After acclimation, 20 fishes (body mass, $M_B = 132.65 \pm 26.33\text{g}$) were exposed
127 for 96 h to either SePM (SeAPM group, $n = 10$) or maintained in clean water (control

128 group, n = 10). Contamination was performed by placing individual tilapia in glass
129 aquaria (200 L) respecting fish:water volume ratio (1:1, OECD 203, 2019) containing
130 SeAPM concentration of 1 g·L⁻¹. After 48 h, 50% of the water was replaced with fresh
131 water containing SeAPM at the appropriate dose. Control group was treated identically
132 but without SeAPM addition. After 96h exposure, fish were anesthetized with
133 benzocaine (100 mg L⁻¹) and blood was sampled for analysis.

134 Arterial blood samples were collected by caudal vessel puncture using syringes
135 bathed in anticoagulant (EDTA 3%) for the hematological analyses; in heparin for
136 respiratory burst activity of leukocytes; in GLISTAB (EDTA 6 g dL⁻¹ and KF 12 g dL⁻¹,
137 Labtest, Sao Paulo, Brazil; code 29) for plasma collection for analyses of stress
138 variables, and without anticoagulant to obtain serum for lysozyme analyses. With the
139 exception of lysozyme and cortisol, all analyzed were done just after blood collection.

140

141 2.4. SeAPM characterization *quantification*

142 The SeAPM was analyzed (Table1) according to standardized methods. We
143 used granulometry, muffle and solubilization with HNO₃/H₂O₂ in a microwave digester
144 to access particle size (Gee and Or, 2002), proportion of organic materials (Goldin,
145 1987; Suguio, 1973), and contents of inorganic material (Environmental Protection
146 Agency – EPA, 3052 - USEPA, 1996). These analyses were carried out at the Brazilian
147 Agricultural Research Corporation – EMBRAPA, Brazil and in the Food Science and
148 Technology Institute of Córdoba (YCITAC - Universidad Nacional de Cordoba,
149 Argentina).

150 Water from experimental aquaria was sampled (5 mL of water collected in the
151 middle of the water column) for quantitative and qualitative description of SeAPM
152 contamination (Table 2). Quantification was performed in a mass spectrometer with

153 inductively coupled plasma (Q-ICPMS, Agilent 7500 Series CX technology) equipped
154 with an ASX-100 autosampler (CETAC-technologies, Omaha, NE, USA). The analysis
155 was carried out at the Food Science and Technology Institute of Córdoba (YCITAC -
156 Universidad Nacional de Cordoba, Argentina).

157

158 *2.5. Stress variables*

159 Induction of a stress response, by exposure to SeAPM, was evaluated as
160 plasma cortisol and glucose titres. The blood collected in anticoagulant and anti-
161 glycolytic GLISTAB bathed syringes was centrifuged for plasma separation. Glucose
162 was analyzed with a commercial kit (Labtest, Sao Paulo, Brazil; code 84). An aliquot
163 of the plasma was stored at -80°C for cortisol analysis by immunoenzymatic assay
164 using a commercial kit (DRG International, Inc., USA; Cortisol ELISA -EIA - 1887).

165

166 *2.6. Hematological and immunological variables*

167 The hematological analyses aimed to evaluate effects of SeAPM exposure on
168 blood oxygen transport capacity and immune function. The hematocrit was read in
169 heparinized hematocrit tubes after centrifuging at 11200 G-force for 15min.
170 Hemoglobin concentration was analyzed with a commercial kit (Latest, Sao Paulo,
171 Brazil; code 43). Erythrocytes were counted in a Neubauer chamber after blood dilution
172 in formaldehyde citrate solution (1: 200). The mean corpuscular volume [MCV = (Ht x
173 10)/number of erythrocyte ($\times 10^6 \mu\text{L}^{-1}$) = fL], mean corpuscular hemoglobin [MCH = (Hb
174 x 10)/erythrocyte number ($\times 10^6 \mu\text{L}^{-1}$) = pg] and mean corpuscular hemoglobin
175 concentration [MCHC = (Hb x 100) Ht = g dL^{-1}] were calculated; and also, refractometry
176 was used to obtain total plasma protein (TPP).

177 Total leukocyte, total thrombocyte and differential leukocyte counts were
178 performed on blood smears stained with May-Grunwald-Giemsa-Wright, according to
179 (Paiva et al., 2013). Differential and total cell counts were performed under a
180 microscope using an oil immersion objective (100x). An aliquot of the blood was
181 heparinized and used to analyze leukocyte respiratory activity (RAL) by a colorimetric
182 assay based on the reduction of nitroblue tetrazolium (NBT) dye inside the phagocyte
183 according (Biller-Takahashi et al., 2013). The optical density of the solution was
184 measured by spectrophotometry (Molecular devices SpectraMax[®] M5) at 540nm. An
185 aliquot of the serum was stored at -80°C to analyze serum lysozyme concentrations
186 (SLC) by a turbidimetric assay, as described by (Demers and Bayne, 1997), with
187 modifications (Zanuzzo et al., 2014).

188

189 *2.7. Statistical analyses*

190 Analyses were performed with Rstudio (version 3.2.2). Data normality and
191 homogeneity were analyzed using SaphiroWilk and Levene's tests, respectively. Data
192 were normally distributed and therefore unpaired T-test was used to compare the
193 respectively and control groups. Data are expressed as mean values \pm standard
194 deviation (SD) and statistical significance was attributed at $P < 0.05$.

195

196 **3. Results**

197 There was 100% of survival following exposures, confirming that the SeAPM
198 dose was sublethal. Metal composition of SeAPM and the consequent concentrations
199 in contaminated water were dominated by iron ores and aluminium, with the emerging
200 elements titanium and cerium to lesser degrees (Table 1 and 2).

201

202
203

Table 1. Particle size analysis and major compounds present in settable atmospheric particulate matter (SeAPM).

Metals (µg/g)		Particle size (%)	
P	<LOD	1000 µm	0.81
Ca	19048.08	500 µm	14.17
Mg	7907.11	250 µm	17.56
Na	148.70	106 µm	25.91
K	844.82	53 µm	11.80
B	20.03	<53µm	29.75
Al	5522.80		
Ti	1108.94		
V	28.22		
Cr	34.53	Compounds (%)	
Mn	554.99	Organic matter	20.14
Fe-56	69199.04	Inorganic matter	79.86 %
Ni	14.74		
Cu	84.79		
Zn	274.86		
As	2.93		
Se	2.25		
Rb	4.92		
Sr	56.63		
Y	17.40		
Zr	43.79		
Nb	2.56		
Ag	0.10		
Cd	1.47		
Sn	15.22		
Ba	1.33		
La	29.90		
Ce	65.49		
Ta	0.00		
W	1.63		
Hg 201	0.32		
Hg 202	0.35		
Pb	50.86		
Bi	1.08		

LOD: limit of detection.

204

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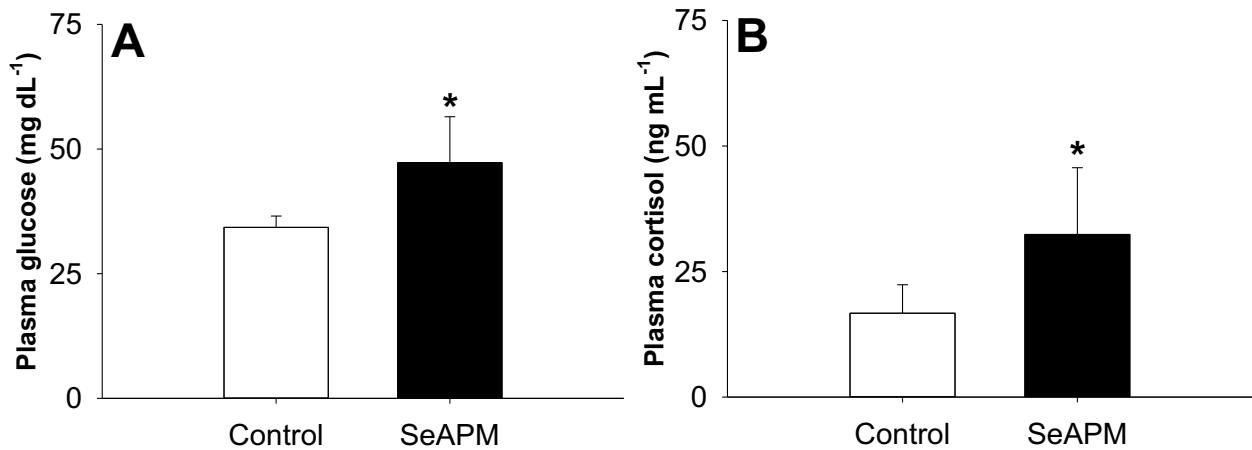
Table 2. Metal concentration (ug L⁻¹) in the experiment water.

Metal	Control	SeAPM
B	<LOD	<LOD
Al	69.73±0.25	553.99±3.23
Ti	<LOD	96.39±1.77
V	1.08±0.008	3.62±0.03
Cr	<LOD	1.45±0.01
Mn	<LOD	37.58±0.17
Fe-56	<LOD	511.05±5.82
Fe-57	<LOD	545.30±2.66
Ni	<LOD	<LOD
Cu	4.97±0.05	15.39±0.17
Zn	9.82±0.22	29.32±0.18
As	<LOD	<LOD
Se	<LOD	<LOD
Rb	10.36±0.04	10.04±0.001
Sr	22.65±0.09	26.85±0.11
Y	<LOD	1.17±0.016
Zr	<LOD	1.84±0.065
Nb	<LOD	<LOD
Ag	0.33±0.004	0.80±0.009
Cd	<LOD	<LOD
Sn	<LOD	<LOD
Ba	0.58±0.001	0.29±0.002
La	<LOD	1.31±0.03
Ce	<LOD	2.93±0.03
Ta	<LOD	<LOD
W	<LOD	<LOD
Hg-201	<LOD	<LOD
H-202	<LOD	<LOD
Pb	0.89±0.001	3.23±0.04
Bi	<LOD	<LOD

208
209
210

Values are mean ± SEM. LOD: limit of detection. Atmospheric particulate matter (APM)

211 Exposure to SeAPM triggered a clear stress response, with an almost doubling
212 of plasma cortisol (94%) associated with an increase in plasma glucose (38%) (Fig 1).



213
 214 **Figure 1. Plasma concentration of glucose (A) and cortisol (B) in Nile tilapia,**
 215 ***Oreochromis niloticus*, exposed to settleable atmospheric particulate matter**
 216 **(SeAPM) for 96 h. * indicates significant difference from the controls ($p < 0.05$).**
 217

218

219 The SeAPM reduced hematocrit (-12%), hemoglobin (-26%) and red blood cell
 220 number (-38%) in the tilapia. The MCHC was also reduced (-17%), whereas MCV
 221 (47%) and MCH (32%) increased (Table 3).

222

223 **Table 3. Hematologic variables in Nile tilapia, *Oreochromis niloticus*, exposed to**
 224 **settleable atmospheric particulate matter (SeAPM) for 96 h: hematocrit;**
 225 **hemoglobin; erythrocyte; mean corpuscular volume (MCV); mean corpuscular**
 226 **hemoglobin concentration (MCHC); mean corpuscular hemoglobin (MCH); total**
 227 **and differential leukocytes counts and thrombocytes.**

Parameters	Control	SeAPM
Blood Cells		
Hematocrit (%)	30.77 ± 1.68	27.04 ± 3.13*
Hemoglobin (g dL ⁻¹)	11.27 ± 1.72	8.32 ± 1.82*
Erythrocytes (×10 ⁶ μL ⁻¹)	2.39 ± 0.23	1.48 ± 0.35*
MCV (fL)	130.08 ± 15.78	190.94 ± 39.28*
MCHC (g dL ⁻¹)	36.87 ± 7.11	30.70 ± 5.32*
MCH (pg)	47.27 ± 6.43	62.18 ± 14.19*
Leukocytes (μL ⁻¹)	69.96 ± 23.07	39.87 ± 14.35*
Thrombocytes (μL ⁻¹)	86.81 ± 25.31	30.12 ± 17.23*

Differential leukocytes

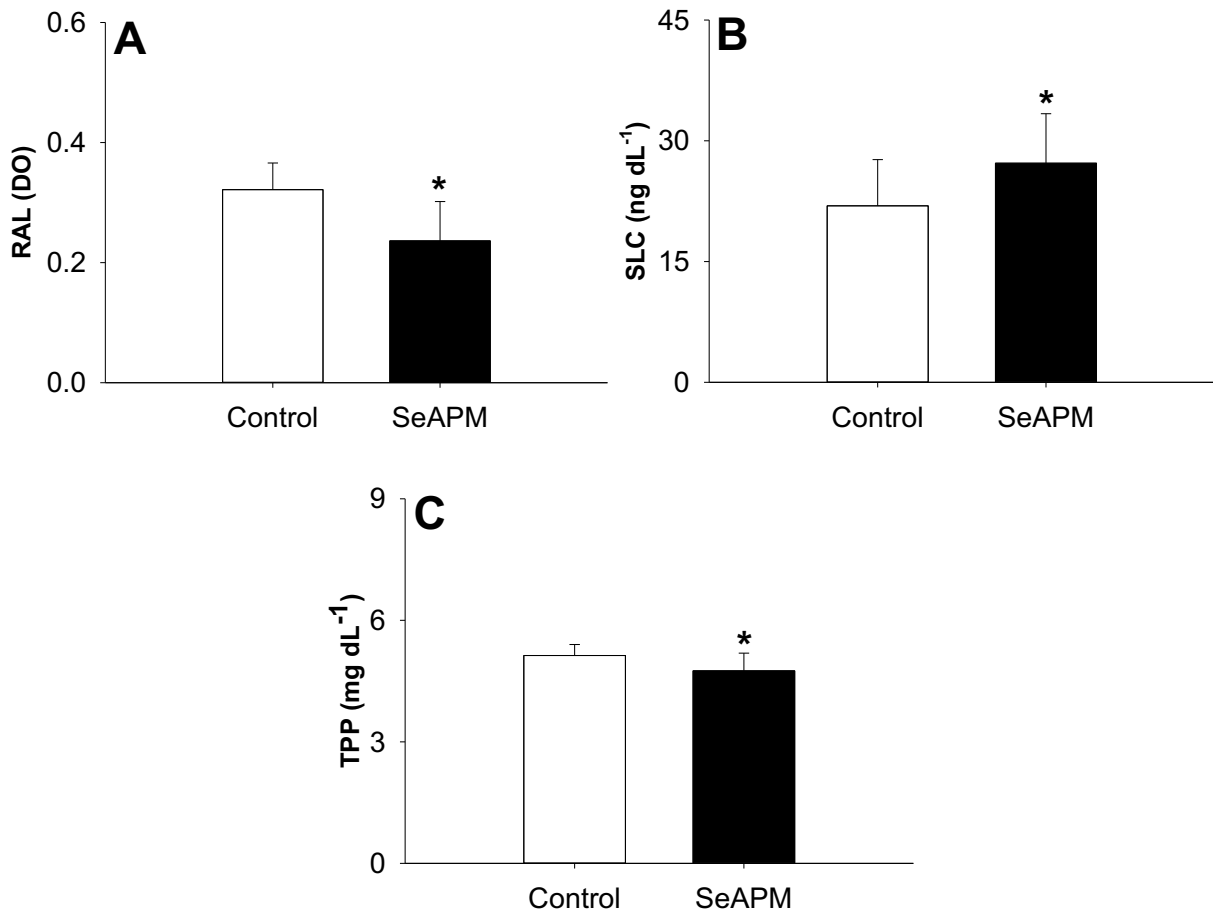
Neutrophils (%)	46.86 ± 18.07	36.92 ± 4.95a
Monocytes (%)	23.42 ± 7.87	15.38 ± 6.02*
Lymphocytes (%)	15.46 ± 5.64	30.83 ± 8.07*

228 Values are mean ± SEM. * indicates significant difference from the controls ($p < 0.05$).
229

230 Immune cell were also affected, fish exposed to SeAPM had lower number of
231 leukocytes (-43%) and thrombocytes (-65%). Considering differential leukocytes, the
232 monocytes were reduced (-34%); in contrast, the lymphocytes increased by almost
233 double (99%) and the neutrophil percentage were similar in SeAPM and control groups
234 (Table 3).

235 After exposure to SeAPM, the tilapia also exhibited significant decrease in TPP
236 (-7%) and RAL (-27%) however, SLC increased (24%) (Fig 2).

237



238
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240

241
242 **Figure 2. Respiratory activity of leukocytes - RAL (A), serum lysozyme**
243 **concentrations - SLC (B) and total plasma protein - TPP (C) in Nile tilapia,**
244 ***Oreochromis niloticus*, exposed to settleable atmospheric particulate matter**
245 **(SeAPM) for 96 h. * indicates significant difference from the controls ($p < 0.05$).**
246

247 **4. Discussion**

248 Exposure to SeAPM caused significant alterations to hematological variables
249 related to oxygen carrying capacity, immune activity and stress, indicating major
250 impacts on fish physiology under routine conditions. Although the SeAPM is sublethal,
251 it involved sensitive physiological systems crucial for maintaining the homeostasis and
252 performance of fish in their natural environment. Unless these short exposure impacts
253 can be properly compensated for, during long term contamination, they could have
254 significant ecological consequences, threatening species survival and fitness.

255 All measured variables were strongly affected by the short-term contamination
256 with SeAPM. Hence, their combined response, including studies of the effect of
257 different levels of contamination, could become useful for monitoring SeAPM presence
258 and toxicity levels in the environment as fish provide potentially valuable tool to
259 evaluate the existence and extent of environmental impacts. There is a need for further
260 dedicated studies, to understand the progression of the contamination and associated
261 damage to fauna. Such research could then support the design of environmental
262 monitoring protocols and risk analyses.

263

264 *SeAPM concentration*

265 SeAPM produced by the industries in the Tubarão Complex (Souza et al., 2018),
266 settles and accumulates over surfaces throughout the city of Vitoria. The SePM
267 comprises a complex mixture of different sized particles that vary in shape and
268 composition. It includes a diversity of organic and inorganic substances (Santos et al,

269 2017, Machado et al, 2018 and Galvão et al 2019). The inorganic components include
270 a wide diversity of metals (Arrivabene et al., 2015), and the amount of SeAPM used is
271 coherent with metal concentrations observed in local estuarine waters (Souza et al.,
272 2013, 2021b and c). The contamination was dominated by Fe and Al, their nominal
273 concentrations were in the range reported for Santa Cruz and Vitoria Bay (Fe, $355 \pm$
274 6.0 and $511 \pm 5.8 \text{ mg}\cdot\text{L}^{-1}$ and Al, 138.9 ± 2.8 and $553.9 \pm 3.2 \text{ mg}\cdot\text{L}^{-1}$, respectively)
275 (Souza et al. 2013). These concentrations are lower than the LC50 reported for
276 freshwater fish (Fe, $1.46\text{-}1.71 \text{ m L}^{-1}$ and Al, $0.095\text{-}235 \text{ mg L}^{-1}$) (Kennedy, 2011). The
277 values were also below to those established as the maximum limits of the Brazilian
278 regulatory agency CONAMA 357/2005 (Fe, $0.3\text{-}5.0 \text{ mg L}^{-1}$ and Al, $0.1\text{-}0.2 \text{ mg L}^{-1}$) for
279 freshwater bodies of classes 1 to 3, referred as protected for aquatic communities,
280 aquaculture, fishing, and supply.

281

282 *Effects of SeAPM on fish physiology*

283 Several different biomarkers have been identified as sensitive indicators of toxic
284 effects in target organisms. They indicate deviation from normal status, function
285 impairment and/or pathological symptomatology (McCarthy and Shugart, 1990).
286 Therefore, they can be important for monitoring the quality of aquatic ecosystems. In
287 the current study, the control data are in accordance with the reference values for these
288 biomarkers in farmed *O. niloticus* (Tavares-Dias and Mariano, 2015) validating the fish
289 group used and the protocol.

290 Based on the observed reductions in erythrocyte count, hemoglobin content and
291 hematocrit, SeAPM exposure has a fast and significant effect on erythrocytes.
292 Erythrocyte destruction decreases hemoglobin content and hematocrit values. Metals
293 and nanoparticles also reduced RBC, hemoglobin content and hematocrit in Nile tilapia

294 (Abdel-Khalek et al., 2016). The authors suggested that such effects could be due to
295 a series of factors: hemolysis of erythrocytes in the blood vessels of the liver and
296 kidneys; rupture of hematopoietic tissues; osmoregulatory dysfunction that increases
297 the erythrocyte destruction rate in hematopoietic organs, or even damage from
298 perfusion through injured gill tissue.

299 The high MCV and MCH, and reduced MCHC, reflected erythrocyte swelling in
300 tilapia exposed to SeAPM, which is related to intracellular osmotic disorders
301 (macrocytic anemia). In studies evaluating the effect of metals on fishes, enlarged red
302 blood cells have been associated with hypoxia that can be caused by gill damage,
303 leading to compromised gas exchange (Abdel-Khalek et al., 2016). The increased
304 MCV with concomitant decreased in RBC, Hb concentrations and MCHC indicated
305 cellular swelling as a mechanism to compensate oxygen transport (Carvalho and
306 Fernandes, 2006). Increases in MCV and MCH and decreases in MCHC levels have
307 also been reported in fish exposed to industrial effluents (Javed et al., 2016; Zutshi et
308 al., 2010). In addition, the changes regarding MCH and MCHC can be attributed to red
309 blood cell hemolysis and a reduction in red blood cell production in hemopoietic
310 tissues. Low MCHC indicates hypochromic anemia, which may relate to malfunction in
311 DNA synthesis (Hoffman R, Benz EJ, Furie B, 2009; Wang et al., 2009). These results
312 are in agreement with general effects of metal contamination reported in various
313 studies on fishes (Cu in *Cyprinus carpio*, *Channa punctatus* and *O. niloticus*; Cd in *C.*
314 *carpio* and *Scyliorhinus canicula*; Hg in *O. niloticus*, *Clarias gariepinus* and *C.*
315 *punctatus*; Pb in *O. niloticus*, *Anguilla anguilla* and *O. mossambicus*; Mn in *C.*
316 *punctatus*, *Tilapia sparmanii* and *O. mossambicus*; Zn in *C. carpio* and *O.*
317 *mossambicus*; Cr in *Saccobranhus fossilis*, *Labeo rohita* and *C. carpio*; Ni in *Colisa*

318 *fasciatus* and *C. gariepinus*) (apud Abdel-Khalek et al., 2016; Javed and Usmani, 2015;
319 Singh et al., 2008).

320 The reduction in oxygen carrying capacity lead to obvious limitations for
321 homeostasis impairing the necessary adjustment to deal with common environmental
322 challenges such as hypoxia, swimming, feeding and reproduction.
323 Therefore, despite sublethal, such damage can lead to imminent ecological problems
324 threaten a long-term population survival and/or exertion of its ecological role. Designed
325 protocols to experimentally investigate SeAPM damage on the resistance to hypoxia,
326 swimming capacity, reproductive process, and specific dynamic action (SDA) should
327 properly address such questions.

328 Stress indicators can reveal whether environmental changes are perceived as
329 harmful, which can influence the capacity of the organism to cope. Our data clearly
330 indicate that SeAPM triggered a stress response in tilapia. Plasma levels of cortisol
331 and glucose are the most widely used stress indicators in fishes (Robertson et al.,
332 1987; Sopinka et al., 2016), and their alteration has been reported after metal
333 contamination in other species, such as zebrafish, *Danio rerio* (Katuli et al., 2014);
334 Atlantic salmon, *Salmo salar* (Farmen et al., 2012); and rainbow trout, *Oncorhynchus*
335 *mykiss* (Wood et al., 1996). They are complementary to functional indices, for example
336 the erythrocyte counts, hemoglobin content and hematocrit measurement (Houston,
337 1997).

338 Furthermore, in our study, the decrease in white blood cell count, monocytes
339 PPT and RAL suggest an immunological suppression. It is well established that the
340 immune system of fishes can be severely affected by various stresses, such as metal
341 pollutants (Ghiasi et al., 2010; Jovanović and Palić, 2012; Witeska and Wakulska,
342 2007). It is possible that a significant decline in leukocyte count may affect RAL.

343 Besides, nanoparticles (especially metallic nanoparticles) can interact with free
344 proteins in plasma resulting in protein conformational changes and decreased activity
345 (Jovanović and Palić, 2012). Additionally, the higher number of lymphocytes may be
346 associated with defense mechanisms after tissue damage. As a relevant window to
347 environment, the gills are likely to be an affected organ reinforcing the relevance of the
348 previous described damage to oxygen cascade. Lymphocytes are particularly involved
349 in the production of antibodies and inflammatory processes (El-Sayed et al., 2007;
350 Sadauskas-Henrique et al., 2011) and consequently, stimulation of lymphopoiesis and
351 increased release of lymphocytes from lymphomyeloid tissue occurs under toxic stress
352 (Seriani et al., 2015). That alteration is directly related to SeAPM stress and reinforces
353 the reported immune umbalance. Similar tendence were observed in *Labeo rohita*,
354 *Achirus lineatus* and *Centropomus parallelus* exposed to metal contamination (Prado
355 et al., 2014; Seriani et al., 2013; Zutshi et al., 2010).

356 Furthermore, the SLC increase further supports the SeAPM impact on immune
357 activity. The level of lysozyme in fish serum is an important parameter to monitor the
358 potential impact of environmental alterations on innate immunity, it is the first line of
359 defense mechanisms (Saurabh and Sahoo, 2008). Exposure to heavy metals affects
360 lysozyme levels, and the nature in complex ways dependent on the stress degree,
361 duration and type (Saurabh and Sahoo, 2008; Yildiz, 2006). It has been suggested that
362 high levels of lysozyme after acute exposure to nanoparticles, such as the reported
363 SeAPM effect, can relate to an inflammatory response (Canesi et al., 2010; Gordon et
364 al., 1979; Jovanović and Palić, 2012; Mir, 1977). Despite that is hard to interpret, that
365 is a major alteration since fish protect themselves from pathogens mainly with the help
366 of innate or nonspecific immunity (Rauta et al., 2014). Thus, the SeAPM related

367 reduction in the innate immunological parameters decreases fish's ability to defend
368 itself against pathogens and infectious diseases.

369 Therefore, using a range of biomarkers, this study demonstrated that exposure
370 to sublethal concentrations of a real SeAPM mixture from the steel industry, causes
371 severe impairments to physiological and immunological performance in a
372 representative fish species. This has important implications for the damage that
373 SeAPM may cause to aquatic ecosystems and the fishery resources that they support.
374 The Nile tilapia provides an good indicator species to monitor levels and impacts of
375 SeAPM contamination in water bodies. After developing a database on this
376 contamination, wild fish monitoring could alert the threat level before the big bounces
377 of animal mortality. Due to the obvious damage, we caution the need for more in-depth
378 analyses involving different species under a range of contamination levels.

379

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