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Natal origin and migration pathways of Mekong catfish (*Pangasius krempfi*) using strontium isotopes and trace element concentrations in environmental water and otoliths

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Abstract

To improve our knowledge of the migration pathway of a highly threatened fish species along the Mekong River, strontium isotope ratios (⁸⁷Sr/⁸⁶Sr) and 18 trace element concentrations were measured in the water and in the otoliths of an anadromous catfish, Pangasius krempfi, to infer its natal origin and potential migration pathways. Water was sampled at 18 locations along the mainstream, tributaries and distributaries of the Mekong River. To check for accuracy and precision, measurements of the 87 Sr $/86$ Sr ratios and trace element concentrations were then compared in two laboratories that use different analytical methods. Differences in trace element concentrations between locations were not significant and could not, therefore, be used to discriminate between migration pathways. However, the Mekong mainstream, tributaries and distributaries could all be discriminated using Sr isotopes. The 87 Sr/ 86 Sr profiles recorded in P. krempfi otoliths showed that there were three contingents with obligate freshwater hatching and variable spawning sites along the Mekong mainstream, from Phnom Penh (Cambodia) to Nong Khai (Thailand) or further. After hatching, the fish migrated more or less rapidly to the Mekong Delta and then settled for most of their lifetime in brackish water. Spawning habitats and migration routes may be threatened by habitat shifts and the increasing number of hydropower dams along the river, especially the contingents born above Khone Falls (Laos). The conservation of P. krempfi, as well as other migratory fish in the Mekong River, requires agreements, common actions and management by all countries along the Mekong River. This study highlighted the importance of using both Sr/Ca and ⁸⁷Sr/⁸⁶Sr ratios to understand life history of anadromous fishes as the

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 87 Sr/ 86 Sr ratio in the water was shown to be less effective than the Sr/Ca ratio in identifying movements between different saline areas.

Introduction

The Mekong River is the main river in South East Asia, it rises in the Tibetan Plateau, crosses six countries (China, Myanmar, Laos, Thailand, Cambodia, and Vietnam), and creates a delta before flowing out to sea. With a mainstream around 4,400 km long and a catchment area of 795,000 km² [1, 2], the Mekong River has the second highest inland fish biodiversity in the world, with more than 1,100 species [3, 4]. Wild fish are an important source of income and consumption for local populations [5]. Of all the major river systems in the world, the Mekong River and its Delta are probably the most affected by global climate change and human activities [6, 7]. Pollution and saline intrusions with rising sea levels due to climate change currently threaten the Delta $[8-10]$, while the whole Mekong River is even more threatened by many dam-building projects that could have strong impacts, especially on fisheries [6]. With the dams currently under construction or planned in the near future on the Mekong mainstream and its tributaries, scientists are considering measures to mitigate the anthropogenic influences on all surrounding socio-ecosystems [6]. The numerous dams being built on the Mekong mainstream will have an negative impact on the migratory behavior of many fish species and on associated fisheries [6, 11]. By changing hydrological flow and habitat connectivity, the presence of dams impacts fish populations, by blocking possible migration pathways and reducing population size and potentially their genetic diversity [6, 12, 13]. Understanding the life history of migratory fish, as well as their key habitats in the Mekong River, has become essential for monitoring and managing fish communities and populations under threat.

It has been estimated that 87% of known fish species in the Mekong River are migratory (anadromous, catadromous, or potamodromous) [14] and that 50% of the catches include long-distance migrants [4]. The migration pathways in the Mekong River have been described based on the hydrological and morphological characteristics of the mainstream and on the feeding behavior of fish in different habitats [14]. In many species, these migration patterns are interconnected or overlapping. Among these patterns, the main migration route includes the mainstream from the Mekong Delta to the Khone Falls at the border between Cambodia and Laos, where annual floods affect large areas, and are essential for fishery productivity [14]. Nevertheless, our understanding of their migratory behavior is speculative, and the location of the spawning grounds and homing cues remain largely unknown.

Fish otoliths (ear stones) are biogenic carbonate structures deposited on a protein matrix, which grow throughout the fish's lifetime with both daily and seasonal growth increments [15]. The otoliths are composed of 99% calcium carbonate, but, during formation, they incorporate trace elements from the water that can provide a detailed history of an individual fish's environment and life events [16, 17]. The incorporation of elements is a complex process involving different physiological barriers. Elements are absorbed from the water through the gills or the intestine into the plasma and the endolymph surrounding the otolith, where they crystallize in the matrix [17]. The process is affected by various environmental and physiological factors, including salinity, temperature, ontogeny, metamorphosis, sex, growth, and diet [18–24]. Incorporation also depends on the element concerned, and there are many differences between them. For example, carbon isotopes and magnesium are related to physiological factors, whereas physiology is weakly related to strontium and lithium, and oxygen isotopes, barium, and manganese are related to past environments [24–26].

The study of fish environmental conditions and physiology is thus made possible by otolith chemistry. Strontium is known as the best proxy for ambient water salinity [20, 27], the Sr/Ca is the ideal variable to estimate fish movements between marine, brackish and fresh waters [28, 29], whereas it is not affected by water temperature [30]. When compared with water concentrations and age estimates, otolith strontium isotope ratios (87 Sr/ 86 Sr) can provide valuable information about the migration pathways of species that pass through different habitats [17, 31, 32]. More specifically, the strontium isotopic ratio $(^{87}Sr)^{86}Sr$) signatures in the water [33] and the otoliths provide evidence for the origin of individual migratory and non-migratory fish [34–39]. Duponchelle et al. [40] recently investigated the natal origin of an Amazonian giant catfish (*Brachyplatystoma rousseauxii*) and by comparing the Sr isotope ratios in otoliths and in the water over a very large spatial scale, these authors identified one of the longest fish migrations in freshwater: more than 4,000 km. However, the Sr isotope ratio is a proxy of natal origin in freshwater [40], and its use in saline areas is limited because the variation of ${}^{87}Sr/{}^{86}Sr$ value is less sensitive to changes in salinity when salinity is higher than 20 [29, 30].

The catfish *Pangasius krempfi* (Fang and Chaux 1949; Siluriformes, Pangasiidae) is one of the most important anadromous fishes in the Mekong River, and is classified as vulnerable in the International Union for Conservation of Nature (IUCN) Red List [41]. This species is widely distributed in the Mekong basin from China to Vietnam, but is primarily found in Laos, Cambodia, Thailand and Vietnam in both brackish waters and freshwaters [42, 43]. This catfish is one of the target species for fishery and aquaculture in the countries along the Mekong where it has high economic value in the local markets [43–46]. Although *P*. *krempfi* represents only around 2% of the total catches by Mekong fisheries, its economic value on landing is three times higher than that of more common species [45]. The domestication of *P*. *krempfi* has been investigated to improve catfish aquaculture in the Mekong Delta [46]. Information on the life history traits of the species is scarce in the literature. *P*. *krempfi* is believed to undertake a long migration from its growing areas in brackish waters in the Mekong delta over the rapids and deep pools of the Khone Falls in Laos for reproduction [44, 47] at the beginning of the rainy season [43]. However, the spawning grounds of *P*. *krempfi* and age at first sexual maturity and reproduction are not yet fully known [41, 43]. After hatching at the Khone Falls, the larvae are assumed to rapidly drift downstream, and juveniles and adults spend almost all their life in the Mekong Delta before migrating back upstream to spawn [14, 44, 47]. Living in the brackish Mekong Delta, this anadromous species is presumed to be sensitive to the construction of dams upstream, which is why it was chosen to infer its natal origin using the Sr isotope ratio in the otolith core $[47]$. The resulting information can be used to identify threats to the migration pathways due to dam construction or habitat changes.

The primary aim of the present study was to identify *P*. *krempfi* spawning grounds and the possible effects of habitat changes (such as dam construction) on this anadromous species. Our specific objectives were to (i) map the Sr isotope ratios (${}^{87}Sr/{}^{86}Sr$) in waters along the Mekong River until Laos, and in some of its main tributaries, (ii) measure the concentrations of trace elements in the waters to identify differences between locations along the river, (iii) estimate the natal origin and migration behavior of *P*. *krempfi* caught in the Mekong Delta. Strontium isotope ratios in the water have never been recorded along the Mekong River for the purpose of tracing migrations. This catfish species was selected as a model because of its particular migratory behavior, and Sr isotope ratios in the water and otoliths were tested as tools for understanding fish migration. This research could provide fundamental data, including strontium isotope values in the Mekong and tributaries, to assign natal origin to larvae, and information on the migration pathway of other fish species caught in the Mekong basin using these isotopes.

Materials and methods

Study area

The Mekong River rises in the Tibetan Plateau in China, and crosses Myanmar, Laos, Thailand, Cambodia, and Vietnam where it forms a gigantic delta before flowing into the sea though nine river mouths. From Laos, the mainstream is supplied by water flowing from three main complex tributaries, Chi-Mun River and Tonle Sap Lake on the right bank, and San-Srepok Rivers that form the Sekong River on the left bank (Fig 1A) [48]. After crossing Phnom Penh city (Cambodia), the Mekong River turns into two distributaries, the Mekong-Tien River and the Bassac-Hau River, which, in Vietnam, form a massive delta that drains into the sea through nine river mouths ($Fig 1B$) [49].

This study was conducted in the large stretch of the Mekong River that crosses four countries (Fig 1), Laos, Thailand, Cambodia, and Vietnam. The 19 sample locations were evenly spaced from the mouth of the river up to a site close to Vientiane, the capital of Laos (Fig 1 and Table 1). The upstream sampling sites began along the Thailand bank of the river in Laos to Cambodia and continued to downstream sites in the Mekong Delta in Vietnam where they

Fig 1. Sampling sites along the Mekong River. (A) Map of sampling sites for water (black circle) and fish (catfish) along the Mekong River, and (B) detailed map of the sites in the Mekong Delta.

Site	Location	Coordinates		Sampling date	Salinity	Distance from the sea (km)
		Latitude	Longitude			
M01	Nong Khai, Thailand	17°49.740'N	102°41.878'E	September 2019	0.0	1,529
M02	Nakhon Phanom, Thailand	17°3.735'N	104°45.268'E	September 2019	0.0	1158
M03	Ubon Ratchathani, Thailand	15°23.823'N	105°32.737'E	September 2019	0.0	903
M04	Khone Fall, Stung Treng, Cambodia	13°34.815'N	106°0.178'E	August 2019	0.0	646
M05	Kampong Cham, Cambodia	$12^{\circ}0.442^{\prime}N$	105°28.461'E	June 2019	0.0	417
M06	Dong Thap, Vietnam	10°24.514'N	105°36.764'E	May 2019	0.0	143
M07	Tien Giang, Vietnam	10°19.488'N	106°0.396'E	May 2018	0.1	90
M08	Chau Thanh, Ben Tre, Vietnam	10°18.699'N	106°26.909'E	April 2018	1.0	39
M09	Binh Dai, Ben Tre, Vietnam	10°16.266'N	106°31.494'E	April 2018	5.9	29
M10	Binh Dai, Ben Tre, Vietnam	10°15.048'N	106°37.698'E	April 2018	10.7	18
M11	Binh Dai, Ben Tre, Vietnam	$10^{\circ}12.114^{\prime}N$	106°44.166'E	April 2018	16.6	5
M12	Binh Dai, Ben Tre, Vietnam	10°10.703'N	106°45.793'E	April 2018	21.3	\overline{c}
M13	Binh Dai, Ben Tre, Vietnam	$10^{\circ}7.254^{\prime}N$	106°48.948'E	April 2018	26.7	$\mathbf{0}$
M14	Vung Tau, Vietnam	10°19.188'N	107°1.494'E	May 2018	30.0	$\mathbf{0}$
MUN	Mun River, Ubon Ratchathani, Thailand	15°18.133'N	105°28.527'E	September 2019	0.0	869
SK	Sekong River, Stung Treng, Cambodia	13°32.735'N	106°0.875'E	June 2019	0.0	646
TS	Tonle Sap, Kampong Thom, Cambodia	12°31.099'N	104°27.334'E	June 2019	0.0	443
HL	Ham Luong River, Ben Tre, Vietnam	$10^{\circ}10.518^{\prime}N$	106°21.480'E	May 2018	0.1	41
cc	Co Chien River, Ben Tre, Vietnam	$10^{\circ}4.561^{\prime}N$	106°16.693'E	May 2018	0.1	41

Table 1. Water samples collected along the Mekong River and its delta.

Sampling locations and date, in situ salinity and distance from the sea are given. Salinity was measured directly in the field using a Castaway® CTD.

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were confirmed as representative of the salinity gradient (Table 1). Fourteen sampling sites belonged from the Mekong mainstream (M01 to M14), three on tributaries (Mun River— MUN, Sekong River—SK, and Tonle Sap—TS), and two on distributaries in the Mekong Delta (Co Chien River—CC and Ham Luong River–HL, in the Tien River), were sampled between April 2018 and September 2019.

Sampling was authorized in Vietnam by the Fisheries Department (Ben Tre Province) for restricted areas. In Cambodia and Thailand, the authors were directly involved in the study, covering the permission required.

Water sampling and analyses

At each sampling site, two replicates of 50 ml surface water were filtered through Minisart (\mathbb{R}) cellulose acetate filters (pore size 0.2 μm, to remove suspended solids) into polypropylene tubes previously rinsed with pure nitric acid and MilliQ water. Each sample was fixed with 0.2 ml of 0.2% ultra-pure nitric acid. Salinity was measured directly in the field at the same time as the sampling using a Castaway \mathbb{R} CTD probe.

To cross-validate the analytical methods, one of each replicate was analyzed in different laboratories. Two types of analysis were carried out on each replicate: trace element concentrations and the strontium isotope ratio (${}^{87}Sr/{}^{86}Sr$). The trace element concentrations (${}^{7}Li, {}^{24}Mg,$ ${}^{31}P, {}^{55}Mn, {}^{66}Zn, {}^{85}Rb, {}^{88}Sr, {}^{118}Sn, {}^{138}Ba$ and ${}^{208}Pb)$ of one replicate from each sampling site were analyzed at the AETE-ISO OREME laboratory (*Analyse des Eléments en Trace dans l'Environnement & ISOtopes–Observatoire de REcherche Méditerranéen de l'Environnement*, University of Montpellier, Montpellier, France) using solution-based inductively coupled plasma mass spectrometry (ICP-MS). Then, the same replicates were analyzed at the SARM laboratory (*Service d'Analyse des Roches et des Minéraux—Centre de Recherches Pétrographiques et Géochimiques*, Nancy, France) to measure Sr isotopes (87Sr and 86Sr) using multi-collector inductively coupled plasma mass spectrometry (MC-ICP-MS). The other replicates were analyzed at IES (Institute of Earth Science, Academia Sinica, Taiwan) to measure both the trace element concentrations (⁷Li, ¹¹B, ²³Na, ²⁴Mg, ²⁷Al, ²⁸Si, ³⁹K, ⁴⁰Ca, ⁵⁵Mn, ⁶³Cu, ⁶⁶Zn, ⁸⁸Sr, ¹³⁸Ba, ²⁰⁸Pb, and 238 U), using high resolution inductively coupled plasma mass spectrometry (HR-ICP-MS), and the Sr isotope ratio, using high-resolution multiple collector inductively-coupled plasma mass spectrometry (HR-MC-ICP-MS). A summary of the analytical procedures is provided in (S1 Fig).

The AETE-ISO OREME laboratory measured trace element concentrations using solutionbased ICP-MS, (7700x; Agilent, Santa Clara, CA, USA) as described in Tran et al. [47]. Samples were filtered with 0.22 mm mesh size to remove suspended particles and diluted by a factor between two and four, depending on their measured salinity, using milli-Q water to ensure that the concentrations of trace elements to be detected were within equipment's measurement limits. Indium (115 In) and Bismuth (209 Bi) were used as internal standards. Concentrations were determined by external calibration using multi-element standard solutions with concentrations in the range 0.25–5 ppb. Polyatomic interference was limited by keeping the oxide level below 1%. The certified material SLRS-6 (National Research Council Canada, Ottawa, ON, Canada) was used as standard, and analytical precisions were generally between 1 and 3% relative standard deviation (RSD).

The SARM laboratory measured the Sr isotope ratios using thermal ionization mass spectrometry (TIMS, ThermoFisher Scientific TRITON Plus). Water samples were evaporated and 1.5–2.0 μg of Sr was loaded onto chromatographic separation columns with a Sr Spec specific resin (modified after Pin et al. [50]) for separation. The Sr was then loaded onto rhenium filaments and measured with a ThermoFisher Scientific TRITON Plus. The strontium isotopic ratios are expressed as ${}^{87}Sr/{}^{86}Sr$ and were normalized to the stable ${}^{86}Sr/{}^{88}Sr$ ratio of 0.1194 to correct for instrumental mass fractionation. Instrument accuracy was assessed by repeated analyses of the standard reference material NIST SRM 987 (National Institute of Standards and Technology, USA), and the blank values were below 300 pg.

The IES laboratory measured the trace element concentrations using HR-ICP-MS (ThermoFisher Scientific, Element XR). As there was a wide range of salinities in the water samples, thus causing matrix effects, the saline waters collected from the estuary and coastal areas were diluted to Na concentration of 10 ppm (around 0.03 salinity) by adding $0.1N$ HNO₃. The freshwater samples were analysed directly. Six in-house standards were prepared with highpurity single element products (high purity standard, U.S.) in a series of different element concentrations and these standards were used to establish calibration curves. A total of 15 trace element concentrations were measured. The analysis of SRM 1640a (the reference material for trace elements in natural water, National Institute of Standards and Technology, USA) resulted in trace element concentrations with an RSD compared to the certified values of better than 10% for all trace elements except Si, Ba and Mn (data in S1 Table). Repeated analysis of SRM 1640a gave a measurement precision better than 0.35 (1 standard deviation - 1SD) in all cases.

The IES laboratory measured the Sr isotope ratios using HR-MC-ICP-MS (ThermoFisher Scientific, Neptune Plus). The Sr element was purified using the procedure developed by Huang et al. [51] and Huang and You [52]. Water samples were passed through Sr resin (EichromTM) and Sr was then eluted with 0.05N HNO₃. The elutant was slowly heated on a hot plate until completely dry. The dried samples were re-dissolved in 0.1M HCl and then diluted to a consistent Sr concentration of 10 ppb for isotope analyses. The blank was less than 5 pg for Sr and the whole procedure with the IAPSO Seawater Standard revealed 100%

recovery of Sr in long-term tests. The sample standard bracketing method was applied to determine the Sr isotope value of the water samples, and the certified international standard, SRM 987 was used as a reference. IAPSO Seawater Standard was also treated as an unknown sample for quality and quantity control, yielding high accuracy (RSD 0.001%) and precision $(1SD 4.2 \times 10^{-6}).$

Otolith sampling and microchemical analysis

A total of 27 individual *Pangasius krempfi* (47.8 to 84.5 cm total length) were collected from March-August 2017 (24 individuals) and in April 2018 (3 individuals) in the Mekong Delta. The fish were collected on landing or in markets in Binh Dai city (18 individuals, coded BD01 to BD18), My Tho city (7 individuals, MT01 to MT07) and Dinh An city (2 individuals, DA01 and DA02) (Fig 1). Samples were collected randomly to get a wide range of lengths and weights. The left (the larger) otolith (*lapillus*) was extracted using plastic forceps from a sagittal head section, cleaned in Milli-Q water, and stored dry in 1.5 ml microtubes. As the right otolith had been used in another study [47], the age and Sr/Ca ratio of the individuals were already known.

The otoliths were prepared at the University of Science, VNU-HCM (Vietnam). They were first photographed whole under reflected light, then embedded in epoxy resin (Creative Life Vietnam, Vietnam), sliced transversally using a Buehler IsoMet Low Speed cutting machine (Buehler, Germany) and polished until the core was reached (S2 Fig). Sections were then attached to a clean microscope slide for further processing. Sr isotopes were measured at the AETE-ISO OREME lab, Montpellier, France) using a Thermo Finnigan Neptune⁺ multicollector inductively-coupled plasma mass spectrometer (MC-ICP-MS) coupled to a 193 nm Analyte G2 Excimer Laser Ablation system (Photon Machines Inc). Spots were ablated at 65 μm diameter at 5 μ m/s. The pulse rate was 7 Hz and the energy density of the beam was 6.0 J/cm². A pre-ablation transect (spot size $85 \mu m$, speed $15 \mu m/s$) was used to clean the sample surface before analysis. A typical analysis comprised a 30 s background measurement and the ablation period required for a transect from the core to the edge. Corrections for Kr and Rb interference and mass bias followed routine procedures using known isotopic ratios [53]. No corrections were applied for interference from doubly charged REE, Ca argides and Ca dimers and polyatomic interference, since numerous studies have shown that Ca argides and dimers have no significant effect on Sr isotopic data using MC-ICP-MS [54]. Ca-P-O and doubly charged ions are also insignificant for material with a high Sr (Sr *>* 300ppm) and low REE contents. Krypton interference $(^{84,86}$ Kr on 84,86 Sr) from the argon tank was corrected for by measuring the background level before the analysis, and then by subtracting the background from the data. Rubidium interference (⁸⁷Rb on ⁸⁷Sr) was corrected for by monitoring ⁸⁵Rb and subtracting the signal at mass 87 amu assuming a natural ${}^{85}Rb/{}^{87}Rb$ of 2.59262 [55]. The ${}^{85}Rb/{}^{87}Rb$ ratio was corrected for mass bias using the mass discrimination factor calculated from Sr, using an exponential law and a natural ${}^{88}Sr/{}^{86}Sr = 8.375209$, and assuming no differential mass discrimination between Sr and Rb. The accuracy and long-term reproducibility of the measurements were checked by analyzing the pressed pellet MACS3 from the USGS (United States Geological Survey, Reston, VA, USA; reference value of ${}^{87}Sr/{}^{86}Sr = 0.70755$) and in-house reference material (Atlantic shell with a reference value of ${}^{87}Sr/{}^{86}Sr = 0.70918$).

Statistical analyses

All data were tested for normality using the Shapiro-Wilk test [56], and for homoscedasticity using Levene's test prior to using parametric or non-parametric tests. Due to the absence of

normality in trace element concentrations, differences in measurement precision and accuracy between the laboratories were compared using the Wilcoxon test.

The ${}^{86}Sr/{}^{87}Sr$ values of the two replicate sets of water samples were normally distributed and were compared using a paired Student's *t-*test. The relationship between these values was calculated using a general linear regression. The correlation between ${}^{87}Sr/{}^{86}Sr$ and water salinity, the distance from the sea and 1/Sr were tested using a Pearson test, and the relationships were then tested using linear or logarithmic regressions based on the R^2 value.

For each sampling site, the mean of the concentrations for each trace element obtained by the two laboratories was used for subsequent analyses. The concentrations of trace elements that were only measured by one laboratory were used directly. Correlations between trace element concentrations and salinity in water were tested by using a Pearson correlation test for Mg and Spearman correlation test for other trace elements. Concentrations of ²⁸Si, ⁶³Cu, ¹³⁸Ba and 238U were only detected in freshwater and were not used to test correlations with salinity. Among these trace elements, only the Sr/Ca and Ba/Ca ratios were tested to determine if they were able identify differences in water chemistry between locations, as mentioned by Fukushima et al. [11].

The ⁸⁷Sr/⁸⁶Sr profiles were calculated for each otolith using moving means of three adjacent values at a given distance from the core to the edge. ANOVA was used to test for differences in otolith 87Sr/86Sr between fish sampled at different locations (Binh Dai, My Tho and Tien Giang), or between years (fish collected in 2017 and 2018). To test for similarity of $87\$ {S}r/86Sr profiles between individuals and for distinct groups of *P. krempfi* based on ⁸⁷Sr/⁸⁶Sr profile, a hierarchical cluster analysis (HCA) was used based on the Euclidean distances between 87 Sr/ 86 Sr profiles and Ward's clustering method ("ade4" package in R). The number of clusters was determined based on the performance of hierarchical cluster analysis. ⁸⁷Sr/⁸⁶Sr profiles were also compared by direct observation to the Sr/Ca ratio already available from the right otolith in a previous publication [47]. All statistical analyses were performed in R [57].

Ethics statement

All the fish samples used in this study were collected dead at fish landings from fishermen or in markets. Thus, all fish were dead at sampling and no ethical approval is required. The sampling in Vietnam was hosted by University of Science–VNUHCM with the permission from Ben Tre Fishery Department and Immigration Department (number 317/ KHTN-QHQT-QLDA on 12/04/2018). Other sampling locations in Cambodia and Thailand

are not included in protected areas or restricted areas and there is no requirement for official permission.

Results

Strontium isotope ratios (87Sr/86Sr) and trace element concentrations in the water

The differences in 87 Sr/ 86 Sr between SARM France and IES Taiwan were well within 2 σ , being less than 0.000022 (Table 2) and were not statistically significant (paired Student's *t-*test, t = 0.347, df = 18, p-value = 0.732). The relationship between ${}^{87}Sr/{}^{86}Sr$ in water analyzed at the two laboratories was linear and highly significant (Fig 2; $R^2 = 0.999$, slope = 0.999). For each sampling site, the mean of the ${}^{87}Sr/{}^{86}Sr$ from the two laboratories was used for subsequent analyses.

The ${}^{87}Sr/{}^{86}Sr$ for water samples from the Mekong River ranged from 0.708934 ± 20 \times 10⁻⁶ to 0.713943 \pm 11 × 10⁻⁶ (Table 2). The ⁸⁷Sr/⁸⁶Sr for the Mekong mainstream ranged from

Table 2. 87Sr/86Sr and Sr concentrations.

 ${}^{87}\text{Sr/}{}^{86}\text{Sr}$ and Sr concentrations (mean \pm S.D.) measured by AETE-ISO OREME and SARM (France) compared with those measured by IES (Taiwan).

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0.709180 ± 6 × 10⁻⁶ at the sea to 0.710750 ± 7 × 10⁻⁶ upstream. In the tributaries, the ⁸⁷Sr/⁸⁶Sr were lower in the Tonle Sap Lake (0.708934 \pm 20 \times 10⁻⁶) and higher in the Sekong River $(0.713943 \pm 11 \times 10^{-6})$ which was higher than any of the ratios in the Mekong mainstream. The other tributary (Mun River–MUN) and the distributaries (Ham Luong—HL and Co Chien River—CC) had ⁸⁷Sr/⁸⁶Sr of 0.710351 ± 2 × 10⁻⁶, 0.710488 ± 11 × 10⁻⁶ and $0.710486 \pm 8 \times 10^{-6}$ respectively, similar to the upper part of Mekong mainstream.

Fig 2. Relationship between 87Sr/86Sr determined by the two laboratories (AETE-ISO OREME and IES). Linear regression between the ⁸⁷Sr/⁸⁶Sr in water samples reported by IES (Taiwan) versus those reported by SARM (France). Black circles indicate sites along the Mekong mainstream, grey triangles indicate sites on tributaries and distributaries.

The ${}^{87}Sr/{}^{86}Sr$ was negatively correlated with salinity (Pearson correlation = - 0.523, pvalue = 0.021). The ⁸⁷Sr/⁸⁶Sr was always much higher in freshwater (Fig 3A) while there was a significant logarithmic regression between ${}^{87}Sr/{}^{86}Sr$ and salinity in more saline waters (R^2 = 0.922, Fig 3B). The ${}^{87}Sr/{}^{86}Sr$ was non-significantly correlated with the distance from the sampling location to the sea (Pearson correlation = 0.449 , p-value = 0.053), but there was a significant logarithmic regression between these two variables along the Mekong mainstream (R^2 = 0.757, Fig 4). The ${}^{87}Sr/{}^{86}Sr$ was divided into five groups depending on their value, salinity and distance from the sea: a saline and brackish water region in Mekong Delta (sites M09 to M14), a freshwater region in the Mekong Delta (site M05 to M08, CC, and HL), upstream regions (sites M01 to M04 and Mun), the Sekong River and Tonle Sap Lake. The relationship between concentrations of ⁸⁷Sr/⁸⁶Sr and 1/Sr in the water was more exponential ($R^2 = 0.408$), but not significant, when all sites were included, the relationship was exponential only if the tributaries and distributaries were excluded ($R^2 = 0.923$, Fig 5), and the correlation between ${}^{87}Sr/{}^{86}Sr$ and $1/Sr$ was significant (Pearson correlation = 0.592, p-value = 0.007).

The concentrations of ${}^{31}P$, ${}^{28}Si$, ${}^{63}Cu$, and ${}^{238}U$ in brackish and marine water samples were below the limit of detection, and were therefore neglected in further analyses. The trace element concentrations did not differ between two analytical methods used in AETE-ISO OREME and IES (Table 3, Wilcoxon test, p-value > 0.05). Most of the trace element concentrations (⁷Li, ¹¹B, 23 Na, 24 Mg, 39 K, 40 Ca, 66 Zn, 85 Rb and 88 Sr) were significantly positively correlated with water salinity (Spearman's correlation, p-value $<$ 0.05), when ²⁷Al, ³¹P, ⁵⁵Mn, ¹¹⁸Sn, ¹³⁸Ba, ²⁰⁸Pb showed no significant correlation (Spearman's correlation and Pearson correlation, pvalue > 0.05) and ²⁸Si, ⁶³Cu, ¹³⁸Ba, ²³⁸U only detected in freshwater region (Table 3). The value of water salinity and trace element concentrations are available in S2 Table.

Correlations between trace element concentrations and salinity were highly significant and positive for ⁷Li, ¹¹B, ²³Na, ²⁴Mg, ³⁹K, ⁴⁰Ca, ⁸⁵Rb and ⁸⁸Sr, whereas for ⁶⁶Zn the correlation was less significant, and no correlation was found for ¹⁹Sn, ²⁷Al ⁵⁵Mn, ¹³⁸Ba and ²⁰⁸Pb (Table 3 and S3 Fig). The relationships between Sr/Ca ratios and Ba/Ca ratios were quite close in the

Fig 3. Water ⁸⁷Sr/⁸⁶Sr versus salinity along the Mekong River. Water ⁸⁷Sr/⁸⁶Sr versus salinity along the Mekong mainstream (dark circles) and tributaries / distributaries (grey triangles); (A) for all sampling locations, and (B) only in saline waters (Mekong Delta) with a logarithmic regression. Labels at each point refer to the sampling sites in Fig 1 and Table 1.

Fig 4. Water 87Sr/86Sr ratio versus distance from the sea along the Mekong River. Water 87Sr/86Sr versus distance from the sea along the Mekong mainstream. Distances were estimated using Google Earth Pro. Labels at each point refer to the sampling sites in Fig 1 and Table 1.

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freshwater parts of the Mekong mainstream compared to values in tributaries (MUN, SK and TS) (Fig 6). Other trace elements showed no or only slight differences among sites indicating their limitations as biotracers in the Mekong mainstream.

Strontium isotope ratios in *Pangasius krempfi* **otoliths**

All the otoliths showed similar ${}^{87}Sr/{}^{86}Sr$ patterns over the lifetime of the fish (Fig 7 and S3 Table): high values at hatching and during the early days of life $(0.7106 \pm 0.0003$ at the core region ~ 30 μm width), corresponding to the ⁸⁷Sr/⁸⁶Sr of waters collected upstream along the Laos–Thailand border and through Cambodia (sites M01 to M04). The ratios then decreased to a more or less variable plateau (0.70928 \pm 7.4 \times 10⁻⁶), corresponding to the ⁸⁷Sr/⁸⁶Sr of the

Fig 5. Water 87Sr/86Sr against 1/Sr. Water 87Sr/86Sr versus 1/Sr along the Mekong River; (A) with and (B) without tributaries and distributaries. Labels at each point refer to the sampling sites in Fig 1 and Table 1. Black circles indicate sites on the Mekong mainstream, grey triangles indicate sites belonging on tributaries and distributaries.

Table 3. Trace element concentrations measured by the two laboratories (AETE-ISO OREME and IES).

Trace element concentrations measured by the two laboratories (AETE-ISO OREME and IES), with p-value (Wilcoxon test) for differences between the two laboratories and correlation between trace element concentrations in the water vs. salinity (Spearman's correlation and Pearson correlation). * significant correlation (p > 0.05).

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brackish waters (site M09 at Binh Dai). Otolith ⁸⁷Sr/⁸⁶Sr never showed values corresponding to water samples collected from Tonle Sap Lake or the Sekong River. There was no difference in ⁸⁷Sr/⁸⁶Sr in the otoliths in fish sampled at locations Binh Dai (BD01 –BD18), My Tho

Fig 6. Ba/Ca ratios versus Sr/Ca ratios in the Mekong mainstream and tributaries. Labels at each point refer to the sampling sites in Fig 1 and Table 1. Black letters correspond to sites on the Mekong mainstream, red letters to sites on tributaries, and blue letters to sites on distributaries.

(MT01 –MT07) and Dinh An (DA01 and DA02) in the Mekong Delta, or between years (BD16 –BD18 were collected in 2018 and the other samples were collected in 2017) (ANOVA, p-value = 0.47 and 0.75, respectively). Comparing the ratios 15 μm from the core with those in the water showed that all individuals were born in the Mekong mainstream and not in tributaries. Nevertheless, the differences in ${}^{87}Sr/{}^{86}Sr$ (Fig 7) at the core pointed to different spawning grounds and different behaviors during the early life stages. The lowest ${}^{87}Sr/{}^{86}Sr$ in the otoliths was lower than the lowest ${}^{87}Sr/{}^{86}Sr$ in saline water that was collected at Vung Tau (site M14), suggesting that the fish may move to saline areas other than our sampling locations once they reach brackish water.

The hierarchical cluster analysis of the general shape of otolith ⁸⁷Sr/⁸⁶Sr profiles produced three main clusters (Fig 7A). One cluster had lower ${}^{87}Sr/{}^{86}Sr$ at the core (1/3 of individuals), close to the ratios at sites M06 and M07 in the freshwater part of the Mekong Delta that fell to reach a plateau corresponding to the ${}^{87}Sr/{}^{86}Sr$ ratios in the saline waters at sites M10 to M14 in the part of the Mekong Delta where the water is brackish (Fig 7B). The second cluster had medium ${}^{87}Sr/{}^{86}Sr$ ratios at the core (1/3 of individuals), corresponding to the water ${}^{87}Sr/{}^{86}Sr$ encountered upstream in Thailand and Cambodia (sites M01 to M04), and variable ratios after reaching brackish water (sites M10 to M14, Fig 7C). In this cluster, after the fish had reached

the Delta, the ${}^{87}Sr/{}^{86}Sr$ sometimes peaked close to the ratio found in the slightly brackish water at M08. The last cluster had higher, variable ratios at the core, higher than any recorded in the water in the Mekong mainstream (1/3 of individuals), falling to a plateau while the fish were in the brackish waters of the delta (ratios close to those at sites M11 to M14, Fig 7D).

In comparison, otolith Sr/Ca profiles measured in our previous study [47] had a very low ratio at the core, a rapid increase to higher ratios, and greater variations during the rest of the lifetime of the fish (Fig 8). While the ${}^{87}Sr/{}^{86}Sr$ profiles showed more variance close to the core, Sr/Ca profiles showed much more variation during the period of their life in brackish waters. Although the Sr/Ca ratio patterns along the otolith transect were the opposite of ${}^{87}Sr/{}^{86}Sr$ profiles, they showed the same migration patterns in the Mekong Delta with several peaks.

The patterns of all individuals suggest that they moved from freshwater at birth, down the Mekong mainstream to coastal, saline regions, during their lifetime. A large part of their lifetime was spent in the brackish and marine water regions in the Mekong Delta, where they were caught. After hatching, they spent only a few months, much less than 1 year (Fig 8), in the freshwaters of the Mekong in Thailand and Cambodia, before moving to the Mekong Delta. None of the individuals showed any evidence of having swum in the Mekong tributaries, such as the Mun River, the Sekong River, or Tonle Sap Lake.

Discussion

In transboundary rivers, such as the Mekong River, migratory fish are threatened by climate changes, pollution, saline intrusions and the construction of dams $[8-10]$. Studying the movement patterns of migratory fish species is thus essential for fishery management and fish

Fig 8. Comparison of 87Sr/86Sr profiles and Sr/Ca profiles for the same individuals. Comparisons of 87Sr/86Sr profiles (solid line) and Sr/Ca profiles (dotted line) for four *P*. *krempfi* individuals. Profiles are shown from the core to the edge of the transverse otolith section. Vertical dotted lines mark the end of each year based on the position of the annual growth increments.

conservation across the Mekong countries [58, 59]. In this study, strontium isotope ratios $({}^{87}\text{Sr}){}^{86}\text{Sr})$ in the water and in fish otoliths were used to identify the migration pathways of one anadromous catfish, *Pangasius krempfi*, from the delta up to the Mekong River. The Sr isotope ratios and trace element concentrations in waters along the river and in some main tributaries were mapped in four countries (Vietnam, Cambodia, Thailand and Laos), to provide the data needed to estimate the origin and migration pathways of fish caught in the Mekong Delta.

Strontium isotope ratios (87Sr/86Sr) and trace element concentrations in the water along the Mekong River

This is the first study mapping the ⁸⁷Sr/⁸⁶Sr ratios along the Mekong River, from the Laos-Thailand border to the delta and the sea, and in the main tributaries (Fig 9). The ${}^{87}Sr/{}^{86}Sr$ at the upper Mekong (0.71064) is below the average for big rivers (0.71107), reported by Peucker-Ehrenbrink et al. [60], and below the average of the Himalayan-Tibetan rivers (0.7127) reported by Richter et al. [61] but similar to that of the Amazon River (0.710–0.712) reported by Duponchelle et al. [40]. The ${}^{87}Sr/{}^{86}Sr$ at the Mekong mouth (0.70919) corresponds to the global average ⁸⁷Sr/⁸⁶Sr for seawater (0.70918, [38]). The strontium concentration and Sr isotope

composition of rivers are largely induced by a mixture of strontium derived from the type of rocks encountered and their age $[62, 63]$. Therefore, the ${}^{87}Sr/{}^{86}Sr$ values measured from the upper Mekong to the mouth could reflect a mixture of sources in different geological regions along the Mekong River, including (1) Himalayan bedrocks in the upper Mekong, (2) the Annamite Range along the Laos–Cambodia—Vietnam border, and (3) primitive volcanic sources, or other tributaries in the Mekong Delta [64, 65]. Seawater, a source of strontium isotopes, could also affect the ${}^{87}Sr/{}^{86}Sr$ in the estuarine regions, as reported in a previous study [66].

The sites sampled in the mainstream (M01, M05) and in the delta (M06 to M14, CC, and HL) are influenced by quaternary sediments, while M02 and M03 are affected by shallow shelf-marine dating from the Cretaceous or Jurassic periods [65]. Concerning the tributaries, Tonle Sap (TS) had a much lower ${}^{87}Sr/{}^{86}Sr$ (0.708934) than M05, the closest Mekong mainstream site (0.710352), even if it belongs to the Phnom Penh Basin region [65]. This may be due to the different sources that affect the sites: Tonle Sap Lake is influenced by ⁸⁷Sr/⁸⁶Sr from the bedrock [65]. On the other hand, the Sekong River (SK) has a much lower ${}^{87}Sr/{}^{86}Sr$ (0.713943) than the nearby mainstream site (M04). The M04 site is influenced by ${}^{87}Sr/{}^{86}Sr$ ratio from the Mun River (Jurassic or Cretaceous Khorat plateau red-bed), while the Sekong River is influenced by a Permo-Triassic volcano-plutonic rock belt [65].

Although the ${}^{87}Sr/{}^{86}Sr$ in surface water reflects bedrock geology and remained constant across seasons and years [37, 67, 68], the ratios are possibly affected by groundwater [69] and rainfall [70–72], and by seawater, depending on the locations. The only slight variation in 87 Sr/ 86 Sr in saline waters limits the usefulness of this ratio to discriminate estuarine and marine waters [29]. In the Mekong Delta, the ${}^{87}Sr/{}^{86}Sr$ showed a logarithmic relationship with salinity from 5 to 30, but the variation was small (0.709174 to 0.709284) due to a short distance between the sites along the salinity gradient. However, the ${}^{87}Sr/{}^{86}Sr$ depended much more on the geology of the upstream origins than on salinity, making it possible to use this ratio to study fish movements in the fresh to estuarine waters along the Mekong River.

Among all the trace elements, the Ba/Ca ratio vs. Sr/Ca ratio was used to distinguish the different areas along the Mekong River because they both provide useable evidence of the origins of the water between the mainstream and tributaries or the different geological regions along the Mekong River [11]. The Ba/Ca ratio *vs*. Sr/Ca ratio in the different tributaries revealed differences that could be used to infer the origin of the water, and, consequently, the movements of fish between them [11, 73, 74]. The differences were particularly marked between the Mekong mainstream and its tributaries [11]. While Humston et al. [73] and Strohm et al. [74] showed that there was strong linear relationship between Sr/Ca and Ba/Ca in otoliths and water in various tributaries in the USA, the changes in otolith Sr/Ca and Ba/Ca did not reflect the seasonal river flow [37]. In the present study, the potential of comparing Sr/Ca and Ba/Ca between water and otoliths to determine the origin of fish origin remains questionable. The $87\text{Sr}/86\text{Sr}$ in the otoliths could be a better indicator and consequently more useful for discerning the natal origin and migration pathways of fish.

Origin and movement of *Pangasius krempfi* **along the Mekong River**

The composition of trace elements in fish otoliths is influenced by different physiological processes that regulate their incorporation [18, 20, 22–24, 75, 76], and may thus limit its usefulness for inferring fish life history. However, some trace element ratios in the otolith, including the Sr/Ca or Sr isotope ratios, could reflect the ambient water conditions such as temperature and salinity [21, 47, 77] that are not strongly affected by biologically factors [28, 29] or seasonalannual flows [78]. Therefore, otolith Sr/Ca and ${}^{87}Sr/{}^{86}Sr$ could be used as evidence to infer past environments the fish encountered during its lifetime.

Sr/Ca profiles of *Pangasius krempfi* otoliths from our previous study [47] had a very low ratio at the core, followed by a rapid increase, and high values during the rest of the fish's life (Fig 8). The fact all individuals were born in freshwater environments was demonstrated, but without identifying the exact origin of the fish. The fact the fish remained in brackish water environments for the rest of their lifetime was also demonstrated. The aim of the present study was to more precisely identify the natal origin using the $87\text{Sr}/86\text{Sr}$ ratio along the Mekong River. Unlike the Sr/ Ca, ${}^{87}Sr/{}^{86}Sr$ was the highest at the core, with more variance. These ${}^{87}Sr/{}^{86}Sr$ confirmed the natal origin in the freshwater environments of the Mekong River. The 87Sr/86Sr support the *P*. *krempfi* migration patterns between freshwater and brackish water in the Mekong Basin over their lifetime that we inferred from the Sr/Ca ratios in our previous study [14, 44, 45].

Combining otolith ${}^{87}Sr/{}^{86}Sr$ and Sr/Ca could provide the best evidence for the natal origin and migration behavior of anadromous species [79]. While otolith ${}^{87}Sr/{}^{86}Sr$ profiles showed the *P*. *krempfi* movements after hatching in freshwater regions, the otolith Sr/Ca profiles indicated possible movements and migrations between different saline environments. The present study demonstrated that *P*. *krempfi* is an anadromous fish, hatches in freshwater regions in the Mekong mainstream along the Laos–Thailand border or in Cambodia, and then migrates at early stages to the Mekong Delta, and spends most of its lifetime in brackish and marine environments. All *P*. *krempfi* caught in the Mekong Delta were born in the upper part of the Mekong mainstream, having migrated more than 1,500 km in the Mekong River mainstream, with cross-border movements at least between Laos-Thailand, Cambodia, and Vietnam.

Comparing the 87Sr/86Sr in the core with those in the water, indicated that *P*. *krempfi* are born in the Mekong mainstream and never in the tributaries. There could be spawning grounds along Laos–Thailand border, or in Cambodia, or even in the upper part of the river in Vietnam. Our results point to several spawning locations, but these were not precisely identified, because the ${}^{87}Sr/{}^{86}Sr$ in the otoliths corresponded to those in different locations in the upper part of the stretch along the Laos-Thailand border. The cluster analysis separated three contingents with distinct ⁸⁷Sr/⁸⁶Sr profiles in their otoliths. One contingent hatched around Kampong Cham (M05, Cambodia), Cao Lanh (M06, Vietnam), Cai Be (M07, Vietnam) or even in the upper part of distributaries (CC and HL), another contingent hatched in the upper region from Khone Fall (Cambodia) to Nong Khai (Thailand). The third contingent had a higher ${}^{87}Sr/{}^{86}Sr$ at the otolith core than that of any water collected in this study with the exception of water in the Sekong River, where the ratio was even higher. These fish could have hatched at upper Nong Khai (Thailand), near Vientiane the capital of Laos.

The main conclusion of the present study is that *P*. *krempfi* does not have only one natal origin. Although the spawning habitat requirements and biology of this species are not fully known, the previous hypothesis was that the species spawns in the Mekong mainstream near the rapids and pool systems around the Khone Falls [43, 44, 80, 81]. The spawning season is assumed to occur between June and August, when fishermen observed specimens with eggs in Laos, but has never been reported in other periods [81]. By quantifying the genetic diversity and structure of *P*. *krempfi* in the Mekong Delta, Duong and Nguyen [82] reported that there were different spawning groups of this species. We, therefore, predicted that Khone Falls is not this species only spawning ground and that the spawning grounds are probably rapids along the Mekong mainstream from Vietnam and Cambodia, to the Laos-Thailand border: spawning grounds could also exist in upper Cao Lanh (Vietnam), from Kandal province to Kampong Cham (Cambodia) [80], at Khone Fall (Laos), or along the Laos–Thailand border of the Mekong River. *P*. *krempfi* breeds in different locations along the Mekong River, but never in tributaries, brackish or saline environments.

After hatching in freshwater, all *P*. *krempfi* migrated to the Mekong Delta and settled in the estuarine area, but the timing of leaving the hatching areas differed between the contingents.

Although some individuals moved directly to the estuary after hatching, this could be due to water currents flushing them before they had grown big enough to swim against the current [44]. Other individuals appeared to move around in their area of origin or freshwater regions, with slightly different ⁸⁷Sr/⁸⁶Sr, before migrating to the Delta. Chan et al. [80] reported twomonth-old juveniles at Mukdahan (Thailand), consistent with the delayed migration recorded for some individuals in our study. Migration to the Mekong Delta probably depends on natural food sources, mainly composed of their favorite leaves and benthic food [44, 81], and subsequently on the abundance of riparian vegetation in the estuarine environments.

The purpose of the migration could be either moving from suitable reproduction habitats to more suitable feeding habitats for growth, or avoiding predators and adverse abiotic conditions upstream [83]. Poulsen et al. [14] proposed that the *P*. *krempfi* caught in the Mekong Delta belong to the upper parts of the Mekong system, and indicated possible migration routes. The marine residence and homing behavior of *P*. *krempfi* were observed by Baran et al. [5] and Hogan et al. [44], who reported that these fish are able to come return to their natal origin for spawning [44]. Two of the contingents settled in the estuarine habitats, but the fish in the other contingents moved frequently between estuarine water and slightly brackish water upstream (M08). These results agree with those of Hogan et al. [44] and Tran et al. [47], who reported several migratory clusters in a single population. During its life in estuarine waters, *P*. *krempfi* can move around different areas, but only occasionally to freshwater. The longest distance traveled between marine and freshwater environments after reaching the Mekong Delta was observed for two individuals who moved 40 km upstream near M08 (My Tho, Tien Giang Province). The other individuals only moved around the brackish and marine environments and did not reach the freshwater areas before capture.

In this study, spawning locations were predicted but not precisely identified, because the 87 Sr/ 86 Sr in the water overlapped in some locations, probably due to mixing between different sources. A larger-scale study of values, sources, and changes in water ${}^{87}Sr/{}^{86}Sr$ in the Mekong River could provide more data for applications of otolith chemistry in this area. The present study was also limited by studying only *P*. *krempfi* captured in the Mekong Delta and no evidence of homing behavior was found because no individual otolith showed a possible return to its freshwater origin during its lifetime. The *P*. *krempfi* analyzed in the present study were either non-homing individuals, or individuals who had not yet reached the reproductive stage, even if the maximum age was 5 years (total length 0.8 m, weight 4.75 kg). Although there have been several studies of the migration patterns of this species based on field observations, fish catches or otolith chemistry [5, 44, 45, 47, 84], the life cycle of *P*. *krempfi* in the Mekong River is still not fully understood. Age at first sexual maturity, age at migration, their behavior during and after spawning remain unknown. Other individuals captured upstream in Thailand and Laos, could provide more information on migrating pathways.

Threats to and conservation of *Pangasius krempfi* **and migratory fish in the Mekong River**

Migratory fish are an important resource for many stakeholders along the Mekong River [45]. Understanding the early life history and the migratory behavior of these fish is crucial for identifying their nursery habitats and their migration pathways for the purpose of conservation and fishery management [85]. This study highlighted the ecological and anthropogenic threats to *P*. *krempfi*, as it is an obligate anadromous species. *P*. *krempfi* has been shown to be highly vulnerable to a combination of stresses including climate change, habitat loss, dam construction, pollution, and overexploitation [5, 12, 86, 87]. Rising sea levels, the combination of severe drought, saltwater intrusion in the coastal Mekong Delta in recent years and the construction of hydropower dams

have reduced feeding habitats for the species at the adult stage [88–91]. Changes in rainfall patterns could also affect the water levels required for breeding and the water levels in migration pathways which, in turn, could affect breeding success [91–93]. Hydropower dams along the Mekong mainstream reduce the availability of rapids and pools, which are possible breeding habitats of this species. Future dams will increase the barriers along the migration pathways of the species, especially for breeding, and will isolate habitats of the required quality [92]. Ultimately, these threats will increase the mortality of adults and juveniles, compromising recruitment, and reducing population fitness [12, 13, 94]. These threats will not only impact anadromous fish, such as *P*. *krempfi*, but also diadromous and potamodromous fish and even non-migratory fish.

Dams in the Mekong basin have more impact on migratory fish species than upstream dams in China, and mainstream dams have a greater environmental impact than dams built on tributaries [12]. If hydropower dams are constructed on the mainstream Mekong in Cambodia and southern Laos (like the Sambor, Stung Treng, Don Sahong, Lat Sua and Ban Koum dams), they will be a major threat to fish communities in the Mekong River, especially anadromous species, such as *P*. *krempfi*, which migrate upstream to complete their life cycle. Dams further upstream and on tributaries will also have local impacts on short-range migratory fish [11, 12]. However, the effects of dam construction on *P*. *krempfi* populations need to be considered carefully as the wide range of breeding locations could reduce the impact of habitat disconnection on the populations. The lower *P*. *krempfi* contingent born around Phnom Penh and the Mekong Delta are hypothesized to not be directly affected in the mainstream because until now, no dam has been constructed between the delta and the sea. It is possible that the upper contingents may adapt to the dams by switching to lower spawning grounds to avoid migrating up or down from the dams.

Protection of hatching habitats, nursery grounds, and environmental connectivity along the migration pathways are key issues for fish conservation. Building fishways, which could mitigate the threats posed by dams and protect fish migration routes, ensuring habitat connectivity, and rehabilitating fisheries are possible solutions to cope with dam construction [94, 95]. However, the fish passages must be designed to suit the specific topographic and hydrological conditions of the river, as well as to respect the ecological and biological conditions required at the different life stages of the fish species [12, 95]. Artificial fish passages are therefore unlikely to be an effective mitigation measure in the Mekong River because of a large number of migration patterns, body sizes, shapes and behavior of migratory fish [96]. Finding a balance between environmental, economic, and social demands are key issues for the sustainable development of the Mekong region [12], and nature-based solutions are the best way to protect and conserve the area and the fish communities.

Conclusion

This study demonstrated the use of ${}^{87}Sr/{}^{86}Sr$ ratios in water and fish otoliths to understand the life history cycle of an anadromous catfish caught in the Mekong River. ${}^{87}Sr/{}^{86}Sr$ has never previously been measured in the Mekong River for this purpose. The ⁸⁷Sr/⁸⁶Sr mapped in this study differed significantly between the Mekong mainstream and its main tributaries, and between different geological regions along the Mekong mainstream. The targeted species *Pangasius krempfi*, caught in the Mekong Delta, hatch in freshwater along the Mekong mainstream, from Phnom Penh (Cambodia) to Nong Khai (Thailand), or even further, before settling in the delta. Spawning habitats and migration pathways are threatened by habitat degradation and by the increasing number of hydropower dams along the river. Thus, the conservation of *P*. *krempfi*, as well as other migratory fish in the Mekong River, requires the agreement of and actions by all the stakeholder countries located along the Mekong River.

Supporting information

S1 [Fig](http://www.plosone.org/article/fetchSingleRepresentation.action?uri=info:doi/10.1371/journal.pone.0252769.s001). Summary analysis process for trace element and 87Sr/86Sr in the water. (TIF)

S2 [Fig](http://www.plosone.org/article/fetchSingleRepresentation.action?uri=info:doi/10.1371/journal.pone.0252769.s002). Otolith of *Pangasius krempfi***.** (A) Whole otolith of *Pangasius krempfi*, (B) the illustration for sliced transversally cutting (white bank) and (C) otolith slice with the laser transect from core to edge (dotted line). (TIF)

S3 [Fig](http://www.plosone.org/article/fetchSingleRepresentation.action?uri=info:doi/10.1371/journal.pone.0252769.s003). A. Correlation between trace element concentrations and salinity in water collected along Mekong River and tributaries. B. Correlation between trace element concentrations and salinity in water collected along Mekong River and tributaries (continuous). (TIF)

S1 [Table.](http://www.plosone.org/article/fetchSingleRepresentation.action?uri=info:doi/10.1371/journal.pone.0252769.s004) Accuracy and precision of trace elemental analyses assessed using SRM 1640a reference material. LR represents low resolution, MR medium resolution and HR high resolution. RSD is calculated by the difference between measured and certified values divided by the certified value and expressed in percentage. (DOCX)

S2 [Table.](http://www.plosone.org/article/fetchSingleRepresentation.action?uri=info:doi/10.1371/journal.pone.0252769.s005) Trace element concentrations. Sampling locations, salinity and trace element concentrations measured by AETE-ISO OREME and IES. (XLSX)

S3 [Table.](http://www.plosone.org/article/fetchSingleRepresentation.action?uri=info:doi/10.1371/journal.pone.0252769.s006) 87Sr/86Sr values in otoliths of *Pangasius krempfi***.** Distance from the core to the analyses points and 87Sr/86Sr values in all otoliths of *Pangasius krempfi*. (XLSX)

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References

- **1.** MRC. State of the Basin Report. Executive summary. Phnom Penh; 2003.
- **2.** Liu S, Lu P, Liu D, Jin P. Pinpointing source of Mekong and measuring its length through analysis of satellite imagery and field investigations. Geo-Spatial Information Science. 2007; 10: 51–56. [https://doi.](https://doi.org/10.1007/s11806-007-0011-6) [org/10.1007/s11806-007-0011-6](https://doi.org/10.1007/s11806-007-0011-6)
- **3.** Rainboth WJ, Vidthayanon C, Mai YD. Fishes of the Greater Mekong Ecosystem with Species List and Photographic Atlas. Ann Arbor, MI, USA: Miscellaneous Publications of the Museum of Zoology, University of Michigan; 2012.
- **4.** Coates D, Poeu O, Suntornratana U, Tung NT, Viravong S. Biodiversity and fisheries in the Lower Mekong Basin. Mekong Development Series No. 2. Mekong River Commission. Phnom Penh: Mekong River Commission; 2003.
- **5.** Baran E, Baird IG, Cans G. Fisheries bioecology at the Khone Falls (Mekong River, Southern Laos). WorldFish Center. Penang, Malaysia; 2005.
- **6.** Stone R. Dam-building threatens Mekong fisheries. Science. 2016; 354: 1084–1085. [https://doi.org/10.](https://doi.org/10.1126/science.354.6316.1084) [1126/science.354.6316.1084](https://doi.org/10.1126/science.354.6316.1084) PMID: [27934711](http://www.ncbi.nlm.nih.gov/pubmed/27934711)
- **7.** Kano Y, Dudgeon D, Nam S, Samejima H, Watanabe K, Grudpan C, et al. Impacts of dams and global warming on fish biodiversity in the Indo-Burma hotspot. PLoS ONE. 2016; 11: 1–21. [https://doi.org/10.](https://doi.org/10.1371/journal.pone.0160151) [1371/journal.pone.0160151](https://doi.org/10.1371/journal.pone.0160151) PMID: [27532150](http://www.ncbi.nlm.nih.gov/pubmed/27532150)
- **8.** Ficke AD, Myrick CA, Hansen LJ. Potential impacts of global climate change on freshwater fisheries. Reviews in Fish Biology and Fisheries. 2007; 17: 581–613. <https://doi.org/10.1007/s11160-007-9059-5>
- **9.** Lauri H, De Moel H, Ward PJ, Räsänen TA, Keskinen M, Kummu M. Future changes in Mekong River hydrology: Impact of climate change and reservoir operation on discharge. Hydrology and Earth System Sciences. 2012; 16: 4603–4619. <https://doi.org/10.5194/hess-16-4603-2012>
- **10.** Nguyen AL, Dang VH, Bosma RH, Verreth JAJ, Leemans R, De Silva SS. Simulated Impacts of Climate Change on Current Farming Locations of Striped Catfish (Pangasianodon hypophthalmus; Sauvage) in the Mekong Delta, Vietnam. Ambio. 2014; 43: 1059–1068. <https://doi.org/10.1007/s13280-014-0519-6> PMID: [24715387](http://www.ncbi.nlm.nih.gov/pubmed/24715387)
- **11.** Fukushima M, Jutagate T, Grudpan C, Phomikong P, Nohara S. Potential effects of hydroelectric dam development in the Mekong River basin on the migration of Siamese mud carp (Henicorhynchus siamensis and H. lobatus) elucidated by otolith microchemistry. PLoS ONE. 2014; 9: e103722. [https://doi.](https://doi.org/10.1371/journal.pone.0103722) [org/10.1371/journal.pone.0103722](https://doi.org/10.1371/journal.pone.0103722) PMID: [25099147](http://www.ncbi.nlm.nih.gov/pubmed/25099147)
- **12.** Ferguson JW, Healey M, Dugan P, Barlow C. Potential effects of dams on migratory fish in the Mekong River: Lessons from salmon in the fraser and Columbia Rivers. Environmental Management. 2011; 47: 141–159. <https://doi.org/10.1007/s00267-010-9563-6> PMID: [20924582](http://www.ncbi.nlm.nih.gov/pubmed/20924582)
- **13.** Junge C, Museth J, Hindar K, Kraabøl M, Vøllestad LA. Assessing the consequences of habitat fragmentation for two migratory salmonid fishes. Aquatic Conservation: Marine and Freshwater Ecosystems. 2014; 24: 297–311. <https://doi.org/10.1002/aqc.2391>
- **14.** Poulsen AF, Poeu O, Viravong S, Suntornratana U, Nguyen TT. Fish migrations of the Lower Mekong River Basin: implications for development, planning and environmental management. Mekong River Commission. Phnom Penh; 2002.
- **15.** Elsdon TS, Wells BK, Campana SE, Gillanders B, Jones CM, Limburg KE, et al. Otolith chemistry to describe movements and life-history parameters of fishes: hypotheses, assumptions, limitations and inferences. Oceanography and Marine Biology: An Annual Review. 2008; 46: 297–330.
- **16.** Panfili J, De Pontual H, Troadec H, Wright P. Manual of fish sclerochronology. Ifremer-IRD coedition, Brest, France; 2002. <https://doi.org/10.1643/OT-03-266>
- **17.** Campana SE. Chemistry and composition of fish otoliths: pathways, mechanisms and applications. Marine Ecology Progress Series. 1999; 188: 263–297.
- **18.** Ruttenberg BI, Hamilton SL, Hickford MJH, Paradis GL, Sheehy MS, Standish JD, et al. Elevated levels of trace elements in cores of otoliths and their potential for use as natural tags. Marine Ecology Progress Series. 2005; 297: 273–281. <https://doi.org/10.3354/meps297273>
- **19.** Hicks AS, Closs GP, Swearer SE. Otolith microchemistry of two amphidromous galaxiids across an experimental salinity gradient: A multi-element approach for tracking diadromous migrations. Journal of Experimental Marine Biology and Ecology. 2010; 394: 86–97. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jembe.2010.07.018) [jembe.2010.07.018](https://doi.org/10.1016/j.jembe.2010.07.018)
- **20.** Wells BK, Rieman BE, Clayton JL, Horan DL, Jones CM. Relationships between Water, Otolith, and Scale Chemistries of Westslope Cutthroat Trout from the Coeur d'Alene River, Idaho: The Potential Application of Hard-Part Chemistry to Describe Movements in Freshwater. Transactions of the American Fisheries Society. 2003; 132: 409–424. [https://doi.org/10.1577/1548-8659\(2003\)132](https://doi.org/10.1577/1548-8659(2003)132<0409:rbwoas>2.0.co;2)<0409: rbwoas>[2.0.co;2](https://doi.org/10.1577/1548-8659(2003)132<0409:rbwoas>2.0.co;2)
- **21.** Elsdon TS, Gillanders BM. Interactive effects of temperature and salinity on otolith chemistry: Challenges for determining environmental histories of fish. Canadian Journal of Fisheries and Aquatic Sciences. 2002; 59: 1796–1808. <https://doi.org/10.1139/f02-154>
- **22.** Sturrock A, Trueman C, Milton J, Waring C, Cooper M, Hunter E. Physiological influences can outweigh environmental signals in otolith microchemistry research. Marine Ecology Progress Series. 2014; 500: 245–264. <https://doi.org/10.3354/meps10699>
- **23.** Javor BJ, Dorval E. Sarability of trace elements in otolith of juvenile Pacific sardine Sardinops sagax. California Cooperative Oceanic Fisheries Investigations Reports. 2016; 57: 109–123.
- **24.** Martino JC, Doubleday ZA, Fowler AJ, Gillanders BM. Identifying physiological and environmental influences on otolith chemistry in a coastal fishery species. Marine and Freshwater Research. 2020. [https://](https://doi.org/10.1071/MF20196) doi.org/10.1071/MF20196
- **25.** Thomas ORB, Swearer SE. Otolith Biochemistry-A Review. Reviews in Fisheries Science & Aquaculture. 2019; 1–32. <https://doi.org/10.1080/23308249.2019.1627285> REVIEW
- 26. Hüssy K, Limburg KE, de Pontual H, Thomas ORB, Cook PK, Heimbrand Y, et al. Trace Element Patterns in Otoliths: The Role of Biomineralization. Reviews in Fisheries Science and Aquaculture. 2020; 1–33. <https://doi.org/10.1080/23308249.2020.1760204>
- **27.** Gillanders BM, Munro AR. Hypersaline waters pose new challenges for reconstructing environmental histories of fish based on otolith chemistry. Limnology and Oceanography. 2012; 57: 1136–1148. <https://doi.org/10.4319/lo.2012.57.4.1136>
- **28.** Hegg JC, Giarrizzo T, Kennedy BP. Diverse early life-history strategies in migratory Amazonian catfish: Implications for conservation and management. PLoS ONE. 2015; 10: 1–20. [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pone.0129697) [journal.pone.0129697](https://doi.org/10.1371/journal.pone.0129697) PMID: [26153984](http://www.ncbi.nlm.nih.gov/pubmed/26153984)
- **29.** Hobbs JA, Lewis LS, Ikemiyagi N, Sommer T, Baxter RD. The use of otolith strontium isotopes (87Sr/ 86Sr) to identify nursery habitat for a threatened estuarine fish. Environmental Biology of Fishes. 2010; 35: 557–569. <https://doi.org/10.1007/s10641-010-9672-3>
- **30.** Chesney EJ, McKee BM, Blanchard T, Chan LH. Chemistry of otoliths from juvenile menhaden Brevoortia patronus: Evaluating strontium, strontium:calcium and strontium isotope ratios as environmental indicators. Marine Ecology Progress Series. 1998; 171: 261–273. [https://doi.org/10.3354/](https://doi.org/10.3354/meps171261) [meps171261](https://doi.org/10.3354/meps171261)
- **31.** Barnett-Johnson R, Ramos FC, Grimes CB, MacFarlane RB. Validation of Sr isotopes in otoliths by laser ablation multicollector inductively coupled plasma mass spectrometry (LA-MC-ICPMS): Opening avenues in fisheries science applications. Canadian Journal of Fisheries and Aquatic Sciences. 2005; 62: 2425–2430. <https://doi.org/10.1139/f05-194>
- **32.** Hegg JC, Fisher CM, Vervoort J. Simultaneous determination of 87Sr/86Sr and trace-element data in otoliths and other sclerochronological hard structures. bioRxiv. 2020; 1–24. [https://doi.org/10.1101/](https://doi.org/10.1101/2020.04.24.060640) [2020.04.24.060640](https://doi.org/10.1101/2020.04.24.060640)
- **33.** Santos R V., Sondag F, Cochonneau G, Lagane C, Brunet P, Hattingh K, et al. Source area and seasonal 87Sr/86Sr variations in rivers of the Amazon Basin. Hydrological Processes. 2015; 29: 187–197. <https://doi.org/10.1002/hyp.10131>
- **34.** Hobbs JA, Yin QZ, Burton J, Bennett WA. Retrospective determination of natal habitats for an estuarine fish with otolith strontium isotope ratios. Marine and Freshwater Research. 2005; 56: 655–660. [https://](https://doi.org/10.1071/MF04136) doi.org/10.1071/MF04136
- **35.** Barnett-Johnson R, Pearson TE, Ramos FC, Grimes CB, Bruce MacFarlane R. Tracking natal origins of salmon using isotopes, otoliths, and landscape geology. Limnology and Oceanography. 2008; 53: 1633–1642. <https://doi.org/10.4319/lo.2008.53.4.1633>
- **36.** Walther BD, Thorrold SR, Olney JE. Geochemical Signatures in Otoliths Record Natal Origins of American Shad. Transactions of the American Fisheries Society. 2008; 137: 57–69. [https://doi.org/10.1577/](https://doi.org/10.1577/t07-029.1) [t07-029.1](https://doi.org/10.1577/t07-029.1)
- **37.** Martin J, Bareille G, Berail S, Pecheyran C, Daverat F, Bru N, et al. Spatial and temporal variations in otolith chemistry and relationships with water chemistry: A useful tool to distinguish Atlantic salmon Salmo salar parr from different natal streams. Journal of Fish Biology. 2013; 82: 1556–1581. [https://](https://doi.org/10.1111/jfb.12089) doi.org/10.1111/jfb.12089 PMID: [23639154](http://www.ncbi.nlm.nih.gov/pubmed/23639154)
- **38.** Brennan SR, Fernandez DP, Zimmerman CE, Cerling TE, Brown RJ, Wooller MJ. Strontium isotopes in otoliths of a non-migratory fish (slimy sculpin): Implications for provenance studies. Geochimica et Cosmochimica Acta. 2015; 149: 32–45. <https://doi.org/10.1016/j.gca.2014.10.032>
- **39.** Hauser M, Duponchelle F, Hermann TW, Limburg KE, Castello L, Stewart DJ, et al. Unmasking continental natal homing in goliath catfish from the upper Amazon. Freshwater Biology. 2020; 65: 325–336. <https://doi.org/10.1111/fwb.13427>
- **40.** Duponchelle F, Pouilly M, Pécheyran C, Hauser M, Renno J-FF, Panfili J, et al. Trans-Amazonian natal homing in giant catfish. Journal of Applied Ecology. 2016; 53: 1511–1520. [https://doi.org/10.1111/](https://doi.org/10.1111/1365-2664.12665) [1365-2664.12665](https://doi.org/10.1111/1365-2664.12665)
- **41.** Baird I. Pangasius krempfi. The IUCN Red List of Threatened Species 2011: e.T181328A7668262. 2011 [cited 24 Feb 2021]. <https://doi.org/10.2305/IUCN.UK.2011-1.RLTS.T181328A7668262.en>
- **42.** Rainboth WL. Fishes of the Cambodian Mekong. FAO Species Identification Field Guide for Fishery Purposes. Food and Agriculture Organization. Rome, Italy: FAO; 1996.
- **43.** Poulsen A, Hortle K, Valbo-Jorgensen J, Chan S, Viravong S, Bouakhamvongsa K, et al. Distribution and Ecology of Some Important Riverine Fish Species of the Mekong River Basin. MRC Technical Paper No. 10. K.G. Hortle SJB and TAMV, editor. Phnom Penh; 2004.
- **44.** Hogan Z, Baird IG, Radtke R, Vander Zanden MJ. Long distance migration and marine habitation in the tropical Asian catfish, Pangasius krempfi. Journal of Fish Biology. 2007; 71: 818–832. [https://doi.org/](https://doi.org/10.1111/j.1095-8649.2007.01549.x) [10.1111/j.1095-8649.2007.01549.x](https://doi.org/10.1111/j.1095-8649.2007.01549.x)
- **45.** Vu A V., Baumgartner LJ, Mallen-Cooper M, Howitt JA, Robinson WA, So N, et al. Diadromy in a large tropical river, the Mekong: more common than assumed, with greater implications for management. Journal of Ecohydraulics. 2020; 1–13. <https://doi.org/10.1080/24705357.2020.1855086> PMID: [33898904](http://www.ncbi.nlm.nih.gov/pubmed/33898904)
- **46.** Trinh TQ, Huynh NH, Thi VT, Nguyen TM, Hoang BQ. Preliminary results of domestication of Pangasius krempfi. Proceedings of 7th Technical Symposium on Mekong Fisheries. Ubon Ratchathani, Thailand; 2005. pp. 217–221.
- **47.** Tran NT, Labonne M, Hoang HD, Panfili J. Changes in environmental salinity during the life of Pangasius krempfi in the Mekong Delta (Vietnam) estimated from otolith Sr: Ca ratios. Marine and Freshwater Research. 2019; 70: 1734–1746. <https://doi.org/10.1071/MF18269>
- **48.** Mekong River Commission. Overview of the Hydrology of the Mekong Basin. Mekong River Commission. Vientiane; 2005.
- **49.** Tri VK. Hydrology and Hydraulic Infrastructure Systems in the Mekong Delta, Vietnam. In: Renaud FG, Kuenzer C, editors. he Mekong Delta System Interdisciplinary Analyses of a River Delta. Springer, Dordrecht; 2012. pp. 49–81.
- **50.** Pin C, Briot D, Bassin C, Poitrasson F. Concomitant separation of strontium and samarium-neodymium for isotopic analysis in silicate samples, based on specific extraction chromatography. Analytica Chimica Acta. 1994; 298: 209–217. [https://doi.org/10.1016/0003-2670\(94\)00274-6](https://doi.org/10.1016/0003-2670(94)00274-6)
- **51.** Huang K-F, You C-F, Chung C-H, Lin I-T. Nonhomogeneous seawater Sr isotopic composition in the coastal oceans: A novel tool for tracing water masses and submarine groundwater discharge. Geochemistry, Geophysics, Geosystems. 2011; 12. <https://doi.org/10.1029/2010gc003372>
- **52.** Huang KF, You CF. Tracing freshwater plume migration in the estuary after a typhoon event using Sr isotopic ratios. Geophysical Research Letters. 2007; 34: 1–5. <https://doi.org/10.1029/2006GL028253>
- **53.** Rosman K. J. R., Taylor P. D. P. Isotopic compositions of the elements. J Phys Chem Ref Data. 1998; 70: 1275–1287. <https://doi.org/10.1351/pac199870010217>
- **54.** Yang YH, Wu FY, Yang JH, Chew DM, Xie LW, Chu ZY, et al. Sr and Nd isotopic compositions of apatite reference materials used in U-Th-Pb geochronology. Chemical Geology. 2014; 385: 35-55. [https://](https://doi.org/10.1016/j.chemgeo.2014.07.012) doi.org/10.1016/j.chemgeo.2014.07.012
- **55.** Jochum KP, Stoll B, Weis U, Kuzmin D V., Sobolev A V. In situ Sr isotopic analysis of low Sr silicates using LA-ICP-MS. Journal of Analytical Atomic Spectrometry. 2009; 24: 1237–1243. [https://doi.org/10.](https://doi.org/10.1039/b905045k) [1039/b905045k](https://doi.org/10.1039/b905045k)
- 56. Avigliano E, Ibañez A, Fabré N, Callicó Fortunato R, Méndez A, Pisonero J, et al. White mullet Mugil curema population structure from Mexico and Brazil revealed by otolith chemistry. Journal of Fish Biology. 2020; 97: 1187–1200. <https://doi.org/10.1111/jfb.14500> PMID: [32799355](http://www.ncbi.nlm.nih.gov/pubmed/32799355)
- **57.** R Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing; 2019. Available: <https://www.r-project.org>
- **58.** Gillanders BM. Otolith chemistry to determine movements of diadromous and freshwater fish. Aquatic Living Resources. 2005; 18: 291–300. <https://doi.org/10.1051/alr:2005033>
- **59.** Carlson AK, Phelps QE, Graeb BDS. Chemistry to conservation: using otoliths to advance recreational and commercial fisheries management. Journal of Fish Biology. 2017; 90: 505–527. [https://doi.org/10.](https://doi.org/10.1111/jfb.13155) [1111/jfb.13155](https://doi.org/10.1111/jfb.13155) PMID: [27704556](http://www.ncbi.nlm.nih.gov/pubmed/27704556)
- **60.** Peucker-Ehrenbrink B, Miller MW, Arsouze T, Jeandel C. Continental bedrock and riverine fluxes of strontium and neodymium isotopes to the oceans. Geochem Geophys Geosyst. 2010; 11: Q03016. <https://doi.org/10.1029/2009GC002869>
- **61.** Richter FM, Rowley DB, DePaolo DJ. Sr isotope evolution of seawater: the role of tectonics. Earth and Planetary Science Letters. 1992; 109: 11–23. [https://doi.org/10.1016/0012-821X\(92\)90070-C](https://doi.org/10.1016/0012-821X(92)90070-C)
- **62.** Mcnutt RH. Strontium Isotopes. In: Cook P.G., Herczeg AL, editors. Environmental Tracers in Subsurface Hydrology. Springer, Boston, MA; 2000. pp. 233–260. https://doi.org/10.1007/978-1-4615-4557-6_8
- **63.** Palmer MR, Edmond JM. Controls over the strontium isotope composition of river water. Geochimica et Cosmochimica Acta. 1992; 56: 2099–2111. [https://doi.org/10.1016/0016-7037\(92\)90332-D](https://doi.org/10.1016/0016-7037(92)90332-D)
- **64.** Liu C, Clift PD, Murray RW, Blusztajn J, Ireland T, Wan S, et al. Geochemical evidence for initiation of the modern Mekong delta in the southwestern South China Sea after 8 Ma. Chemical Geology. 2017; 451: 38–54. <https://doi.org/10.1016/j.chemgeo.2017.01.008>
- **65.** Steinshouer DW, Qiang J, McCabe PJ, Ryder RT. Maps showing geology, oil and gas fields, and geologic provinces of the Asia Pacific region. USGS Open File Report. 1997.
- **66.** Shao Y, Farkaš J, Holmden C, Mosley L, Kell-Duivestein I, Izzo C, et al. Calcium and strontium isotope systematics in the lagoon-estuarine environments of South Australia: Implications for water source mixing, carbonate fluxes and fish migration. Geochimica et Cosmochimica Acta. 2018; 239: 90–108. <https://doi.org/10.1016/j.gca.2018.07.036>
- **67.** Muhlfeld CC, Thorrold SR, McMahon TE, Marotz B. Estimating westslope cutthroat trout (Oncorhynchus clarkii lewisi) movements in a river network using strontium isoscapes. Canadian Journal of Fisheries and Aquatic Sciences. 2012; 69: 906–915. <https://doi.org/10.1139/F2012-033>
- **68.** Walther BD, Thorrold SR. Inter-annual variability in isotope and elemental ratios recorded in otoliths of an anadromous fish. Journal of Geochemical Exploration. 2009; 102: 181–186. [https://doi.org/10.1016/](https://doi.org/10.1016/j.gexplo.2008.10.001) [j.gexplo.2008.10.001](https://doi.org/10.1016/j.gexplo.2008.10.001)
- **69.** Goldstein SJ, Jacobsen SB. Nd and Sr isotopic systematics of river water suspended material: implications for crustal evolution. 1988; 87: 249–265.
- **70.** Herut B, Starinsky A, Katz A. Strontium in rainwater from Israel: Sources, isotopes and chemistry. Earth and Planetary Science Letters. 1993; 120: 77–84. [https://doi.org/10.1016/0012-821X\(93\)90024-4](https://doi.org/10.1016/0012-821X(93)90024-4)
- **71.** Chadwick OA, Derry LA, Vitousek PM, Huebert BJ, Hedin LO. Changing sources of nutrients during four million years of ecosystem development. Nature. 1999; 397: 491–497. [https://doi.org/10.1038/](https://doi.org/10.1038/17276) [17276](https://doi.org/10.1038/17276)
- **72.** Vengosh A, Spivack AJ, Artzi Y, Ayalon A. Geochemical and boron, strontium, and oxygen isotopic constraints on the origin of the salinity in groundwater from the Mediterranean Coast of Israel. Water Resources Research. 1999; 35: 1877–1894. <https://doi.org/10.1029/1999WR900024>
- **73.** Humston R, Priest BM, Hamilton WC, Bugas PE. Dispersal between Tributary and Main-Stem Rivers by Juvenile Smallmouth Bass Evaluated Using Otolith Microchemistry. Transactions of the American Fisheries Society. 2010; 139: 171–184. <https://doi.org/10.1577/t08-192.1>
- **74.** Strohm DD, Budy P, Crowl TA. Matching watershed and otolith chemistry to establish natal origin of an endangered desert Lake Sucker. Transactions of the American Fisheries Society. 2017; 146: 732–743. <https://doi.org/10.1080/00028487.2017.1301994>
- **75.** Panfili J, Darnaude AM, Vigliola L, Jacquart A, Labonne M, Gilles S. Experimental evidence of complex relationships between the ambient salinity and the strontium signature of fish otoliths. Journal of Experimental Marine Biology and Ecology. 2015; 467: 65–70. <https://doi.org/10.1016/j.jembe.2015.03.007>
- **76.** Correia AT, Hamer P, Carocinho B, Silva A. Evidence for meta-population structure of Sardina pilchardus in the Atlantic Iberian waters from otolith elemental signatures of a strong cohort. Fisheries Research. 2014; 149: 76–85. <https://doi.org/10.1016/j.fishres.2013.09.016>
- **77.** Gillanders BM, Munro AR. Relations between water chemistry, otolith chemistry and salinity of a hypersaline system: implications for determining past environmental history of fish. CSIRO. 2009.
- **78.** Kennedy BP, Blum JD, Folt CL, Nislow KH. Using natural strontium isotopic signatures as fish markers: Methodology and application. Canadian Journal of Fisheries and Aquatic Sciences. 2000; 57: 2280– 2292. <https://doi.org/10.1139/f00-206>
- **79.** Milton DA, Chenery SR. Movement patterns of barramundi Lates calcarifer, inferred from ⁸⁷Sr/⁸⁶Sr ratios in otoliths, indicated non-participation in spawning. Marine Ecology Progress Series. 2005; 301: 279–291. <https://doi.org/10.3354/meps301279>
- **80.** Chan S, Chhuon CK, Sintavong V, Kongpeng B, Ubolratana S, Noppanum Y, et al. Fish migrations and spawning habits in the Mekong Mainstream—a survey using local knowledge. AMFC Technical Report. Vientiane, Lao; 1999.
- **81.** Roberts TR, Baird IG. Traditional fisheries and fish ecology on the Mekong River at Khone waterfalls in southern Laos. The Natural History Bulletin of the Siam Society. 1995; 43: 219–262.
- **82.** Duong YT, Nguyen VT. Genetic diversity of Pangasius krempfi in the Mekong River estuaries. Can Tho University Journal of Science. 2019; 11: 81–88. <https://doi.org/10.22144/ctu.jen.2019.027>
- **83.** Wootton RJ. Fish Ecology. First. Springer Science+Business Media Dordrecht; 1992. [https://doi.org/10.](https://doi.org/10.1007/978-94-011-3832-1) [1007/978-94-011-3832-1](https://doi.org/10.1007/978-94-011-3832-1)
- **84.** Vu VA, Nguyen ND, Hidas E, Nguyen MN. Vam Nao deep pools: a critical habitat for Pangasius krempfi and other valuable species in the Mekong Delta, Vietnam. Asian Fisheries Science. 2009; 22: 631– 639.
- **85.** Beck MW, Heck KL, Able KW, Childers DL, Eggleston D. B., Gillanders BM, et al. The Identification, Conservation, and Management of Estuarine and Marine Nurseries for Fish and Invertebrates. Bioscience. 2001; 51: 633–641. [https://doi.org/10.1641/0006–3568\(2001\)051\[0633:TICAMO\]2.0.CO;2](https://doi.org/10.1641/00063568(2001)051[0633:TICAMO]2.0.CO;2)
- **86.** Dugan PJ, Barlow C, Agostinho AA, Baran E, Cada GF, Chen D, et al. Fish migration, dams, and loss of ecosystem services in the mekong basin. Ambio. 2010; 39: 344–348. [https://doi.org/10.1007/s13280-](https://doi.org/10.1007/s13280-010-0036-1) [010-0036-1](https://doi.org/10.1007/s13280-010-0036-1) PMID: [20799685](http://www.ncbi.nlm.nih.gov/pubmed/20799685)
- **87.** Strayer DL, Dudgeon D. Freshwater biodiversity conservation: recent progress and future challenges. Journal of the North American Benthological Society. 2010; 29: 344–358. [https://doi.org/10.1899/08-](https://doi.org/10.1899/08-171.1) [171.1](https://doi.org/10.1899/08-171.1)
- **88.** IFRC. Vietnam—Drought and Saltwater Intrusion. Operation update report. Vietnam; Available: [https://](https://reliefweb.int/report/viet-nam/vietnam-drought-and-saltwater-intrusion-emergency-plan-action-epoa-dref-operation-0) [reliefweb.int/report/viet-nam/vietnam-drought-and-saltwater-intrusion-emergency-plan-action-epoa](https://reliefweb.int/report/viet-nam/vietnam-drought-and-saltwater-intrusion-emergency-plan-action-epoa-dref-operation-0)[dref-operation-0](https://reliefweb.int/report/viet-nam/vietnam-drought-and-saltwater-intrusion-emergency-plan-action-epoa-dref-operation-0)
- **89.** CGIAR. The drought and salinity intrusion in the Mekong River Delta of Vietnam. CGIAR Research Centers in Southeast Asia. Available: [https://cgspace.cgiar.org/handle/10568/75633%0Apapers3://](https://cgspace.cgiar.org/handle/10568/75633%0Apapers3://publication/uuid/1F44F90C-A3AD-4202-B9EE-EDB184F85B37) [publication/uuid/1F44F90C-A3AD-4202-B9EE-EDB184F85B37](https://cgspace.cgiar.org/handle/10568/75633%0Apapers3://publication/uuid/1F44F90C-A3AD-4202-B9EE-EDB184F85B37)
- **90.** Nguyen NA. Historic drought and salinity intrusion in the Mekong Delta in 2016: Lessons learned and response solutions. Vietnam Journal of Science, Technology and Engineering. 2017; 59: 93–96. [https://doi.org/10.31276/vjste.59\(1\).93](https://doi.org/10.31276/vjste.59(1).93)
- **91.** Reist JD, Wrona FJ, Prowse TD, Power M, Dempson JB, King JR, et al. An Overview of Effects of Climate Change on Selected Arctic Freshwater and Anadromous Fishes. Ambio. 2006; 35: 381–387. [https://doi.org/10.1579/0044-7447\(2006\)35\[381:aooeoc\]2.0.co;2](https://doi.org/10.1579/0044-7447(2006)35[381:aooeoc]2.0.co;2) PMID: [17256642](http://www.ncbi.nlm.nih.gov/pubmed/17256642)
- **92.** Mcnamara A, Atkinson J, Froy H, Khela S, Smith R, Peet J, et al. Climate Change Vulnerability of Migratory Species. Species Assessments. ZSL, UNEP/CMS Secretariat. Bonn, Germany; 2010.
- **93.** Baran E, So N, Leng SV, Arthur R, Kura Y. Relationships between bioecology and hydrology among Tonle Sap fish species. 2007.
- **94.** Baumgartner LJ, Barlow C, Mallen-Cooper M, Boys C, Marsden T, Thorncraft G, et al. Achieving fish passage outcomes at irrigation infrastructure; a case study from the Lower Mekong Basin. Aquaculture and Fisheries. 2021; 6: 113–124. <https://doi.org/10.1016/j.aaf.2018.12.008>
- **95.** Schmutz S, Mielach C. Review of Existing Research on Fish Passage through Large Dams and its Applicability to Mekong Mainstream Dams. MRC Technical Paper No. 48. Phnom Penh, Cambodia; 2015.
- **96.** ICEM. MRC Strategic Environmental Assessment (SEA) of hydropower on the Mekong mainstream: summary of the final report. Ha Noi, Vietnam; 2010.