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Response to Mepolizumab Treatment in Patients with Severe Eosinophilic Asthma and Atopic Phenotypes

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Purpose: Improved understanding of characteristics that may influence treatment response across phenotypes may help guide treatment decisions.

Patients and Methods: This was a post hoc analysis of MENSA, a multicenter, randomized, double-blind, placebo-controlled trial (NCT01691521). Patients aged ≥12 years with severe eosinophilic asthma received mepolizumab (75 mg intravenously or 100 mg subcutaneously) or placebo, plus standard of care, every 4 weeks for 32 weeks. Outcomes assessed were the annualized rate of clinically significant exacerbations and change from baseline in Asthma Control Questionnaire (ACQ)-5 score. Subgroup analyses were performed by baseline blood eosinophil count (<150, ≥150–300, ≥300 cells/µL) within atopic subgroups (non-atopic [specific immunoglobulin E <0.35 kU/L], atopic [≥0.35–17.5 kU/L], strongly atopic [≥17.5 kU/L]), and by house dust mite (HDM) sensitivity.

Results: Of 576 patients analyzed, 272 were non-atopic, 181 were atopic and 94 were strongly atopic; 29 had missing atopy data. In patients with blood eosinophil counts ≥300 cells/µL, mepolizumab versus placebo reduced clinically significant exacerbations by 74%, 43% and 25% in the non-atopic, atopic and strongly atopic subgroups. Similar reductions were observed in all atopic subgroups in other blood eosinophil count categories where there were sufficient patient numbers for analysis, except for non-atopic patients with baseline blood eosinophil counts of <150 cells/µL. Improvements in ACQ-5 scores of −0.75, −0.73 and −0.78 with mepolizumab versus placebo were observed in non-atopic, atopic and strongly atopic patients with blood eosinophil counts ≥300 cells/µL; consistent improvements in ACQ-5 were not observed in patients with blood eosinophil counts <150 or ≥150–300 cells/µL. Reductions in clinically significant exacerbations with mepolizumab versus placebo were also observed irrespective of sensitivity to HDMs.

Conclusion: Mepolizumab was associated with a trend for reductions in clinically significant exacerbations and improved asthma control versus placebo in patients with severe eosinophilic asthma, irrespective of atopic status or HDM sensitivity.

Keywords: allergens and epitopes, eosinophils, asthma, asthma treatments, biologics

Introduction
Severe asthma is a heterogeneous disease comprising different and overlapping phenotypes driven by distinct pathophysiological processes.1 These phenotypes are distinguished by clinical characteristics, physiological measures and biomarker expression.2,3 Two such phenotypes, severe allergic asthma and severe eosinophilic asthma, are driven by the type 2 immune response.4 Severe allergic asthma is characterized by an early age of disease onset, high serum immunoglobulin...
E (IgE), specific allergen sensitization, and eosinophilic inflammation, while patients with severe eosinophilic asthma typically have a later age of onset, eosinophilic inflammation and recurrent exacerbations, although there is overlap in the clinical presentation of these phenotypes.\(^2\,^3\)

Advances in our understanding of the biological mechanisms underlying severe asthma have provided several disease biomarkers and therapeutic targets. For example, it is established that the airway epithelium is the first line of defense against inhaled allergens and plays a major role in the resulting immune response.\(^5\) Stimulation of the airway epithelium results in the release of alarmins that leads to the recruitment and activation of innate and adaptive immune cells, and the release of inflammatory mediators such as IgE, interleukin (IL)-4, IL-5 and IL-13.\(^5\) As a result, several biologic therapies have been developed that target these mediators. For example, omalizumab, a humanized monoclonal anti-IgE antibody has been developed and approved for use as add-on therapy in patients with moderate to severe allergic asthma,\(^6\,^7\) and mepolizumab, an anti-IL-5 monoclonal antibody, is indicated for use in patients with severe eosinophilic asthma.\(^8\,^9\)

Decisions regarding the use of add-on biologic therapy in patients with severe asthma are based on a phenotypic assessment, and can also include an assessment of biomarkers (such as IgE levels, eosinophil counts and FeNO).\(^1\,^10\) In some cases, assessment of sensitization to particular allergens is also considered.\(^1\,^10\) Historically, allergen sensitization has been a hallmark assessment of allergic asthma. However, based on phenotypic and clinical classifications, predictors of patient response to omalizumab are very similar to those predicting a response to mepolizumab,\(^11\,^12\) and it has been reported that approximately 30% of patients eligible for mepolizumab are also eligible for omalizumab.\(^11\) With the increasing availability of targeted therapies that neutralize different mediators of inflammation, further understanding of patient characteristics that may influence treatment response across phenotypes is needed to inform prescribing choices.

The previously reported MENSA study (GSK ID: MEA115588; NCT01691521) evaluated the efficacy of mepolizumab in patients with severe eosinophilic asthma, and found that the rate of clinically significant exacerbations was significantly reduced and asthma control was improved with mepolizumab compared with placebo.\(^13\)

The objective of this post hoc, subgroup analysis was to compare the effect of mepolizumab across different atopic phenotypes and baseline blood eosinophil count categories in patients from the MENSA study. Subgroup analyses were also performed by sensitivity to particular allergens, including the two common species of house dust mites (HDMs), *Dermatophagoides pteronyssinus* and *Dermatophagoides farinae*.

**Patients and Methods**

**Study Design**

This was a post hoc, subgroup analysis of MENSA, full details of which have been reported previously.\(^13\) In brief, MENSA was a multicenter, randomized, placebo-controlled, double-dummy, double-blind, parallel-group trial. Following a 1–6-week run-in period, patients were randomized (1:1:1) to receive mepolizumab 75 mg intravenously (IV), mepolizumab 100 mg subcutaneously (SC) or placebo, in addition to standard of care, every 4 weeks for 32 weeks. The primary study was conducted in accordance with the ethical principles of the Declaration of Helsinki, International Council for Harmonization Good Clinical Practice, and the applicable country-specific regulatory requirements. The local institutional review board or ethics committee at each study center oversaw trial conduct and documentation (see Supplementary Material for further details). All patients provided written informed consent.

**Patients**

MENSA enrolled patients \(\geq 12\) years of age with severe eosinophilic asthma (blood eosinophil count: \(\geq 300\) cells/\(\mu\)L in the previous year or \(\geq 150\) cells/\(\mu\)L at screening) and a history of \(\geq 2\) exacerbations in the year prior to enrolment despite regular treatment with high-dose inhaled corticosteroids (ICS) in the 12 months prior to screening, plus additional controller medication(s) for \(\geq 3\) months prior to screening. Patients with biologic use within 130 days or five half-lives of administration at screening were excluded.

**Post Hoc Endpoints and Assessments**

The primary outcome for this analysis was the annualized rate of clinically significant exacerbations, defined as a worsening of asthma requiring systemic corticosteroids for \(\geq 3\) days and/or emergency room visit and/or hospitalization. The secondary outcome was the change from
baseline in Asthma Control Questionnaire (ACQ)-5 score at study end.

At baseline, specific IgE titers to *D. pteronyssinus* (HDM 1), *D. farinae* (HDM 2), dog dander, cat dander, *Alternaria alternata* and cockroach were measured. Sensitivity to these allergens was classified into seven categories: class 0 (IgE <0.35 kU/L); class 1 (IgE 0.35–0.70 kU/L); class 2 (IgE 0.71–3.50 kU/L); class 3 (IgE 3.51–17.50 kU/L); class 4 (IgE 17.60–50.00 kU/L); class 5 (IgE 51.00–100.00 kU/L); class 6 (IgE >100.00 kU/L). Post hoc subgroup analyses were performed by baseline blood eosinophil count within atopy strength categories. Patients were categorized as non-atopic (class 0), atopic (classes 1, 2 and 3) or strongly atopic (classes 4, 5 and 6). Within these atopic strength categories, patients were further grouped according to baseline blood eosinophil count (<150 cells/μL, 150–<300 cells/μL or ≥300 cells/μL).

Additional post hoc subgroup analyses were performed by HDM sensitivity and sensitivity to additional allergens. Based on the baseline IgE measurements, patients were categorized as HDM-sensitive (classes 1–6 to *D. pteronyssinus* or *D. farinae*) or HDM non-sensitive (class 0 to *D. pteronyssinus* and *D. farinae*). Within HDM sensitivity categories, patients were further grouped according to their sensitivity to additional allergens (ie, ≥1, ≥2 or ≥3 additional allergens; *A. alternata*, cockroach, cat dander or dog dander).

**Statistical Analysis**

Patients who received ≥1 dose of study drug were included in the analysis and formed the modified intent-to-treat (mITT) population.

Due to similar bioequivalence and efficacy, the mepolizumab doses (75 mg IV/100 mg SC) were combined in this analysis. Analysis of the number of exacerbations was performed using separate negative binomial models for each subgroup, with covariates of treatment group, baseline maintenance oral corticosteroid (OCS) therapy, region, exacerbations in the year prior to the study and baseline percent predicted forced expiratory volume in 1 s (FEV1). Analysis of change from baseline in ACQ-5 score was performed using mixed model repeated measures for each subgroup, with covariates of baseline ACQ-5, region, baseline maintenance OCS therapy, exacerbations in the year prior to the study, baseline percent predicted FEV1, treatment group and visit, plus interaction terms for visit by baseline and visit by treatment group.

**Results**

**Patient Population**

This post hoc analysis was conducted in the mITT population from the MENSA trial, which included 576 patients, of whom 385 (66.8%) received mepolizumab and 191 (33.2%) received placebo; 75 patients had previously received omalizumab. A total of 272 patients were non-atopic (class 0), 181 were atopic (class 1, 2 and 3) and 94 were strongly atopic (class 4, 5 and 6); atopy data were missing for 29 patients. Blood eosinophil count data were missing for an additional seven patients. Of the 576 patients in the overall population, 202 (35%) patients were HDM-sensitive and 345 (60%) were not; HDM sensitivity data were missing for 29 patients. Baseline demographics and disease characteristics by atopy status are shown in Table 1. In the groups categorized as strongly atopic (classes 4, 5 and 6), atopic (classes 1, 2 and 3) and non-atopic (class 0) the mean number of exacerbations in the year prior to the study was similar (3.7, 3.5 and 3.8, respectively) and geometric mean baseline blood eosinophil counts were also similar (280, 310 and 300 cells/μL, respectively) (Table 1).

**Clinically Significant Exacerbations**

Compared with placebo, mepolizumab treatment resulted in reductions in the annual rate of clinically significant exacerbations in all atopy subgroups where an analysis could be performed (ie, baseline blood eosinophil count within atopy subgroups), with the exception of non-atopic patients with baseline blood eosinophil counts of <150 cells/μL where the rate was numerically higher with mepolizumab (Figure 1). Comparing across atopy subgroups in patients with baseline blood eosinophil counts ≥300 cells/μL, reductions in clinically significant exacerbations were numerically higher in the non-atopic subgroup (74%) compared with the atopic (43%) and strongly atopic (25%) subgroups (Figure 1). Reductions were similar in non-atopic and atopic patients with blood eosinophil counts ≥150–300 cells/μL. In the placebo group, annual exacerbation rates increased with increasing baseline blood eosinophil count in the non-atopic subgroups, while there was no obvious trend in patients who were atopic or strongly atopic.

In addition, mepolizumab versus placebo numerically reduced the annual rate of clinically significant exacerbations consistently across all HDM sensitivity subgroups where an analysis could be performed (Figure 2). Within these subgroups, annual exacerbation rates were similar for patients...
sensitive to HDMs (1.71 event/year) versus those not sensitive to HDMs (1.66 event/year) in the placebo group.

**Asthma Control**

Overall, ACQ-5 scores were consistently numerically improved with mepolizumab versus placebo in patients with blood eosinophil counts ≥300 cells/μL compared with lower counts, with treatment differences numerically similar irrespective of atopic status (Figure 3). In patients with a baseline blood eosinophil count <150 and ≥150–300 cells/μL, numerical improvements in ACQ-5 score with mepolizumab versus placebo were only observed in non-atopic but not in atopic or strongly atopic subgroups (Figure 3). ACQ-5 score was also consistently numerically improved with mepolizumab versus placebo irrespective of sensitivity to HDMs and other allergens. Patients sensitive to HDM including those sensitive to ≥1 and ≥2 additional allergens demonstrated numerically smaller reductions in ACQ-5 score compared with patients not sensitive to HDM (Figure 4).

**Discussion**

In this post hoc subgroup analysis of the MENSA study, mepolizumab reduced the rate of clinically significant exacerbations in patients with asthma and a baseline blood eosinophil count ≥150 cells/μL, regardless of atopic status and strength, and in HDM-sensitive and non-sensitive patients. Improvements in asthma control were also shown with mepolizumab versus placebo in all patients with baseline blood eosinophil counts ≥300 cells/μL and in non-atopic patients with baseline blood eosinophil counts of <300 cells/μL, as well as in HDM-sensitive and non-sensitive patients. These results demonstrate the efficacy of mepolizumab across a range of eosinophil thresholds, irrespective of atopic status, and highlight that among patients with severe eosinophilic asthma, clinically important benefits with mepolizumab treatment may be achieved across a broad range of atopic phenotypes. These findings are also consistent with a recent real-world study, which demonstrated that mepolizumab improved exacerbation frequency, OCS use, symptom control and lung function for patients with severe eosinophilic asthma, irrespective of allergic status.14

Patients with severe asthma often have comorbidities, which increase disease burden and the risk of exacerbations.15 Exacerbations in turn have a substantial impact on health-related quality of life for patients with severe asthma, and are associated with considerable healthcare costs.16–18 Therefore, it is important to select an appropriate add-on therapy to help reduce exacerbations and
improve asthma control. We observed that mepolizumab versus placebo provided reductions in clinically significant exacerbations in non-atopic, atopic and strongly atopic patients. Additionally, in non-atopic individuals, a trend towards greater reductions with mepolizumab over placebo in clinically significant exacerbation rates with increasing blood eosinophil count was observed. As such, mepolizumab may be an important treatment option for patients who

**Figure 1** Rate ratios for clinically significant exacerbations by atopy/baseline blood eosinophil count status. Atopy was defined according to sensitization to any one of the following allergens: *A. alternata*, cockroach, HDM, cat dander and dog dander. Non-atopic: class 0 (IgE <0.35 kU/L); atopic: classes 1, 2 and 3 (IgE ≥0.35–17.5 kU/L); strongly atopic: classes 4, 5 and 6 (IgE >17.5 kU/L).

**Abbreviations:** CI, confidence interval; HDM, house dust mite; IgE, immunoglobulin E; NE, rate ratio could not be estimated due to insufficient patient numbers.

**Figure 2** Rate ratios for clinically significant exacerbations by HDM sensitivity. HDM-sensitive (classes 1–6: IgE ≥0.35 kU/L to *D. farinae* and/or *D. pteronyssinus*) or HDM non-sensitive (class 0: IgE <0.35 kU/L to *D. farinae* and/or *D. pteronyssinus*). Within these categories, patients were grouped according to sensitivity to other allergens (*A. alternata*, cockroach, cat dander or dog dander; classes 1–6: ≥0.35 kU/L) to ≥1, ≥2 or ≥3 additional allergens.

**Abbreviations:** CI, confidence interval; HDM, house dust mite; IgE, immunoglobulin E; NE, rate ratio could not be estimated due to insufficient patient numbers.
remained uncontrolled on other biologics, including omalizumab. Indeed, a recent study found that in patients with severe eosinophilic asthma and an overlapping allergic phenotype who were uncontrolled with omalizumab therapy, switching from omalizumab to mepolizumab was associated with clinically significant improvements in asthma control, health

### Figure 3
Change from baseline in ACQ-5 score at Week 32 by atopy and baseline blood eosinophil count categories. Atopy was defined according to sensitization to any one of the following allergens: *A. alternata*, cockroach, HDM, cat dander and dog dander. Non-atopic: class 0 (IgE <0.35 kU/L; atopic: classes 1, 2 and 3 (IgE 0.35–17.5 kU/L); strongly atopic: classes 4, 5 and 6 (IgE 17.6–>100 kU/L).

**Abbreviations:** ACQ, asthma control questionnaire; CI, confidence interval; HDM, house dust mite; IgE, immunoglobulin E; LS, least squares; NE, rate ratio could not be estimated due to insufficient patient numbers.

### Figure 4
Change from baseline in ACQ-5 score at Week 32 by HDM sensitivity. HDM-sensitive (classes 1–6: IgE ≥0.35 kU/L to *D. farinae* and/or *D. pteronyssinus*) or HDM non-sensitive (class 0: IgE <0.35 kU/L to *D. farinae* and/or *D. pteronyssinus*). Within these categories, patients were grouped according to HDM sensitivity (classes 1–6: ≥0.35 kU/L) and to additional allergens (ie ≥1, ≥2 or ≥3 additional allergens; *A. alternata*, cockroach, cat dander or dog dander).

**Abbreviations:** ACQ, asthma control questionnaire; CI, confidence interval; HDM, house dust mite; IgE, immunoglobulin E; LS, least squares.

### Table

<table>
<thead>
<tr>
<th>HDM-sensitivity and baseline eosinophil count</th>
<th>Placebo</th>
<th>Mepolizumab</th>
<th>Favors mepolizumab</th>
<th>Favors placebo</th>
<th>Treatment difference (95% CI)</th>
</tr>
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<tbody>
<tr>
<td>&lt;150 cells/μL Non-atopic</td>
<td>14</td>
<td>-0.46</td>
<td>27</td>
<td>-0.77</td>
<td>-0.31 [-1.00, 0.38]</td>
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<td>Aortic</td>
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<td>-0.60</td>
<td>25</td>
<td>-0.43</td>
<td>0.17 [-0.61, 0.95]</td>
</tr>
<tr>
<td>Strongly atopic</td>
<td>3</td>
<td>NE</td>
<td>13</td>
<td>NE</td>
<td>NE</td>
</tr>
<tr>
<td>≥150–300 cells/μL Non-atopic</td>
<td>20</td>
<td>-0.63</td>
<td>44</td>
<td>-0.66</td>
<td>-0.22 [-0.71, 0.27]</td>
</tr>
<tr>
<td>Aortic</td>
<td>13</td>
<td>-1.19</td>
<td>20</td>
<td>-0.65</td>
<td>0.35 [-0.45, 1.14]</td>
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<tr>
<td>Strongly atopic</td>
<td>12</td>
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<td>14</td>
<td>-0.61</td>
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<tr>
<td>≥300 cells/μL Non-atopic</td>
<td>46</td>
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<td>90</td>
<td>-1.15</td>
<td>-0.75 [-1.06, -0.43]</td>
</tr>
<tr>
<td>Aortic</td>
<td>30</td>
<td>-0.26</td>
<td>61</td>
<td>-1.00</td>
<td>-0.73 [-1.19, 0.28]</td>
</tr>
<tr>
<td>Strongly atopic</td>
<td>15</td>
<td>-0.43</td>
<td>22</td>
<td>-1.21</td>
<td>-0.78 [-1.30, -0.26]</td>
</tr>
</tbody>
</table>

**Note:** Treatment difference, mepolizumab - placebo (95% CI)

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**Figure 3** Change from baseline in ACQ-5 score at Week 32 by atopy and baseline blood eosinophil count categories. Atopy was defined according to sensitization to any one of the following allergens: *A. alternata*, cockroach, HDM, cat dander and dog dander. Non-atopic: class 0 (IgE <0.35 kU/L; atopic: classes 1, 2 and 3 (IgE 0.35–17.5 kU/L); strongly atopic: classes 4, 5 and 6 (IgE 17.6–>100 kU/L).

**Abbreviations:** ACQ, asthma control questionnaire; CI, confidence interval; HDM, house dust mite; IgE, immunoglobulin E; LS, least squares; NE, rate ratio could not be estimated due to insufficient patient numbers.

**Figure 4** Change from baseline in ACQ-5 score at Week 32 by HDM sensitivity. HDM-sensitive (classes 1–6: IgE ≥0.35 kU/L to *D. farinae* and/or *D. pteronyssinus*) or HDM non-sensitive (class 0: IgE <0.35 kU/L to *D. farinae* and/or *D. pteronyssinus*). Within these categories, patients were grouped according to HDM sensitivity (classes 1–6: ≥0.35 kU/L) and to additional allergens (ie ≥1, ≥2 or ≥3 additional allergens; *A. alternata*, cockroach, cat dander or dog dander).

**Abbreviations:** ACQ, asthma control questionnaire; CI, confidence interval; HDM, house dust mite; IgE, immunoglobulin E; LS, least squares.

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status and exacerbation rate, with no new safety issues reported.\textsuperscript{19}

We did not observe any impact of mepolizumab on clinically significant exacerbations in non-atopic patients with baseline blood eosinophil counts of <150 cells/μL, although there was some improvement in asthma control (as shown by the improvement in ACQ-5 score) with mepolizumab versus placebo in this subset of patients. In addition, although the largest improvement in clinically significant exacerbations with mepolizumab versus placebo was seen in the atopic subgroup with baseline blood eosinophil counts <150 cells/μL, this is likely due to the relatively high exacerbation rate observed in the patients receiving placebo and the low number of patients in the placebo group (N=9). Therefore, overall our data provide further support for the use of 150 cells/μL as a threshold for mepolizumab treatment in patients with severe asthma.\textsuperscript{20,21}

We noted that the largest improvements in clinically significant exacerbation rates with mepolizumab versus placebo in patients with baseline blood eosinophil counts ≥300 cells/μL were in patients who were non-atopic compared with atopic and strongly atopic. This may indicate that eosinophils are a primary contributing factor to exacerbations in atopic patients with blood eosinophil counts ≥300 cells/μL, and that other non-IL-5 related mechanisms may be contributing factors to exacerbation etiology at lower blood eosinophil counts.\textsuperscript{22} Similarly, patients sensitive to HDM including those sensitive to ≥1 and ≥2 additional allergens demonstrated numerically smaller reductions in ACQ-5 score compared with patients not sensitive to HDM, which may also suggest that atopy is a more important driver of symptoms in HDM-sensitive patients. However, further work will be needed to verify these findings, particularly because these trends were not consistently observed across subgroups or endpoints.

The presence of both IgE and eosinophilia are indicative of an asthma phenotype driven by type 2 inflammation, which in turn may be triggered by a number of stimuli, including IgE-mediated sensitivity to various allergens.\textsuperscript{23} The allergens selected for atopy assessments in the current study were chosen based on their effects on patients with asthma owing to their perennial nature, and their use in other published studies.\textsuperscript{24} We also felt it was important to determine the effect of HDM allergy on mepolizumab efficacy, given its widespread impact on patients with allergic disease and the association of specific allergen immunotherapy for HDM allergy with improvement in asthma outcomes.\textsuperscript{25,26} Therefore, the HDM species, \textit{D. pteronyssinus} and \textit{D. farinae}, were selected for this study based on their global prevalence, perennial nature and frequent assessment in a clinical setting.\textsuperscript{25,27} Both species have also been shown to have substantial allergenic homology.\textsuperscript{28} Importantly, both reductions in clinically significant exacerbations and improvements in ACQ-5 score were consistent across HDM-sensitive and non-sensitive subgroups, independent of sensitivity to the additional allergens tested. These results confirm that selection of mepolizumab as an add-on treatment for patients with severe eosinophilic asthma is associated with a reduction in exacerbation rates and improved asthma symptom control, regardless of sensitivity to the common HDM allergen.

This analysis has several limitations which require consideration. First, the post hoc nature of the analysis and the fact that the number of patients was low in some of the subgroups should be kept in mind when interpreting the findings. Second, we did not account for seasonal variation in the frequency of exacerbations or assess the polysensitization status of patients to seasonal allergens. Although many patients with severe asthma experience frequent exacerbations throughout the year, some experience stable periods punctuated by significant exacerbations at particular times of the year.\textsuperscript{29} Third, it should be noted that atopy was measured using blood biochemistry only, the results of which may have been discordant with the clinical characteristics displayed by patients. Indeed, it has been suggested that reporting of asthma worsening in a clinical setting is a better predictor of response to omalizumab than IgE levels.\textsuperscript{30,31} Finally, as in previous post hoc analyses, we combined different doses and routes of administration of mepolizumab in this post hoc analysis, based on the similar reductions in exacerbation rate with the bioequivalent IV and SC doses shown in the original study.\textsuperscript{13}

**Conclusions**

Reductions in clinically significant exacerbations and improvements in asthma control with mepolizumab versus placebo were generally consistent across the atopic subgroups in which an analysis could be performed. This suggests that mepolizumab is suitable for use in patients with severe asthma and an eosinophilic phenotype, irrespective of atopic status and HDM sensitivity. As such, these results provide valuable information for clinicians treating patients with severe asthma across a range of phenotypes.
Abbreviations
ACQ, Asthma Control Questionnaire; CI, confidence interval; FEV₁, forced expiratory volume in 1 second; FeNO, fractional exhaled nitric oxide; HDM, house dust mite; ICS, inhaled corticosteroid; IgE, immunoglobulin E; IL, interleukin; IV, intravenously; LS, least squares; mITT, modified intent-to-treat population; NE, rate ratio could not be estimated due to insufficient patient numbers; OCS, oral corticosteroid; SC, subcutaneously.

Data Sharing Statement
Anonymized individual participant data and study documents for the parent study can be requested for further research from www.clinicalstudydatarequest.com.

Ethics Approval and Informed Consent
This was a post hoc analysis of anonymized patient data from study MEA115588/NCT01691521. The primary study was conducted in accordance with the Declaration of Helsinki, the International Council on Harmonization Good Clinical Practice guideline, and the applicable country-specific regulatory requirements. The local institutional review board or ethics committee at each study center oversaw trial conduct and documentation. All patients provided written informed consent.

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Author Contributions
All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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Disclosure
CMP, SM and SWY are all employees and shareholders of GSK. FCA is a former employee of GSK, holds GSK stocks/shares and is currently employed by Avillion US, Inc. MI has received fees from Actelion, AstraZeneca, Bayer, Berlin Chemie, Boehringer Ingelheim, Chiesi, CSL-Behring, Grifols, GSK, MSD, Novartis and Teva. JAD has received personal fees from AstraZeneca, GSK, Novartis and Sanofi-Aventis, has been an investigator for AstraZeneca, GSK, Novartis, Sanofi-Aventis, Grifols, BioCryst, CSL Ltd and Equilum, and holds shares in CSL Ltd. AB has received fees from GSK, Regeneron, Chiesi, AstraZeneca, Teva, Novartis and Boehringer Ingelheim and been an investigator for Novartis and AstraZeneca. The authors report no other conflicts of interest in this work.

References


