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Comprehensive spatial distribution OPEN of tropical fsh assemblages from multifrequency acoustics and video fulfls the island mass efect framework

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Tropical marine ecosystems are highly biodiverse and provide resources for small-scale fsheries and tourism. However, precise information on fsh spatial distribution is lacking, which limits our ability to reconcile exploitation and conservation. We combined acoustics to video observations to provide a comprehensive description of fsh distribution in a typical tropical environment, the Fernando de Noronha Archipelago (FNA) off Northeast Brazil. We identified and classified all acoustic **echoes into ten fsh assemblage and two triggerfsh species. This opened up the possibility to relate the diferent spatial patterns to a series of environmental factors and the level of protection. We provide the frst biomass estimation of the black triggerfsh** *Melichthys niger***, a key tropical player. By comparing the efects of euphotic and mesophotic reefs we show that more than the depth, the most important feature is the topography with the shelf-break as the most important hotspot. We also complete the portrait of the island mass efect revealing a clear spatial dissymmetry regarding fsh distribution. Indeed, while primary productivity is higher downstream, fsh concentrate upstream. The comprehensive fsh distribution provided by our approach is directly usable to implement scientifcgrounded Marine Spatial Planning.**

Tropical marine ecosystems hold major biodiversity hotspots 1 and provide a significant share of global fish catch². Meanwhile, they are increasingly threatened by anthropic pressure including overfishing, global change, invasive species introduction, habitats destruction and pollution 3 . In particular, on-going global ocean warming is expected to severely afect species distribution, abundance and extinction rates but also trophic interactions and entire food webs balance^{4,5}. These threats are critical especially for human populations that rely heavily on marine resources and depend on small-scale fsheries (SSF) or tourism for their livelihoods such as tropical developing states or small tropical islands⁶⁻⁸.

Tropical coastal environments form a mosaic of interconnected mega-habitats extending from the shoreline to the open ocean. This complex structure greatly influences the dynamics of fish assemblages⁹. In recent years, mesophotic reef ecosystems (MREs) have gained attention^{10–14}, not least because their depth may offer protection from anthropic stressors14,15. MREs occur in tropical and subtropical regions and are characterized by the presence of light-dependent corals and associated fauna at depths below 30–40 m extending to 150 m in areas with high water clarity^{12,13,16}. MREs are known hot spots of tropical fish diversity and host fish communities ecologically distinct from shallow water reefs¹⁷. The mesophotic zone usually encompasses the shelf-break, a transition area from shelf to ocean characterised by a rapid change in the topography with a steep slope. The

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stifness of the slope is associated with turbulent mixing enhancing primary productivity and therefore attracting prey and predators^{18–20}. It concentrates diverse fishing resources over a relatively narrow area, sustaining important multispecific reef fisheries²¹⁻²⁴. However, so far few study actually quantified the relative importance of mesophotic reefs for fsh and/or in comparison to euphotic reefs, in particular because consistent observations extending from the shoreline to the shelf-break are lacking²⁵.

Oceanic islands and shallow seamounts also act as topographic anomalies that trigger complex physical processes increasing primary production and concentrating higher trophic levels. Tis phenomenon, known as the Island Mass Effect (IME²⁶) is originated by the turbulence created by the island bathymetry, which uplift nutrientrich water into the photic zone, enhancing primary production²⁷. Oceanic islands and shallow seamounts are important environments for maintaining local biodiversity and non-resident migrating top predatory species²⁸. IME aggregative effect on top predators supports commercial, artisanal and recreational fisheries^{29,30}, which play an important role in the local socio-economic life of insular populations31. So far, most studies on the IME focused on physical-biogeochemical processes^{32,33}. They showed that primary productivity is most enhanced on the leeward side of islands^{30,34}. However, since fewer studies focused on higher trophic levels, the response of fish is generally depicted as symmetrical around islands²⁷. No studies, for instance, determined if fish follow the pattern of primary productivity and concentrate downstream of islands.

Yet, this kind of knowledge is essential to assist decision making in conservation policies to protect biodiversity and the sustainability of fshing and diverse marine uses. Protective management is generally achieved through the creation of Marine Protected Areas (MPAs) delineating permitted and non-permitted zones according to pre-defined management objectives³⁵. However, in some cases, the consequences of establishing MPAs are not adequately thought out, and a poorly planned MPA can be detrimental for local populations that rely on marine resources³⁶. Indeed the decision support tools used to design MPAs rely on available data. To coherently manage the use of maritime space and achieve ecological, economic and social objectives, Marine Spatial Planning (MSP) is increasingly used as a strategic alternative aiming at integrating MPAs in a broader context³⁷. MSP is a complex process requiring the use of optimization solvers that ultimately requires large quantities of spatially explicit cross-disciplinary knowledge and data (ecological, legal, social, economic)³⁸. One of the main challenges to improve knowledge of tropical ecosystems and their resources and implement MSP is thus the data collection³⁹.

Comprehensive monitoring is required to provide ground information for sustainable management^{40,41}. Fish assemblage data are often used to help understanding how human activities influence marine ecosystems^{42,43} or as a measure of ecosystem health⁴⁴ and as a basis for managerial decisions⁴⁵. A variety of methods is used to assess tropical fsh populations, including fshing gears or visual observations, each presenting its own pros and cons⁴⁶. Fishery-dependent methods provide long time-series, wide spatiotemporal coverage but are biased by, among other, gear selectivity⁴⁷. Scientific fish catches are more reliant but have a limited spatiotemporal coverage47*.* Tropical reef fsh communities are also classically described via direct in situ observations through diver-based underwater visual census (UVC)⁴⁸. Scuba diving is constrained by a set of limitations including underwater time and maximal diving depth and visibility^{49–51}. As a result, most UVC-based studies are restricted to near shore shallow waters and provide punctual small-scale information whereas species richness and patterns of distribution is heavily influenced by the range of the sampling area⁵². To overcome part of these limitations and bias, underwater video techniques are increasingly used, whether stationary or towed, remotely operated or autonomous, baited or not⁵³. The use of video increases sampling range and is more time efficient than diverbased observation⁵⁴ but each technique has different limitations and combining underwater video and other sampling methods is therefore recommended⁵⁵.

Active acoustics, in particular multi-frequency, is a powerful tool allowing simultaneous and continuous observation of the distribution of a variety of marine communities and abiotic characteristics^{56–58}. Acoustics range of observation reaches several hundreds of meters below the surface, which allows prospecting the pelagic domain59. However, the ability to discriminate acoustically among taxa remains coarse and works best in relatively low-diversity temperate systems with a few well-defined and acoustically distinct groups⁵⁶. Acoustic species discrimination remains challenging in highly diverse tropical ecosystems. Moreover, acoustics methods needs to be coupled with other observational methods to perform species identifcation, classically extractive one such as trawls and nets⁶⁰. But trawling can be destructive and is not always possible to operate in topographically complex environments or in MPAs^{61–63}. To lift out this lock, the combination of acoustic methods with non-extractive optical methods has emerged. These methods were mostly applied in temperate water^{64–69} whereas to date, only few studies focused on tropical waters $70-72$.

In this context, we combine multifrequency acoustic and video observation to provide a comprehensive vision of fish distribution around a tropical oceanic marine ecosystem. The study area, Fernando de Noronha Archipelago (FNA) located \sim 350 km off the coast of Brazil (Fig. 1), is a typical low productivity and high biodiversity system^{34,73,74}, representative of tropical ecosystems. Like many other tropical small islands, the local population of FNA relies on artisanal fsheries for protein income75 and the economic activity is mainly based on tourism. Tourism generates demographic pressure and all its externalities, amplifying the demand for fsh but also enhancing marine related activities such as recreational fishing or diving^{76,77}. Beside, FNA is protected by a series of legal instruments regulating the uses of the marine environment and marine resources. Indeed, FNA is bathing in an Environmental Protection Area (EPA) where sustainable use of marine resources and tourism is allowed. The EPA includes a smaller no-take MPA, the "National Marine Park of Fernando de Noronha (PARNAMAR)", covering about 70% of the main island and the coastline from the shore to the 50 m isobaths⁷⁸.

On the base of three surveys combining multi-frequency active acoustics and underwater videos, we propose to address the questions identifed above. Specifcally our work includes a series of objectives. First, we aim at providing a comprehensive description of the distribution of the acoustic fsh biomass and fsh assemblages in FNA. Second, we propose to perform the biomass estimation of the most observed fsh, the black triggerfsh *Melichthys niger*. Tird, we propose to quantify the relative importance of mesophotic reefs for fsh in comparison

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Figure 1. Fernando de Noronha archipelago (FNA) (03°51′S, 32°25′W). The blue line delimits the no take MPA PARNAMAR. Acoustic transects are depicted by light grey (FAROFA 1), dark grey (FAROFA 2) and black lines (FAROFA 3); video transects by red dashed lines and video stations by red dots. The black dashed lines depict the 50 and 300 m isobaths. The insert on the main map show the PARNAMAR hatched in blue and the black solid line separates the leeward side (L) and the windward side (W). Map was created by the authors using Matlab R2018b [\(https://fr.mathworks.com/](https://fr.mathworks.com/)) and m_map mapping package 84 .

to euphotic reefs. Four, on this background, we propose to complete the portrait of the IME. Finally, we propose to discuss how the comprehensive information gained by such approach can be usable to implement scientifcgrounded MSP.

Material and methods

Data were collected during three 'Fish Acoustics around Fernando de Noronha' (FAROFA) surveys performed onboard a 10-m-long sport fshing boat (see Supplementary Fig. S1 online) in September 15–21, 2017 (FAROFA179), April 17–23, 2018 (FAROFA2⁸⁰) and April 15–22, 2019 (FAROFA3⁸¹). The first survey was conducted during the dry season (August to January) while the two others during the rainy season (March to July). Data were collected during daytime while prospecting over the continental shelf, shelf-break and near ofshore area (Fig. 1). Video annotations and a degraded resolution of acoustic raw data are published in SEANOE open source platform^{82,83}.

Video data. To identify species and bottom habitat characteristics, four diferent optical systems were deployed: (i) a towed video camera; (ii) a video camera fxed on the transducer support close to the surface; (iii) a video camera deployed vertically; and (iv) a remotely operated vehicle (ROV) (see Supplementary Fig. S1 online). The towed video camera and the video camera fixed on the transducer were used to capture videos along transects. Both provided a view of the water column and allowed for substratum identifcation in shallow water. The towed video was set on a downrigger to deepen the camera, which looked downwards and dragged with a 15 m long rope at a vessel speed of 1.5 m s^{−1}. The videos captured from the camera fixed on the transducer support were especially useful in very shallow waters. Vertical videos and ROV were deployed during stations whose location was driven by observations of important quantity of fsh on the echogram with the purpose of species identification (Fig. 1). Vertical videos were made using a camera fitted on a fishing line. The ROV was operated from the vessel with live stream video.

Towed, fxed and vertical videos were captured with a GoPro TM Hero3+ operating in HD at 1080 p and 30 frames per second during FAROFA 1 and 2 and 60 frames per second during FAROFA 3. ROV videos were performed using a Blue Robotics BlueROV2 system operating in HD at 1080 p and 30 frames per second. To synchronize acoustic and video observations form the towed video, a delay of 9.7 s was subtracted to the video time to adjust with the echosounder time. Each video was annotated using the Solomon Coder software⁸⁵ to identify and enumerate observed species and sediment characteristics classed over nine types (Table 1 and see Supplementary Table S1 online). For the species censuses, we used the maximum number of individuals (MaxN) of a given species present in a single video frame86. In video stations, the MaxN was directly used on each frame to avoid double counting of individuals. In video transects, the MaxN over 3 s of record was used. To estimate the abundance of each taxon observed by video, we used the sum of the maximum values of the MaxN of each video (TMaxN).

Composition	Code
Sand	Sa
Large rock + algae	LrAl
Sand + unknown	SaUn
Sand + algae	Sa Al
$Sand + stone + algebra$	SaStAl
Sand + large rock + algae	SaLrAl
Sand + rhodolith + algae	SaRhAl
$Sand + coral + rhodolith + algebra$	SaCoRhAl
$Sand + stone + cord + rhodolith$	SaStCoRhAl

Table 1. Sediment composition description and code.

Figure 2. Difference between the mean $log10(s_A + 1)$ of different surveys in the selected pixels of 100 m, (a) between surveys, and (**b**) within a given survey. s_A in m^2 nm⁻²; F1, F2, F3: FAROFA 1, 2 and 3, respectively.

Acoustic data. Acoustic data were collected continuously throughout the survey with two SIMRAD EK80 echosounders connected to two 7° split beam transducers centred on the frequencies of 70 and 200 kHz and operated simultaneously in narrow band (continuous wave) transmission. Transducers were attached with a stainless-steel pole to the port side of a 10-m-long sport fishing boat. The ping rate was set to 'maximum' for a maximum acquisition range of 100 m (over the continental shelf) and to 1 ping s−1 of the continental shelf, where the maximum acquisition range was set to 400 m. Vessel speed was ~2.5 m s^{−1} during acquisition of acoustic data and pulse duration was set at 1.024 s. Acoustic data were converted to HAC fles using Hermes software^{87,88}. Processing was completed using the Matecho tool⁸⁹ and Movies3D software⁸⁸. Details on acquisition and calibration parameters as well as on acoustic pre-processing steps from data acquisition i.e., data conversion, bottom detection, filtering and manual cleaning are available in Salvetat et al.⁹⁰.

Acoustic data processing. To discriminate fsh echoes from other organisms, multifrequency approaches, generally rely on the property that swimbladder-bearing fsh have, well beyond the resonance of their swimbladder, high and homogenous backscattering response to frequency⁹¹. To discriminate between scatters attributed to fsh (fsh-like) and those originated by other organisms (no-fsh), e.g. gelatinous and crustaceans, we developed an approach based on thresholds on (i) volume backscattering strength Sv (Sv, in dB re 1 m⁻¹; see⁹² for acoustic definitions), (ii) the bi-frequency sum of Sv, and (iii) the variance of Sv. See Supplementary methods and Supplementary Figs. S2 and S3 online for a detailed description of the methodology. To study the horizontal distribution of fish-like and no-fish echoes, we used the Nautical Area Scattering Coefficient (NASC or s_A , in m^2 nm⁻²)⁹², an index of acoustic biomass, for each ping integrated over the water column. Since fsh-like and no-fsh data were highly correlated at the two frequencies, only the 70 kHz echograms were used for further analyses.

Combining surveys. The three FAROFA surveys were conducted at different seasons and years. To determine whether the surveys could be combined to provide a more comprehensive picture of fsh distribution, we verified if, locally, s_A values were sufficiently similar between surveys with regards to the natural variability observed within surveys (see Supplementary Fig. S4 online). Punctual comparisons were not possible given that observations of the diferent surveys were not located at the same geographical points. We thus selected the pixels containing observations from different surveys at a pixel sizes of 100 m (Fig. 2). The inter-survey comparison was based on the difference between the mean $log10(s_A + 1)$ of different surveys in the selected pixels. Meanwhile,

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Figure 3. Horizontal distribution of the $log10(s_A + 1)$ of fish-like (**a**) and no-fish (**b**) echoes along transects. Geostatistical interpolation of fish-like (**c**) and no-fish (**d**) echoes. The black dashed lines depict the 50 m isobath. Maps were created by the authors using R [\(https://www.R-project.org/\)](https://www.R-project.org/) and RGeostats package¹¹².

the intra-survey variability was computed by selecting records at least one hour apart within the same grid cell. The inter-survey differences of fish-like $log10(s_A + 1)$ were centred on 0 (Fig. 2a) whatever the pair of surveys considered or the pixel size, and were comparable to intra-survey ones. Based on these results indicating a strong spatial stability in the horizontal fish-like $log10(s_A + 1)$ distribution around FNA over years and seasons, we combined the data from the three surveys to provide a comprehensive spatial coverage.

Spatial distribution. To interpolate the horizontal distribution of fish-like and no-fish acoustic biomass outside acoustics transects around FNA, we applied a tailored geostatistical approach. Spatial interpolation was adapted to face the fact that the domain area was elliptical with radial transects (Fig. 3a,b). The geographic reference system was thus irrelevant to describe the orientation between observations. For instance, North–South did not mean the same thing in different parts of the survey area. The relevant orientations to consider were rather parallel or perpendicular to the coast, which required projecting the data in a system conformal to these two main orientations.

To unfold the sampling area, we covered the domain by a series of trapezes that were then aligned and resized one by one so that the distance perpendicular to the coast ranged from 0 (coast line) to 1 (ofshore border of the trapezes), and the distance parallel to the coast ranged from 0 (beginning of the frst trapeze chosen conventionally) to the sum of the length of the bases of the trapezes (Supplementary Fig. S5 online). This projection was bijective so that we could move back and forth between the geographical space and the unfolded space. In particular, the s_A and the kriging grid cells whose coordinates were defined in the geographical space were projected in the unfold space to compute their variogram and their kriging values. To avoid border efects at the edge of the unfolded system, the starting and ending trapezes were duplicated prior to kriging. So, the interpolation of the left side of the first trapeze was made taking into account the data of the last trapeze also. Interpolations were performed over regular cells of 55 m longitude \times 44 m latitude.

Fish assemblages. Despite the simultaneous acquisition of acoustic and video, except for two triggerfish species, it was not possible to allocate each fsh-like echo to a given species. However, consistent fsh assemblages with characteristic echotypes were observed on echograms (Table 2). To attribute each fish-like acoustic scatter to an assemblage, we labelled all fsh-like echoes. Label assignment, hereafer called "labelling", was based on video observation and the presence of characteristic structures in echograms. Video footages made it possible to identify the species observed simultaneously by the cameras and the echosounder. This experience was then used to label the echograms not monitored by videos. For the three cruises, 70 kHz fsh-like echograms were labelled manually by the same operator using the software Matecho⁸⁹, which allows drawing polygons to encompass scatters corresponding to a given assemblage. All fsh-like echoes inside a polygon were allocated to a given assemblage. In total ten assemblages were defned (Table 2). In addition, two species, the black triggerfsh *Melichthys niger* and the ocean triggerfsh *Canthidermis sufamen* could be identifed on echograms due to their characteristic shoal shape. The black triggerfish forms large loose shoals occupying the whole water column distributed over the shelf from 6 to 40 m depth exhibiting diferent body orientations. Ocean triggerfsh form smaller looser shoal generally found on deeper depth (\sim 17 to 70 m) close to the shelf-break. The fish-like s_A of each label corresponding to the 10 assemblages and the two triggerfsh species was echointegrated over the water column by 25 m-long elementary sampling distance unit (ESDU).

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Table 2. Description of the fsh assemblages and two fsh species defned from echotypes (surrounded by a blue dashed line). ⁽¹⁾Barros (2020)⁹³. ⁽²⁾Sazima et al. (2005)⁹⁴. ⁽³⁾Soto (2001)⁹⁵. ⁽⁴⁾Krajewski and Floeter (2011)⁹⁶. ⁽⁵⁾Sazima et al. (2006)⁹⁷.

Black triggerfish biomass estimation. The black triggerfish was particularly abundant in observations. The availability of target strength measurement for this species⁹⁰ opened the field for estimating its biomass. To account for the strong dissymmetry of s_A histograms, we used a non-linear geostatistical approach^{98–100}. Observations were reduced to 5 classes of acoustic biomass, i.e. null, small, medium, large and very large densities, corresponding to the classes 0,]0–33%], [33–66%], [66–95%], [95–100%], respectively. Each interval was coded by an indicator variable, the frst one being nothing but the presence/absence. Tis coding translated the univariate approach of s_A into a multivariate approach (five disjunctive indicator variables, that reduced to four as they sum to one). This became a real issue given the very large number of s_A data available. To solve this problem, the five spatially mutually correlated indicators variables were transformed into five factors called Min-max Autocorrelation Factors—MAF101. Tese factors are linear combinations of the input georeferenced variables, and are uncorrelated at null and at short distances. Assuming that MAFs were also uncorrelated for larger geographical distances, allowed using them independently from the others. In this context, we performed the global estimation of each of the five MAFs over the study polygon by global kriging⁹⁹ and then recombined them to get the kriging estimates of the mean overall s_A together with its estimation variance¹⁰².

Using all black triggerfsh labels, we delineated its area of main presence, concentrated in the east side of FNA. In this area, based on the estimation of the mean overall acoustic value described above, we estimated the biomass by Eq. (1):

Biomass = *mean overall*
$$
s_A \times
$$
 Surface \times W/ $\left(1852^2 \times 4\pi \times 10^{\frac{TS}{10}}\right)$ (1)

where the *Surface* corresponds to the total surface of the delineated area (in m²), *W* is the triggerfish mean weight estimated at 485 g for 27.8 cm long black triggerfsh and *TS* is the target strength of the black triggerfsh at 70 kHz (*TS* = −39.3 dB for 27.8 cm long black triggerfish, the mean size during the surveys⁹⁰).

Environmental drivers. We investigated the relationships between the fsh-like and no-fsh acoustic biomasses to a series of categorical environmental variables:

- Wind/current exposure: FNA was categorised in two sides, leeward and windward, according to the exposition to main winds and currents. Indeed, FNA is under the trade wind regime and washed by the central branch of the South Equatorial Current (cSEC) that both flow from east to west^{103,104}.
- Depth strata: data were classifed in two euphotic (upper euphotic: 0–20 m; lower euphotic: 20–40 m) and three mesophotic (upper mesophotic: 40–60 m; mid-mesophotic: 60–80 m and lower mesophotic: 80–100 m) depth strata using the acoustically-detected bottom-depth.
- MPA: data were compared inside and outside the PARNAMAR in the same depth range $(0-50 \text{ m})$. The area outside the MPA belongs to the multiple use Environmental Protection Area (EPA).
- Sediment type: the nine sediment types extracted from video observation (Table 1).

We used both univariate and multivariate statistics to relate the distribution of the acoustic biomasses $log10(s_A + 1)$ of fish-like and no-fish data as well as the acoustic biomasses of the ten assemblages plus the two triggerfsh species to the environmental factors. To seek for signifcant diferences in acoustic biomass according to each environmental factor, we used a non-parametric Kruskal Wallis test followed by pairwise Wilcoxon tests since the distribution of the data did not follow a normal distribution.

Classification And Regression Trees $(CART^{105})$ were used to explore the relationships between the fish-like acoustic biomass and the fsh assemblages (plus the two triggerfsh species), and environmental variables. In the first case, we used the rpart package¹⁰⁶ while we used the diet package¹⁰⁷ in the second case. The diet package, originally designed to study diet composition, allows a non-parametrical exploratory and predictive approach for identifying complex relationships between environmental variables and assemblages composition. Classifcation tree using the diet package uses a bootstrap technique similar to Breiman¹⁰⁸ and Kuhnert et al.¹⁰⁹. The diet package also allows visualizing the bagged predictions by mapping the predictions110. Both trees were pruned to the smallest cross-validated relative error¹⁰⁵

In addition to the categorical environmental factors, we added two continuous explicative variables, the nofsh acoustic biomass and the bottom local slope (absolute value of the diference between the frst and the last depth in a given ESDU relative to the length of the ESDU expressed in %). All statistical analyses were performed with R^{111} .

All statistical analyses were performed twice, including and not the sediment types. Indeed, the sediment types were extracted from video observation and were thus not available for all ESDU but mostly restricted to the shallow areas where videos observations are available and the sediment observable.

Results

Video data. In total, 49h51 of video footage were acquired. Complete information on species identifcation and sediment classifcation are available at<https://doi.org/10.17882/76019>. Video footage allowed the identifcation of 47 taxa (Table 3) from 29 families including one turtle (*Chelonia mydas*), one dolphin (*Stenella longirostris*), six elasmobranchs including four sharks (*Carcharhinus falciformis, C. perezi, Sphyrna lewini* and *Ginglymostoma cirratum*) and two stingrays (*Aetobatus narinari* and *Hypanus americanus*). We identifed thirty-four osteichthyes fsh species at specie level, four at gender level (*Harengula* sp., *Seriola* spp., *Caranx* spp., *Tunnus* spp.) and three at family level (*Gobidae* sp., *Bothidae* sp., *Ostraciidae* sp.). For 12 species, only a single individual was recorded. 49,189 fish were recorded using the TMaxN. The most abundant specie, *Harengula* sp. (TmaxN 23529, 47.8%) was observed forming large schools in 2 videos, while the second most abundant species, the black triggerfsh *Melichthys niger* (TMaxN 21416, 43.5%) was the fsh observed in more videos (48). Afer those two species, the relative abundance falls with only three species with abundance above 1% of the total abundance: the sergeant major *Abudefduf saxatilis* (TMaxN 1495, 3%), the brown chromis *Chromis multilineata* (TmaxN 636, 1.3%), the ocean triggerfsh *Canthidermis sufamen* (TMaxN 540, 1.1%). Afer *M. niger* that occurred in 48 videos the species observed in more videos were the barracuda *Sphyraena barracuda* (45), the oceanic triggerfish *Canthidermis sufflamen* (39), the black jack *Caranx lugubris* (22), the blue runner *Caranx crysos* (13), the sergeant major *A. saxatilis* (12), and the dog snapper *Lutjanus jocu* (12).

Table 3. List of species observed in video footages. *TMaxN* sum of the maximum number of individuals of a given species present in a single video frame, *TV* towed video, *TS* transducer support, *VP* vertical profle, *RV* ROV. The numbers in parenthesis associated to video types indicates the number of videos in which each species was observed.

Figure 4. Regression tree on values of the fish-like acoustic biomass ($log10(s_A + 1)$) performed on the complete data set (a) and the dataset limited to the locations where sediment was observed (b). The values inside the each leaf is the mean fish-like acoustic biomass ($log10(s_A + 1)$) and the corresponding percentage of ESDU.

Beside the fsh species, we observed gelatinous (salps, siphonophores and ctenophores), fsh larvae (including leptocephalus) and fragments of pelagic algae. These, and likely crustaceans that could not be observed in videos, are the main components of the 'no-fsh' acoustic data.

Fish-like and no fish acoustic biomass. The geostatistical interpolation of fish-like s_A reveals a heterogeneous distribution with the presence of several hotspots, mostly on the windward (east) side of FNA, in the vicinity of the shelf-break (Fig. 3c). The lowest fish-like acoustic biomass was observed on the north-western side, in particular at the mid-shelf. The pattern was different for the no-fish acoustic biomass that was concentrated off the shelf-break with a main aggregation on the leeside (Fig. 3d).

The regression tree relating the fish-like acoustic biomass to the environmental parameters without considering the sediment (Fig. 4a) reveals that the main driving factor is the wind exposure with much higher acoustic biomass per ESDU windward (log10(s_A + 1) = 1.2 m² nm⁻²) than leeward (0.58 m² nm⁻²). A second split occurs in the windward side with higher acoustic biomass (1.3 vs. 0.35 m² nm⁻²) when the slope is greater than 0.08% meaning that fsh-like acoustic biomass is very low in fat areas. When considering the reduced database (restricted to the neritic zone) containing sediment (Fig. 4b), the frst explicative variable is the sediment. The most complex sediment (SaStCoRhAl) encompasses a much higher biomass (2.3 m² nm⁻²) than the others (0.93 m² nm^{−2}). For sediments other than SaStCoRhAl, the next splits are wind exposure, sediment types and no-fish biomass with the highest fish-like biomass $(1.9 \text{ m}^2 \text{ nm}^{-2})$ distributed windward where the no-fish acoustic biomass ranges between 2.3 and 1.6 m² nm^{−2} and over the more complex sediments (SaCoRhAl, SaLrAl, LrAl, SaUn).

Univariate analyses provide some additional information (see Supplementary Fig. S6 online). Fish-like and no-fsh acoustic biomasses signifcantly varied according to wind and current exposure, protection levels, sediment types, and bottom depth strata. Indeed fsh-like acoustic biomass was signifcantly lower on the leeward even if some ESDU encompassed very high acoustic biomass (log10(s_A + 1) up to 6.8 m² nm^{−2}) in presence of small pelagic schools distributed in the upper mesophotic zone (40–60 m) (see "Fish assemblages"). On the opposite, the no-fish acoustic biomass was slightly (but significantly) lower windward (2.1 m² nm⁻²) than leeward $(2.2 \text{ m}^2 \text{ nm}^{-2})$.

The type of sediment encompassing the highest acoustic biomass was by far the most complex one, SaSt-CoRhAl (mean $log10(s_A + 1) = 2.16$ m² nm⁻²), followed by SaCoRhAl (1.14 m² nm⁻²), SaLrAl (1.11 m² nm⁻²), LrAl (1.06 m² nm⁻²), SaStAl (0.99 m² nm⁻²), SaRhAl (0.95 m² nm⁻²), while the less complex habitats Sa (0.78 m² nm⁻²), SaAl (0.77 m² nm⁻²) and SaUn (0.70 m² nm⁻²), presented the lowest mean acoustic biomass and a strong dominance of zero values. The no-fish acoustic biomass did not present any clear association with the sediment complexity since the higher acoustic biomass was associated to SaUn (mean $log10(s_A + 1) = 1.94$ m² nm⁻²), followed by SaStCoRhAl (1.93 m² nm⁻²), SaCoRhAl (1.86 m² nm⁻²), SaStAl and Sa (1.77 m² nm⁻²), SaAl and SaRhAl $(1.66 \text{ m}^2 \text{ nm}^{-2})$, SaLrAl $(1.30 \text{ m}^2 \text{ nm}^{-2})$, and LrAl $(0.69 \text{ m}^2 \text{ nm}^{-2})$.

Figure 5. Violin plot containing boxplot representing the median (horizontal line), interquartile range, whiskers and outlying points of the acoustic biomass of individual assemblages (in $log10(s_A + 1)$) by ESDU, their cumulative sum (red triangles) and the percentage of ESDU with presence of each assemblages (blue diamonds).

Fish-like acoustic biomass was significantly higher (mean $log10(s_A + 1) = 1.14$ m² nm⁻²) in the mid-mesophotic zone (60–80 m) that encompasses the upper edge of the shelf-break than the lower euphotic (1.07 m² nm^{−2}), followed by the upper mesophotic (0.92 m² nm^{−2}), the upper euphotic (0.87 m² nm^{−2}) and the lower mesophotic $(0.85 \text{ m}^2 \text{ nm}^{-2})$. The no-fish acoustic biomass significantly increased with the bottom depth. It was higher in the lower mesophotic strata (mean $\log 10(s_A + 1) = 2.49$ m² nm⁻²) where dense and strong layers of gelatinous were observed, than in the mid-mesophotic (2.25 m² nm⁻²), followed by the upper mesophotic (2.08 m² nm⁻²), the lower euphotic (1.84 m² nm⁻²), and the upper euphotic zone (1.37 m² nm⁻²).

Finally, fish-like acoustic biomass was significantly higher inside (mean $log10(s_A + 1) = 1.18$ m² nm⁻²) than outside (0.46 m² nm⁻²) the no-take zone. Although less marked, the same trend was observed for the no-fish acoustic biomass (1.88 m² nm⁻² vs. 1.75 m² nm⁻²).

Fish assemblages. Fish-like echoes were assigned to ten assemblages and two triggerfish species (Table 2). Video observations allowed a good identifcation of the species present for most of the groups. However, the composition of four groups (bottom weak fsh detection, sand fsh, loose school, shelf-break school) could not be fully validated by the videos.

Small pelagic school presented the highest total acoustic biomass, followed by the black triggerfsh (Fig. 5). The percentage of ESDU with presence of a given assemblage also mostly followed the trends in acoustic biomass with some notable exceptions. Small pelagic fsh school that encompassed the highest acoustic biomass was only observed in 1% of ESDU (Fig. 5), indicating that they were concentrated within some large schools. On the opposite, *M. niger* was the most frequently observed assemblage (in 9.9% of ESDU) followed by bottom weak fsh detection (9.4%) that ranked eighth in terms of total acoustic biomass.

Fish assemblages presented specifc spatial patterns of distribution (Fig. 6). Four assemblages (bottom small fsh school, bottom weak fsh detection, individual demersal and mix reef fsh) presented the most comprehensive distributions over the shelf, all around FNA. Te other assemblages associated to the shelf were loose school and sand fsh, mostly distributed close to the coast and *M. niger*, mainly distributed on the windward side of FNA. Small pelagic schools were distributed both over the shelf and at the shelf-break. The other groups were mostly associated to the shelf-break, with shelf-break schools and shelf-break large fsh distributed all over FNA while small pelagic and predators and *C. sufflamen* were mostly distributed on the windward side.

The regression tree relating the fish-like acoustic biomass to the environmental parameters without considering the sediment (Fig. 7a,c) reveals that the main driving factor is depth strata, discriminating between areas shallower (upper and lower euphotic, upper mesophotic) and deeper than 60 m depth (mid and lower mesophotic). Bottom depths shallower than 60 m correspond to the shelf where neritic assemblages dominate: bottom small fsh school, mix reef fsh, bottom weak fsh detection, *M. niger*. Pelagic assemblages logically dominate in deeper areas: small pelagics and predators, shelf-break large fsh and shelf-break school. Over the shelf, the next splits of the trees are depth strata, wind exposure, MPA protection and s_A no-fish. Mix reef fish constituted 79% of the assemblages on the upper euphotic strata, it was also the dominant group (27%) in the lower euphotic and upper mesophotic strata on the leeward side outside the MPA. *M. niger* dominates (37%) on lower euphotic and upper mesophotic strata on the windward side with low no-fsh acoustic biomass. On the pelagic side, shelf-break large fsh dominated (29%) on the mid-mesophotic zone 60–80 m and small pelagic and predators (47%) on the lower mesophotic zone 80–100 m. When considering the reduced database containing sediment information (Fig. 7b,d), the frst explicative variable was the sediment. On SaStCoRhAl sediment, *M. niger* dominates (73%)*.* For sediments other than SaStCoRhAl, the next splits of the trees are sediment, no-fsh acoustic biomass, wind

 $32.50^{\circ}W$ 32.45 °W 32.40 °W 32.50°W 32.45 °W 32.40 °W 32.50°W 32.45 °W 32.40 °W 32.50°W 32.45 °W 32.40 °W 32.35 °W Longitude

Figure 6. Spatial distribution represented by coloured points of the fish assemblages. The black dotted line depict the 50 m isobath. Maps were created by the authors using R [\(https://www.R-project.org/\)](https://www.R-project.org/) and ggmap¹¹³.

exposure, MPA protection and depth strata. On the sediments LrAl, SaAl, SaRhAl, SaLrAl, SaStAl 52% of the assemblages belong to mix reef fsh.

Univariate analyses showed that the percentage of space occupation was substantially higher on the windward side for half of the assemblages (*C. sufamen,* bottom weak fsh detection, shelf-break large fsh, shelf- break school, small pelagics and predators, *M. niger*) (Supplementary Fig. S7 online). The highest acoustic biomass of all groups corresponded to small pelagics school in the leeward side, followed by *M. niger* in the windward side and mix reef fsh in the leeward side.

All assemblages (Supplementary Fig. S7 online) varied substantially according to the bottom depth strata. The acoustic biomass percent of presence were higher in the upper and lower euphotic strata for mix fish and *M. niger*, respectively. The acoustic biomass and percent of presence of demersal assemblages, sand fish and mix reef fsh, decreased with depth. Te opposite occurred for pelagic groups shelf-break large fsh, shelf-break school, small pelagics and predator, *C. sufflamen* that peaked at mid and lower mesophotic depths, except for small pelagic school that presented highest acoustic biomass in the upper mesophotic zone (40–60 m). Bottom weak fsh detection and bottom small fsh school, individual demersal fsh were distributed and presented higher acoustic biomass either on the lower euphotic or upper mesophotic.

Sediment identifcation was only possible on shallow water and represented a small portion of the data. In this dataset, some assemblages presented clear association with one sediment (Supplementary Fig. S7 online). In particular mix reef fsh were strongly related with SaLrAl, *M. niger* with SaStCoRhAl and bottom small fsh school with Sa. Mix reef fsh and *M. niger,* although showing a higher occurrence on a particular substrate, were the two only groups that appeared on all sediment types.

The no-take MPA effect was clear on fish distribution as six assemblages (bottom weak fish detection, shelfbreak large fsh, bottom small fsh school, shelf-break school, small pelagics and predators, *M. niger*) were more present and had higher acoustic biomass inside the MPA (Fig. 8). *C. sufamen* was virtually absent outside the no-take MPA. The five remaining assemblages (sand fish, loose school, individual demersal fish, mix reef fish and small pelagic school), were all more present and presented higher acoustic biomass outside than inside the no-take MPA.

Figure 7. Regression tree (**a**,**b**) and associated prediction map (**c**,**d**) performed on the acoustic biomass (in $\log 10(s_A + 1)$) of fish assemblages according to the environmental parameters with the complete data set (**a**,**c**) and the dataset limited to the locations with observed sediment (b,d). The fish assemblages identified at each terminal node are those with the highest proportion composition in percent. The composition in assemblage percent for each terminal node is represented by the histogram beneath it. Covariates used to develop the tree were depth strata (*up_eu* upper euphotic, *lo_eu* lower euphotique, *up_meso* upper mesophotic, *mid-meso* mid-mesophotic, *lo_meso* lower mesophotic), position (wind exposure: windward or leeward), protection level (MPA: in or out) sediment (see Table 1 for sediments codes), fish-like and no-fish acoustic biomass (s_A). Fish assemblages abbreviations were, *BSF* bottom small fsh, BWF: bottom weak detection, IDF: individual demersal fsh, MNI: *M. niger*, MRF: mix reef fsh, SBS: shelf-break school, SLF: shelf-break large fsh, SPP: small pelagics and predators. Trees were made by the authors using diet package¹⁰⁷. Maps were created by the authors using R $(https://www.R-project.org/)$ $(https://www.R-project.org/)$ and $ggmap¹¹³$.

Figure 9. Interpolated distribution of the black triggerfish distributed in 5 classes of values of s_A. The black dashed lines represent the acoustic transects. Map was created by the authors using R ([https://www.R-project.](https://www.R-project.org/) [org/\)](https://www.R-project.org/) and RGeostats package112. Created with Adobe Illustrator sofware (<https://www.adobe.com>) by graphic designer Pierre Lopez.

Black triggerfish biomass. The black triggerfish, the second group in terms of fish acoustic biomass, was mostly concentrated in the East side of FNA. In this area (Fig. 9), its actual biomass was estimated to 700 tonnes (19 g m−2) with an estimated CV of 40%. Its distribution inside the area was heterogeneous and organised in patches.

Discussion

By combining acoustic and video observations, we provided for the frst time, a comprehensive vision of tropical fish distribution from the near-shore line to off the shelf-break with a description of (i) fish diversity, (ii) horizontal fish-like and no-fish distribution, and (iii) a focus on the black triggerfish. The data gathered also provided important evidence to (iv) revisit the Island Mass Efect (IME), and (v) give some insights for Marine Spatial Planning (MSP).

Fish diversity. In about 50 h of video footage, we recorded 47 fish species of 29 families (Table 3). This is only a small fraction of the ichthyofauna of FNA that, with a total of 250 species and 77 families, harbours the greatest richness of marine fish among the oceanic islands of the South Atlantic^{96,114}. Indeed our objective was not to perform an exhaustive description of the fsh diversity but to capture the fsh composing the bulk of the biomass. Still, compared with other visual census techniques we observed a similar number of families $(27-28^{96,115,116})$ but much fewer species $(50-66)^{96,115,116}$. Only, Soto⁹⁵ depicted much more families (68) and species (167) but his inventory included pelagic species combining visual census, fsheries surveys and literature records. The most diverse families (Serranidae, Labridae and Pomacentridae) are underrepresented in our study since we focused on pelagic and demersal species that can be assessed with acoustics and did not put eforts on flming benthonic and cryptobenthic communities.

In our study, two species, the tropical sardine *Harengula* sp. and the black triggerfsh *M. niger* accounted for more than 90% of the fish recorded. This confirm the fact that the biodiversity in FNA is represented by few very abundant species¹¹⁷. *M. niger* was the second most abundant and the most often observed species. Such results differ from other studies in FNA. Indeed, most studies^{96,115,118} report *Thalassoma noronhanum*, *Haemulon chrysargyreum* and *Stegastes rocasensis* as dominant species. The difference between our and other studies is likely due to our extensive depth coverage compared with other that mostly focused on shallow (< 20 m) waters. Only Schmid et al.116 observed a dominance of *M. niger* but their study was performed using baited remote underwater video stations and black triggerfish are voracious bold species¹¹⁹. Even if far form exhaustive, our video records likely provide a robust picture of the main pelagic and demersal species present in FNA and a robust information to complement and identify acoustic observations.

Fish echotypes are known to provide a heuristic description of fish species, assemblages or communities^{120,121}. Combining video observations with the scrutinising of acoustic echograms allowed for the identifcation of consistent fsh assemblages (Table 2). We acknowledge that, in some cases, some fsh echotraces may not have been correctly assigned to the proper fish assemblage. However, since these assignations were sustained by \sim 50 h of video footage we are confdent that potential mis-assignation should not have signifcantly impacted our results.

Comprehensive tropical fish distribution. The algorithm we applied on acoustic data allowed discriminating between fish-like and no-fish echograms. The access to simultaneous video observation and the care taken to validate the algorithm with these images makes us confdent that the fsh-like echograms do indeed correspond to fsh.

The strong stability in acoustic fish biomass distribution between FAROFA surveys (Fig. 2), through years and seasons that allowed merging the data from the three surveys, suggests a bottom-up structuration of fsh assemblage. Most fsh observed by video and acoustics are demersal and pelagic. Demersal fsh are classically associated with typical habitat in terms of sediment, benthic composition, structural complexity or depth^{24,122-12} (see also "Revisiting the Island mass efect (IME)"), which may explain site-fdelity. In the same way, except for *Harengula* sp., the pelagic fish species were concentrated at the shelf-break, a known hotspot for pelagic fish⁷⁵. Seasonal variations (e.g. rainy vs. dry season) do not imply signifcant environmental changes. Indeed, the seasonal variation gradient sea surface temperature is minimal (varying from 26.5 to 28 °C)³⁴ and does not seem to signifcantly impact the distribution of the fsh as observed by acoustic. A similar result with no change regarding the season was found in fish predator diet¹²⁵.

The comprehensive spatial coverage we achieved allows providing the first map of fish acoustic biomass around FNA (Fig. 3). Such a picture cannot be completed with classic methods based on visual census or fshing operations (in systems where fshing activity is allowed). More generally, this is the frst comprehensive highresolution map of fsh distribution of a tropical system from the near-shore to the shelf-break. To our knowledge at least one example of map of tropical fsh biomass was produced from acoustic data covering a fraction of the U.S. Virgin Islands¹²⁶. However, this study did not use simultaneous video observation limiting the skills of species identifcation. By combining acoustics and video, we provide the spatial distribution of all acoustically detected fsh (Fig. 3) but also of a variety of fsh assemblages (Fig. 6). Our results reveal a strong heterogeneity in the distribution of fish acoustic biomass with the presence of hotspots. This reinforces the fact that punctual observations may miss hotspots in comparison with our extensive continuous sampling. The comprehensive maps we provide have several benefts since they can help defning areas for further sampling strategies, and are key elements for management in particular for implementing $\mathrm{MSP^{6,38,77}}.$

Black triggerfsh: a key tropical player. In phase with video observations, the black triggerfsh was the second most important species in terms of acoustic biomass and the one occupying most space. In its main zone of distribution, we estimated the black triggerfsh biomass to 700 tonnes (i.e. 19 g m−2). *M. niger* is one of the very few reef fish with a circumtropical distribution¹²⁷. It is known to form large shoals of more than one hundred individuals and has been reported to exhibit high densities around remote oceanic islands such as Ascension Island^{128,129}, Clipperton Atoll¹³⁰, Trinidad Island¹³¹, Johnson atoll and Porto Rico¹²⁷ or St Peter and St Paul's Rocks^{132,133}. *M. niger* thrives at colonizing and maintaining high population levels at remote location¹²⁷. Tis is probably due to its long pelagic phase that enables its settlement in remote location and its high plasticity

in resource use. Indeed, *M. niger* broad omnivory gives him the potential to forage opportunistically on a variety of prey, including pelagic algae remains or dolphin vomit and faeces⁹⁷. In addition, isolated oceanic Islands such as FNA, are impoverished and the functional richness is low^{96,134,135}. *M. niger* has the ability to take advantage of an empty niche as demonstrated by Mendes et al.119 at St Peter and St Paul Archipelago where it endorses the functional role of opportunistic grazing herbivore. FNA lacks of large herbivore that are represented by few small Scaridae of the gender *Sparisoma*136 and roving herbivore represented by few Kyphosidae and Acanthuridae. In our study we observed one species of parrotfsh *Sparisoma amplum* (3 individuals in 2 videos), one species of sea chub *Kyphosus sectatrix* (183 individuals in 4 videos) and one surgeon fsh *Acanthurus chirurgus* (11 individuals in 1 videos). Tose species are found with higher abundance in southern location of the Brazilian coast such as Abrolhos, Bahia for parrot fish and surgeon fish, or even higher latitude such as Arvoredo Island, Santa Catarina and Arraial do Cabo, Rio de Janeiro for sea chubs¹³⁴.

The black triggerfish is mostly distributed on the northeast side of the Island. This windward pattern of distribution is facilitated by its high swimming abilities⁹⁶. The windward side of FNA also concentrates the more complex sediment, containing reef algal-vermetid barriers along rocky shorelines. *M. niger* directly benefts from this sediment complexity as it lays its eggs in the sand, feeds on epilithic algal matrix (EAM) and sleeps in rocky reef137. Adult, *M. niger* is strictly reef-associated as it sleeps in the same hole every night, which attests to a high level of site-fdelity138. If the bulk of the *M. niger* occurred in the selected area for biomass estimation, we also observed *M. niger* on the leeward side close to the shore where large rocks occur. In this case, *M. niger* were juvenile associated with other reef fsh (e.g. surgeons fsh, sergeants). Although abundant, easily catchable, edible and appetent, the black triggerfsh is not a commercial species and is only consumed occasionally.

Revisiting the Island mass effect (IME). The IME describes well the higher primary and secondary productivity on the leeside due to turbulent mixing and advection created by eddies on the wake of islands^{139–141}. However, the IME on tertiary productivity remains quite unknown or is described as isotrope around the island²⁷. Our data allows us to better describe the IME and propose a new conceptual figure (Fig. 10) with an asymmetrical response of the fsh distribution.

Horizontal fsh distribution patterns were very diferent regarding the position of the archipelago face to the main wind/current fows (windward vs leeward). Overall, fsh were much more abundant on the windward side. Tis is true for most assemblages, especially for pelagic fsh and triggerfshes. According to the IME, an overall increase in island-related production is expected. Tis efect is expected to be more marked on the leeward side due to hydrodynamic retention processes^{27,29,34,142,143}. The no-fish acoustic biomass, mostly composed of zooplankton (including gelatinous) was indeed higher on the leeward, in particular of the shelf where dense layers of gelatinous were observed. Lessa et al.74 observed a more important concentration of larvae on the west side (leeward) of FNA. Thus, leeward side primary productivity enhancement³⁴ (see also vertical profiles of chlorophyll concentration performed during the FAROFA surveys showing higher concentration in the leeward side; [https://doi.org/10.17882/70647\)](https://doi.org/10.17882/70647) may participate in fsh larvae recruitment where the leeside fulfls the conditions of Bakun triade: (i) nutrient enrichment, (ii) concentration of larval food distributions, and (iii) local retention of eggs and larvae 144,145.

If the IME is well described in terms of turbulent processes and further primary productivity 30 , there is a lack of fne scale information on the consequences in terms of fsh distribution. We reveal that most pelagic fsh are concentrated windward facing the main fow, where the productivity is expected to be lower. Tis study does not allow concluding about the mechanisms explaining this feature, but we can propose at least three hypotheses. First, the current flow reaching the island topography likely concentrates the flux of particles¹⁴³ including zooplankton, favouring the feeding behaviour of medium size planktivorous pelagic fsh (e.g. *C. sufamen*) or intermediate predators that feed on small fsh, shrimps or invertebrates (e.g. *Caranx crysos*) and are targeted by top predators (e.g. large Scombridae and Carangidae, Istiophoridae and sharks). The aggregation of planktivorous fsh on the windward side of island has been observed by Hamner et al.146 that described a "wall of mouth" composed by planktivorous fish picking up zooplankton before it reaches the reef^{146,147}. Second, the water is much more turbid leeward than windward where visual predators concentrate. Tird, a behavioural pattern consisting in facing the current (reotrope) to keep associated to the island and avoid advection may also play a role.

Interestingly, most demersal assemblages were also more abundant on the windward side. Structural habitat complexity is known to be an important factor for fish richness and abundance^{24,148,149} and higher acoustic biomass was associated with the more complex sediment SaStCoRhAl, a mosaic of diferent substrates. Tis may be a consequence of turbulent processes. Indeed, on the windward side, the strong oceanic flow flush soft sediments, favouring the development of complex habitats that concentrate fish¹⁴⁸. On the other hand, soft sediments deposit leeward, forming sandy habitats that are less populated¹⁴⁸. Wind and current exposure influence the underwater landscape with the windward side, characterized by extensive reef barriers along rocky shorelines, and the leeward side, mainly composed of descending slopes along a rocky shoreline with large scattered boulders and small reefs scattered on sandy habitats¹⁵⁰. This difference in sediment between the windward leeward due to the wind and wave exposure is a common feature observed elsewhere. e.g., in the Madeira Archipelago¹³⁹.

IME efect is thus not just an enhancement of primary productivity though physical process in the leeward side of islands but it also drives the fsh distribution by shaping the habitats. Wind and current cause erosion and a transformation of the shoreline and sediment distribution which also impacts the distribution of fsh. Tis results in an anisotropic distribution of fsh with schools of small pelagic and associated predators fourishing at the shelf-break on the windward side taking advantage of the vertical mixing, the current and the clear water.

Depth strata. Fish acoustic biomass and assemblage composition varied according to the depth strata. The highest overall fish acoustic biomass was observed on the mid mesophotic zone (60–80 m) that corresponds to

Figure 10. Synthetic representation of the island mass effect as illustrated by the case of Fernando de Noronha.

the shelf-break. By providing a cross-shore along-depth acoustic biomass estimations of demersal and pelagic fish, we confirm the importance of the bathymetry¹⁵¹ and quantify and rank the fish distribution among depth strata. Specifcally, the shelf-break was a hot spot for fve assemblages (Fig. 6). Indeed, at the shelf-break, we

observed the steady presence of shelf-break schools as well as small pelagics and predators. It is in this area, in particular at the windward, that SSF targeting both demersal and pelagic species concentrate¹⁵². With our results, it is difcult to determine whether the depth and associated light conditions (euphotic vs mesophotic) or the structural characteristic of the vertical zones are the main drivers of diversity and abundance. Indeed, in our case the mid mesophotic zone matches the shelf-break. This zone has specific characteristics. It presents a steep slope associated to high structural complexity¹⁸. In addition, by being at the interface of neritic and oceanic domains it concentrates organisms from both domains153. Mesophotic reefs associated to the shelf-break are thus hot spots for marine life. However, our result do not confrm that mesophotic reefs per se (independently of the shelfbreak) concentrate more life than euphotic ones. Despite their importance, shelf edge reefs are seldom included in marine protected area network, in particular in Brazil23. Shelf-breaks should thus be considered as important area for biodiversity conservation²⁴.

Insights for marine spatial planning. MSP is a natural extension of practices including integrated coastal management and multi-use MPA management³⁶. MPAs are regarded by many marine scientists as a major management tool needed to tackle fsheries collapse and regular loss of marine biodiversity154–156. Data and analysis should be central to decision-making. The data we provide here have the advantage of being comprehensive. We show that fsh acoustic biomass was signifcantly higher inside than outside the no-take zone (Supplementary Fig. S6 online) indicating a potential efect of the MPA. Such positive efect was witnessed by Ilarri et al.115 on shallow-reef fsh communities. Here, by providing a comprehensive covering of the entire FNA we provide much more robust estimates. If most assemblages had higher acoustic biomass inside than outside the no-take MPA, it is difficult to fully unravel between protection effects and habitat characteristics. Indeed the no-take zone encompasses the entire windward shelf that is characterised by higher bottom habitat complexity while the no-take MPA area encompasses the area close to the port that is susceptible to enhance the productivity through the eutrophication and the presence of rocks and structures. In all cases, with the objective to protect fsh biomass and assemblages, the no-take zone seems overall well designed. It covers most of the shelf, protecting reef fsh that exhibit high densities in near shore shallow waters areas that classically sufer great anthropic pressure linked to multiple use activities including artisanal and recreational fshing. However, the no-take zone stops at the 50 m isobaths and thus leaves the shelf-break unprotected. As an important zone for biodiversity, foraging and spawning, protecting the shelf-break could favour species reliant on shelf-break mesophotic reefs. However, since most SSF operate at the shelf-break it is important to let a sufficient portion of the shelf-break open to fsheries, in particular for those targeting pelagic species that temporally use FNA as a shelter during their long journey.

Conclusion

By combining multifrequency acoustic data and video, we provide the frst comprehensive description of demersal and pelagic fsh distribution of a tropical ecosystem. We also provide the frst biomass estimation of the black triggerfsh *Melichthys niger*, a key tropical player. More generally, we pictured the distribution of a variety of fsh assemblages and related their spatial patterns to biotic and abiotic environmental features. Comparing the efects of euphotic and mesophotic reefs we show that more than the depth, the most important feature is the topography with the shelf-break as the most important hotspot. Beyond, this approach allowed us to revisit the IME. We completed the IME portrait and revealed that it is an asymmetric process regarding fsh distribution. Indeed, while primary productivity is mostly enhanced in the leeward, higher trophic levels concentrate on the windward side. We also tested for the impact of the no-take MPA of Fernando de Noronha Archipelago on the distribution and acoustic biomass of demersal and pelagic fshes. Tis MPA protects the most complex habitats that shelter the highest fsh diversity and biomass. Still, an important hot spot, the shelf-break, currently unprotected could be partly included given that sufficient space is left for fishing activities, in particular for pelagics. Maintaining pelagic small-scale fsheries in FNA is indeed socially and economically important. More generally, describing fish distribution and associated environmental features is the first step toward understanding how fish communities are spatially structured and is a necessary step to conduct MSP and operate relevant protection policies.

Data availability

Video annotations, CTD data and a degraded resolution of acoustic data are available online on SEANOE Sea scientifc open data publication site [\(https://doi.org/10.17882/76019](https://doi.org/10.17882/76019);<https://doi.org/10.17882/71024>, [https://](https://doi.org/10.17882/70647) doi.org/10.17882/70647).

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References

- 1. Bowen, B. W., Rocha, L. A., Toonen, R. J. & Karl, S. A. Te origins of tropical marine biodiversity. *Trends Ecol. Evol.* **28**, 359–366 (2013).
- 2. Lam, V. W. *et al.* Climate change, tropical fsheries and prospects for sustainable development. *Nat. Rev. Earth Environ.* **1**, 440–454 (2020).
- 3. Halpern, B. S. *et al.* Recent pace of change in human impact on the world's ocean. *Sci. Rep.* **9**, 1–8 (2019).
- 4. Capitani, L., de Araujo, J. N., Vieira, E. A., Angelini, R. & Longo, G. O. Ocean warming will reduce standing biomass in a tropical western atlantic reef ecosystem. *Ecosystems* <https://doi.org/10.1007/s10021-021-00691-z>(2021).
- 5. Lima, L. S. *et al.* Potential changes in the connectivity of marine protected areas driven by extreme ocean warming. *Sci. Rep.* **11**, 1–12 (2021).
- 6. Sale, P. F. *et al.* Transforming management of tropical coastal seas to cope with challenges of the 21st century. *Mar. Pollut. Bull.* **85**, 8–23 (2014).
- 7. Dunstan, P. K. *et al.* How can climate predictions improve sustainability of coastal fsheries in Pacifc Small-Island Developing States?. *Mar. Policy* **88**, 295–302 (2018).
- 8. Martins, I. M. & Gasalla, M. A. Perceptions of climate and ocean change impacting the resources and livelihood of small-scale fshers in the South Brazil Bight. *Clim. Change* **147**, 441–456 (2018).
- 9. Moura, R. L. *et al.* Spatial patterns of benthic megahabitats and conservation planning in the Abrolhos Bank. *Cont. Shelf Res.* **70**, 109–117 (2013).
- 10. Lesser, M. P., Slattery, M. & Leichter, J. J. Ecology of mesophotic coral reefs. *J. Exp. Mar. Biol. Ecol.* **375**, 1–8 (2009).
- 11. Bryan, D. R., Kilfoyle, K., Gilmore, R. G. Jr. & Spieler, R. E. Characterization of the mesophotic reef fsh community in south Florida, USA. *J. Appl. Ichthyol.* **29**, 108–117 (2013).
- 12. Fukunaga, A., Kosaki, R. K., Wagner, D. & Kane, C. Structure of Mesophotic Reef Fish Assemblages in the Northwestern Hawaiian Islands. *PLoS One* **11**, e0157861 (2016).
- 13. Kahng, S., Copus, J. M. & Wagner, D. Mesophotic coral ecosystems. In *Marine Animal Forests* (eds Rossi, S. *et al.*) 1–22 (Springer International Publishing, Paris, 2016). https://doi.org/10.1007/978-3-319-17001-5_4-1.
- 14. Rocha, L. A. *et al.* Mesophotic coral ecosystems are threatened and ecologically distinct from shallow water reefs. *Science* **361**, 281–284 (2018).
- 15. Bongaerts, P. *et al.* Deep reefs are not universal refuges: Reseeding potential varies among coral species. *Sci. Adv.* **3**, e1602373 (2017).
- 16. Rosa, M. R. *et al.* Mesophotic reef fsh assemblages of the remote St. Peter and St. Paul's Archipelago, Mid-Atlantic Ridge, Brazil. *Coral Reefs* **35**, 113–123 (2016).
- 17. Medeiros, A. P. *et al.* Deep reefs are not refugium for shallow-water fsh communities in the southwestern Atlantic. *Ecol. Evol.* **11**, 4413–4427 (2021).
- 18. Reid, D. G. SEFOS—Shelf edge fsheries and oceanography studies: An overview. *Fish. Res.* **50**, 1–15 (2001).
- 19. Heyman, W. D. & Kjerfve, B. Characterization of transient multi-species reef fsh spawning aggregations at Gladden Spit, Belize. *Bull. Mar. Sci.* **83**, 531–551 (2008).
- 20. Paxton, A. B. *et al.* Four decades of reef observations illuminate deep-water grouper hotspots. *Fish Fish.* **22**, 749–761. [https://](https://doi.org/10.1111/faf.12548) doi.org/10.1111/faf.12548 (2021).
- 21. Frédou, T. & Ferreira, B. P. Bathymetric trends of northeastern Brazilian snappers (Pisces, Lutjanidae): Implications for the reef fshery dynamic. *Braz. Arch. Biol. Technol.* **48**, 787–800 (2005).
- 22. Longhurst, A. R. & Pauly, D. *Ecologia dos oceanos tropicais* (Edusp, 2007).
- 23. Olavo, G., Costa, P. A., Martins, A. S. & Ferreira, B. P. Shelf-edge reefs as priority areas for conservation of reef fsh diversity in the tropical Atlantic. *Aquat. Conserv. Mar. Freshw. Ecosyst.* **21**, 199–209 (2011).
- 24. Eduardo, L. N. *et al.* Identifying key habitat and spatial patterns of fsh biodiversity in the tropical Brazilian continental shelf. *Cont. Shelf Res.* **166**, 108–118 (2018).
- 25. Silva, M. B., Rosa, R. S., Menezes, R. & Francini-Filho, R. B. Changes in reef fsh assemblages in a cross-shelf euphotic-mesophotic gradient in tropical SW Atlantic. *Estuar. Coast. Shelf Sci.* **259**, 107465 (2021).
- 26. Doty, M. S. & Oguri, M. Te island mass efect. *ICES J. Mar. Sci.* **22**, 33–37 (1956).
-
- 27. Gove, J. M. *et al.* Near-island biological hotspots in barren ocean basins. *Nat. Commun.* **7**, 1–8 (2016). 28. Letessier, T. B. *et al.* Remote reefs and seamounts are the last refuges for marine predators across the Indo-Pacifc. *PLoS Biol.* **17**, e3000366 (2019).
- 29. Heywood, K. J., Barton, E. D. & Simpson, J. H. Te efects of fow disturbance by an oceanic island. *J. Mar. Res.* **48**, 55–73 (1990). 30. Signorini, S. R., McClain, C. R. & Dandonneau, Y. Mixing and phytoplankton bloom in the wake of the Marquesas Islands.
- *Geophys. Res. Lett.* **26**, 3121–3124 (1999). 31. Henry, G. W. & Lyle, J. M. National recreational and indigenous fshing survey (2003).
- 32. Coutis, P. F. & Middleton, J. H. Flow-topography interaction in the vicinity of an isolated, deep ocean island. *Deep Sea Res. Part Oceanogr. Res. Pap.* **46**, 1633–1652 (1999).
- 33. Cardoso, C., Caldeira, R. M. A., Relvas, P. & Stegner, A. Islands as eddy transformation and generation hotspots: Cabo Verde case study. *Prog. Oceanogr.* **184**, 102271 (2020).
- 34. Tchamabi, C. C., Araujo, M., Silva, M. & Bourlès, B. A study of the Brazilian Fernando de Noronha island and Rocas atoll wakes in the tropical Atlantic. *Ocean Model* **111**, 9–18 (2017).
- 35. Motta, F. S. *et al.* Efects of marine protected areas under diferent management regimes in a hot spot of biodiversity and cumulative impacts from SW Atlantic. *Reg. Stud. Mar. Sci.* **47**, 101951 (2021).
- 36. Agardy, T., di Sciara, G. N. & Christie, P. Mind the gap: Addressing the shortcomings of marine protected areas through large scale marine spatial planning. *Mar. Policy* **35**, 226–232 (2011).
- 37. Shucksmith, R. J. & Kelly, C. Data collection and mapping—Principles, processes and application in marine spatial planning. *Mar. Policy* **50**, 27–33 (2014).
- 38. Quefelec, B. *et al.* Marine spatial planning and the risk of ocean grabbing in the tropical Atlantic. *ICES J. Mar. Sci.* **78**, 1196–1208 (2021).
- 39. Rubio-Cisneros, N. T. *et al.* Poor fsheries data, many fshers, and increasing tourism development: Interdisciplinary views on past and current small-scale fsheries exploitation on Holbox Island. *Mar. Policy* **100**, 8–20 (2019).
- 40. Samhouri, J. F., Haupt, A. J., Levin, P. S., Link, J. S. & Shuford, R. Lessons learned from developing integrated ecosystem assessments to inform marine ecosystem-based management in the USA. *ICES J. Mar. Sci.* **71**, 1205–1215 (2014).
- 41. Long, R. D., Charles, A. & Stephenson, R. L. Key principles of marine ecosystem-based management. *Mar. Policy* **57**, 53–60 (2015) .
- 42. Hewitt, J. E., Anderson, M. J. & Trush, S. F. Assessing and monitoring ecological community health in marine systems. *Ecol. Appl.* **15**, 942–953 (2005).
- 43. Caselle, J. E., Rassweiler, A., Hamilton, S. L. & Warner, R. R. Recovery trajectories of kelp forest animals are rapid yet spatially variable across a network of temperate marine protected areas. *Sci. Rep.* **5**, 1–14 (2015).
- 44. Díaz-Pérez, L. *et al.* Coral Reef Health Indices versus the Biological, Ecological and Functional Diversity of Fish and Coral Assemblages in the Caribbean Sea. *PLoS One* **11**, e0161812 (2016).
- 45. Topor, Z. M., Rasher, D. B., Dufy, J. E. & Brandl, S. J. Marine protected areas enhance coral reef functioning by promoting fsh biodiversity. *Conserv. Lett.* **12**, e12638 (2019).
- 46. Pennino, M. G. *et al.* Fishery-dependent and -independent data lead to consistent estimations of essential habitats. *ICES J. Mar. Sci.* **73**, 2302–2310 (2016).
- 47. Hilborn, R. & Walters, C. J. *Quantitative Fisheries Stock Assessment: Choice, Dynamics and Uncertainty* (Springer Science and Business Media, 2013).
- 48. Bohnsack, J. A. & Bannerot, S. P. A stationary visual census technique for quantitatively assessing community structure of coral reef fshes (1986).
- 49. Jones, R. S. & Tompson, M. J. Comparison of Florida reef fsh assemblages using a rapid visual technique. *Bull. Mar. Sci.* **28**, 159–172 (1978).
- 50. Kimmel, J. J. A new species-time method for visual assessment of fshes and its comparison with established methods. *Environ. Biol. Fishes* **12**, 23–32 (1985).
- 51. Michalopoulos, C., Auster, P. J. & Malatesta, R. J. A comparison of transect and species-time counts for assessing faunal abundance from video surveys. *Mar. Technol. Soc. J.* **26**, 27–31 (1992).
- 52. Gray, J. S., Ugland, K. I. & Lambshead, J. Species accumulation and species area curves: A comment on Scheiner (2003). *Glob. Ecol. Biogeogr.* **13**, 473–476 (2004).
- 53. Mallet, D. & Pelletier, D. Underwater video techniques for observing coastal marine biodiversity: A review of sixty years of publications (1952–2012). *Fish. Res.* **154**, 44–62 (2014).
- 54. Langlois, T. J. *et al.* Cost-efcient sampling of fsh assemblages: Comparison of baited video stations and diver video transects. *Aquat. Biol.* **9**, 155–168 (2010).
- 55. Logan, J. M., Young, M. A., Harvey, E. S., Schimel, A. C. G. & Ierodiaconou, D. Combining underwater video methods improves efectiveness of demersal fsh assemblage surveys across habitats. *Mar. Ecol. Prog. Ser.* **582**, 181–200 (2017).
- 56. Koslow, J. A. The role of acoustics in ecosystem-based fishery management. *ICES J. Mar. Sci.* **66**, 966–973 (2009).
57. Bertrand, A. *et al.* Broad impacts of fine-scale dynamics on seascape structure from zooplankton
- 57. Bertrand, A. *et al.* Broad impacts of fne-scale dynamics on seascape structure from zooplankton to seabirds. *Nat. Commun.* **5**, $1-9(2014)$
- 58. Benoit-Bird, K. J. & Lawson, G. L. Ecological insights from pelagic habitats acquired using active acoustic techniques. *Annu. Rev. Mar. Sci.* **8**, 463–490 (2016).
- 59. Sutton, T. T. Vertical ecology of the pelagic ocean: Classical patterns and new perspectives. *J. Fish Biol.* **83**, 1508–1527 (2013).
- 60. McClatchie, S., Torne, R. E., Grimes, P. & Hanchet, S. Ground truth and target identifcation for fsheries acoustics. *Fish. Res.* **47**, 173–191 (2000).
- 61. Cappo, M., Speare, P. & De'ath, G. Comparison of baited remote underwater video stations (BRUVS) and prawn (shrimp) trawls for assessments of fsh biodiversity in inter-reefal areas of the Great Barrier Reef Marine Park. *J. Exp. Mar. Biol. Ecol.* **302**, 123–152 (2004) .
- 62. Harvey, E. S., Cappo, M., Butler, J. J., Hall, N. & Kendrick, G. A. Bait attraction afects the performance of remote underwater video stations in assessment of demersal fsh community structure. *Mar. Ecol. Prog. Ser.* **350**, 245–254 (2007).
- 63. Fitzpatrick, B. M., Harvey, E. S., Heyward, A. J., Twiggs, E. J. & Colquhoun, J. Habitat specialization in tropical continental shelf demersal fsh assemblages. *PLoS One* **7**, e39634 (2012).
- 64. Rooper, C. N., Hof, G. R. & De Robertis, A. Assessing habitat utilization and rockfsh (*Sebastes* spp.) biomass on an isolated rocky ridge using acoustics and stereo image analysis. *Can. J. Fish. Aquat. Sci.* **67**, 1658–1670 (2010).
- 65. Jones, D. *et al.* Evaluation of rockfsh abundance in untrawlable habitat: Combining acoustic and complementary sampling tools (2012)
- 66. O'Driscoll, R. L. *et al.* Species identifcation in seamount fsh aggregations using moored underwater video. *ICES J. Mar. Sci.* **69**, 648–659 (2012).
- 67. Fernandes, P. G., Copland, P., Garcia, R., Nicosevici, T. & Scoulding, B. Additional evidence for fsheries acoustics: Small cameras and angling gear provide tilt angle distributions and other relevant data for mackerel surveys. *ICES J. Mar. Sci.* **73**, 2009–2019 (2016).
- 68. Gastauer, S., Scoulding, B. & Parsons, M. An unsupervised acoustic description of fsh schools and the seabed in three fshing regions within the Northern Demersal Scalefsh Fishery (NDSF, Western Australia). *Acoust. Aust.* **45**, 363–380 (2017).
- 69. Blanluet, A. *et al.* Characterization of sound scattering layers in the Bay of Biscay using broadband acoustics, nets and video. *PLoS One* **14**, e0223618 (2019).
- 70. Campanella, F. & Taylor, J. C. Investigating acoustic diversity of fsh aggregations in coral reef ecosystems from multifrequency fshery sonar surveys. *Fish. Res.* **181**, 63–76 (2016).
- 71. Domokos, R. On the development of acoustic descriptors for semi-demersal fsh identifcation to support monitoring stocks. *ICES J. Mar. Sci.* **78**, 1117–1130 (2021).
- 72. Villalobos, H. *et al.* A practical approach to monitoring marine protected areas: An application to El Bajo Espíritu Santo Seamount near La Paz, Mexico. *Oceanography* **34**, 32–43 (2021).
- 73. Hazin, F. H., Zagaglia, J. R., Broadhurst, M. K., Travassos, P. E. P. & Bezerra, T. R. Q. Review of a small-scale pelagic longline fishery off northeastern Brazil. *Mar. Fish. Rev.* 60, 1-8 (1998).
- 74. Lessa, R. P. *et al.* Distribution and abundance of ichthyoneuston at seamounts and islands of north-eastern Brazil. *Arch. Fish. Mar. Res.* **47**, 239–252 (1999).
- 75. Dominguez, P. S., Zeineddine, G. C., Rotundo, M. M., Barrella, W. & Ramires, M. A pesca artesanal no arquipélago de Fernando de Noronha (PE). *Bol. Inst. Pesca* **42**, 241–251 (2014).
- Lopes, P. F. M., Mendes, L., Fonseca, V. & Villasante, S. Tourism as a driver of conflicts and changes in fisheries value chains in Marine Protected Areas. *J. Environ. Manag.* **200**, 123–134 (2017).
- 77. Outeiro, L., Rodrigues, J. G., Damásio, L. M. A. & Lopes, P. F. M. Is it just about the money? A spatial-economic approach to assess ecosystem service tradeofs in a marine protected area in Brazil. *Ecosyst. Serv.* **38**, 100959 (2019).
- 78. Garla, R. C., Chapman, D. D., Wetherbee, B. M. & Shivji, M. Movement patterns of young Caribbean reef sharks, Carcharhinus perezi, at Fernando de Noronha Archipelago, Brazil: The potential of marine protected areas for conservation of a nursery ground. *Mar. Biol.* **149**, 189–199 (2006).
- 79. Bertrand, A. FAROFA 1 cruise. *RV TUBARAO Tigre.* <https://doi.org/10.17600/18001399> (2017).
- 80. Bertrand, A. FAROFA 2 cruise. *RV TUBARAO Tigre.* <https://doi.org/10.17600/18001411> (2018).
- 81. Bertrand, A. FAROFA 3 cruise. *RV TUBARAO Tigre.* <https://doi.org/10.17600/18001381> (2019).
- 82. Bertrand, A. *et al.* Acoustic data from FAROFA surveys, 2017-09-15 to 2019-04-22.<https://doi.org/10.17882/71024>(2020).
- 83. Salvetat, J. *et al.* Underwater video observations from FAROFA surveys, 2017-09-15 to 2019-04-22. [https://doi.org/10.17882/](https://doi.org/10.17882/76019) [76019](https://doi.org/10.17882/76019) (2020).
- 84. Pawlowicz, R. M_Map: A mapping package for MATLAB, version 1.4 m (computer sofware) (2020).
- 85. Péter, A. *Solomon Coder: Te Concept of Behavioral Elements, Categories and the Representation of Data in Solomon Coder* (2019). 86. Priede, I. G., Bagley, P. M., Smith, A., Creasey, S. & Merrett, N. R. Scavenging deep demersal fshes of the Porcupine Seabight,
- north-east Atlantic: Observations by baited camera, trap and trawl. *J. Mar. Biol. Assoc. U. K.* **74**, 481–498 (1994). 87. McQuinn, I. H. *et al. Description of the ICES HAC standard data exchange format, version 1.60* (Conseil international pour
- l'exploration de la mer, 2005). 88. Trenkel, V. M. *et al.* Overview of recent progress in fsheries acoustics made by Ifremer with examples from the Bay of Biscay. *Aquat. Living Resour.* **22**, 433–445 (2009).
- 89. Perrot, Y. *et al.* Matecho: An open-source tool for processing fsheries acoustics data. *Acoust. Aust.* **46**, 241–248 (2018).
- 90. Salvetat, J. *et al.* In situ target strength measurement of the black triggerfsh *Melichthys niger* and the ocean triggerfsh *Canthidermis sufamen*. *Mar. Freshw. Res.* **71**, 1118–1127 (2020).
- 91. Lavery, A. C. *et al.* Determining dominant scatterers of sound in mixed zooplankton populations. *J. Acoust. Soc. Am.* **122**, 3304–3326 (2007).
- 92. MacLennan, D. N., Fernandes, P. G. & Dalen, J. A consistent approach to defnitions and symbols in fsheries acoustics. *ICES J. Mar. Sci.* **59**, 365–369 (2002).
- 93. Barros, M. J. G. Analises da Ictiofauna marinha e habitats associados atraves de videos subaquatica. (Universidade Federal de Pernambuco, 2020).
- 94. Sazima, C., Bonaldo, R. M., Krajewski, J. P. & Sazima, I. Te Noronha wrasse: A jack-of-all-trades follower. *Aqua J. Ichthyol. Aquat. Biol.* **9**, 97–108 (2005).
- 95. Soto, J. M. R. Peixes do arquipélago Fernando de Noronha. *Mare Magnum* **1**, 147–169 (2001).
- 96. Krajewski, J. P. & Floeter, S. R. Reef fsh community structure of the Fernando de Noronha Archipelago (Equatorial Western Atlantic): Te infuence of exposure and benthic composition. *Environ. Biol. Fishes* **92**, 25 (2011).
- 97. Sazima, I., Sazima, C. & da Silva-Jr, J. M. Fishes associated with spinner dolphins at Fernando de Noronha Archipelago, tropical Western Atlantic: An update and overview. *Neotropical Ichthyol.* **4**, 451–455 (2006).
- 98. Petitgas, P. Use of a disjunctive kriging to model areas of high pelagic fsh density in acoustic fsheries surveys. *Aquat. Living Resour.* **6**, 201–209 (1993).
- 99. Chiles, J.-P. & Delfner, P. *Geostatistics: Modeling Spatial Uncertainty* Vol. 497 (Wiley, 2009).
- 100. Bez, N. & Braham, C.-B. Indicator variables for a robust estimation of an acoustic index of abundance. *Can. J. Fish. Aquat. Sci.* **71**, 709–718 (2014).
- 101. Switzer, P. Min/max autocorrelation factors for multivariate spatial imagery. *Comput. Sci. Stat.* (1985).
- 102. Bez, N. Global estimation based on indicators factorization (2021).
- 103. Assunção, R. V., Silva, A. C., Martins, J. & Montes, M. F. Spatial-temporal variability of the thermohaline properties in the coastal region of Fernando de Noronha Archipelago, Brazil. *J. Coast. Res.* **75**, 512–517 (2016).
- 104. da Silva, A. C. *et al.* Surface circulation and vertical structure of upper ocean variability around Fernando de Noronha archipelago and Rocas atoll during spring 2015 and fall 2017. *Front. Mar. Sci.* **8**, 598101 (2021).
- 105. Breiman, L., Friedman, J., Olshen, R. & Stone, C. Classifcation and regression trees. *Wadsworth Int. Group* **37**, 237–251 (1984).
- 106. Therneau, T., Atkinson, B., Ripley, B. & Ripley, M. B. Package 'rpart'. Available Online Cran Ma Ic Ac Ukwebpackagesrpartrpart Pdf Accessed 20 April 2016 (2015).
- 107. Kuhnert, P. M., Dufy, L. M., Young, J. W. & Olson, R. J. Predicting fsh diet composition using a bagged classifcation tree approach: A case study using yellowfn tuna (*Tunnus albacares*). *Mar. Biol.* **159**, 87–100 (2012).
- 108. Breiman, L. Bagging predictors. *Mach. Learn.* **24**, 123–140 (1996).
- 109. Kuhnert, P. M., Henderson, A.-K., Bartley, R. & Herr, A. Incorporating uncertainty in gully erosion calculations using the random forests modelling approach. *Environmetrics* **21**, 493–509 (2010).
- 110. Kuhnert, P. M. & Mengersen, K. Reliability measures for local nodes assessment in classifcation trees. *J. Comput. Graph. Stat.* **12**, 398–416 (2003).
- 111. R Core Team. *R: A language and environment for statistical computing* (2020).
- 112. ParisTech, M. ARMINES: RGeostats: The Geostatistical R Package (2020).
- 113. Kahle, D. J. & Wickham, H. ggmap: Spatial visualization with ggplot2. *R J* **5**, 144 (2013).
- 114. Pimentel, C. R. *et al.* Mesophotic ecosystems at Fernando de Noronha Archipelago, Brazil (South-western Atlantic), reveal unique ichthyofauna and need for conservation. *Neotropical Ichthyol.* **18** (2020).
- 115. Ilarri, M. I., Souza, A. T. & Rosa, R. S. Community structure of reef fshes in shallow waters of the Fernando de Noronha archipelago: Efects of diferent levels of environmental protection. *Mar. Freshw. Res.* **68**, 1303–1316 (2017).
- 116. Schmid, K. *et al.* First fsh fauna assessment in the Fernando de Noronha Archipelago with BRUVS: Species catalog with underwater imagery. *Biota Neotropica* **20** (2020).
- 117. de Araújo, M. E. *et al.* Diversity patterns of reef fsh along the Brazilian tropical coast. *Mar. Environ. Res.* **160**, 105038 (2020).
- 118. Krajewski, J. P., Floeter, S. R., Jones, G. P. & Leite, F. P. Patterns of variation in behaviour within and among reef fsh species on an isolated tropical island: Infuence of exposure and substratum. *J. Mar. Biol. Assoc. U. K.* **91**, 1359–1368 (2011).
- 119. Mendes, T. C., Quimbayo, J. P., Bouth, H. F., Silva, L. P. & Ferreira, C. E. Te omnivorous triggerfsh *Melichthys niger* is a functional herbivore on an isolated Atlantic oceanic island. *J. Fish Biol.* **95**, 812–819 (2019).
- 120. Petitgas, P. & Levenez, J. J. Spatial organization of pelagic fsh: Echogram structure, spatio-temporal condition, and biomass in Senegalese waters. *ICES J. Mar. Sci.* **53**, 147–153 (1996).
- 121. Burgos, J. M. & Horne, J. K. Characterization and classifcation of acoustically detected fsh spatial distributions. *ICES J. Mar. Sci.* **65**, 1235–1247 (2008).
- 122. Russ, G. R. Grazer biomass correlates more strongly with production than with biomass of algal turfs on a coral reef. *Coral Reefs* **22**, 63–67 (2003).
- 123. Friedlander, A. M. & Parrish, J. D. Habitat characteristics afecting fsh assemblages on a Hawaiian coral reef. *J. Exp. Mar. Biol. Ecol.* **224**, 1–30 (1998).
- 124. Munday, P. L. Does habitat availability determine geographical-scale abundances of coral-dwelling fshes?. *Coral Reefs* **21**, 105–116 (2002).
- 125. Martins, K. *et al.* Assessing trophic interactions between pelagic predatory fsh by gut content and stable isotopes analysis around Fernando de Noronha Archipelago (Brazil), Equatorial West Atlantic. *J. Fish Biol.* **99**, 1576–1590 (2021).
- 126. Costa, B., Taylor, J. C., Kracker, L., Battista, T. & Pittman, S. Mapping reef fsh and the seascape: Using acoustics and spatial modeling to guide coastal management. *PLoS One* **9**, e85555 (2014).
- 127. Kavanagh, K. D. & Olney, J. E. Ecological correlates of population density and behavior in the circumtropical black triggerfsh *Melichthys niger* (Balistidae). *Environ. Biol. Fishes* **76**, 387–398 (2006).
- 128. Lubbock, R. Te shore fshes of Ascension Island. *J. Fish Biol.* **17**, 283–303 (1980).
- 129. Price, J. H. & John, D. M. Ascension Island, South Atlantic: A survey of inshore benthic macroorganisms, communities and interactions. *Aquat. Bot.* **9**, 251–278 (1980).
- 130. Robertson, D. R. & Allen, G. R. Zoogeography of the shorefsh fauna of Clipperton Atoll. *Coral Reefs* **15**, 121–131 (1996).
- 131. Gasparini, J. L. & Floeter, S. R. Te shore fshes of Trindade Island, western south Atlantic. *J. Nat. Hist.* **35**, 1639–1656 (2001).
- 132. Lubbock, R. & Edwards, A. The fishes of Saint Paul's rocks. *J. Fish Biol.* 18, 135-157 (1981)
- 133. Feitoza, B. M., Rocha, L. A., Luiz-Júnior, O. J., Floeter, S. R. & Gasparini, J. L. Reef fshes of St. Paul's Rocks: New records and notes on biology and zoogeography. *Aqua* **7**, 61–82 (2003).
- 134. Ferreira, C. E. L., Floeter, S. R., Gasparini, J. L., Ferreira, B. P. & Joyeux, J. C. Trophic structure patterns of Brazilian reef fshes: A latitudinal comparison. *J. Biogeogr.* **31**, 1093–1106 (2004).
- 135. Floeter, S. R. *et al.* Atlantic reef fsh biogeography and evolution. *J. Biogeogr.* **35**, 22–47 (2008).
- 136. Morais, R. A., Ferreira, C. E. L. & Floeter, S. R. Spatial patterns of fsh standing biomass across Brazilian reefs. *J. Fish Biol.* **91**, 1642–1667 (2017).
- 137. Walsh, W. J. Patterns of recruitment and spawning in Hawaiian reef fshes. *Environ. Biol. Fishes* **18**, 257–276 (1987).
- 138. Walsh, W. J. *Aspects of Nocturnal Shelter, Habitat Space, and Juvenile Recruitment in Hawaiian Coral Reef Fishes* (University of Hawaii, 1984).
- 139. Caldeira, R. M. A., Groom, S., Miller, P., Pilgrim, D. & Nezlin, N. P. Sea-surface signatures of the island mass efect phenomena around Madeira Island, Northeast Atlantic. *Remote Sens. Environ.* **80**, 336–360 (2002).
- 140. Martinez, E. & Maamaatuaiahutapu, K. Island mass efect in the Marquesas Islands: Time variation. *Geophys. Res. Lett.* **31**, 18 (2004).
- 141. Messié, M. *et al.* Te delayed island mass efect: How islands can remotely trigger blooms in the oligotrophic ocean. *Geophys. Res. Lett.* **47**, e2019GL085282 (2020).
- 142. de Souza, C. S., da Luz, J. A. G., Macedo, S., de Montes, M. J. F. & Mafalda, P. Chlorophyll *a* and nutrient distribution around seamounts and islands of the tropical south-western Atlantic. *Mar. Freshw. Res.* **64**, 168–184 (2013).
- 143. Travassos, P., Hazin, F. H., Zagaglia, J. R., Advíncula, R. & Schober, J. Thermohaline structure around seamounts and islands off North-Eastern Brazil. *Arch. Fish. Mar. Res.* **47**, 211–222 (1999).
- 144. Bakun, A. Ocean triads and radical interdecadal variation: Bane and boon to scientifc fsheries management. in *Reinventing fsheries management* 331–358 (Springer, 1998).
- 145. Agostini, V. N. & Bakun, A. 'Ocean triads' in the Mediterranean Sea: Physical mechanisms potentially structuring reproductive habitat suitability (with example application to European anchovy, *Engraulis encrasicolus*). *Fish. Oceanogr.* **11**, 129–142 (2002).
- 146. Hamner, W. M., Jones, M. S., Carleton, J. H., Hauri, I. R. & Williams, D. M. Zooplankton, planktivorous fsh, and water currents on a windward reef face: Great Barrier Reef, Australia. *Bull. Mar. Sci.* **42**, 459–479 (1988).
- 147. Valenzuela, J., Bellwood, D. & Morais, R. Ontogenetic habitat shifs in fusiliers (Lutjanidae): Evidence from Caesio cuning at Lizard Island, Great Barrier Reef. *Coral Reefs* **40**, 1687–1696 (2021).
- 148. Curley, B. G., Kingsford, M. J. & Gillanders, B. M. Spatial and habitat-related patterns of temperate reef fsh assemblages: Implications for the design of Marine Protected Areas. *Mar. Freshw. Res.* **53**, 1197–1210 (2002).
- 149. Ferrari, R. *et al.* Habitat structural complexity metrics improve predictions of fsh abundance and distribution. *Ecography* **41**, 1077–1091 (2018).
- 150. Maida, M. & Ferreira, B. P. Coral reefs of Brazil: An overview. in *Proceedings of the 8th International Coral Reef Symposium* Vol. 1 74 (Smithsonian Tropical Research Institute Panamá, 1997).
- 151. Pittman, S. J., Costa, B. M. & Battista, T. A. Using lidar bathymetry and boosted regression trees to predict the diversity and abundance of fsh and corals. *J. Coast. Res.* **2009**, 27–38 (2009).
- 152. Costa, T. Análise comportamental e distribuição da atividade pesqueira no Arquipelágo de Fernando de Noronha (Nordeste, BR) baseada em dados de GPS. (Universidade Federal Rural de Pernambuco, 2019).
- 153. Spalding, M. D. *et al.* Marine ecoregions of the world: A bioregionalization of coastal and shelf areas. *Bioscience* **57**, 573–583 (2007).
- 154. Claudet, J., Pelletier, D., Jouvenel, J.-Y., Bachet, F. & Galzin, R. Assessing the efects of marine protected area (MPA) on a reef fsh assemblage in a northwestern Mediterranean marine reserve: Identifying community-based indicators. *Biol. Conserv.* **130**, 349–369 (2006).
- 155. Caveen, A. J., Gray, T. S., Stead, S. M. & Polunin, N. V. C. MPA policy: What lies behind the science?. *Mar. Policy* **37**, 3–10 (2013).
- 156. Hernández, C. M. *et al.* Evidence and patterns of tuna spawning inside a large no-take Marine Protected Area. *Sci. Rep.* **9**, 1–11 (2019).

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Author contributions

J.S. prepared and curated data, conducted analyses, designed and implemented methods and prepared the initial manuscript draf; N.B. designed and implemented geostatistical methods and visualisations; J.H., C.L. and G.R. prepared and curated data; G.R. designed and implemented acoustic methods; M.S. designed and implemented statistical methods; A.B., A.L.D., P.T. conceived the study, secured funding; A.B. oversaw the project; J.S., N.B., J.H., A.L.D., C.L., G.R., M.S., P.T., G.V. and A.B. reviewed the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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