Trophic ecology of a blooming jellyfish (Aurelia coerulea) in a Mediterranean coastal lagoon
Raquel Marques, Delphine Bonnet, Claire Carré, Cécile Roques, Audrey M. Darnaude

To cite this version:
Raquel Marques, Delphine Bonnet, Claire Carré, Cécile Roques, Audrey M. Darnaude. Trophic ecology of a blooming jellyfish (Aurelia coerulea) in a Mediterranean coastal lagoon. Limnology and Oceanography, Association for the Sciences of Limnology and Oceanography, 2021, 66 (1), pp.141-157. 10.1002/lno.11593 . hal-03411044

HAL Id: hal-03411044
https://hal.umontpellier.fr/hal-03411044
Submitted on 17 Nov 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.
Trophic ecology of a blooming jellyfish (*Aurelia coerulea*) in a Mediterranean coastal lagoon

*Raquel MARQUES*¹, *Delphine BONNET*¹, *Claire CARRÉ*¹, *Cécile ROQUES*¹, *Audrey M. DARNAUDE*¹

¹ MARBEC, Univ. Montpellier, CNRS, Ifremer, IRD, Montpellier, France

**Corresponding author:**

marques.rfs@gmail.com / raquel.marques@umontpellier.fr

CC093, Place Eugène Bataillon, 34095 Montpellier Cedex 05, France

Tel: +33769370312

**Co-authors contact:**

delphine.bonnet@umontpellier.fr; claire.carre@ird.fr; Cecile.Roques@cnrs.fr;

Audrey.Darnaude@cnrs.fr

**Running head:** Trophic ecology of jellyfish in a coastal lagoon

**Key words:** Stable Isotopes, Diet, Trophic niche, Trophic competition, Population dynamics, Scyphistomae, Medusae, Oysters
Abstract

The current lack of knowledge on the trophic ecology of scyphozoans, particularly at the benthic stage, prevents a full understanding of the controls on many jellyfish blooms. The blooming scyphozoan (*Aurelia coerulea*) completes its entire life cycle in the Thau lagoon (southern France), where the annual population dynamics of both its benthic and pelagic stages have been described. This offered an exceptional framework to investigate the trophic processes regulating jellyfish populations over time. To this aim, stable isotopic signature analysis ($\delta^{13}$C and $\delta^{15}$N) was used to infer the diet of both *A. coerulea* scyphistomae and medusae over one year. These results were matched with medusae gut content analysis and with the monthly abundances of local plankton groups. Lastly, the isotopic signatures of *A. coerulea* scyphistomae and medusae were compared with those of the oysters (*Crassostrea gigas*) cultivated in the lagoon to evaluate the potential interspecific trophic competition. The results revealed two seasonal shifts in the trophic niche of *A. coerulea* and substantial overlap between the diets of its benthic and pelagic stages. Conversely, trophic niche overlaps with the oysters were restricted, suggesting a limited impact of the local jellyfish bloom on shellfish production. Phytoplankton, microzooplankton, mesozooplankton, and sedimentary organic matter were all important food sources during critical periods of *A. coerulea* life-cycle. However, microzooplankton abundance was found to be key for the production of buds by the scyphistomae and, therefore it is likely to control the benthic population size and, thereby, to modulate the intensity of its annual bloom in Thau.
Introduction

Due to the impact of their conspicuous blooms on coastal ecosystems functioning and economic activities, jellyfish have received increasing scientific attention during the last decades (Purcell 2012). In particular, the ecological drivers of jellyfish mass occurrences have been investigated, revealing a complex interaction of natural (e.g. Condon et al. 2012) and anthropogenic (e.g. Purcell 2012) causes. However, uncovering the drivers of blooms is particularly challenging for most scyphozoan blooming species because their life-cycle comprises a benthic (scyphistomae) and a pelagic (ephyrae and medusae) phase (e.g. Lucas 2001). Therefore, bloom formation is a joint consequence of the production of pelagic ephyrae by the benthic scyphistomae and of their survival and growth into medusae. As a result, the ecology of both life stages controls bloom intensity.

Bottom-up processes within food webs often play a key role in ecological systems functioning and are amongst the most important drivers of jellyfish blooms (Boero et al. 2008). Food quality and availability are known to control the production of ephyrae by the scyphistomae (Han and Uye 2010; Ikeda et al. 2017) and to modulate the growth rate of medusae (Ishii and Båmstedt 1998). This supports the need for comprehensive studies on the trophic ecology of both life stages in the field. Yet, although information is growing on the trophic ecology of medusae (e.g. Javidpour et al. 2016; Milisenda et al. 2018), the diet of jellyfish scyphistomae is still poorly known.

Jellyfish from the Aurelia genus are present globally in coastal areas and are among the most common scyphozoans that form blooms (Mills 2001). Large accumulations of Aurelia spp. have been reported all around the world, including in the Mediterranean, where they occur mainly in protected waters and semi-enclosed seas (Mills 2001). Their medusae have been described as zooplanktivorous, with a dominance of mesozooplankton, especially copepods, in their diet (e.g. Ishii and Tanaka 2001; Lo and Chen 2008). However, while
microzooplankton and benthic food sources have been considered for long as negligible food sources for jellyfish, recent findings based on new techniques (such as stable isotope analysis) suggest the opposite (Javidpour et al. 2016). In laboratory studies, newly hatched *Artemia* sp. are usually provided as food (e.g. Han and Uye 2010, Hubot et al. 2017), but the few studies regarding the diet of *Aurelia* sp. scyphistomae in the wild suggest that they eat a mix of phytoplankton (Huang et al. 2015), microzooplankton (Kamiyama 2013) and small mesozooplankton species (e.g. copepods, cladocerans, gelatinous zooplankton; Östman 1997).

Considering the critical role of scyphistomae in the formation of scyphozoans blooms, it is urgent to specify natural prey preferences in *Aurelia* species and the potential trophic competition among their benthic and pelagic stages to understand blooms formation in this genus and evaluate their ecological consequences.

Situated along the North-western Mediterranean coast, the Thau lagoon offered an exceptional framework for this. Indeed, this lagoon presents the rare particularity to harbour a complete resident population of *Aurelia coerula* (Bonnet et al. 2012; Marques et al. 2015a), which allows investigating the trophic processes that regulate its population dynamics at both stages. The scyphistomae of *A. coerula* are widespread in the lagoon, fixed mainly on biofouling organisms that grow on anthropogenic structures (predominantly on oysters and mussels; Marques et al. 2015a). They are present all year round, with a peak of coverage in the Spring (April) and lower densities in the Summer and Autumn (Marques et al. 2019).

Ephyrae appear in the early winter (November – December) and give rise to adult medusae at the beginning of the Spring (April – May), generating the annual jellyfish bloom, which persists until June – July (Bonnet et al. 2012; Marques et al. 2015b). Because no clear link was found between the abundance of mesozooplankton in the lagoon and the benthic population dynamics of *A. coerula*, it was suggested that other food sources might sustain
the species local production (Marques et al. 2019). Nevertheless, further confirmation is still required in this regard.

Coastal lagoons are usually very productive environments, where high continental inputs in nutrients and particulate organic matter sustain high and diversified primary and secondary productions (Nixon et al. 1995). This benefits the whole food web and enhances the growth of lagoon predators like juvenile fish (Escalas et al. 2015). In Thau, it also supports a massive shellfish production: ~10% of the Pacific oysters Crassostrea gigas produced in France come from the lagoon, with a yearly shellfish production of 15 000 tons (Mongruel et al. 2013).

In this context, the present work not only aimed to describe the trophic ecology of both the benthic and the pelagic life-stages of A. coerulea in Thau and assess its influence on critical periods of population dynamics of this jellyfish (e.g. peak of bud production, strobilation, and medusae growth), but also to evaluate whether A. coerulea medusae and scyphistomae compete for food with the Pacific oysters reared in the lagoon. For this, we combined medusae gut content assessments with stable isotopes analysis. This latter technique has been increasingly used to study the structure and transfer of organic matter within coastal food webs (Layman et al. 2012) and has recently allowed uncovering the diet, trophic levels, and trophic interactions of different jellyfish species (Fleming et al. 2015; Javidpour et al. 2016; Milisenda et al. 2018). Using it to explore the changes in A. coerulea diet during a full annual cycle should allow assessing whether its benthic and pelagic stages occupy the same trophic niche than the oysters cultivated in the lagoon. This strongly contributes to a better understanding of the impacts of A. coerulea blooms on the local shellfish production.

**Material and Methods**

**Study site**
The Thau lagoon is a semi-enclosed marine coastal lagoon of 75 km$^2$ area, connected to the Mediterranean Sea by three narrow channels (Fig. 1). It is relatively shallow, with mean and maximum depths of 4 and 10 m, respectively (except for a localized depression of 24 m). The local tidal range (< 1m) is weak, so water residence time in the lagoon is globally high (1—4 months) and strongly influenced by seasonal strong wind events (Millet and Cecchi 1992). The lagoon environment parameters show strong seasonal variations, characteristic of temperate regions, with temperature and salinity at their lowest in the winter (with minimum values of 7.6 and 35.0, respectively) and at their highest in the summer (with maximum values of 25.8 ºC and 39.6, respectively; Marques et al 2019). The lagoon mainly receives water from the Sète canal that connects it to the Mediterranean Sea and from several small intermittent rivers that drain its catchment area (290 km$^2$, Plus et al. 2006). These later dry out between May and September and show occasional flash floods in the wet season (Fouilland et al. 2012). As a result, marine conditions prevail in the lagoon, the annual influence of the freshwater coming from the watershed being highly dependent on the intensity of rainfall events during the winter (Plus et al. 2006). With regards to anthropogenic influence, the lagoon is under multiple pressures due to the presence of the touristic city of Sète and many small villages and agriculture fields on its coastline. Shellfish farming is the most important economic activity on the lagoon (Mongruel et al. 2013): around 20% of its surface is occupied by farms, mainly in the northern and north-western parts (Fig. 1).

**Sampling**

For this study, the jellyfish and their potential food sources were sampled in the eastern part of the lagoon, at two close sites where the benthic and the pelagic population dynamics of *A. coerulea* had been previously described (Bonnet et al. 2012; Marques et al. 2015b; Marques et al. 2019). Both sites (benthic sampling site: 43°25’31.1"N; 03°42’0.9"E
and pelagic sampling site: 43°23′59.1″N; 03°36′37.2″E; Fig. 1) are located on soft-bottom sediments punctuated by sparse seagrass meadows and are strongly influenced by marine water influxes due to their proximity to the Sète channel, which connects the lagoon to the Mediterranean Sea.

*A. coerulea* scyphistomae were collected monthly on a partially submerged boat present at the benthic sampling site (see Marques et al. 2019 for more details), over an entire calendar year (from January 2017 to January 2018). For this, mussel shells with sizeable aggregates of scyphistomae attached on their underside surface (three per sampling date) were collected directly on the surface of the boat by SCUBA diving. They were brought to the laboratory in ambient water and placed in 0.2-µm-filtered seawater (ca. 20°C) for about 2h to ensure all scyphistomae had empty guts. Fifty individual scyphistomae were then collected under a dissecting microscope (Olympus SZ40; Olympus KL 1500 LCD), using needles and tweezers to carefully detach them, and preserved in cryotubes at –30°C.

The pelagic ephyrae of *A. coerulea* are usually present in the lagoon from November to April (Bonnet et al. 2012; Marques et al. 2015b). However, because stable isotope analysis requires pooling high numbers of these small organisms to be applicable, sampling for this life stage in this work was successful in January 2018 only. The ephyrae were collected near the water surface at the pelagic sampling site, by horizontal towing, using a modified WP2 plankton net (1.2 m long, 50-cm opening, and 200-µm mesh). In the laboratory, they were picked and kept for ca. 2h in filtered seawater to allow for complete gut evacuation. Then 50 individuals were pooled per sample and preserved at –30°C.

*A. coerulea* medusae (i.e., pelagic individuals with bell diameter > 1 cm), were collected every two weeks at the pelagic sampling site, from March to June 2017, i.e., over the entire period of their presence in the lagoon. They were collected in surface waters using hand nets and transported to the laboratory in ambient water. Five individual medusae were then...
randomly selected and prepared for stomach content analysis. For this, they were each
partially dried on a paper towel to remove excess water, measured (bell diameter in cm),
weighted (total wet weight in g), and individually preserved in 4% buffered formaldehyde.
The remaining medusae were kept for ca. 2 h in 0.2 µm filtered seawater (ca. 20ºC) to empty
their guts. Three of them were then placed on a paper towel for about 1 minute (30 s on each
side) to remove excess water, weighed, and measured. As bell tissue is the most suitable body
part for stable isotope analysis in jellyfish (D’Ambra et al. 2014), gonads, oral arms, and
gastric pouches were removed from each medusa. The remaining individual bell tissues were
preserved separately at -30ºC. In March 2017, due to the small size of the medusae (ca. 2 cm
bell diameter), eight complete individuals were pooled per replicate before preservation at -
30ºC.

For this work both the plankton and the sedimentary organic matter of the lagoon were also
sampled as they both constitute potential food sources for *A. coerulea*. Samples for these two
components were collected at pelagic and benthic sampling sites, respectively, on the same
sampling dates as *A. coerulea* medusae and scyphistomae collection. Within the plankton, the
fraction larger than 200 µm, that between 60 and 200 µm and that between 20 and 60 µm
were assumed to be composed mainly by mesozooplankton, microzooplankton, and
phytoplankton, respectively. Mesozooplankton samples were collected near the surface, by
horizontal towing, using a modified WP2 plankton net (length: 1.2 m; opening area: 50 cm;
mesh size: 200 µm). Once in the laboratory, each sample was filtered through a 60 µm mesh
sieve to eliminate excess water and divided into five subsamples. Microzooplankton and
phytoplankton samples were also collected by horizontal towing near the surface, but using a
phytoplankton net (length: 1 m; opening area: 30 cm; mesh size: 20 µm). Once in the
laboratory, each sample was filtered through a 200-µm sieve. The size fraction > 200 µm was
discarded. The remaining sample was then separated into the two size fractions,
corresponding to microzooplankton and phytoplankton, using a 60 µm sieve, and then divided into 5 subsamples. For each plankton size fraction, the subsamples were collected separately on pre-combusted (500°C for 24h) Whatman GF/F filters. Two filters of each plankton component were acidified with 1% HCl and triple rinsed with distilled water to remove inorganic carbon, which can bias C stable isotope results (Yokoyama et al. 2005). The remaining non-acidified filters were used for N stable isotope analysis, since sample acidification may affect the stable isotope signature for this element (Pinnegar and Polunin 1999). All samples were preserved at –30°C until further analysis. For sedimentary organic matter, the first 2 cm of the sediment were collected by SCUBA diving at the benthic monitoring site. Samples (2 replicates) were carefully scrutinized to eliminate any large organisms, sediment inorganic particles, or vegetal debris, before preservation at –30°C.

To investigate the trophic interactions between A. coerulea and the local oysters, both wild and cultivated individuals of Crassostrea gigas were sampled seasonally from October 2017 to August 2018, including during the peak of the jellyfish bloom (which occurred in June in 2018). Wild oysters (mean size: 11.5 ± 2.0 cm) were collected by SCUBA diving at the benthic monitoring site, while the cultivated ones (mean size: 11.9 ± 1.0 cm) were obtained from the shellfish producer Huitres-Bouzigues.com. Immediately after their removal from the lagoon, the oysters were transported to the laboratory in ambient water, measured and carefully dissected to collect their adductor muscle. The muscle tissues were then rinsed with distilled water and preserved separately at –30°C until further analysis.

In situ abundance of plankton in the Thau lagoon

Phytoplankton, microzooplankton and mesozooplankton samples were collected at the pelagic monitoring site, every two weeks from January to June 2017 and monthly onwards, until December 2017. For phytoplankton, 10 to 20L of surface water were collected, filtered with a
15-µm-mesh net, and preserved with 2% buffered formaldehyde. For microzooplankton, a subsample of 30 ml of surface water was preserved with 2% buffered formaldehyde (to estimate ciliates' abundance) and one of 110 ml was preserved with Lugol’s solution (to estimate heterotrophic flagellates' abundance). Phytoplankton and microzooplankton species were identified and counted using sedimentation chambers and an inverted microscope (Olympus IX70) following the Utermöhl method (Utermöhl 1958). Mesozooplankton samples were collected near the surface by horizontal towing using a modified WP2 plankton net (1.2 m long, 50-cm opening, and 200-µm mesh). Samples were immediately preserved in 4% buffered formaldehyde until further analysis in the laboratory. Mesozooplankton abundance was determined by counting organisms under a dissecting microscope (Olympus SZX7 – ILLT). The diversity of mesozooplankton was not assessed.

Gut content analyses

To evaluate the diet of A. coerulea medusae, their gastric pouches, oral arms, and the preserving solution were examined under a dissecting microscope (Olympus SZX7 – ILLT). Although A. coerulea medusae were present in the lagoon from March, most individuals exhibited empty guts during this month. Therefore, gut content analysis was only performed on the medusae collected between April and June. For this, only complete exoskeletons were considered for prey identification. This was done to the lowest possible taxonomic level, although the level of exoskeleton digestion often precluded prey identification down to the species level. The importance of each prey in the diet was expressed by the following indices: (i) the frequency of occurrence (in %), which represents the percentage of medusae with the prey i in their guts among all those that had non-empty guts; (ii) the index of relative importance (in %), representing the percentage of prey i in relation to the total number of prey.
items found in the non-empty guts; and (iii) the mean abundance of prey $i$ in non-empty guts (in ind. medusae$^{-1}$).

**Stable isotope analysis**

All filters containing plankton (phytoplankton, microzooplankton, and mesozooplankton) were oven-dried at 60°C for 48h and the biological material was gently scraped off the filter surface. Samples for the sedimentary organic matter, the oysters, $A. coerulea$ medusae, scyphistomae, and ephyrae were freeze-dried for 48h and ground to a fine powder using a mortar and pestle. The sedimentary organic matter samples were divided into two subsamples. One half was used directly for N stable isotope analysis. The remaining subsample was acidified with 1% HCl to remove carbonates before C stable isotope analysis, rinsed several times with distilled water, and oven-dried at 70°C.

Stable isotopic analyses for biological samples were performed using a PDZ Europa ANCA-GSL elemental analyser interfaced with a PDZ Europa 20-20 isotope ratio mass spectrometer (Sercon Ltd., Cheshire, UK). Measurements of $\delta^{13}C$ and $\delta^{15}N$ signatures were performed each on 1.5 to 4 mg of dry samples, with exception of the medusae, for which ca. 10 mg of dry sample was required for successful analysis, after salt content correction, based on dry weight and ash-free dry weight relationships (Lucas et al. 1994; Pitt et al. 2009). Sedimentary organic matter samples (of ca. 55 mg each) were analysed using an Elementar Vario EL Cube or Micro Cube elemental analyser (Elementar Analysensysteme GmbH, Hanau, Germany) interfaced to a PDZ Europa 20-20 isotope ratio mass spectrometer (Sercon Ltd., Cheshire, UK). Calibration was performed against NIST Standard Reference Materials (IAEA-600, USGS-40, USGS-41, USGS-42, USGS-43, USGS-61, USGS-64, and USGS-65). Isotope ratios of all samples were expressed as parts per thousand (‰) differences from the internal
reference standards (glutamic acid, alfalfa flour, nylon 6, bovine liver, and enriched alanine) using the following equation:

\[ \delta X = \left[ \left( \frac{R_{\text{sample}}}{R_{\text{standard}}} \right) - 1 \right] \times 1000 \]

where \( X \) is the \(^{13}\)C or \(^{15}\)N and \( R \) is the corresponding ratio, \(^{13}\)C/\(^{12}\)C or \(^{15}\)N/\(^{14}\)N. As the lipid content of organisms affects their \( \delta^{13} \)C signatures, \( \delta^{13} \)C correction is required when C:N is higher than 3.5 (Post et al. 2007). Therefore, the \( \delta^{13} \)C values obtained for A. coerulea scyphistomae and medusae (mean C:N 3.7 ± 0.1 and 3.9 ± 0.6, respectively) and for the mesozooplankton (mean C:N of 6.9 ± 3.0) were corrected (\( \delta^{13} \)C\(_{\text{corr}} \)) according to the equations proposed by D’Ambra et al. (2014) for jellyfish:

\[ \delta^{13}C_{\text{corr}} = \delta^{13}C_{\text{initial}} - 9.43 + 2.69 \times C:N \]

and by Syväranta and Rautio (2010) for zooplankton:

\[ \delta^{13}C_{\text{corr}} = \delta^{13}C_{\text{initial}} + 7.95 \times \left( \frac{C:N - 3.8}{C:N} \right) \]

Relationship between benthic population dynamics and plankton abundance

Data on A. coerulea benthic population dynamics were obtained from Marques et al. (2019). Generalized linear models (using linear and logistic regressions, without interactions) were employed to assess the respective contributions of the absolute abundances of the non-averaged phytoplankton, the microzooplankton and the mesozooplankton (after logarithmic transformation \( \ln (x+1) \)) to temporal trends in the scyphistomae density (% coverage) and in the proportion of the scyphistomae producing buds. The models were validated by examination of residuals versus fitted values plots (Zuur et al. 2009).

Determination of Isotopic Niche Periods
To reveal potential shifts in the trophic niches of *A. coerulea* scyphistomae and medusae during the year and identify the periods when they present unchanging isotopic signatures (hereafter "Isotopic Niche Periods"), a cluster analysis was performed on the monthly mean isotopic values of both life stages (Jain 2010). For this, partitioning algorithms, based on the k-means clustering method, were applied using the package “factoextra” (Kassambara and Mundt 2017). The k-means approach subdivides the data into a set of k groups so that the sum of squares from the data points to the center of each group is minimized (Kassambara and Mundt 2017). This clustering approach allowed to identify the successive isotopic niche periods for both life stages, providing the basis for identifying their successive sources of organic matter during the year.

**Assessment of potential intra- and interspecific trophic competition**

Our sampling design allowed for reliable estimation of the potential intraspecific trophic competition between *A. coerulea* scyphistomae and medusae within each isotopic niche period. However, because oyster and jellyfish samples were collected in different years (except for one isotopic niche period), the trophic competition between the two species was only investigated globally, assuming that interannual variability in the trophic niche in the two species is negligible. In both cases, the Bayesian framework proposed by Jackson et al. (2011) for evaluating trophic competition was used. For this, Bayesian multivariate normal distributions were first fitted to the isotopic signatures of all organisms. Then, the overlap between their trophic niches was calculated based on maximum likelihood fitted ellipses, using the function “maxLikOverlap” from the R package “SIBER” (Jackson et al. 2011).

**Determination of jellyfish diet using Stable isotope analysis**
Differences in isotopic signatures (δ\text{13}C and δ\text{15}N) among the main local potential food sources (phytoplankton, microzooplankton, mesozooplankton, and sedimentary organic matter) were tested by a PERMANOVA (Anderson 2017) on the log10 transformed Bray-Curtis distance matrix (−δ\text{13}C and δ\text{15}N), made using the package “vegan” (Oksanen et al. 2019), followed by pairwise comparisons made using the “pairwiseAdonis” package in R (Martinez Arbizu 2019). Sources with no significant differences were grouped for subsequent analyses.

Diet compositions for A. coerulea scyphistomae and medusae within each isotopic niche period were assessed using Bayesian mixing models developed specifically for stable isotope analysis (“MixSIAR” package, Stock and Semmens 2016). By generating the probability distributions of all potential mixing solutions with the associated confidence intervals (based on 300 000 chain length), this approach allows identifying the most likely contribution for each food source. MixSIAR further provides a graphical user interface that allows investigation of the contributions of multiple food sources to the diet of target predators, considering not only the isotopic signatures (δ\text{13}C and δ\text{15}N) of the sources and the predators but also the uncertainties and variability around these estimates. Finally, the method allows us to use different isotopic fractionation factors at each trophic level. As previously performed in other studies on jellyfish diet (e.g. Morais et al. 2017), the fractionation values applied here for both A. coerulea life stages were those proposed by Vander Zanden and Rasmussen (2001): for δ\text{13}C we used 0.47 ± 1.23 ‰ in all cases, while for δ\text{15}N we used 2.52 ± 2.5 ‰ and 3.23 ± 0.41 ‰ according to the type of food consumed (plant vs. animal, respectively). Like Fleming et al. (2015) and Milisenda et al. (2018), we did not use the fractionation values reported by D’Ambra et al. (2014), since they are very distinct from those mostly used in the literature (Vander Zanden and Rasmussen 2001; Post 2002) and they still require further laboratory corroboration (D’Ambra et al. 2014).
The basal tissue turnover rate for Aurelia sp. is of ca. 1‰ day\(^{-1}\) for \(\delta^{13}\)C and 2‰ day\(^{-1}\) for \(\delta^{15}\)N, and it takes 18 to 20 days for the tissues of this jellyfish to reach the stable isotopic equilibrium with the food ingested (D’Ambra et al. 2014). To account for such turnover rates, MixSIAR models were run by isotopic niche period, but jellyfish signatures at a given sampling date were matched with those recorded one month earlier for all potential food sources.

**Results**

**Medusae gut contents**

Among the 25 medusae collected for gut content analysis from April to June 2017, 21 had food in their guts. The bell diameter of these individuals did not vary significantly over time (ANOVA, \(F_2 = 1.4, p\)-value = 0.2), remaining at ca. 8.5 cm. Overall, gut content composition predominantly consisted of mesozooplankton (>88%). Microzooplankton (mainly tintinnids) and phytoplankton (mainly diatoms and dinoflagellates) represented only 8 and 4% of the total prey identified, respectively, and they were only found in the guts in April and May (Fig. 2, Table 1, Supplementary Table 1). In April, phytoplankton and microzooplankton occurred in 20 and 60% of the guts analysed, but their relative importance and abundances were still low (< 7.5% and < 2.2 ind.medusa\(^{-1}\), respectively, Table 1). In May, frequency of occurrence increased for phytoplankton (33%) and slightly decreased for microzooplankton (56%) and showed a growing trend of their relative importance for both groups (5.6 and 12.5%, respectively, Fig. 2). Indeed, in May, microzooplankton relative importance was higher than some mesozooplankton organisms, like the “other crustaceans” group, which includes cladocerans and ostracods (10.0%; Table 1, Supplementary Table 1). Masses of unidentified organic matter were also recurrently observed over the entire study period.
Twenty-four different taxa of mesozooplankton were identified in the guts of the medusae, but, among them, copepods and nauplii (from cirripeds and copepods) dominated. They occurred in 40 to 88.9% of the guts analysed and represented up to 46.3% of the prey identified (in June, Table 1). The maximum average abundance of mesozooplankton organisms in the guts (26.2 ± 35.4 ind.medusae⁻¹) was recorded in April when non-crustacean taxa (mainly gastropod veliger), and copepods represented more than 80% of the prey identified in medusae gut contents (Table 1). Nauplii (index of relative importance = 31.3%) were the most important mesozooplanktonic prey in the guts in May, while in June, copepods dominated (index of relative importance = 46.3%, Table 1).

Prey availability and relationship with benthic population dynamics

Prey abundances in A. coerulea medusae gut contents did not reflect their availability in the water column. The abundances of phytoplankton, microzooplankton, and mesozooplankton in the lagoon all showed high intra-annual variability (Fig. 3), with respective peaks in January (25 138 ± 34 047 cell.L⁻¹) and May (35 794 ± 18 374 and cell.L⁻¹), in February, April, and September (> 6 200 cell.L⁻¹) and in June (90 895 ± 107 072 ind.m⁻³). Thus, when A. coerulea medusae were present in the water column, the planktonic community was mainly dominated by phytoplankton in April and May, and by mesozooplankton in March and June, while the microzooplankton showed consistently lower abundances despite a small peak in April. In terms of species composition, the most abundant phytoplanktonic and microzooplanktonic taxa in the water column during the study period were Chaetoceros sp. and Strombidium sp., respectively (Supplementary Table 2). Mesozooplankton diversity was not assessed, but Acartia sp. are recurrently the most abundant taxa in Thau (Boyer et al. 2013).

Annual variations in scyphistomae coverage, which peaked in April (11.6 ± 3.7 %, Marques et al. 2019), were positively correlated with the non-averaged abundance of phytoplankton.
fluctuations in the mean percentage of scyphistomae producing buds, which varied between 0.4 ± 0.7% in November and 25.2 ± 7.3% in September (Marques et al. 2019, Fig. 4), were positively correlated with variations in non-averaged microzooplankton abundance (Generalized linear models, t-value = 2.97, p-value = 0.01, Table 2). In turn, fluctuations in the mean percentage of scyphistomae producing buds, which varied between 0.4 ± 0.7% in November and 25.2 ± 7.3% in September (Marques et al. 2019, Fig. 4), were positively correlated with variations in non-averaged microzooplankton abundance (Generalized linear models, t-value = 2.97, p-value = 0.01, Table 2).

Temporal variation of A. coerulea isotopic signatures

δ\(^{13}\)C and δ\(^{15}\)N signatures showed significant temporal variation for both the scyphistomae and medusae (one-way PERMANOVA, Pseudo-F\(_{11}\) = 22.7, p-value < 0.01 and Pseudo-F\(_{3}\) = 38.6, p-value = 0.001, respectively), but differences between life stages were never significant during the period of medusae presence, from March to June (one-way PERMANOVA, Pseudo-F\(_{1}\) = 1, p-value = 0.4). The mean bell diameter of the medusae used for stable isotope analysis, showed a sharp increase between March (1.0 ± 0.3 cm) and June (8.9 ± 1.1 cm), with an estimated overall growth of 0.8 mm.day\(^{-1}\). Over this period, medusae δ\(^{13}\)C signatures increased progressively from −23.4 ± 0.1‰ to −19.4 ± 0.5‰, while their δ\(^{15}\)N signatures remained stable for the first three months (at ca. 8.1‰), and increased (to a maximum at 8.9 ± 0.3‰) only in June (Fig. 5). For the scyphistomae, minimum δ\(^{13}\)C values were registered at the beginning of the study period (in January 2017, mean: −23.4 ± 0.1‰). The δ\(^{13}\)C signatures then increased to reach maximum values in June, July, and August (> −19.4‰) before decreasing again until January 2018 (−22.3 ± 0.4‰). The δ\(^{15}\)N signatures of scyphistomae showed a similar temporal trend, with low values at the beginning and the end of the study period (8.3 ± 0.1‰ and 8.0 ± 0.4‰ in January 2017 and 2018, respectively), and maximum values in July and August (>9‰). The minimum values of δ\(^{15}\)N signatures, though, were observed in February 2017 (7.1 ± 0.5‰). The average δ\(^{13}\)C and δ\(^{15}\)N signatures of the ephyrae collected in January 2018 (bell diameter of 0.21 ± 0.1 cm) were of −22.8 ± 0.1‰ and 8.5 ±
0.3‰, respectively. They did not differ significantly from those of the scyphistomae collected at the same sampling time (T-test, t = −1.9, df = 2.3 p-value = 0.2 and t = 1.2, df = 2.6, p-value = 0.3, for δ^{13}C and δ^{15}N respectively).

The clustering analysis revealed three distinct groups of isotopic signatures among the monthly values obtained for all life stages of *A. coerulea* (Fig. 6) allowing to identify three isotopic niche periods during the year.: Period 1 gathered the δ^{13}C and δ^{15}N signatures of all life stages from December to April, irrespective of the year (2017 or 2018). Period 2 reflected the signatures of both the medusae and the scyphistomae from June to August. Period 3 corresponded to the signatures of the scyphistomae from September to November, together with the signatures of the medusae and the scyphistomae in May. However, May showed a particular sharp shift in δ^{13}C and δ^{15}N reflecting the rapid transition from the isotopic niche of period 1 to that of period 2 and, therefore, it was not included in any isotopic niche period.

**Monthly variability of organic matter sources signatures**

δ^{13}C and δ^{15}N signatures varied significantly according to the organic matter source and the month (significant interaction, PERMANOVA, Pseudo-$F_{17} = 23.1$, p-value < 0.01; Fig. 7). For carbon signatures, minimum δ^{13}C values for phytoplankton, microzooplankton and mesozooplankton (of −24.7 ± 0.3, −23.3 ± 0.1 and −23.7 ± 0.0 ‰, respectively) were all observed in March. A sharp increase in δ^{13}C was observed in the following months, with maximums in May for mesozooplankton (−18.8 ± 0.2 ‰) and in November for phytoplankton (−19.0 ± 0.0 ‰) and microzooplankton (19.9 ± 0.1 ‰). Concerning nitrogen signatures, mesozooplankton was the organic matter source with the highest δ^{15}N values, ranging from 7.3 ± 0.3 (in May) to 8.4 ± 0.0‰ (in March). Minimum δ^{15}N values were also observed in May for the phytoplankton and the microzooplankton (at 5.8 ± 0.5 ‰ and 6.0 ± 0.3 ‰,
respectively) but, for these two organic matter sources, maximum values were observed in July (at 6.7 ± 0.3 ‰ and 7.4 ± 0.2 ‰, respectively). Moreover, another peak in δ¹⁵N (at 6.7 ± 0.0 ‰) was observed in February for the phytoplankton. For the sedimentary organic matter, both the δ¹³C and δ¹⁵N signatures decreased from March (–18.9‰ and 5.8‰, respectively) to April (–20.7‰ and 5.5‰, respectively), remaining constant afterwards.

Contribution of organic matter sources to A. coerulea isotopic signatures

Since the δ¹³C and δ¹⁵N signatures of the phytoplankton and the microzooplankton were not significantly different (PERMANOVA post-hoc test, Pseudo-F₁ = 5.7, adjusted p-value = 0.17) these two organic matter sources were pooled as Small Plankton group in the mixing models used to assess the diet of A. coerulea. The remaining sources were included individually in the models (Table 3). The contribution of each source was found to vary according to the isotopic niche periods and the life stage of A. coerulea considered (Fig. 8).

For the scyphistomae, the model suggested a dietary shift from small plankton consumption in period 1 (93.3%) to a diet based on a mix of benthic (36.6% of sedimentary organic matter) and pelagic (39.3% of mesozooplankton and 24.4% of small plankton) sources in period 2. The same occurred in period 3, although the small plankton was the main food source (69.2%), and sedimentary organic matter contribution decreased (27.0%). For the medusae, small plankton was the only food source (100%) in period 1, but the diet changed in period 2, including mainly sedimentary organic matter (64.3%) and mesozooplankton (32.3%). As the isotopic signatures of the ephyrae collected in January 2018 were very similar to those of the scyphistomae in the same period, their diet probably mainly consist of small plankton organisms.

Intra- and interspecific competitions for the food resources
Intraspecific isotopic niche overlap was substantial during the whole period of co-occurrence of the benthic and pelagic stages of *A. coerulea* in the lagoon (March to June; Fig. 9). Indeed, although the percentage of niche overlap was higher in period 1 (41.5%) than in period 2 (only 9.9%), the isotopic niche of the medusae entirely overlaid that of the scyphistomae in period 2. Similarly, although only three ephyrae samples were analysed in this study (all from January 2018), their isotopic signatures were close to those observed for the scyphistomae in period 1, suggesting high (although not quantifiable) trophic niche overlap among these two life stages.

In Thau, interspecific trophic competition between *A. coerulea* and bivalves was observed, although limited. The δ13C and δ15N signatures of the oysters from the lagoon varied from –25.6 to –18.5 ‰ and from 8.4 to 9.4‰, respectively (Fig. 10). Significant differences in isotopic signatures were observed between cultivated and wild individuals (PERMANOVA, Pseudo-F11 = 12.4, p-value < 0.01; Fig. 10), with the former showing significantly higher δ13C (–19.7 ± 0.9 ‰) and lower δ15N (8.6 ± 0.3 ‰) signatures on average than the later (–20.1 ± 0.4 ‰ and 9.2 ± 0.3 ‰, respectively). Interspecific isotopic niche overlaps were limited (<30%) and lower than that between cultivated and wild oysters (35.4%). Interspecific isotopic niche overlap was more important between cultivated oysters and *A. coerulea* medusa stage (29.1%). However, if we assume that the seasonal shifts in isotopic signatures are consistent among years for both the jellyfish and the oysters, the trophic competition for food should only occur at a limited period of the year and only with the medusae stage. Indeed, only the signatures recorded in period 2 were responsible for the interspecific niche overlap observed among *A. coerulea* medusae and cultivated (21.8%) or wild (21.1%) oysters.

**Discussion**
To our knowledge, this is the first study to investigate the trophic ecology of both the benthic and the pelagic stages of a jellyfish species (*A. coerulea*) in association with its in situ population dynamics and the plankton availability. The results obtained offer the unprecedented opportunity to identify the bottom-up processes regulating *A. coerulea* populations, contributing to our understanding of the formation of its blooms.

*Trophic ecology of the pelagic stages of *A. coerulea***

Ephyrae were only collected once during the study period and their isotopic signature was similar to that of scyphistomae at the same time, indicating major importance of the small planktonic organic matter (i.e. phytoplankton and microzooplankton) in their diet. This result will have to be confirmed because, in Thau, *A. coerulea* ephyrae are mainly released in November, but strobilation continues until April (Marques et al. 2019). Therefore, we cannot exclude that the ephyrae caught in January 2018 had been released just a few days or weeks before their collection and therefore still had the isotopic signature of the scyphistomae that produced them. Moreover, because of their very low growth rate during the winter (< 0.1 mm.day\(^{-1}\), Marques et al. 2015b), the ephyrae caught in January might not have yet incorporated the signature of the prey ingested after their release (Frazer et al. 1997). Nevertheless, phytoplankton, microzooplankton (such as rotifers) and suspended particulate organic matter have all been previously identified as important food sources for ephyrae (Båmstedt et al. 2001; Zheng et al. 2015) so our findings are in agreement with the literature. The results from medusae gut contents analysis support previous reports describing *A. coerulea* medusae as mesozooplanktivorous, feeding mainly on copepods and nauplii (mainly of cirripeds). Indeed, *Aurelia* spp. medusae have been suggested to prey mainly on mesozooplankton and to have higher clearance rates and selective preferences for crustacean prey such as copepods, cirriped nauplii, and cladocerans (Hansson 2006; Lo and Chen 2008).
Phytoplankton and microzooplankton also contributed to the diet of *A. coerulea* medusae in Thau, but only during their first two months of growth and with low relative importance. Indeed, *Aurelia* spp. diet often echoes prey local abundances in their environment (e.g. Ishii and Tanaka 2001), which might explain these results, since the abundance of microzooplankton and phytoplankton in the lagoon were higher in April and May. Yet, variations of prey availability in Thau were not entirely reflected in *A. coerulea* medusae diet, since mesozooplankton represented consistently more than 80% of the prey identified in their guts, despite its lower in situ abundance in this period. Although gut content analyses provided important qualitative information on the diet of jellyfish medusae, conclusions regarding the importance of each prey type for their growth, at longer time scales, should be drawn with caution, due to the bias associated with this technique. The digestion time of mesozooplankton in the medusae guts might vary between 1 and 5h, depending on medusa size, temperature, and prey type (Ishii and Tanaka 2001; Martinussen and Båmstedt 2001), with smaller prey being digested faster (Martinussen and Båmstedt 2001). Therefore, gut content analysis often leads to an overestimation of the importance of hard and big prey in the diet, such as crustaceans. This might have contributed to a general overlook of the potential relevance of the lower trophic levels to the diet of jellyfish (Javidpour et al. 2016). Indeed, in Thau, the diet composition of *A. coerulea* medusae differed between gut content and stable isotope analyses. The later approach underlined not only the importance of the phytoplankton and microzooplankton (pooled as small plankton) for the diet of *A. coerulea* medusae in Thau but also that of the sedimentary organic matter.

The diet of the *A. coerulea* medusae varied over time. In general, the $\delta^{13}$C (−23.4 to −19.4‰) and $\delta^{15}$N (8.1 to 8.9‰) values found for the *A. coerulea* medusae stage were in the range of the values published by Fleming et al. (2015) (−20.3 to −18.1 for $\delta^{13}$C and 8.5 to 11.8 for $\delta^{15}$N) and D’Ambra et al. (2013) (−20.5 ± 0.3‰ and 7.2 ± 0.4‰ on average for $\delta^{13}$C and
δ^{15}N, respectively). However, intra-annual fluctuations in medusae isotopic signatures revealed a significant shift in May, with an increase of ca. 3.5 and 1‰ for δ^{13}C and δ^{15}N, respectively. This separates two distinct periods of stable isotopic signatures: period 1, during medusae growth from March to April, and period 2, in June, when they reproduce before the collapse of the bloom. This variation in the isotopic signature might indicate a rapid ontogenic shift in the diet of the medusae, reflecting the change from small plankton to mesozooplankton and sedimentary organic matter sources. A similar shift in the trophic niche was also shown for Aurelia aurita in Northern Ireland, where medusae fed on higher trophic levels by the end of their growing period (Fleming et al. 2015). Temporal variations in isotopic signatures of predators might also reflect analogous changes in the isotopic signatures at the base of the food webs (Post 2002). In this study, the values of the assessed organic matter sources agree with those previously reported in Thau (Pernet et al. 2012) and other north-western Mediterranean coastal lagoons (Dierking et al. 2012; Escalas et al. 2015) but revealed significant fluctuations over time. In Thau, ^{13}C-depleted coastal inputs are dependent on the rainfall, which was high in March and low in April (http://www.meteofrance.fr/climat-passe-et-futur/bilans-climatiques/bilan-2017. Accessed 27 Jul 2019), likely contributing to the variation in the δ^{13}C isotopic signatures of the lower trophic levels and then reflected in those of A. coerulea medusae. However, similar trends were not observed for δ^{15}N isotopic signatures, which showed a decreasing trend for most organic matter sources in May, contrasting with an increasing trend for medusae in June. This underlines that the observed isotopic niche shift for A. coerulea medusae was not only a reflection of temporal fluctuations in the signatures of their prey but likely induced by a significant change in their diet. Finally, our results highlight the importance of sedimentary organic matter (64.3%) in the diet of A. coerulea medusae, as previously observed for A. aurita in the Kiel Fjord (Javidpour et al. 2016). Like most shallow marine ecosystems, the Thau lagoon is recurrently subjected to
sediment resuspension, triggered by river floods and strong wind activity (Fouilland et al. 2012). With this regard, the unidentified masses of organic matter found in the guts of the medusae were probably aggregates of re-suspended sedimentary organic matter.

**Trophic ecology of the benthic stage of A. coerulea**

The temporal variability of the δ¹³C and δ¹⁵N signatures of *A. coerulea* scyphistomae suggested two significant intra-annual shifts in their diet and identified three different isotopic niche periods. The diet of scyphistomae was mostly based on small plankton during period 1, included all available food sources during period 2 and changed to a mix of pelagic (i.e., small plankton) and sedimentary organic matter during period 3. These seasonal variations agree with those of the availability of planktonic food sources in the lagoon, following the high abundances of phytoplankton and microzooplankton in periods 1 and 3 and that of mesozooplankton in period 2 (i.e., in June). Our results agree with the few existing reports on the diet of jellyfish scyphistomae, which suggested that they feed on small mesozooplankton species (e.g. copepods, cladocerans, and cirripeds nauplii; Östman 1997; Ikeda et al. 2017), as well as on microzooplankton and phytoplankton (dinoflagellates, ciliates, rotifers, and diatoms; Kamiyama 2013; Wang et al. 2015; Huang et al. 2015). However, as for medusae, the temporal variation in scyphistomae isotopic signatures might also reflect the origin of the carbon and nitrogen inputs in the lagoon (Post 2002). Indeed, fluctuations in δ¹³C and δ¹⁵N values might reflect the stronger contribution of terrestrial inputs to the basis of the food web, after rainy events in period 1 (Vizzini et al. 2005; Pernet et al. 2012a) and the exceptionally low terrestrial inputs from June onwards (periods 2 and 3), due to a very dry summer and autumn in 2017 (> 80% loss of rainfall when compared with the mean between 1981 – 2010 in October, http://www.meteofrance.fr/climat-passe-et-futur/bilans-climatiques/bilan-2017. Accessed 27 Jul 2019). Furthermore, it might also be affected by the higher influence of
wastewater effluent in the lagoon during dry periods (Perrin and Tournoud 2009), which is suggested to induce an enrichment of $\delta^{15}$N signatures, as in other coastal lagoons (Vizzini et al. 2005; Dierking et al. 2012; Escalas et al. 2015). Yet, the skewed temporal pattern of the scyphistomae isotopic signatures when compared with their sources further confirm a seasonal variation in their diet.

The increase in mesozooplankton consumption during period 2, when the abundance of this prey is maximal, is not surprising. Higher abundances of this type of prey (especially of newly hatched *Artemia* sp.) is recognized to induce better performances of scyphistomae (i.e., growth, asexual reproduction, and strobilation) in laboratory experiments (e.g. Ikeda et al. 2017; Hubot et al. 2017). However, our results further highlight the prominent role of the lower trophic levels in the feeding and benthic population dynamics of the species in Thau.

Although we were not able to precisely quantify the relative importance of phytoplankton and microzooplankton in the diet of *A. coerulea* scyphistomae, they both appear to be important. Phytoplankton cells are seemingly insufficient to support scyphistomae basic metabolic rates at high temperatures (20ºC) and for long periods (Wang et al. 2015; Huang et al. 2015), but they provide a suitable alternative source of energy for their survival and asexual reproduction at low temperatures (Huang et al. 2015; Wang et al. 2015). Therefore, peaks in phytoplankton abundances during period 1 probably support *A. coerulea* scyphistomae survival over the winter and early spring. Similarly, the significant positive correlation between the abundance of microzooplankton and the percentage of scyphistomae producing buds suggests that this type of prey promotes the buds production, ultimately driving the benthic population density.

Indeed, buds production by scyphistomae of *Aurelia aurita* has been previously shown to increase when reared on a ciliate-based diet rather than on the larger *Artemia* prey (Kamiyama 2013). Interestingly, although more buds were produced in April in the lagoon (due to high scyphistomae density) the peak of the percentage of scyphistomae producing buds, as well as
the maximum number of buds per scyphistoma, were registered in September (Marques et al. 2019), co-occurring with high abundances of microzooplankton in the lagoon.

Lastly, as for medusae, our results highlight the importance of the sedimentary organic matter in the diet of A. coerulea scyphistomae in Thau. This does not come as a major surprise because re-suspended sediments were often observed on the scyphistomae samples collected in situ. Sedimentary organic matter is usually composed by a mixture of microphytobenthos, heterotrophic microorganisms (bacteria, ciliates, protozoans, nematodes) and detritus, classically associated and re-suspended with sediment (Dubois et al. 2007), which might provide a suitable source of food for jellyfish benthic stages (Östman 1997).

Intra- and interspecific competition

The benthic scyphistomae and the pelagic medusae of A. coerulea, although inhabiting different habitats, appeared to share, at least partially, the same organic matter sources in the lagoon. During period 1, their high isotopic niche overlap, and the results of the mixing models, indicate that both stages feed on phytoplankton and/or microzooplankton. In period 2, despite a lower isotopic niche overlap, the trophic niche of the medusae entirely covers that of the scyphistomae. This suggests that during large medusae blooms and under food limitation conditions, intraspecific competition for food might occur in the lagoon, with possible detrimental impacts on the scyphistomae population.

One of the main concerns regarding the presence of A. coerulea in Thau is the potential competition for food with the oysters produced in the lagoon, in particular during the medusae blooms and due to the overspread distribution of scyphistomae (Marques et al. 2015a). However, our results suggest only a limited trophic niche overlap. Although oysters and A. coerulea stages were not collected in the same year (except in period 3) we assumed that the isotopic signature of the oysters mostly varies intra-annually (Pernet et al. 2012). If this is
true, our results indicate that interspecific competition for food only potentially occurs between *A. coerulea* medusae and oysters (cultivated and wild) in period 2. During this period, sedimentary organic matter was an important source in the diet of *A. coerulea* medusae and also reported as part of the diet of oysters (Dubois and Colombo 2014). This might explain the isotopic niche overlap, although restricted, between these two organisms at this period. The limited interspecific trophic competition between the *A. coerulea* and the oysters might result from their different filtration and particle retention mechanisms, as previously suggested for other suspension-feeding species co-occurring with oysters (Dubois and Colombo 2014). Indeed, *A. coerulea* medusae are cruising predators, capturing their prey using locally generated flow currents (Dabiri et al. 2005) and the scyphistomae use a passive ambush strategy (Huang et al. 2015), contrasting with the true filter-feeding strategy of the oysters (Dubois et al. 2007; Dubois and Colombo 2014). The different mechanisms to capture prey, likely promoted the selection and ingestion of different organic matter sources, reducing the trophic competition for the same type of prey. Phytoplankton (especially diatoms) is the main source of food for oysters (Dupuy et al. 2000; Pernet et al. 2012). In situ feeding experiments showed that the consumption of *Aurelia* sp. medusae on micro- and mesozooplankton organisms released the predation pressure from these secondary producers on the lower trophic levels, boosting phytoplankton biomass and bacterial production (Turk et al. 2008). Therefore, it is possible that the blooms of *A. coerulea* medusae might even be advantageous for the production of oysters in the lagoon, via a top-down cascade effect on the microbial community.

**Bottom-up control of the *A. coerulea* population dynamics**

In the Thau lagoon, the winter and early spring are critical periods for the formation of the *A. coerulea* bloom (Marques et al. 2019). The production of ephyrae occurs between
November and April, with two main peaks: in November (during period 3) due to a high percentage of the scyphistomae strobilating (despite their low densities), and in February–March (during period 1), when this percentage is low but the density of scyphistomae is high (Marques et al. 2019). As they grow to become medusae, the magnitude of the bloom is thus, tightly dependent on the accumulated production of ephyrae, their survival, and growth rate.

In period 1, phytoplankton and microzooplankton are the main sources of food for both the ephyrae and scyphistomae of *A. coerulea*. This stresses the role of the lower trophic levels in the formation of the local jellyfish blooms: they promote higher levels of scyphistomae and ephyrae survival and they boost the production of buds, leading to higher scyphistomae densities and ephyrae production. In summer (during period 2) both *A. coerulea* life stages change their diet to a mix of all sources (except small plankton for medusae). This is particularly important because it supports the peak of the bloom, following high growth rates of medusae, as well as their sexual reproduction (Fig. 11; Marques et al. 2015b). It is also during this period that scyphistomae coverage declines (Fig. 11, Marques et al. 2019). Our results suggest a potentially high intraspecific trophic competition between scyphistomae and medusae, especially during this period. Therefore, the high abundance and high predation pressure of the medusae might lead to the reduction of food availability for scyphistomae and could contribute to the reduction of their coverage. During the following dry season (i.e., period 3), a bacteria-based food web prevails in the lagoon, with internal regeneration of nitrogen, due to the absence of terrestrial freshwater inputs in the lagoon (Chapelle et al. 2000). This likely supports the peaks of microzooplankton abundance since these organisms are recognized as important bacterivorous (Rassoulzadegan and Sheldon 1986).

Microzooplankton appear to have a critical role as a source of food for scyphistomae, which, in period 3, would sustain not only the noticed peak of buds production in September but also
the main strobilation period in November (Marques et al. 2019), i.e., the first peak of ephyrae production and the initial foundation of the subsequent jellyfish bloom in the Thau lagoon.

**Limitation of the study**

Although stable isotope analysis is a powerful tool to assess the trophic ecology of predators, the MixSIAR results should be considered with caution. Indeed, mixing models always provide a solution but their results might not always be biologically relevant: their precision decreases with the number of introduced organic matter sources and depends greatly on the accuracy of their signatures (Dubois et al. 2007).

In this work, we used a turnover time of one month for both jellyfish life stages, following the results reported for *Aurelia* sp. (18 – 20 days, D’Ambra et al. 2014). If inaccurate, this might have significantly biased the MixSIAR results for each isotopic niche period because the set of organic matter source signatures matching with those of the jellyfish might be incorrect. Moreover, despite the frequency of sampling for organic matter sources during our study, some periods of the year (e.g. July – September) were less represented in the database. Given the intra-annual variability in the isotopic signatures of the plankton component, we cannot fully exclude that this sampling gap slightly biased our results.

The implementation of different isotopic fractionation values in the mixing models also drastically modify their final results. In our study, using the fractionation values proposed by D’Ambra et al. (2014) would result in a higher contribution of mesozooplankton to the diet in both stages of *A. coerulea*. However, the values from D’Ambra et al. (2014) are very different from those typically reported in the literature (e.g. Vander Zanden and Rasmussen 2001; Post 2002), leading to unrealistic trophic levels (Fleming et al. 2015; Milisenda et al. 2018).

Furthermore, the temperature (which is highly variable in Thau), the feeding condition, the
sexual maturity (e.g. Barnes et al. 2007), and, probably, the life stage might also affect fractionation and turnover values.

**Conclusion**

Knowledge of the trophic ecology and population dynamics of jellyfish is imperative to understand the main environmental drivers of blooms. With this regard, the Thau lagoon offered an exceptional framework to study both benthic and pelagic trophic interactions and to uncover the main organic matter sources supporting key periods of the *A. coerulea* life cycle. In particular, we highlight the role of phytoplankton and microzooplankton in supporting scyphistomae survival and asexual reproduction, that of mesozooplankton and sedimentary organic matter for the growth of medusae, as well as the possible negative influence of intraspecific competition on the benthic population dynamics. Moreover, we demonstrate that the interspecific trophic competition between *A. coerulea* and the commonly cultivated oyster *C. gigas* is likely to be limited, at least in the Thau lagoon, and therefore, we advocate that *A. coerulea* blooms have a seemingly restricted impact on the local shellfish production.
References:


Kassambara A, Mundt F (2017) factoextra: Extract and Visualize the Results of Multivariate Data Analyses. 1–74. R package version 1.0.5


to the fat of the matter: Models, methods and assumptions for dealing with lipids in

Purcell JE (2012) Jellyfish and Ctenophore Blooms Coincide with Human Proliferations and
marine-120709-142751

Rassoulzadegan F, Sheldon RW (1986) Predator-prey interactions of nanozooplankton and
bacteria in an oligotrophic marine environment. Limnol Oceanogr 31:1010–1029. doi:
10.4319/lo.1986.31.5.1010

10.5281/zenodo.47719

Syväranta J, Rautio M (2010) Zooplankton, lipids and stable isotopes: importance of seasonal,
latitudinal, and taxonomic differences. Can J Fish Aquat Sci 67:1721–1729. doi:
10.1139/F10-091

Turk V, Lučić D, Flander-Putrle V, Malej A (2008) Feeding of *Aurelia* sp. (Scyphozoa) and
links to the microbial food web. Mar Ecol 29:495–505. doi: 10.1111/j.1439-
0485.2008.00250.x

Utermöhl H (1958) Zur Vervollkommnung der quantitativen Phytoplankton-Methodik. SIL

reproduction in *Aurelia* sp.1 polyps. Hydrobiologia 754:169–178. doi: 10.1007/s10750-
014-2020-4

diet-tissue isotopic fractionation in estuarine macrobenthos. Mar Ecol Prog Ser 296:115–
128. doi: 10.3354/meps296115


Acknowledgment:

We would like to thank Solenn Soriano, Nicolas Nouguier, and Remy Valdes for their technical support during SCUBA dives and fieldwork. We also thank Sandrine Crochemore and Sébastien Colantoni for their assistance during laboratory samples preparation. We thank Simon Julien from huitres-bouzigues.com for providing the cultivated oyster samples.

Plankton diversity and abundance data are part of a long-term monitoring program on microbial communities in the Thau lagoon funded by Observatoire des Sciences de l’Univers (OREME). Data are available on (https://data.oreme.org/plankton/plankton_thau_home). The laboratory analysis of samples to determine stable isotope signatures was performed at the Stable Isotope Facility at the University of California, Davis, USA. The authors declare that they have no conflict of interests.
Table 1: Frequency of occurrence (FO), index of relative importance (IRI) and mean abundance of prey items found in *A. coerulea* medusae gut contents during the period of its presence in the Thau lagoon. Numbers in parenthesis are the number of medusae with prey items analyzed.

<table>
<thead>
<tr>
<th>Prey</th>
<th>FO (%)</th>
<th>IRI (%)</th>
<th>Abundance (± SD) (ind.medusae⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Apr</td>
<td>May</td>
<td>Jun</td>
</tr>
<tr>
<td></td>
<td>(5)</td>
<td>(9)</td>
<td>(8)</td>
</tr>
<tr>
<td><strong>Phytoplankton</strong></td>
<td>20.0</td>
<td>33.3</td>
<td>0.0</td>
</tr>
<tr>
<td><strong>Microzooplankton</strong></td>
<td>60.0</td>
<td>55.6</td>
<td>0.0</td>
</tr>
<tr>
<td><strong>Mesozooplankton (total)</strong></td>
<td>80.0</td>
<td>88.9</td>
<td>100</td>
</tr>
<tr>
<td>- <em>Copepods</em></td>
<td>40.0</td>
<td>66.7</td>
<td>87.5</td>
</tr>
<tr>
<td>- <em>Nauplii (copepods and cirripeds)</em></td>
<td>60.0</td>
<td>88.9</td>
<td>62.5</td>
</tr>
<tr>
<td>- <em>Other crustaceans</em></td>
<td>20.0</td>
<td>55.6</td>
<td>50.0</td>
</tr>
<tr>
<td>- <em>Non-crustaceans</em></td>
<td>60.0</td>
<td>66.7</td>
<td>25.0</td>
</tr>
</tbody>
</table>
Table 2: Parameters of the generalized linear models used to assess correlations between the benthic population dynamics variables (scyphistomae coverage and scyphistomae producing buds) with the abundance [ln (x+1)] of phytoplankton (cell L\(^{-1}\)), microzooplankton (cell L\(^{-1}\)) and mesozooplankton (ind m\(^{-3}\)).

<table>
<thead>
<tr>
<th></th>
<th>Estimate</th>
<th>Std. Error</th>
<th>t value</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Scyphistomae coverage (%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(Intercept)</td>
<td>-0.17</td>
<td>0.07</td>
<td>-2.46</td>
<td>0.03</td>
</tr>
<tr>
<td>Phytoplankton</td>
<td>0.02</td>
<td>0.01</td>
<td>2.97</td>
<td>0.01</td>
</tr>
<tr>
<td>Microzooplankton</td>
<td>0.01</td>
<td>0.01</td>
<td>2.10</td>
<td>0.05</td>
</tr>
<tr>
<td>Mesozooplankton</td>
<td>0.00</td>
<td>0.00</td>
<td>0.02</td>
<td>0.98</td>
</tr>
<tr>
<td>Scyphistomae producing buds (%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(Intercept)</td>
<td>-3.82</td>
<td>0.35</td>
<td>-10.95</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Phytoplankton</td>
<td>-0.01</td>
<td>0.03</td>
<td>-0.47</td>
<td>0.64</td>
</tr>
<tr>
<td>Microzooplankton</td>
<td>0.26</td>
<td>0.03</td>
<td>10.19</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Mesozooplankton</td>
<td>0.00</td>
<td>0.02</td>
<td>-0.09</td>
<td>0.93</td>
</tr>
</tbody>
</table>
Table 3: Stable δ¹³C and δ¹⁵N isotope signatures (mean ± SD) of *A. coerulea* and organic matter sources used in MixSIAR model for each isotopic niche period. Sources A are the values of organic matter sources used for scyphistomae models, including all data, while Sources B are the values of organic matter sources collected from February to May, used for medusae models. *n* is the number of samples used to calculate each mean. SP: small plankton; Mesoz.: mesozooplankton; SOM: sedimentary organic matter.

| Source Type | Period 1 | | Period 2 | | Period 3 | |
|-------------|----------|--------|----------|--------|----------|
|              | δ¹³C     | δ¹⁵N   | n        | δ¹³C     | δ¹⁵N   | n        | δ¹³C     | δ¹⁵N   | n        |
| **Scyphistomae** | -22.8 (0.4) | 8.0 (0.5) | 18 | -19.3 (0.2) | 9.0 (0.1) | 9 | -21.1 (0.3) | 8.5 (0.4) | 9 |
| **Medusae** | -23.4 (0.7) | 8.1 (0.3) | 13 | -19.4 (0.5) | 8.9 (0.3) | 7 |
| **Sources A** | | | | | | | | | |
| SP | -22.1 (2.0) | 6.5 (0.3) | 18 | -20.6 (0.8) | 6.2 (0.7) | 22 | -21.0 (0.9) | 6.7 (0.3) | 6 |
| Mesoz. | -22.9 (0.9) | 8.0 (0.4) | 9 | -19.2 (0.7) | 7.4 (0.3) | 12 | -20.1 (0.1) | 7.5 (0.0) | 3 |
| SOM | -20.2 (0.9) | 5.5 (0.3) | 6 | -20.6 (0.1) | 5.4 (0.2) | 6 | -20.7 (0.0) | 5.3 (0.0) | 2 |
| **Sources B** | | | | | | | | | |
| SP | -23.3 (0.9) | 6.4 (0.3) | 12 | -20.9 (0.5) | 5.8 (0.3) | 15 |
| Mesoz. | -23.4 (0.3) | 8.2 (0.2) | 6 | -18.8 (0.2) | 7.3 (0.3) | 9 |
| SOM | -18.9 (0.0) | 5.8 (0.1) | 2 | -20.5 (0.0) | 5.6 (0.0) | 2 |
Fig. 1: Map of the Thau lagoon showing the location of the benthic (star) and pelagic (dot) sampling stations for this study. Shaded areas represent urban zones and grey points represent oyster farms.
Fig. 2: Index of relative importance of the three main prey groups found in the guts of *A. coerulea* medusae and the bell diameter of all individuals collected for gut content analysis.
Fig. 3: Temporal variability of phytoplankton ($\times 10^3$ cell L$^{-1}$), microzooplankton ($\times 10^3$ cell L$^{-1}$), and mesozooplankton ($\times 10^3$ ind m$^{-3}$) abundance collected in the Thau lagoon during the study period. All values represent monthly mean ± SD. In June 2017 (*), the mean (± SD) of mesozooplankton abundance was 90,895 ± 107,072 ind m$^{-3}$. 
Fig. 4: Temporal variability of the *A. coerulea* benthic population dynamics and the abundance of plankton in the Thau lagoon during the study period (adapted from Marques et al. 2019). Black lines represent the percentage of scyphistomae coverage (i.e., an indicator of population size) and the percentage of the scyphistomae producing buds. Each point represents replicate means and vertical lines are SD (see Marques et al 2019 for further information). Coloured lines represent the non-averaged abundance (after logarithmic transformation) of phytoplankton (cell L\(^{-1}\)), microzooplankton (cell L\(^{-1}\)), and mesozooplankton (ind m\(^{-3}\)).
Fig. 5: Temporal variability of $\delta^{13}$C (a) and $\delta^{15}$N (b) of *A. coerulea* scyphistomae, medusae, and ephyrae in Thau. All values represent monthly means ± SD. Background colours represent the different isotopic niche periods (periods 1, 2, and 3 in blue, green, and yellow, respectively; see Fig. 6). May represents a transitional period and it was not included in any isotopic niche period.
Fig. 6: Time trajectory of the evolution of the isotope signature, averaged by month, from *A. coerulea* scyphistomae (a) and medusae (b). Letters represent months (from January 2017 to January 2018). Coloured points represent isotopic niche periods defined after cluster analysis: period 1 is from January to April 2017 and from December 2017 to January 2018; period 2 is from June to August 2017 and period 3 is from September to November 2017. May represents the transition between periods 1 and 2 and was therefore not included in any isotopic niche period.
Fig. 7: Monthly variability of the δ¹³C (a) and δ¹⁵N (b) of the organic matter sources collected in this study. SOM: sedimentary organic matter.
Fig. 8: Proportion of the contribution of each organic matter source to the diet of *A. coerulea* scyphistomae and medusae during the different isotopic niche periods. The proportion was calculated using MixSIAR mixing models. The points indicate the median and the horizontal bars represent 75% and 95% Bayesian credibility intervals. SOM: sedimentary organic matter, SP: small plankton, Mesoz.: mesozooplankton.
Fig. 9: Biplot of isotope values of *A. coerulea* ephyrae, medusae, and scyphistomae. Ellipses indicate their isotopic niche in the Thau lagoon (as 95% confidence ellipse of the bivariate means), during the different isotopic niche periods. Grey areas and associated values indicate the percentage of overlap, when observed.
Fig. 10: Biplot of isotope values of *A. coerulea* ephyrae, medusae, scyphistomae, and oysters (*C. gigas*). Ellipses indicate their isotopic niche in the Thau lagoon, considering the whole study period (as 95% confidence ellipse of the bivariate means). Dark and light grey areas indicate niche overlap between *A. coerulea* and wild or cultivated oysters, respectively. Associated values on the graph indicate the percentage of overlap with medusae (in red) and scyphistomae (in black). The shape of points represents isotopic niche periods (period $t$: transitional period, i.e., samples collected in May).
Fig. 6: Scyphistomae coverage (in black, from Marques et al. 2019) and medusae bell diameter of the individuals collected for stable isotope analysis in this study (in red). The arrows indicate the main periods of sexual and asexual reproduction of *A. coerulea* (after Marques et al. 2015b, 2019). The background colours represent the isotopic niche periods (periods 1, 2, and 3 in blue, green, and yellow, respectively).