

First simultaneous assessment of macro- and meiobenthic community response to juvenile shellfish culture in a Mediterranean coastal lagoon (Thau, France)

Élise Lacoste, Fehmi Boufahja, Corinne Pelaprat, Patrik Le Gall, Tom Berteaux, Gregory Messiaen, Serge Mortreux, Jocelyne Oheix, Vincent Ouisse, Emmanuelle Roque d'Orbcastel, et al.

▶ To cite this version:

Élise Lacoste, Fehmi Boufahja, Corinne Pelaprat, Patrik Le Gall, Tom Berteaux, et al.. First simultaneous assessment of macro- and meiobenthic community response to juvenile shellfish culture in a Mediterranean coastal lagoon (Thau, France). Ecological Indicators, 2020, 115, pp.106462. 10.1016/j.ecolind.2020.106462. hal-03411034

HAL Id: hal-03411034 https://hal.umontpellier.fr/hal-03411034

Submitted on 20 May 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



1 First simultaneous assessment of macro- and meiobenthic community response to 2 juvenile shellfish culture in a Mediterranean coastal lagoon (Thau, France) 3 Élise Lacoste^{a,b*}, Fehmi Boufahja^c, Corinne Pelaprat^d, Patrik Le Gall^a, Tom Berteaux^a, 4 5 Gregory Messiaen^a, Serge Mortreux^a, Jocelyne Oheix^a, Vincent Ouisse^a, Emmanuelle Roque d'Orbcastela, Nabila Gaertner-Mazounib, Marion Richarda 6 7 8 ^a MARBEC, Univ Montpellier, CNRS, Ifremer, IRD, Sète, France 9 ^b EIO-UMR 241, Université de la Polynésie française, Tahiti, Polynésie française 10 ^c Laboratory of Biomonitoring of the Environment, Coastal Ecology and Ecotoxicology Unit, Carthage 11 University, Faculty of Sciences of Bizerte, Zarzouna 7021, Tunisia 12 ^dBenthId, Benthos Identification, Escaudes, France 13 *Corresponding author: Élise Lacoste. UMR-241 EIO/ MARBEC: eliz.lacoste@gmail.com 14 15 Present address: Station Ifremer de Sète, Avenue Jean Monnet, 34200 Sète, France 16 17 18 **Abstract** Shellfish aquaculture has the potential to alter benthic assemblage composition with 19 20 subsequent modifications of ecosystem functioning. While the impacts of aquaculture on the 21 taxonomic structure of macro- and — to a lesser extent — meiofauna, have been widely 22 studied, the functional changes of these communities remain relatively unknown. The recent 23 development of biological trait analysis (BTA) has made it possible to produce information 24 about how ecosystem functioning could change across specific terrestrial or aquatic system. 25 In the present study, we used a BTA in parallel with standard taxonomic analysis to evaluate 26 how well the two approaches detected the potential influence of juvenile oyster culture on the 27 benthic community in the French Mediterranean Thau Lagoon. Two sites were sampled under 28 farm structures and compared with two reference sites beyond the influence of farming. This 29 study is the first detailed parallel description of macro- and meiofauna (nematodes). A total of 30 118 and 41 taxa were determined for macrofauna and nematodes respectively. Some taxa 31 were more abundant or exclusively observed under farm structures such as *Chaetozone gibber*

32 and Neanthes acuminata for macrofauna and Daptonema fallax and Anticoma eberthi for

33 nematodes.

34

35

36

37

38

39

40

41

Overall, our results indicate that biological traits and functional indices both detected the

impact of shellfish culture on benthic assemblages, whereas macrofauna taxonomic indices

revealed no difference. Our results thus suggest that trophic and ecological groups are

particularly good indicators of the effects of shellfish culture. This study confirms the

relevance of the functional approach, and more generally of multi-index approaches, to detect

the influence of aquaculture on benthic communities. Further work is required to test multiple

traits in different regions and under different systems, but this work paves the way for

environmental impact assessment using a trait based approach.

42

43

Key-words: Environmental Impact Assessment; Aquaculture; Benthic indicators; Trait based

Approach; Macrofauna; Meiofauna

45

46

48

49

50

51

52

53

54

55

56

44

1. Introduction

47 Coastal ecosystems are facing drastic environmental changes due to many human pressures

that threat the integrity of communities and may have major impacts on ecosystem

functioning and its ability to sustain ecosystem services (Halpern et al. 2008, Cardinale et al.

2012). In shallow costal ecosystems, benthic communities play a central ecological role in

benthic-pelagic coupling through their feeding and movements that promote nutrient and

oxygen exchange throughout the sediment layer. Thus, any alteration of these communities

may have implications for ecosystem structure and performance (Thrush et al. 2006,

Kristensen et al. 2014) and understanding how they respond to disturbance is crucial to

predict future changes and help maintain and improve the quality of marine ecosystems. To

date, most studies have focused on the response of benthic macrofaunal communities to

disturbances (Pratt et al. 2015, Dimitriou et al. 2017, Ellis et al. 2017). Recently, interest has been growing in the study of meiofauna, since it has been shown that these organisms may modulate biological interactions in the sediment layer (Bonaglia et al. 2014, Lacoste et al. 2018a) and play a significant role in benthic ecosystem processes and services (Schratzberger and Ingels 2017). Due to their widespread distribution, short life and reproductive cycles, high abundance and diversity, meiofauna are thus beginning to be used as biological indicators for the assessment and monitoring of aquatic ecosystems (Semprucci et al. 2015, Zeppilli et al. 2015, Bianchelli et al. 2018). Bivalve aquaculture has various environmental impacts (McKindsey et al. 2011, Brenner et al. 2014) including the accumulation of biodeposits on the bottom under culture sites that may results in local organic enrichment. Such disturbance typically affect benthic communities according to the Pearson and Rosenberg (1978) model of organic enrichment with a progressive increase in the abundance of enrichment-tolerant species in the vicinity of farms, whereas communities directly under the farming structures are dominated by a few opportunistic species (Callier et al. 2007, 2009, McKindsey et al. 2011). Several studies have reported lower richness and/or abundance of organisms, as well as differential community composition of macrofauna (Christensen et al. 2003, Harstein and Rowden 2004, Lacoste et al. 2018b), or meiofauna (Mirto et al. 2000, Mahmoudi et al. 2008, Netto and Valgas 2010) under farming structures compared with reference sites. One notable feature of these studies is the focus on structural attributes of the benthos (e.g. taxonomic composition, number of species) whereas functional changes are disregarded. Only a few studies have used a functional approach to investigate the impact of interactions between the environment and fish (Papageorgiou et al. 2009, Dimitriadis and Koutsoubas 2011), or mussel (Lacoste et al. 2019) aquaculture.

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

Growing awareness that changes in biodiversity may modify ecosystem functioning (e.g. Loreau et al. 2001, Naeem et al. 2009) has led to the recognition that impacts on the functional diversity of benthic assemblages, as opposed to their structural features, should be better quantified (e.g. Strong et al. 2015, Cernansky 2017). Due to functional redundancies in communities, observed changes in species composition may be buffered (Walker 1992, Snelgrove 1998) such that structural impacts on benthic assemblages are not always matched by their functional counterparts (Bolam 2012). Otherwise, trophic pathways (indicator of ecosystem functioning) may be impacted without apparent changes of the overall benthic diversity (Dubois et al. 2007). Thus, functional-based approaches, including biological trait analysis and the computation of functional diversity (FD) indices, provide useful proxies to investigate the effect of human disturbances at the ecosystem-functioning level (Mouillot et al. 2006, 2013) and to anticipate the potential loss of ecosystem services associated with biodiversity erosion (Cardinale et al. 2012). Moreover, this approach can be used to compare communities that have no species in common but do share traits. Thau Lagoon is a Mediterranean marine ecosystem of notable economic importance due to the many ecosystem services it supports, in particular, the significant oyster farming activity (about 7 000 tons per year, near 10% of the national production). A few authors have described changes in macrofaunal communities in relation to oyster culture (Mazouni et al. 1996, Duport et al. 2007, Thouzeau et al. 2007), and one study showed spatial variation of meiofauna abundance (Guelorget et al. 1994). But to date, no list has been published of meiofauna assemblages in the lagoon, even though these communities may respond more rapidly to biodeposition than macrofauna do and could be good indicators of lagoon ecological status. Potential associated functional changes in macro- and meiofauna remain unknown. Thus, in a global context of increasing intensity and frequency of climatic and anthropogenic disturbances, there is a need to describe and predict changes in functional

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

community structure to anticipate potential losses of ecosystem services associated with biodiversity modifications in coastal areas such as Thau Lagoon.

The aims of this study were therefore 1/ to document macrobenthic and meiobenthic (nematode) communities in relation to oyster culture in Thau Lagoon and 2/ to compare the response of some selected taxonomic and functional attributes to disturbances caused by shellfish culture for macro- and meiofauna assemblages. To our knowledge, this is the first time the impact of bivalve aquaculture on macro- and meiobenthic communities has been studied simultaneously using a traditional taxonomic based approach coupled with a functional approach.

2. Materials and Methods

2.1. Study area and sampling strategy

Thau Lagoon is a French Mediterranean semi-enclosed shallow lagoon (mean depth 4 m) covering an area of 68 km² (Fig. 1). It is the most important French Mediterranean shellfish farming zone with an annual production of about 7 330 tons of oysters in 2016 (DDTM34 2017) reared on spatially organized structures, called "tables". Each table is 50 × 12 m long and supports between 800 and 1200 suspended nets or ropes holding juvenile or adult oysters. Sixteen tables form a 'block' and the blocks are separated from each other by a distance of 150m. Annual variations in water temperature and salinity in the lagoon are large, varying from 3 °C to 30 °C and from 33 to 40 PSU respectively (Messiaen et al. 2019).

The study was conducted in a dedicated shellfish-farming zone in the western part of the lagoon (Fig. 1). As a part of a research program (see Richard et al. 2017, 2019), the study focused on juvenile production, whose impact on ecosystem has never previously been evaluated in Thau Lagoon. Samples were collected at two sites in the farmed zone directly below juvenile tables (TO and TE sites) and at two additional sites located at a distance of

200 m outside the farming zone (CO and CE sites) (Fig. 1). Some characteristics of the four sites are given in Table 1, the main difference being algal cover (*Halopytis* sp.) which was greater at TO and CO sites. Sampling was done during three surveys in 2015 (September 29–October 2; October 28–November 4 and December 1–December 4, hereafter referred to as October, November and December) covering the whole juvenile production period. Each date, three replicate sediment cores (9 cm diameter, 10 cm depth) were randomly collected by scuba divers at each site. The samples were preserved in 10% buffered formalin and later transferred into 70% alcohol.

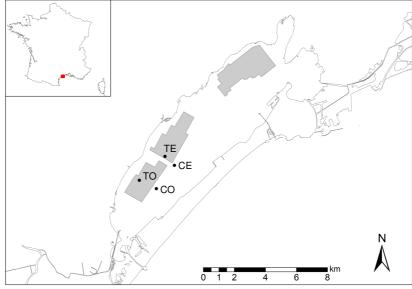


Figure 1. Location of the four sampling sites in Thau lagoon. The two T sites (TE & TO) were directly under the tables and the two reference sites (CE & CO) were located at a distance of 200 m from the shellfish farming area.

Table 1. Summary description of the 4 sampled sites

	Refere	ence sites	Farm sites		
	CO	CE	ТО	TE	
Depth	3.7 m	4.2 m	3.6 m	4.2 m	
Bottom observation					
Culture system	-	-	Juvenile oysters ca. 1.1 t per table*	Juvenile oysters ca. 1.4 t per table	

2.2. Abundance and species richness

Back to the laboratory, the content of each core sample was passed through a 1mm mesh sieve. Macrofauna in the > 1mm fraction was sorted and identified to the lowest taxonomic resolution possible, up to species in most cases. The < 1 mm fraction was decanted over a 40-µm sieve. All nematodes were sorted under a stereo-microscope, transferred to glycerol through a series of ethanol-glycerine solutions, mounted on glycerine slides (Somerfield et al. 1994) and identified to the lowest taxonomic level possible using the generic keys of Platt and Warwick (1983, 1988) and Warwick et al. (1998) and species descriptions in the literature.

Abundance (N) and species richness (S) of macrofauna and nematodes were characterized. The Shannon-Wiener diversity index (H') and Pielou's eveness (J') were calculated for each site. Taxa that were observed only once were removed from analyses (Clarke and Warwick 2001); all results are expressed per core (63 cm²).

2.3. Functional analysis

The common step in all functional ecology studies is to characterize the functional strategy of each species of interest by identifying a relevant combination of functional traits (Violle et al. 2007). There is currently no consensus on the methodology for selecting the most appropriate traits for a given study (Marchini et al. 2008). As the aim of the present study was to use traits as a proxy for assemblage response to shellfish culture, we decided to focus on the biological traits and ecological groups of macrofauna that have already been shown to be good indicators of organic enrichment in this specific context (Borja et al. 2000, Lacoste et al. 2019). To our knowledge, this is the first study to simultaneously evaluate the functional response of both macrofauna and meiofauna (nematodes) to bivalve aquaculture. We thus selected nematodes biological traits that have been shown to respond to organic enrichment

(Alves et al. 2013, Kandratavicius et al. 2018), with the assumption that they are comparable with those of macrofauna. Thus, selected traits for both assemblages were related to trophic guilds, ecological strategy, and interaction with sediments (Table 2). Each selected trait was subdivided into several modalities (Table 2). Organisms identified up to species level were classified in functional groups based on existing literature (e.g. Wieser 1953, Thistle et al. 1995, Queiros et al. 2013, WoRMS Editorial Board 2018). When reliable information was not available, data from the closest phylogenetic neighbor were used or the taxon was excluded from subsequent analysis. All nematodes and 44 macrofaunal (accounting for 91% of all individuals identified) taxa were thus retained for functional analysis. The subsequent 'trait per taxon' matrix was combined with the 'taxa per sample' matrix (abundance) to create a 'trait per sample' matrix. The relative abundance of the functional groups was then calculated by summing abundance across all taxa allocated to each functional group at a given site per sampling date. Finally, the two matrices were used to calculate functional indices (functional richness: FRic, functional eveness: FEve and functional dispersion: FDis) as outlined by Villéger et al. (2008) and Laliberté and Legendre (2015) for both macrofauna and nematodes. Using only categorical traits, FRic was defined as the number of unique trait combinations (equivalent to singular species). Although its results are ambiguous (Semprucci et al. 2015), the Index of Trophic Diversity (ITD, Heip et al. 1985) is popular and widely used for nematodes. We thus calculated ITD for both nematodes and macrofauna, using their respective trophic guilds, with the aim of evaluating its consistency in the case of disturbances caused by shellfish culture. The index was calculated following Heip et al. (1985): ITD= $\Sigma \theta^2$, where θ is the percentage contribution of each feeding type. Thus, a low ITD value corresponds to high trophic diversity (each group accounting for the same percentage).

196

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

Table 2. List of biological traits and respective categories selected for macrofauna and nematodes.

	Category	Level	Code
	Feeding group	Filter/suspension-feeder	F
		Grazer/Detritivorous	Gr
		Deposit-feeder	D
		Subsurface deposit-feeder	SubD
		Carnivorous	C
	Ecological group	Very sensitive	I
		Tolerant	II
		Indifferent	III
		2 ^d -order opportunistic	IV
Macrofauna		1 st -order opportunistic	V
	Mobility	Fixed in a tube	I
		Limited movement	II
		Slow movement	III
		Free movement (burrow system)	IV
	Reworking type	Epifauna	Е
		Surficial modifier	S
		Biodiffusers	В
		Upward conveyors	UC
		Upward/downward conveyors	UDC
	Feeding groups	Selective deposit-feeder	1A
		Non selective deposit-feeder	1B
		Epigrowth feeder	2A
		Predators/omnivores	2B
	Cp-score	Opportunistics	2
	•		3
		to	4
Nematodes		Persisters	5
	Adult shape	Long/thin	Lt
	•	Slender	Sl
	Amphid shape	Circular	Cr
	- •	Indistinct	Id
		Pocket-like	Pk
		Rounded or elongate loop	REL
		Spiral	Sp

2.4. Statistical analysis

The effect of Date × Condition (Farm vs Reference) on univariate measurements of benthic communities (taxonomic and functional indices) was tested using linear mixed effect models (IMEM; modelling normally distributed responses) with site as a random factor nested in

Condition. The effect of factors was considered as significant when the null vs main effect models (using maximum likelihood estimation) differed significantly (ANOVA, p < 0.05). Assumptions were assessed visually by examination of residual plots (Quinn and Keough 2002, Bates et al. 2015). All mixed effect models were constructed in R using *lme4* (Bates et al. 2015) and Tukey's HSD tests as implemented in library *emmeans* (Lenth 2019) were used as post hoc tests to compare conditions at each date if IMEM showed a significant effect of the interaction. Variation in multivariate assemblage structure (abundance data) was visualized using model-based ordination (generalized linear latent variable model) with the R package *gllvm* (Niku et al. 2019) and evaluated using multivariate linear modelling with the R package *mvabund* (Wang et al. 2012), assuming a negative binomial distribution. Species that contributed significantly to differences among conditions were identified using the *p.uni* argument that allows univariate 'species-by-species' comparisons in *mvabund*. Variation in assemblage functional composition between dates and conditions ('traits by site' matrix) was also compared using the same multivariate approach.

3. Results

- 221 3.1. Taxonomic indices
- 222 A total of 2 679 organisms belonging to 118 taxa were identified in the macrofaunal
- community in this study. Annelida (36.7%) and Mollusca (35.5%) were the most widely
- 224 represented group followed Arthropoda (25.8%). The mollusk Musculus subpictus alone
- represented 24.2% of the total sample, whereas we found a single individual of 42 taxa, which
- were subsequently removed from analyses.
- 227 A total of 778 nematodes belonging to 41 taxa were identified. Two taxa, *Paracomesma*
- 228 dubium and Spirina gerlachi, represented 34% of the organisms recorded and 6 taxa were

represented by a single individual. As this is the first time the nematodes have been described for Thau Lagoon, a complete list of taxa is provided in Table S1.

Table 3 lists the values of each index averaged per Condition (for all sampling dates). Statistical results are those of the Condition effect (pairwise comparison), except in case of a significant effect of Date \times Condition interaction. Although there was a trend of more taxa and greater abundance of macrofauna at reference sites, the Condition effect was not statistically significant (Table 3). The effect of Condition on the diversity indices was neither significant (Table 3).

A similar pattern was observed for nematodes, with the greatest abundance and richness values observed at reference sites (Table 3). IMEM revealed a significant effect of Date \times Condition on S, H' and J', with contrasts indicating higher values of S and H' in December at reference sites (S = 11 \pm 3 and H = 2.1 \pm 0.1) compared to farm sites (S = 4 \pm 1

and $H = 1.3 \pm 0.2$).

Table 3. Macrofauna and nematodes characteristics (mean \pm SE, n = 3) under juvenile oyster tables (Farm sites) and beyond the farming zone (Reference sites). Abundance (number of individuals, N) and richness (number of taxa, S), Shannon diversity (H') and Pielou's eveness (J'). Statistical effect of factors (Fc) Condition (Cd: Farm vs Reference) or Date \times Condition interaction (D \times Cd). *Significant difference at the 0.05 level.

		Farm	Reference	Fc	df	L-ratio	p
Macrofauna	N	40±8	106±16	Cd	3	5.592	0.133
	\mathbf{S}	13±1	21±2	Cd	3	2.941	0.401
	н'	2.1±0.1	2.3±0.1	Cd	3	2.740	0.480
	J'	0.8 ± 0.1	0.8 ± 0.1	Cd	3	5.791	0.122
Meiofauna	N	12±1	31±5	Cd	3	5.962	0.113
	\mathbf{S}	6±1	10±1	D×Cd	5	12.356	0.030*
	н'	1.5±0.1	1.9±0.1	D×Cd	5	20.704	<0.001*
	J'	0.9±0.1	0.8 ± 0.1	D×Cd	5	12.538	0.028*

3.2. Community structure

The model-based projection of macrofauna community structure (Fig. 2A) showed a clear distinction between conditions. The multivariate test confirmed the significant difference between communities at the farm vs at the reference sites (Table 4). Univariate comparisons were significant for five taxa; three of which were more abundant at the farm sites (Chaetozone gibber, Notomastus sp. and Neanthes acuminata) and two that were observed only at reference sites (*Phascolion strombus* and *Pusillina lineolata*). The ordination based on nematodes abundance (Fig. 2B) also showed a distinction between communities from the two conditions (Farm vs Reference), confirmed by the significance of the multivariate test (Table 4). Differences between sites within conditions were less clear relative to macrofauna as there was a major overlap of symbols representing samples from different sites (especially TO and TE) in the ordination graph (Fig. 2B). Nematode assemblages also varied among dates (Table 4). Pairwise comparisons indicated that assemblages differed significantly between October and November (p = 0.041). Among species that best explained variation among sites, Paracomesoma dubium, Anticomopsis longicaudata and Spirina gerlachi, were more abundant at the two reference sites. Conversely, one taxon was more abundant under farming structures than at the reference sites (Daptonema fallax) and three taxa were present only at the two farm sites (Anticoma eberthi, Eurystomina sp. and Metalinhomoeus sp.).

268269

270

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

Table 4. Results of the GLM testing the effect of Date, Condition and their interaction on macrofauna and nematode assemblages composition (abundance). *Significant difference at the 0.05 level.

		df	Dev.	p
Macrofauna	Condition	1	341.3	0.001*
	Date	2	214.7	0.109
	Condition \times Date	2	121.2	0.089
Meiofauna	Condition	1	141.83	0.001*
	Date	2	125.58	0.019*
	Condition \times Date	2	78.07	0.055

²⁷²

²⁷³

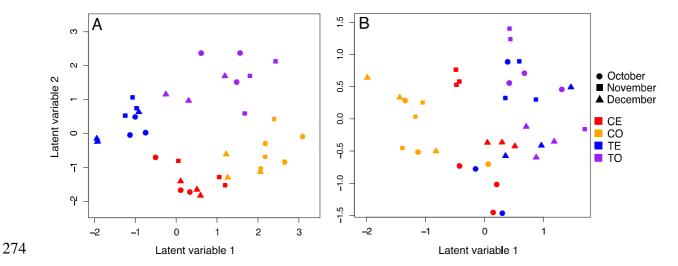


Figure 2. Model based biplot (unconstrained ordination) of abundance data for (A) macrofauna and (B) nematodes. Samples (cores) are shown in colors indexed by Site and Date (symbols).

3.3. Functional analysis

3.3.1 Macrofauna functional composition

Figure 3 shows the relative abundance of each biological trait. Although there was some variability within conditions and between dates, macrofauna at the reference sites was mainly characterized by filter-feeder organisms (ca 50%: Fig. 3), which are mostly sensitive to organic enrichment (ecological group I, Fig. 3). The proportion of filter-feeders was lower in the farming zone, whereas the proportion of deposit feeders and grazers increased (Fig. 3). These communities were dominated by animals that are mostly tolerant to organic enrichment or first-order opportunistic (ecological groups III and IV: Fig. 3). Mobility and reworking types did not allow us to clearly distinguish farming and reference sites (Fig. 3), except that upward-downward conveyors (UC and UDC: Fig. 3) appeared to be more abundant at the farm sites. The high abundance of surficial reworkers (S) at site TO (Fig. 3) was mainly due to the presence of many Arthropoda, also belonging to grazers (Gr) and ecological group I, and the the bivalve *Musculus subpictus*. These taxa were mainly found in association with the algae *Halopithys* sp. The GLM indicated a significant effect of Condition on the functional

composition of assemblage (df = 1, Dev. = 225.45, p < 0.001), with no significant effect of the interaction (Date × Condition, df = 2, Dev. = 48.39, p = 0.393).

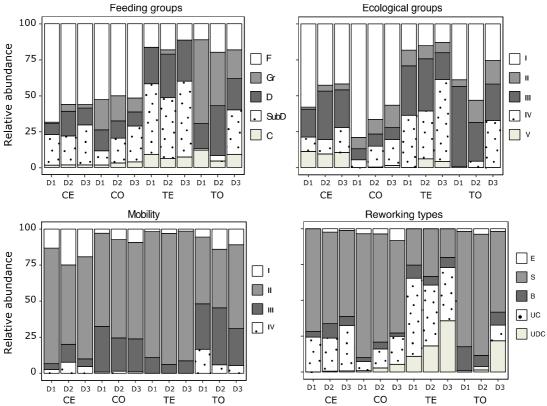


Figure 3. Relative abundance of the different macrofauna functional groups at the farm sites (TE, TO) and at the reference sites (CE, CO) on the three sampling dates (D1: October, D2: November, D3: December). Feeding groups: F: filter-feeder, Gr: grazer, D: deposit-feeder, SubD: subsurface deposit-feeder, C: carnivorous; Ecological groups from very sensitive (I) to opportunistic (V); Mobility from fixed (I) to burrower (IV); Reworking types: E: epifauna, S: surficial modifier, B: biodiffuser, UC: upward conveyor, UDC: up-/downward conveyor.

3.3.2. Nematode functional composition

The most representative feeding group at reference sites was epigrowth feeders (2A) whereas the percentage of detritivores (1B) was increased at the farm sites (Fig. 4). Overall, the community at the two reference sites was dominated by taxa with a cp score of 3 (Fig. 4) and amphids with a spiral shape (Sp: Fig. 4). At the two farm sites, the percentage of opportunistic nematodes (cp score of 2: Fig. 4) with circular amphid (Cr) increased (Fig. 4). The adult shape did not clearly differ betweenfarming zone sites and those outside the farming zone (Fig. 4). The high percentage of long/thin adults (lt) at site TO in September was mainly due

to the presence of *Metalinhomoeus longiseta*. The GLM indicated a significant effect of Condition on assemblage functional composition (df = 1, Dev. = 154.49, p < 0.001), with no significant effect of the interaction (Date × Condition, df = 2, Dev. = 48.22, p = 0.212).



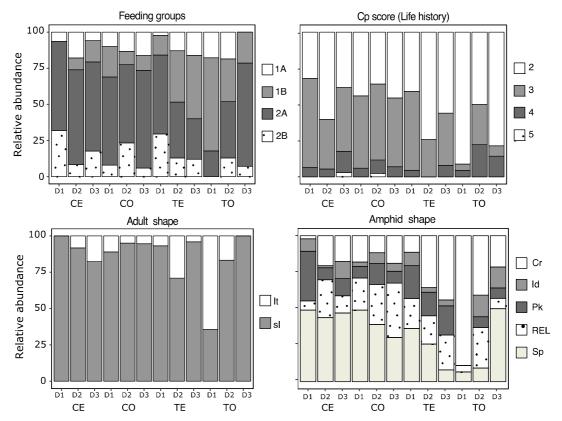


Figure 4. Relative abundance of the different nematode functional groups at the farm sites (TE, TO) and at the reference sites (CE, CO) on the three sampling dates (D1: October, D2: November, D3: December). Feeding groups: 1A: selective or 1B: non selective deposit-feeder, 2A: epigrowth feeder, 2B: predators; Life history: opportunistic (2) to persister (5); Adult shape: lt: long/thin, sl: slender; Amphid shape: Cr: circular, Id: indistinct, Pk: pocket-like, REL: rounded or elongate, Sp: spiral.

3.3.2. Functional indices

The lowest FRic values of both macrofauna and nematodes were recorded under the tables (TE and TO) (Table 5). The Date × Condition interaction had a significant effect for nematodes, with contrasts indicating significantly higher FRic values at reference sites in December (9.2 \pm 1.6) compared with the farm sites (4 \pm 0.6). Macrofauna FEve was higher under the tables in November (0.72 \pm 0.06) and December (0.72 \pm 0.04) than at the reference sites (0.57 \pm 0.04 and 0.56 \pm 0.04 respectively in November and December) whereas no

difference was found for nematodes. The Date \times Condition interaction had a significant effect on nematodes FDis but with no difference between conditions each date.

The ITD for macrofauna was significantly lower at the farm sites (Table 5) indicating that these sites presented a higher trophic diversity than did reference sites. For nematodes, the Date × Condition interaction had a significant effect on ITD but no difference was found between conditions at each date.

Table 5. Macrofauna and nematodes functional characteristics (mean \pm SE, n = 3) at sites under the juvenile oyster tables (Farm sites) and outside the farming zone (Reference sites). (FRic: functional richness, FEve: functional evenness, FDis: functional dispersion, ITD: Index of Trophic Diversity). Statistical effect of factors (Fc) Condition (Cd: Farm vs Reference) or Date \times Condition interaction (D \times Cd) *Significant difference at the 0.05 level.

		Farm	Reference	Fc	df	L-ratio	p
Macrofauna	FRic	8±1	12±1	D×Cd	5	10.657	0.05
	FEve	0.7 ± 0.1	0.5 ± 0.1	D×Cd	5	11.805	<0.05*
	FDis	0.4 ± 0.1	0.4 ± 0.1	Cd	3	1.520	0.677
	ITD	0.4 ± 0.1	0.7 ± 0.1	Cd	3	9.237	<0.05*
Meiofauna	FRic	6±1	9±1	D×Cd	5	12.326	<0.05*
	FEve	0.7 ± 0.1	0.7 ± 0.1	Cd	3	2.481	0.479
	FDis	0.3 ± 0.1	0.4 ± 0.1	D×Cd	5	13.69	0.017*
	ITD	0.5 ± 0.1	0.5 ± 0.1	D×Cd	5	13.043	<0.05*

4. Discussion

4.1. First description of nematodes in Thau Lagoon

With the exception of marine benthic ecosystems in Italy and Tunisia (Semprucci et al. 2008, Boufahja et l. 2014), there is a serious gap in knowledge concerning the Mediterranean meiofauna. This is particularly true for coastal lagoons, for which the spatial and temporal distribution of meiofauna, including free-living nematodes, is not well documented or is underestimated, despite they often greatly contribute to the biodiversity of coastal areas (Balsamo et al. 2012). Nematodes can provide critical information on the functioning of ecosystems and have already been proposed as potential indicators of anthropogenic

disturbance (Schratzberger et al. 2004, Danovaro et al. 2008, Moreno et al. 2011, Pusceddu et al. 2011). Our study is the first to provide a complete list of nematode taxa observed in the French Mediterranean Thau lagoon. Compared to the few studies that have documented other Mediterranean coastal areas, the number of taxa reported here from Thau Lagoon is similar but abundances were in the lower range (Mirto et al. 2000, Danovaro et al. 2004, Mahmoudi et al. 2008). The community structure showed many similarities with checklists of Italian and Tunisian lagoons, published respectively by Semprucci et al. (2008) and Boufahja et al. (2014). Ten of the common species found in Thau Lagoon have also been found in Italy (e.g. Anticoma eberthi, Longicyatholaimus longicaudatus) and 18 species were recorded in both Thau Lagoon and Tunisia (e.g. Spirinia gerlachi, S. parasitifera, Thalassironus britannicus, for details, see Table S1). These shared records could be linked to common environmental parameters among areas and to water exchanges between the two Mediterranean basins, which make large-scale dispersal between Tunisia, Italy and France possible. The checklist of nematodes resulting from this study could be the basis to further documenting meiobenthic diversity in Mediterranean coastal areas, with the aim to advance our understanding of its spatio-temporal variability.

369

370

371

372

373

374

375

376

377

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

4.2. Response of macrofaunal and nematode assemblages to shellfish culture

Overall, community structure (taxonomic and functional) differed between conditions, with lower abundance and richness (some significant, some not) at farm sites, as expected from previous studies (Mirto et al. 2000, Christensen et al. 2003, Callier et al. 2008). The presence of opportunistic species under the farm tables (e.g. *Neanthes acuminata*, *Prionospio fallax*, *Daptonema* sp.) whose abundance is favored under organically enriched conditions, showed that biodeposition by juvenile oysters impacts the benthic environment below farm structures in the study area. However, the high and similar species evenness values (J) at the farm and

reference sites revealed the absence of strongly dominant species that would be expected in an organically enriched environment (Pearson and Rosenberg 1978). The organic enrichment due to the presence of juvenile oysters is likely low compared with that of adult oysters, but the continuous exploitation of the area for farming may have modified community characteristics compared with outside of the farming zone. The results of the analysis of nematodes revealed differences in both univariate indices and community structure at the farm and reference sites. Our results for nematodes are in agreement with the study of Mirto et al. (2000) who reported decreased meiofauna richness and abundance in aquaculture sites relative to reference areas (Mirto et al. 2000). Inversely, a study in Bizerta Lagoon indicated increased nematode abundance at farm sites relative to control sites (Mahmoudi et al. 2008). The latter authors suggested that increased abundance was due to differential tolerance to organic matter, decreased competition, and a subsequent competitive release of more resistant species due to sediment organic enrichment. However, other factors (biotic and abiotic) could also explain the distribution of species since, for example, Paracomesma dubium was more abundant at farm sites in the study by Mahmoudi et al. (2008), whereas in our study, we observed a higher abundance at reference sites. Temperature variations may have contributed to explain temporal changes since when significant, the effect of Condition was mostly detected in December. In this study, macrofaunal taxonomic indices failed to detect a significant effect of shellfish culture whereas the multivariate approach did, as already reported in other studies (Grant et al. 2012, Lacoste et al. 2018a). Previous studies in Thau Lagoon produced conflicting results on the impact of aquaculture on macrofauna, ranging from low (Mazouni et al. 1996, Duport et al. 2007) to severe with a completely defaunated site in a deep shellfish area (Thouzeau et al. 2007). These authors highlighted the concomitant role of depth, algal cover and aquaculture in community composition. Our study also suggests that algal cover played an

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

important role, at least for the macrofauna community. Some arthropods, (e.g. *Gammarella* sp., *Cymodoce truncata*) were found attached to the algae *Halopithys* sp., so the TO and CO sites had specific assemblages structure due to these taxa.

The general trend thus supports the hypothesis that juvenile oysters slightly influenced both macrobenthic and nematode communities, although the variability between sites within conditions may have masked statistical differences in some indicators. Although this may require a labor-intensive sampling effort, it is recommended to sample several farm and reference sites to disentangle the influence of aquaculture on benthic communities, if such an influence exists, from the natural variability. This is particularly important in transitional waters as coastal lagoons where high natural variability may exist due to rapid environmental changes.

4.3. Insights from the functional approach

To date, the effect of aquaculture on benthic communities has usually been evaluated using traditional taxonomic approaches (Lacoste et al. 2020). Here, we demonstrate that some species traits can be used to predict the response of macro- and meiofauna to disturbances caused by shellfish culture since the distribution of traits and functional indices performed well in distinguishing between the farm and reference sites.

For both assemblages, trophic group and ecological group/life history were the traits that best responded to the presence of oyster tables. As reported in other studies, the presence of aquaculture facilities often decreases the abundance and diversity of filter-feeding organisms (Forrest and Creese 2006, Dubois et al. 2007, Fabi et al. 2009) whereas increased organic supply in farm zones favors the presence of non-selective deposit feeders. Following the classification of Grall and Glemarec (1997) and Borja et al. (2000), farming zones favored the presence of tolerant and opportunistic taxa whereas reference sites were dominated by

sensitive organisms. Among the meiofauna, epigrowth feeders were the most abundant organisms at reference sites and were replaced by non-selective deposit feeders in the farming zone. The reduction in the abundance of epigrowth feeders at the farm sites may be due to excluding competition with other groups but we can also hypothesize a lower abundance of diatoms under oyster tables due to a shading effect (McKindsey et al. 2011). Amphids are complex sensorial organs of nematodes used in the search of food and partners, and are useful for taxonomical identification. To date, there is no information linking amphids to a particular type of marine environment, except Semprucci et al. (2018), who described a potential link between inconspicuous amphids and rich food sources areas. Accordingly, our results suggest that the more intense and regular food supply at farm sites may limit the advantage of large amphids (REL, Sp) and hence that nematodes with smaller amphids (Cr) are more abundant in farming zones. However, this preliminary result requires further investigation to highlight potential links between morphological features of nematodes and their marine habitat. Functional diversity (FD) indices were computed for macrofauna and nematodes to evaluate the relevance of these indices in the case of aquaculture-induced disturbances. Nematode FRic was reduced at the farm sites and FD indices revealed the influence of aquaculture on macrofauna assemblages that was not detected using taxonomic indices. This confirms the usefulness of the functional approach, and more generally of multi-index approaches to detect the influence of farming (Borja et al. 2009, Lacoste et al. 2019). Interestingly, macrofauna FD indices indicated a higher trophic diversity and functional evenness at farm sites whereas no changes were detected at the richness level. The actual level of disturbance may therefore have shifted community composition that promoted a more even use of the trophic niche, which, in turn, may have increased the productivity of the system (Mason et al. 2005). Compared with the taxonomic approach, functional analysis not only offers the opportunity to

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

study the response of communities to the environment but also the subsequent impact of the changes on ecosystem properties (Lavorel and Garnier 2002, Lacoste et al. 2020), which is essential to evaluate the sustainable exploitation of aquaculture systems.

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

453

454

455

4.4 Limitation and future outlook of the functional approach

This work provides insights into the functional response of benthic communities to aquaculture-induced disturbance. We proposed the use of four selected traits that may respond both for macrofauna and nematode assemblages, that we assumed referred to similar behavior. The choice of biological traits may strongly influence the calculation of functional indices (Petchey and Gaston 2006). It was not our objective to test the most appropriate traits to use from a wide selection of possible choices. Rather we wished to highlight the potential of functional analysis to evaluate the impact of aquaculture using various kinds of benthic organisms (macrofauna and nematodes) and traits that have already been shown to respond to organic enrichment (Borja et al. 2000, Alves et al. 2013, Kandratavicius et al. 2018, Lacoste et al. 2019). Future experiments that examine multiple traits in different regions and systems will be needed to evaluate if functional analysis is a reliable tool to assess the benthic impacts of aquaculture. The number of taxa and the completeness and accuracy of the species trait data play a significant role in the estimation of functional trait metrics (Pakeman and Quested 2007, Pakeman 2014). In this exploratory study, we chose to use the maximum number of taxa for which traits were available. However, the inclusion of arthropods, for example, is questionable since it appears that these organisms were mostly associated with *Halopithys* sp. algae and are very mobile, so they could also have been classified as epifauna instead of infauna and disregarded. When we excluded them from the functional analysis (data not

shown), we obtained differences in the representation of relative abundances based on traits,

but no difference with respect to FD indices.

Although further studies are needed to identify the traits that react most consistently in a given manner or the taxonomic groups to considered, this work provides interesting information on the potential of the trait based approach to detect disturbances caused by aquaculture. Along with theoretical work, we think that further studies encompassing different kinds of disturbance across ecosystems will enable the implementation of a framework intended to include the functional approach in benthic monitoring programs.

Conclusion and Perspectives

This study documented the macrobenthic and the nematode communities in relation with juvenile oyster culture in Thau Lagoon. The trait-based approach we propose in this work shows the feasibility of detecting the impact of shellfish culture on the functional structure of both macro- and meiofauna communities in coastal lagoons, although further studies are needed to identify general patterns, within and across biogeographical regions and aquaculture systems. Combining taxonomic and functional approaches may add relevant ecological information and help obtain a clearer understanding of the functioning of benthic communities that are impacted by anthropogenic activities. For aquaculture, as well as other human-induced disturbance, the functional approach will undoubtedly enhance our ability to monitor activities and aid compliance with conservation strategies such as the Water Framework Directive Strategy (EC 2000), which requires an understanding of both how function varies spatially and how it responds to human disturbances. Several gaps that represent many research opportunities should however be addressed to 'operationalize' such approach. As examples i) new tools such as eDNA would allow to get rapid identification of

organisms once taxonomic validation of database will be improved, ii) further description of benthic organisms biological and ecological traits should be completed to feed databases.

503

504

Acknowledgements

- This work is a contribution to the MORTAFLUX program, funded by the Scientific Direction
- of Ifremer and by the EC2CO BIOHEFECT action (Coordinator: M. Richard). The
- 507 PostDoctoral fellowship of É. Lacoste was funded by the Ifremer and the University of
- 508 French Polynesia. Ifremer and IRD funded two missions for Dr Fehmi Boufahja in Sète in
- 509 2019 to facilitate nematode determination analysis and French-Tunisian exchanges.
- Macrofauna analysis were funded by the MARBEC UMR. The authors thank Nicolas
- 511 Cimiterra for the realization of the map (Figure 1). Many thanks to Geneviève Guillouet and
- 512 Zoely Rakotomonga-Rajaonah for their administrative help, and Daphne Goodfellow for
- 513 revising the English of the manuscript. We finally thank the anonymous reviewers for the
- 514 helpful comments on a previous version of this manuscript.

515

516 References

- 517 Alves, A.S., Adão, H., Ferrero, T.J., Marques, J.C., Costa, M.J., Patrício, J., 2013. Benthic
- meiofauna as indicator of ecological changes in estuarine ecosystems: The use of
- nematodes in ecological quality assessment. Ecol. Indic. 24, 462–475.
- 520 Balsamo, M., Semprucci, F., Frontalini, F., Coccioni, R., 2012. Meiofauna as a tool for
- marine ecosystem biomonitoring. In: Cruzado A (ed) Marine Ecosystems. InTech,
- 522 Rijeka, p 77-104
- Bates, D., Mächler, M., Bolker, B., Walker, S., 2015. Fitting Linear Mixed-Effects Models
- using lme4. J. Stat. Softw. 67. Bianchelli, S., Buschi, E., Danovaro, R., Pusceddu, A.,
- 525 2018. Nematode biodiversity and benthic trophic state are simple tools for the
- assessment of the environmental quality in coastal marine ecosystems. Ecol. Indic. 95,
- 527 270–287.
- 528 Bolam, S.G., 2012. Impacts of dredged material disposal on macrobenthic invertebrate
- communities: A comparison of structural and functional (secondary production) changes

- at disposal sites around England and Wales. Mar. Pollut. Bull. 64, 2199–2210.
- Bolam, S.G., Garcia, C., Eggleton, J., et al., 2017. Differences in biological traits composition
- of benthic assemblages between unimpacted habitats. Mar. Environ. Res. 126, 1–13.
- Bonaglia, S., Nascimento, F.J.A., Bartoli, M., Klawonn, I., Brüchert, V., 2014. Meiofauna
- increases bacterial denitrification in marine sediments. Nat. Commun. 5, 5133.
- Borja, A., Franco, J., Perez, V., 2000. A Marine Biotic Index to establish the ecological
- quality of soft-bottom benthos within European estuarine and coastal environments. Mar.
- 537 Pollut. Bull. 40, 1100–1114.
- Borja, Á., Rodríguez, J. G., Black, K., Bodoy, A., Emblow, C., Fernandes, T. F., ... &
- Papageorgiou, N., 2009. Assessing the suitability of a range of benthic indices in the
- evaluation of environmental impact of fin and shellfish aquaculture located in sites across
- 541 Europe. Aquaculture, 293(3-4), 231-240.
- Boufahja, F., Vitiello, P., Aissa, P., 2014. More than 35 years of studies on marine nematodes
- from Tunisia: a checklist of species and their distribution. Zootaxa 3786(3):269-300.
- Brenner, M., Fraser, D., Van Nieuwenhove, K., O'Beirn, F., Buck, B. H., Mazurié, J., ... &
- Flimlin, G., 2014. Bivalve aquaculture transfers in Atlantic Europe. Part B:
- environmental impacts of transfer activities. Ocean Coast. Manag. 89, 139-146.
- 547 Callier, M.D., McKindsey, C.W., Desrosiers, G., 2007. Multi-scale spatial variations in
- benthic sediment geochemistry and macrofaunal communities under a suspended mussel
- 549 culture. Mar. Ecol. Prog. Ser. 348, 103–115.
- 550 Callier, M.D., McKindsey, C.W., and Desrosiers, G. 2008. Evaluation of indicators used to
- detect mussel farm influence on the benthos: two case studies in the Magdalen Islands,
- Eastern Canada. Aquaculture, 278: 77-88.
- 553 Callier, M.D., Richard, M., McKindsey, C.W., Archambault, P., Desrosiers, G., 2009.
- Responses of benthic macrofauna and biogeochemical fluxes to various levels of mussel
- biodeposition: An in situ "benthocosm" experiment. Mar. Pollut. Bull. 58, 1544–1553.
- Cardinale, B.J., Duffy, J.E., Gonzalez, A. et al., 2012. Biodiversity loss and its impact on
- 557 humanity. Nature 489, 326–326.
- 558 Cernansky, R., 2017. The biodiversity revolution. Nature 546, 22–24.
- Christensen, P.B., Glud, R.N., Dalsgaard, T., Gillespie, P., Bondo, P., 2003. Impacts of
- longline mussel farming on oxygen and nitrogen dynamics and biological communities
- of coastal sediments. Aquaculture 218, 567–588.
- Clarke, K., Warwick, R.M., 2001. Change in marine communities. An approach to statistical
- analysis and interpretation. Prim. Plymouth, UK 1–176.

- Danovaro, R., Gambi, C., Dell'Anno, A., Corinaldesi, C., Fraschetti, S., Vanreusel, A., Vincx,
- M., Gooday, A.J., 2008. Exponential decline of deep-sea ecosystem functioning linked to
- benthic biodiversity loss. Curr. Biol. 18, 1–8.
- Danovaro, R., Gambi, C., Luna, G.M., Mirto, S., 2004. Sustainable impact of mussel farming
- in the Adriatic Sea (Mediterranean Sea): Evidence from biochemical, microbial and
- meiofaunal indicators. Mar. Pollut. Bull. 49, 325–333.
- 570 DDTM 34 Direction Départementale des Territoires et de la Mer, 2017. Bilan de la
- production conchylicole dans le département de l'Hérault en 2016.
- 572 http://www.herault.gouv.fr/content/download/24437/176354/file/Bilan-
- 573 d%C3%A9clarations-production-2016.pdf, Accessed date: 17 avril 2020
- 574 Dimitriadis, C., Koutsoubas, D., 2011. Functional diversity and species turnover of benthic
- invertebrates along a local environmental gradient induced by an aquaculture unit: The
- 576 contribution of species dispersal ability and rarity. Hydrobiologia 670, 307–315.
- 577 Dimitriou, P.D., Papageorgiou, N., Karakassis, I., 2017. Response of benthic macrofauna to
- eutrophication in a mesocosm experiment: Ecosystem resilience prevents hypoxic
- 579 conditions. Front. Mar. Sci. 4, 1–10.
- Dubois, S., Marin-Léal, J.C., Ropert, M., Lefebvre, S., 2007. Effects of oyster farming on
- macrofaunal assemblages associated with *Lanice conchilega* tubeworm populations: A
- trophic analysis using natural stable isotopes. Aquaculture 271, 336–349.
- Duport, E., Gilbert, F., Poggiale, J.C., Dedieu, K., Rabouille, C., Stora, G., 2007. Benthic
- macrofauna and sediment reworking quantification in contrasted environments in the
- Thau Lagoon. Estuar. Coast. Shelf Sci. 72, 522–533.
- Ellis, J.I., Clark, D., Atalah, J., Jiang, W., Taiapa, C., Patterson, M., Sinner, J., Hewitt, J.,
- 587 2017. Multiple stressor effects on marine infauna: Responses of estuarine taxa and
- functional traits to sedimentation, nutrient and metal loading. Sci. Rep. 7, 1–16.
- 589 European, Council, 2000. Directive 2000/60/EC of the European Parliament and of the
- Council of 23 October 2000 establishing a framework for Community action in the field
- of water policy.
- Fabi, G., Manoukian, S., Spagnolo, A., 2009. Impact of an open-sea suspended mussel culture
- on macrobenthic community (Western Adriatic Sea). Aquaculture 289, 54–63.
- 594 Forrest, B.M., Creese, R.G., 2006. Benthic impacts of intertidal oyster culture, with
- consideration of taxonomic sufficiency. Environ. Monit. Assess. 112, 159–176.
- 596 Grall, J., Glémarec, M., 1997. Using biotic indices to estimate macrobenthic community
- 597 perturbations in the Bay of Brest. Est. Coast. Shelf. Sc. 44, 43-53.

- 598 Grant, C., Archambault, P., Olivier, F., & McKindsey, C. W., 2012. Influence of 'bouchot'
- mussel culture on the benthic environment in a dynamic intertidal system. Aquaculture
- 600 Env Int, 2(2), 117-131.
- 601 Guelorget, O., Perthuisot, J.P., Lamy, N., Lefebvre, A., 1994. Structure et organisation de
- l'étang de Thau d'après la faune benthique (macrofaune, meiofaune). Relations avec le
- 603 confinement. Oceanol. Acta 17:105-11
- Halpern, B.S., Walbridge, S., Selkoe, K.A., et al., 2008. A global map of human impact on
- marine ecosystems. Science. 319, 948–952.
- Hartstein, N.D., Rowden, A.A., 2004. Effect of biodeposits from mussel culture on
- macroinvertebrate assemblages at sites of different hydrodynamic regime. Mar. Environ.
- 608 Res. 57, 339–357.
- Heip, C., Vincx, M., Vranken, G., 1985. The Ecology of Marine Nematodes. Oceanogr. Mar.
- 610 Biol. 23, 399–489.
- Huston, M., 1979. A general hypothesis of species diversity author. Am Nat 81-101.
- Kandratavicius, N., De Ward, C.P., Venturini, N., Giménez, L., Rodriguez, M., Muniz, P.,
- 613 2018. Response of estuarine free-living nematode assemblages to organic enrichment: an
- experimental approach. Mar. Ecol. Prog. Ser. 602, 117–133.
- Khedhri, I., Atoui, A., Ibrahim, M., Afli, A., Aleya, L., 2016. Assessment of surface sediment
- dynamics and response of benthic macrofauna assemblages in Boughrara Lagoon (SW
- Mediterranean Sea). Ecol. Indic. 70, 77–88.
- Kristensen, E., Delefosse, M., Quintana, C.O., Flindt, M.R., Valdemarsen, T., 2014. Influence
- of benthic macrofauna community shifts on ecosystem functioning in shallow estuaries.
- 620 Front. Mar. Sci. 1, 1–14.
- Lacoste, É., Drouin, A., Weise, A.M., Archambault, P., McKindsey, C.W., 2018a. Low
- benthic impact of an offshore mussel farm in Îles-de-la-Madeleine, eastern Canada.
- 623 Aquac. Environ. Interact. 10, 473–485.
- Lacoste, É., Piot, A., Archambault, P., McKindsey, C.W., Nozais, C., 2018b. Bioturbation
- activity of three macrofaunal species and the presence of meiofauna affect the abundance
- and composition of benthic bacterial communities. Mar. Environ. Res. 136, 62–70.
- Lacoste, É., Weise, A.M., Lavoie, M., Archambault, P., McKindsey, C.W., 2019. Changes in
- infaunal assemblage structure influence nutrient fluxes in sediment enriched by mussel
- biodeposition. Sci. Total Environ. 692, 39–48.
- Lacoste, É., McKindsey, C.W., Archambault, P., 2020. Biodiversity-Ecosystem Functioning
- (BEF) approach to further understanding aquaculture-environment interactions with

- application to bivalve culture and benthic ecosystems. Rev. Aquacult.
- 633 https://doi.org/10.1111/raq.12420
- 634 Laliberté, E., Legendre, P., Shipley, B., 2015. FD: measuring functional diversity from
- multiple traits, and other tools for functional ecology. R Packag. Version 1.0-12.
- 636 Lavorel, S., Garnier, E., 2002. Predicting changes in community composition and ecosystem
- functioning from plant traits: Revisiting the Holy Grail. Funct. Ecol. 16(5)-545-556
- 638 Lenth, R., 2019. emmeans: Estimated Marginal Means, aka Least-Squares Means. R package
- version 1.3.2. https://CRAN.R-project.org/package=emmeans
- 640 Loreau, M., Hector, A., 2001. Partitioning selection and complementarity in biodiversity
- experiments. Nature 412, 72–6.
- Mahmoudi, E., Hedfi, A., Essid, N., Beyrem, H., Aïssa, P., Boufahja, F., Vitiello, P., 2008.
- Mussel-farming effects on Mediterranean benthic nematode communities. Nematology
- 644 10, 323–333.
- Marchini, A., Munari, C., Mistri, M., 2008. Functions and ecological status of eight Italian
- lagoons examined using biological traits analysis (BTA). Mar. Pollut. Bull. 56, 1076-
- 647 1085.
- Mason, N.W.H., Mouillot, D., Lee, W.G., Wilson, J.B., 2005. Functional richness, functional
- evenness and functional divergence: The primary components of functional diversity.
- 650 Oikos 111, 112–118.
- Mazouni, N., Gaertner, J.C., Deslous-Paoli, J.M., Landrein, S., Geringer D'Oedenberg, M.,
- 652 1996. Nutrient and oxygen exchanges at the water-sediment interface in a shellfish
- farming lagoon (Thau, France). J. Exp. Mar. Bio. Ecol. 205, 91–113.
- Mckindsey, C., Archambault, P., Callier, M.D., Olivier, F., 2011. Influence of suspended and
- off-bottom mussel culture on the sea bottom and benthic habitats: a review. Can. J. Zool.
- 656 89, 622–646.
- Messiaen G., Mortreux S., Le Gall P., Crottier A., Lagarde F., 2019. Marine environmental
- station database of Thau lagoon. SEANOE. https://doi.org/10.17882/52404
- 659 Mirto, S., La rosa, T., Danovaro, R., Mazzola, A., 2000. Microbial and meiofaunal response
- to intensive mussel-farm biodeposition in coastal sediments of the Western
- Mediterranean. Mar. Pollut. Bull. 40, 244–252.
- Mouillot, D., Graham, N.A.J., Villéger, S., Mason, N.W.H., Bellwood, D.R., 2013. A
- functional approach reveals community responses to disturbances. Trends Ecol. Evol. 28,
- 664 167–177.
- Mouillot, D., Spatharis, S., Reizopoulou, S., Laugier, T., Sabetta, L., Basset, A., Chi, T. Do,

- 2006. Alternatives to taxonomic-based approaches to assess changes in transitional water
- communities. Aquat. Conserv. Mar. Freshw. Ecosyst. 16, 469–482.
- Moreno, M., Semprucci, F., Vezzulli, L., Balsamo, M., Fabiano, M., & Albertelli, G., 2011.
- The use of nematodes in assessing ecological quality status in the Mediterranean coastal
- 670 ecosystems. Ecol Ind, 11(2), 328-336.
- Naeem, S., Bunker, D. E., Hector, A., Loreau, M., & Perrings, C. (Eds.), 2009. Biodiversity,
- ecosystem functioning, and human wellbeing: an ecological and economic perspective.
- Oxford University Press. Oxford.
- Netto, S.A., Valgas, I., 2010. The response of nematode assemblages to intensive mussel
- farming in coastal sediments (Southern Brazil). Environ. Monit. Assess. 162, 81–93.
- Niku, J., Brooks, W., Herliansyah, R., Hui, F.K.C., Taskinen, S., Warton, D.I., 2018. gllvm:
- Generalized Linear Latent Variable Models. R package version 1.1.0.
- Pakeman, R.J., 2014. Functional trait metrics are sensitive to the completeness of the species'
- trait data? Methods Ecol. Evol. 5, 9–15.
- Pakeman, R.J., Quested, H.M., 2007. Sampling plant functional traits: What proportion of the
- species need to be measured? Appl. Veg. Sci. 10, 91–96.
- Papageorgiou, N., Sigala, K., Karakassis, I., 2009. Changes of macrofaunal functional
- composition at sedimentary habitats in the vicinity of fish farms. Estuar. Coast. Shelf
- 684 Sci. 83, 561–568.
- Pearson, T.H., Rosenberg, R., 1978. Macrobenthic succession in relation to organic
- 686 enrichment and pollution of the marine environment. Oceanogr. Mar. Biol. Annu. Rev.
- 687 16, 229–311.
- Petchey, O.L., Gaston, K.J., 2006. Functional diversity: back to basics and looking forward.
- 689 Ecol. Lett. 9, 741–758.
- 690 Platt, H.M., Warwick, R.M., 1983. Free living marine nematodes. Part I. British Enoplids.
- Synopses of the British Fauna No. 28, Cambridge University Press, Cambridge.
- 692 Platt, H.M., Warwick, R.M., 1988. Free-living marine nematodes. Part II. British
- 693 Chromadorids. Synopsis of the British fauna (New Series) No. 38.
- Pratt, D.R., Lohrer, A.M., Thrush, S.F., Hewitt, J.E., Townsend, M., Cartner, K., Pilditch,
- 695 C.A., Harris, R.J., Van Colen, C., Rodil, I.F., 2015. Detecting subtle shifts in ecosystem
- functioning in a dynamic estuarine environment. PLoS One 10, 1–16.
- Pusceddu, A., Bianchelli, S., Gambi, C., Danovaro, R., 2011. Assessment of benthic trophic
- status of marine coastal ecosystems: Significance of meiofaunal rare taxa. Estuar. Coast.
- 699 Shelf Sci. 93, 420–430.

- Queiros, A.M., Birchenough, S.N.R., Bremner, J., et al., 2013. A bioturbation classification of
- European marine infaunal invertebrates. Ecol. Evol. 3, 3958–3985.
- 702 Quinn, G.P., Keough, M.J., 2002. Experimental Design and Data Analysis for Biologists,
- 703 Cambridge University press. Cambridge.
- 704 Richard, M., Bec, B., Vanhuysse, C., et al., 2019. Changes in planktonic microbial
- components in interaction with juvenile oysters during a mortality episode in the Thau
- lagoon (France). Aquaculture 503, 231–241.
- Richard, M., Bourreau, J., Montagnani, C., Ouisse, V., Le Gall, P., Fortune, M., Munaron, D.,
- Messiaen, G., Callier, M.D., Roque d'Orbcastel, E., 2017. Influence of OSHV-1 oyster
- mortality episode on dissolved inorganic fluxes: An ex situ experiment at the individual
- 710 scale. Aquaculture 475, 40–51.
- 711 Schratzberger, M., Ingels, J., 2017. Meiofauna matters: The roles of meiofauna in benthic
- ecosystems. J. Exp. Mar. Bio. Ecol. 1–14.
- Schratzberger, M., Whomersley, P., Kilbride, R., Rees, H.L., 2004. Structure and taxonomic
- 714 composition of subtidal nematode and macrofauna assemblages at four stations around
- 715 the UK coast. J. Mar. Biol. Assoc. UK 84, 315–322.
- 716 Semprucci, F., Cesaroni, L., Guidi, L., Balsamo, M., 2018. Do the morphological and
- functional traits of free-living marine nematodes mirror taxonomical diversity? Mar.
- 718 Environ. Res. 135, 114–122.
- 719 Semprucci, F., Losi, V., Moreno, M., 2015. A review of Italian research on free-living marine
- nematodes and the future perspectives on their use as Ecological Indicators (EcoInds).
- 721 Mediterr. Mar. Sci. 16, 452–465.
- 722 Semprucci, F., Sandulli, R., de Zio Grimaldi, S., 2008. ADENOPHOREA NEMATODI
- 723 MARINI. Checklist della Fauna Marina Italiana. Biol. Mar. Medit. 15 (suppl.), 184-209.
- Snelgrove, P.V.R., 1998. The biodiversity of macrofaunal organisms in marine sediments.
- 725 Biodivers. Conserv. 7, 1123–1132.
- Somerfield, P.J., Gee, J.M., Warwick, R.M., 1994. Soft sediment meiofaunal community
- structure in relation to a long-term heavy metal gradient in the Fal Estuary system. Mar.
- 728 Ecol. Prog. Ser. 105, 79–88.
- 729 Strong, J.A., Andonegi, E., Bizsel, K.C., et al., 2015. Marine biodiversity and ecosystem
- function relationships: The potential for practical monitoring applications. Estuar. Coast.
- 731 Shelf Sci. 161, 46–64.
- Thistle, D., Lambshead, P.J.D., Sherman, K.M., 1995. Nematode tail-shape groups respond to
- environmental differences in the deep sea. Vie et Milieu 45(2)-105 :116.

- Thouzeau, G., Grall, J., Clavier, J., Chauvaud, L., Jean, F., Leynaert, A., Longphuirt, S.,
- Amice, E., Amouroux, D., 2007. Spatial and temporal variability of benthic
- biogeochemical fluxes associated with macrophytic and macrofaunal distributions in the
- 737 Thau lagoon (France). Estuar. Coast. Shelf Sci. 72, 432–446.
- 738 Thrush, S.F., Hewitt, J.E., Gibbs, M., Lundquist, C., Norkko, A., 2006. Functional role of
- large organisms in intertidal communities: Community effects and ecosystem function.
- 740 Ecosystems 9, 1029–1040.
- Villéger, S., Mason, N.W.H., Mouillot, D., 2008. New multidimensional functional diversity
- indices for a multifaceted framework in functional ecology. Ecology 89, 2290–2301.
- Violle, C., Navas, M.L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., Garnier, E., 2007.
- Let the concept of trait be functional! Oikos 116, 882–892.
- Walker, B.H., 1992. Biodiversity and Ecological Redundancy. Conserv. Biol. 6, 18–23.
- Wang, Y., Naumann, U., Wright, S.T., Warton, D.I., 2012. Mvabund an R package for
- model-based analysis of multivariate abundance data. Methods Ecol. Evol. 3, 471–474.
- Warwick, R.M., Platt, H.M. and Somerfield, P.J., 1998. Free-living Marine Nematodes. Part
- 749 III. British Monhysterida. Synopses of the British Fauna, vol. 53. Field Studies Council,
- 750 Shrewsbury.

757

- 751 Wieser, W., 1953. Die Bezichung swischen Mundho" hlengestalt, Erna"hrungsweise und
- Vorkommen bei freilebenden marinen Nematoden. Arkive for Zoologi 4, 439–484.
- Horton, T., Kroh, A., Ahyong, S. et al., 2019. World Register of Marine Species. Available
- from http://www.marinespecies.org at VLIZ. Accessed 2019-10-15. doi:10.14284/170
- 755 Zeppilli, D., Sarrazin, J., Leduc, D., et al., 2015. Is the meiofauna a good indicator for climate
- change and anthropogenic impacts? Mar. Biodivers. 45, 505–535.