



HAL
open science

Biotic community and landscape changes around the Eocene–Oligocene transition at Shapaja, Peruvian Amazonia: Regional or global drivers?

Pierre-Olivier Antoine, Johan Yans, Angélica Aliaga Castillo, Narla Stutz, M. Alejandra Alejandra Abello, Sylvain Adnet, Michele Andriolli Custódio, Aldo Benites-Palomino, Guillaume Billet, Myriam Boivin, et al.

► To cite this version:

Pierre-Olivier Antoine, Johan Yans, Angélica Aliaga Castillo, Narla Stutz, M. Alejandra Alejandra Abello, et al.. Biotic community and landscape changes around the Eocene–Oligocene transition at Shapaja, Peruvian Amazonia: Regional or global drivers?. *Global and Planetary Change*, 2021, 202, pp.103512. 10.1016/j.gloplacha.2021.103512. hal-03229171

HAL Id: hal-03229171

<https://hal.umontpellier.fr/hal-03229171>

Submitted on 18 May 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Biotic community and landscape changes around the Eocene–Oligocene transition at Shapaja,**
2 **Peruvian Amazonia: regional or global drivers?**

3 –
4 Pierre-Olivier Antoine*¹, Johan Yans², Angélica Aliaga Castillo^{3,4}, Narla Stutz^{1,5}, M. Alejandra
5 Abello^{6,7}, Sylvain Adnet¹, Michele Andrioli Custódio^{8,9}, Aldo Benites-Palomino^{3,10}, Guillaume
6 Billet¹¹, Myriam Boivin¹², Fabiany Herrera¹³, Carlos Jaramillo^{1,4,14}, Camila Martínez^{4,15}, Federico
7 Moreno^{4,16}, Rosa E. Navarrete¹⁷, Francisco Ricardo Negri¹⁸, Francisco Parra^{17,19}, François Pujos^{7,20},
8 Jean-Claude Rage¹¹, Ana Maria Ribeiro^{5,21}, Céline Robinet⁶, Martin Roddaz^{8,9}, Julia V. Tejada-
9 Lara^{1,3}, Rafael Varas-Malca^{3,22}, Roberto Ventura Santos⁸, Rodolfo Salas-Gismondi^{3,22}, and Laurent
10 Marivaux¹

11
12 ¹ Institut des Sciences de l'Évolution de Montpellier, Univ Montpellier, CNRS, IRD, EPHE, Montpellier,
13 France

14 ² University of Namur, Department of Geology, Institute of Life, Earth and Environment, ILEE, 61 rue de
15 Bruxelles, 5000 Namur, Belgium

16 ³ Departamento de Paleontología de Vertebrados, Museo de Historia Natural - Universidad Nacional Mayor
17 de San Marcos, Lima, Peru

18 ⁴ Smithsonian Tropical Research Institute, Unit 0948, APO AA 34002, Balboa, Ancon, 0843-03092, Panama

19 ⁵ Programa de Pós-Graduação em Geociências, Universidade Federal do Rio Grande do Sul, Av. Bento
20 Gonçalves, 9500, 91501-970 Porto Alegre, Brazil

21 ⁶ Unidades de investigación Anexo Museo, Facultad de Ciencias Naturales y Museo (UNLP). Av. 60 y 122.
22 B1900FWA, La Plata, Argentina

23 ⁷ Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Argentina

24 ⁸ Laboratório de Geocronologia, Instituto de Geociências, Universidade de Brasília, Brasília, DF 70910-000,
25 Brazil

26 ⁹ Géosciences-Environnement Toulouse, Université de Toulouse; UPS (SVT-OMP); CNRS; IRD; 14 Avenue
27 Édouard Belin, F-31400 Toulouse, France

28 ¹⁰ Paläontologisches Institut und Museum, Universität Zürich, Karl-Schmid-Straße 4, Zürich, 8006,
29 Switzerland

30 ¹¹ Sorbonne Universités, CR2P, MNHN, CNRS, UPMC-Paris 6, Muséum national d'Histoire naturelle, CP 38, 8
31 rue Buffon, 75231 Paris cedex 05, France

32 ¹² Instituto de Ecorregiones Andinas (INECOA), Universidad Nacional de Jujuy, CONICET, IdGyM, Av. Bolivia
33 1661, San Salvador de Jujuy 4600, Jujuy, Argentina

34 ¹³ Chicago Botanic Garden, 1000 Lake Cook Rd, Glencoe, IL 60022, USA

35 ¹⁴ Department of Geology, Faculty of Sciences, University of Salamanca, Salamanca, Spain

36 ¹⁵ Departamento de Ciencias Biológicas, Facultad de Ciencias, Universidad EAFIT, Medellín, Colombia

37 ¹⁶ Earth & Environmental Sciences, University of Rochester, 227 Hutchison Hall, University of Rochester,
38 Rochester, NY, 14627, USA.

39 ¹⁷ Paleosedes E.U. Tv 27 n°57-49 Campin, Bogotá, Colombia

40 ¹⁸ Laboratório de Paleontologia, Universidade Federal do Acre, Cruzeiro do Sul, Brésil

41 ¹⁹ Laboratorio de Bioquímico-Estratigrafía, Facultad de Geociencias, Universidad Nacional de Colombia,
42 Bogotá, Colombia

43 ²⁰ Instituto Argentino de Nivología, Glaciología y Ciencias Ambientales (IANIGLA), CCT-CONICET-Mendoza,
44 Av. Ruiz Leal s/n, Parque Gral. San Martín, 5500 Mendoza, Argentina

45 ²¹ Museu de Ciências Naturais, Secretaria do Meio Ambiente e Infraestrutura do Rio Grande do Sul, Av.
46 Salvador França, 1427, 90690-000, Porto Alegre, Brazil.

47 ²² BioGeoCiencias Lab, Facultad de Ciencias y Filosofía/CIDIS, Universidad Peruana Cayetano Heredia, Lima,
48 Perú

49
50
51 *Corresponding author: pierre-olivier.antoine@umontpellier.fr
52

53 Keywords. Andes-Amazonia transition; Pozo System demise; paleontology; sedimentary geology;
54 chemostratigraphy; paleoenvironments.

55

56 Abstract

57

58 Since 2012, we have investigated a stratigraphic section encompassing the late Eocene–earliest
59 Oligocene interval at Shapaja (Tarapoto area, Peruvian Amazonia, *ca.* 7°S), through paleontological
60 and geological fieldwork. The measured sedimentary series (120 m-thick [West] plus 90 m-thick
61 [East]), assigned to the upper member of the Pozo Formation, records fluvial micro-conglomeratic
62 lenses intercalated with floodplain and evaporite-rich fine red deposits, estuarine/coastal-plain
63 tidally-influenced fine sandstones, and oxbow lake nodule-rich blue clays. This sedimentary shift
64 coincides locally with the demise of the large Eocene coastal-plain wetland known as Pozo System.
65 The late Eocene–early Oligocene Shapaja section was extensively sampled for chemostratigraphy
66 ($\delta^{13}\text{C}$ on dispersed organic matter and pedogenic carbonate nodules), which in turn allowed for
67 refining the location of the Eocene-Oligocene Transition (EOT) and other climatic events
68 recognized at a global scale (i.e., Oi-1 and Oi-1a). The section has yielded nine fossil localities with
69 plant remains (leaves, wood, charophytes, and palynomorphs), mollusks, decapods, and/or
70 vertebrates (selachians, actinopterygians, lungfishes, amphibians, sauropsids, and mammals),
71 documenting ~130 distinct taxa. Four localities of the upper member of the Pozo Formation at
72 Shapaja predate the EOT, one is clearly within the EOT, while four are earliest/early Oligocene in
73 age. The small leaf impressions found along the Shapaja section could be indicative of dry and/or
74 seasonal conditions for this region throughout and after the EOT. Monkeys, indicative of tropical
75 rainforest environments, are only recorded in a latest Eocene locality (TAR-21). Two biotic
76 turnovers are perceptible in the selachian, metatherian, and rodent communities, well before the
77 EOT [~35–36 Ma] and a few hundred thousand years after the EOT [~33 Ma]. The latter turnover
78 seems to be primarily related to a global sea-level drop (ichthyofauna: marine-littoral elements
79 replaced by obligate freshwater taxa) and/or the onset of a drier and more seasonal climate in
80 early Oligocene times (terrestrial components). Changes in the structure of the Shapaja
81 paleocommunities were mostly locally driven by the flexural subsidence during the late Eocene,
82 and then globally driven by the earliest Oligocene climatic deterioration.

83

84 1. Introduction

85

86 The Eocene-Oligocene transition (EOT, *ca.* 33.9 million years ago) was one of the most
87 dramatic episodes of climatic, environmental, and biotic change recorded throughout the
88 Cenozoic era. It corresponds to a phase of oceanic reorganization, global cooling (*ca.* 4°C), and to
89 onset of semi-permanent ice sheets on Antarctica, which led to a noticeable drop in global sea
90 level (Mudelsee et al., 2014; Miller et al., 2020). Basically, the EOT marks the rapid shift from the
91 Paleocene–Eocene “greenhouse” Earth to the “icehouse” Earth during the earliest Oligocene. The
92 prevailing proposed mechanisms of that climatic deterioration include a decrease in the
93 concentration of atmospheric CO₂ (e.g., Zachos et al., 2001, 2008; DeConto & Pollard, 2003; Pagani
94 et al., 2005; Westerhold et al., 2020) and the opening of Southern Ocean gateways (Tasmania Sea
95 and Drake Passage) that established the Antarctic circumpolar current (e.g., Toumoulin et al.,
96 2020). Important changes occurred in floras and faunas around the world, reflecting the shift from
97 warm to colder/drier climatic regimes (e.g., Miller et al., 1991, Coxall and Pearson, 2007; Dupont-
98 Nivet et al., 2007; Hutchinson et al., 2018). This climatic transition consisted of alternating phases
99 of expansion and contraction of Antarctic ice sheets, reflected in successive episodes of sea level
100 rise and fall (Miller et al., 2020). Accordingly, the EOT was an interval of substantial ecological
101 reorganization, biotic turnovers, dispersals, extinction, provincialism, and speciation in many

102 biological groups, with patterns varying from landmass to landmass and across latitudes (e.g.,
103 [Stehlin, 1909](#); [Savage & Russell, 1983](#); [Janis, 1993](#); [Meng & McKenna, 1998](#); [Marivaux et al., 2005](#),
104 [2017a,b](#); [Coxall & Pearson, 2007](#); [Seiffert, 2007](#); [Liu et al., 2009](#); [Goin et al., 2010](#); [Beard et al.,](#)
105 [2017](#); [Godfrey et al., 2020](#)).

106 Continental proxies crossing the EOT, however, are somewhat sparse and they seem to be
107 much blurrier than in the marine records (e.g., [Berggren & Prothero, 1992](#); [Coxall & Pearson,](#)
108 [2007](#); [Zachos et al., 2008](#); [Tramoy et al., 2016](#); [Pound & Salzmann, 2017](#); [Westerhold et al., 2020](#)).
109 Furthermore, continental fossil records often lack the accurate time resolution that allows for a
110 precise correlation of biotic events and pulses with the earliest Oligocene glacial maxima, as
111 recorded by marine records. Also, compared with northern continents (e.g., [Coxall & Pearson,](#)
112 [2007](#); [Zanazzi et al., 2007](#)), much less is known about the tempo and intensity of biotic and
113 environmental changes that occurred across the EOT in tropical-equatorial areas of Africa (e.g.,
114 [Seiffert, 2007](#); [Beard et al., 2017](#); [Marivaux et al., 2017a,b](#); [Benammi et al., 2019](#)), South Asia
115 ([Marivaux et al., 2002, 2005](#); [Ni et al., 2016](#)), and more markedly, of South America ([Jaramillo et](#)
116 [al., 2006](#); [Hoorn et al., 2010](#)). Surprisingly, although with regionally-heterogeneous floras, no
117 drastic change in paleo-biome distribution, terrestrial isotopes and phytolith assemblages has
118 been detected between the late Eocene and the early Oligocene in the Southern cone of South
119 America (e.g., [Dunn et al., 2015](#); [Kohn et al., 2015](#); [Pound & Salzmann, 2017](#); [Barreda & Palazzesi,](#)
120 [in revision, this issue](#)). In contrast, substantial changes are recorded on palynomorphs at lower
121 latitudes, especially from north-east South America (Colombia and Venezuela), with a rapid phase
122 around the EOT, likely linked to the onset of the Antarctic glaciation ([Jaramillo et al., 2006](#)). At
123 tropical-equatorial latitudes, it seems that the global cooling provoked both a sustained drop in
124 precipitation and increased seasonality. So far, based only on vegetation proxies, the aridification
125 resulted in a decrease of plant morpho-diversity and in the opening of terrestrial environments,
126 hitherto dominated by evergreen rainforests ([Jaramillo et al., 2006](#); [Hoorn et al., 2010](#)).
127



128
 129 **Fig. 1.** Location (A) and geological map (B) of the studied area, around the Río Huallaga–Río Mayo
 130 confluence, with fossil localities of interest, at Shapaja, San Martín Department, Peruvian
 131 Amazonia. TAR-67 and TAR-66 localities (Pozo Sands; late middle–early late Eocene), significantly
 132 older than all other TAR- localities (based on stratigraphy, lithology, and distinct mammalian
 133 faunas). EOT, Eocene–Oligocene transition. Ideograms denote fossil content for the locality TAR-
 134 13, which is situated outside the measured sections illustrated in Figure 2.

135
 136 Here, we describe nine plurispecific fossil localities originating from Shapaja, San Martín
 137 Department, at *ca.* 7° S in Peruvian Amazonia, and distributed along a 120 + 50 m-thick section
 138 (Figs. 1, 2; see Geological setting for more details). This stratigraphic interval belongs to the upper
 139 member of the Pozo Formation [Fm.] (Pozo Shales) and encompasses the late Eocene–earliest
 140 Oligocene interval (Fig. 2). The concerned fossil localities yielded plant remains (leaves, seeds,
 141 wood, charophytes, and palynomorphs), mollusks, decapods, and/or vertebrates (selachians,
 142 actinopterygians, lungfishes, amphibians, turtles, crocodylomorphs, and mammals. Decapods and
 143 rodents originating from some of the concerned localities (TAR-20, TAR-21, TAR-22, TAR-13, and
 144 TAR-01) were previously studied by Klaus et al. (2017) and Boivin et al. (2018, 2019a, 2019b),
 145 respectively.

146 In the absence of radioisotopic datings so far available in the Shapaja section (e.g., through
 147 Ar/Ar ages on volcanoclastic sediments or U/Pb ages on detrital zircons), this section was
 148 extensively sampled for chemostratigraphy ($\delta^{13}\text{C}$ on dispersed organic matter and pedogenic
 149 carbonate nodules) in order to refine the local stratigraphy and to accurately locate the EOT, as

150 well as hypothetical hints of early Oligocene glaciations (Fig. 2; [Zachos et al., 2001, 2008;](#)
151 [Vandenberghe et al., 2012;](#) [Westerhold et al., 2020](#)). Indeed, C-isotopes on both organic matter
152 and pedogenic nodules have been judged as reliable material for chemostratigraphy in many
153 sections and paleoenvironments over the world, including terrestrial settings (e.g., [Gröcke et al.,](#)
154 [1999;](#) [Magioncalda et al., 2004;](#) [Noiret et al., 2016](#) and references therein). Ultimately, we also aim
155 to test the occurrence of biotic turnovers related to the EOT and subsequent climatic events,
156 through the analysis of potential shifts in terrestrial and aquatic communities along the concerned
157 section.

160 2. Regional setting

162 Thick Cretaceous–Cenozoic sections crop out continuously along river banks and newly-cut roads
163 in the San Martín Department. [Hermoza et al. \(2005\)](#) and [Roddaz et al. \(2010\)](#) provided a synthetic
164 overview of Cenozoic tectono-sedimentary history of Northern Peruvian foreland basin, recently
165 refined by [Eude et al. \(2015\)](#) and [Moreno et al. \(2020\)](#). The studied section, overhanging the
166 confluence between the Ríos Mayo and Huallaga (Fig. 1), was considered as documenting the
167 sandstone-dominated Chambira Fm., *i.e.* allegedly late Oligocene–early Miocene in age, in the
168 Utcucarca geological map, without any chronostratigraphic or biostratigraphic constraints
169 ([Sánchez Fernández et al., 1997](#)). Nevertheless, given that floodplain and fine-grained detrital
170 deposits (shales) are fully dominating, the concerned sequence might be referable as a whole to
171 the upper member of the underlying Pozo Fm. instead (“Upper Pozo (shale) member”; see
172 [Hermoza et al., 2005;](#) [Roddaz et al., 2010](#)). This shale-dominated member, further characterized
173 by shallow marine/littoral components, most probably spans the late Eocene–late Oligocene
174 interval in the northern Peruvian foreland basin ([Roddaz et al., 2010;](#) [Antoine et al., 2016](#)). Similar
175 deposits assigned to the Upper Pozo Shale member recently yielded various fossil remains near
176 Juanjuí, 80 km further to the SW (TAR-47, TAR-49, and TAR-50 localities; [Assemat et al., 2019](#)). In
177 contrast, the base of the Chambira Fm. (late Oligocene) overlies unconformably the sand-
178 dominated lower member of the Pozo Fm. (Pozo Sands, late middle–late Eocene) in the Ucayali
179 Basin, near Contamana ([Antoine et al., 2016](#)).

180 The fossil-yielding stratigraphic subsections at Shapaja are i) 120 m-thick, with a general
181 N80-110 direction and 20-30° W dipping for the western part and ii) 90 m-thick, with a general
182 N160 direction and 45° W dipping for the eastern part. A reverse fault delimits both sections, with
183 the eastern section overriding the western one (Fig. 1). A ~10 m-thick overlap is hypothesized
184 between the eastern and the western sections, on lithostratigraphical grounds. On both sides, the
185 stratigraphic sequence is dominated by fine-grained sediments (red shales and siltites), with
186 gypsum veins distributed obliquely with respect to the general dipping, and alternating with fine-
187 to coarse-grained yellow sandstones, with oxidized leaf litters (TAR-74, TAR-20, TAR-21sup, and
188 TAR-22 localities) and clay pebble-rich grey microconglomerates (TAR-72, TAR-21, and TAR-73
189 localities). Two lenses of carbonate nodule-rich blue clays further occur in the upper part of the
190 western section (TAR-13 and TAR-01 localities). Given the general geometry of deposits and
191 thrust, all nine successive fossil localities are distributed within these coarse-grained bodies, with
192 the following fossil locality sequence: TAR-74, TAR-20, TAR-72, TAR-21, TAR-21sup, TAR-13, TAR-
193 TAR-22, TAR-73, and TAR-01 (Fig. 1C). Lower in the eastern series two more fossil-yielding localities
194 were found, TAR-67 and TAR-66. These two localities are assignable to the lower member of the
195 Pozo Fm. (Pozo Sands) and they document a noticeably-earlier stratigraphic interval (late middle-
196 early late Eocene; Fig. 1). Their detailed analysis falls beyond the scope of the current work.

197
198

3. Material and Methods

The fossil localities studied here were discovered in 2012–2018 and investigated in 2012–2019 (see SI). Samplings for chemostratigraphical analyses were performed in 2018.

3.1 Chemostratigraphy

A total of 168 samples were collected in both western and eastern sections at Shapaja (137 and 31 samples, respectively). Carbon isotope analyses on dispersed organic matter ($\delta^{13}\text{C}_{\text{org}}$) were performed on 121 samples (Table 1). Samples not considered for organics had too low carbon content and/or major traces of roots, deep oxidation/weathering, preventing robust isotopic analyses. Carbon isotope analyses on pedogenic nodules ($\delta^{13}\text{C}_{\text{nod}}$) were performed on 92 samples, typically in red/brown pedogenic marls and siltstones (Table 1). In the Shapaja section, C-isotope analyses on bulk (carbonates) were not performed due to the fact that secondary fluids most likely perturbed the primary signal.

Organic matter of the sediments was isolated, following the procedure described in [Yans et al. \(2010\)](#) refined by [Storme et al. \(2012\)](#). The dispersed organic carbon isotope analyses ($\delta^{13}\text{C}_{\text{org}}$) are based on powdered rock samples of about 3 to 10 g, acidified in 25% HCl solution during 2 hours in order to remove carbonate. Soluble salts were removed by repetitive (1-6) centrifuging (4,000 revolutions per minute) with deionized water until a neutral sediment was obtained. Finally, residues were dried at 35° C and powdered again. Analyses were performed with an elemental analyzer (Carlo-Erba 1110) connected online to a ThermoFinnigan Delta V Plus mass spectrometer at the University of Erlangen. Accuracy and reproducibility of the analyses were checked by replicate analyses of international standards USGS40 and USGS41. The reproducibility of analyses is within 0.2 ‰ (1 σ).

Pedogenic nodules ($\delta^{13}\text{C}_{\text{nod}}$) were crushed and powders reacted with 100% phosphoric acid at 70° C using a Gasbench II[®] connected to a ThermoFinnigan V Plus[®] massspectrometer at the University of Erlangen. Isotopic compositions are calibrated with standards NBS19 for $\delta^{13}\text{C}_{\text{nod}}$.

Reproducibilities are within +/- 0.05 ‰ (1 σ). Each sample was analyzed 1 to 3 times. Organic $^{13}\text{C}/^{12}\text{C}$ values are normalized to the international VPDB standard (Vienna Pee Dee Belemnite).

Total organic content (%TOC) was measured with an elemental analyzer LECO[®] (CS-200). The CaCO₃ (%) content of the samples was measured with a Bernard Calcimeter.

3.2 Fossil plants

The macrofossil plant material of the Shapaja section corresponds to impressions of non-monocot angiosperm leaves and fruits, which were studied and stored at the Paleontological Collection of the Museo de Historia Natural de la Universidad Nacional Mayor de San Marcos Lima, Peru (DPV-MHN-UNMSM; inventory numbers PVMUSM-60 to -83). Fossil leaves were organized and described by morphotypes following the method proposed by [Peppe et al. \(2008\)](#). Each morphotype has a two-letter prefix (PZ) based on the formation name (Pozo Formation) plus a number starting from one.

Fossil seeds were recovered by screenwashing (see additional details below) from samples TAR-74, TAR-20, TAR-21, TAR-13, TAR-22, TAR-73, and TAR-01. The specimens are mostly preserved as silicified and/or iron-rich locule casts, and less commonly as carbonized remains.

Taxonomic affinities of the leaf, fruit, and seed morphotypes were proposed when possible based on the presence of taxonomically-informative characters (shape, size, margin type or venation patterns in leaves), extant material available for comparison, and the level of completeness of the description of each morphotype.

3.3 Other fossil remains (charophytes, mollusks, decapods, and vertebrates)

248 Large mollusk and vertebrate specimens uncovered were prepared mechanically in the field and at
249 lab. Small vertebrate specimens, decapod claws, and micromollusks were concentrated by screen-
250 washing *ca.* 2,000 kg of raw sediment, with 2 mm, 1 mm, and 0.7 mm meshes (12–762 kg per
251 locality; see SI for more details), using highly-diluted hydrogen peroxide, in 2012–2019. This
252 protocol also allowed for obtaining charophyte oogonia and seeds, as well as carbonized and
253 silicified wood chunks. Except when mentioned, all concerned specimens are permanently housed
254 in the DPV-MHN-UNMSM.

255 The palynological preparation was carried out at the Stratigraphy Research Institute (IIES) at
256 the Universidad de Caldas, Manizales, Colombia, following the [Traverse's \(2007\)](#) standard
257 procedure (37% hydrochloric acid, Hydrofluoric acid at 70% and density separation using Zinc
258 Chloride). Samples were mounted using Canada balsam.

259

260

261 4 Results

262

263 4.1 Chemostratigraphy

264 Carbon isotopic values on dispersed organic matter ($\delta^{13}\text{C}_{\text{org}}$) range from -26.9 to -21.0 ‰ (Table 1).
265 These data are in good agreement with the expected $\delta^{13}\text{C}$ values on organics at the Eocene-
266 Oligocene interval (see [Sarkar et al., 2003](#); [Benammi et al., 2019](#)). Carbon isotopic values on
267 pedogenic nodules ($\delta^{13}\text{C}_{\text{nod}}$) range from -5.7 ‰ to -11.8 ‰ (Table 1).

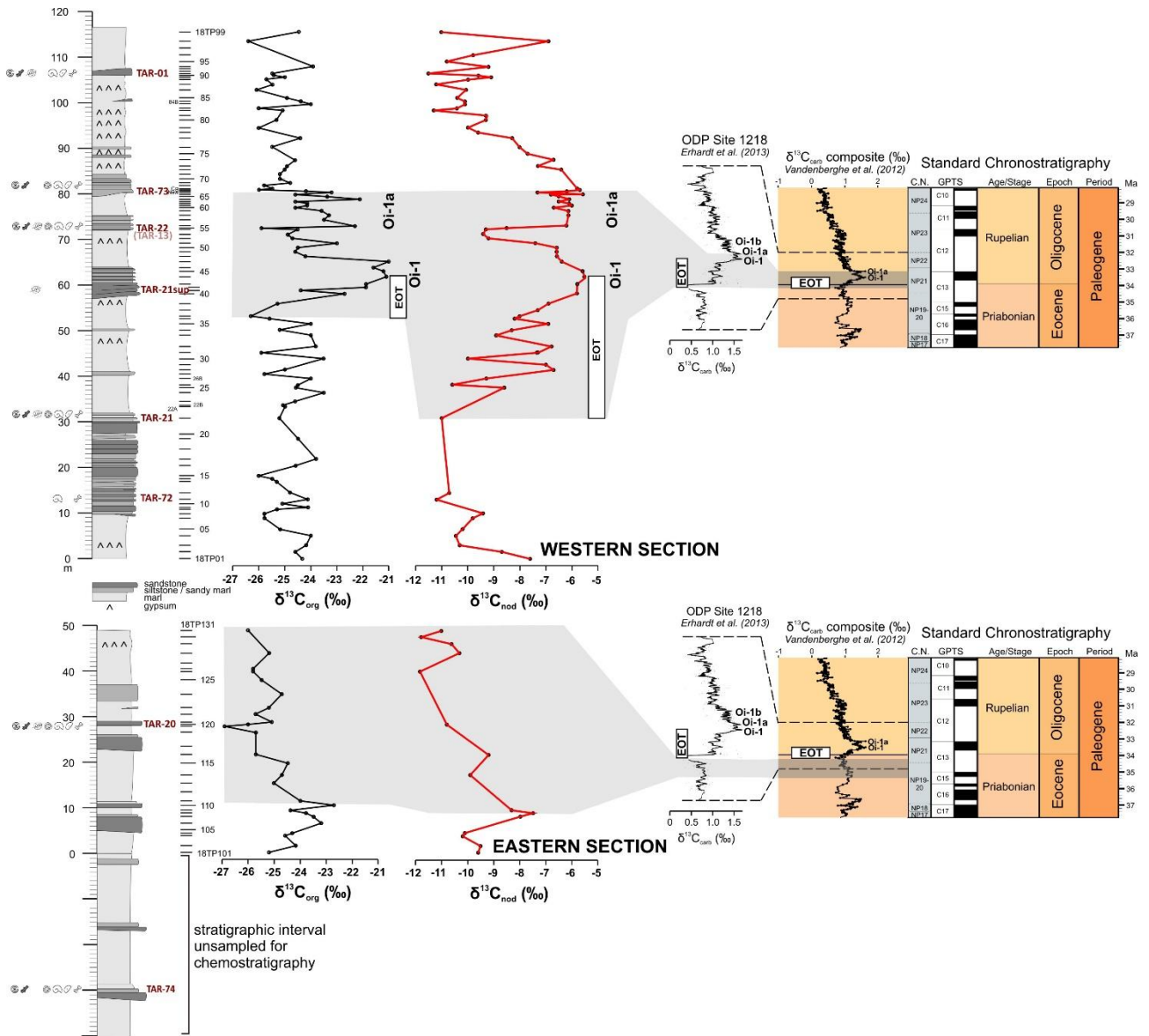
268 We provide a detailed description of all the isotopic results in SI. The most relevant
269 chemostratigraphic features are as follows: in the Eastern section, C-isotope values on both
270 organics and pedogenic nodules show (from base to top) a short positive trend followed by a quite
271 long negative trend (Fig. 2; Table 1). In the Western section, two main positive excursions are
272 observed. This pattern is particularly obvious for C-isotope data on pedogenic nodules (Fig. 2;
273 Table 1).

274 TOC content is very low for all the samples (around 0.05 %), whereas the CaCO₃ contents much
275 more satisfactorily range from 5 % to 66 % depending on the samples (Table 1).

Section	Sample	Height (m)	CaCO ₃ (%)	$\delta^{13}\text{C}_{\text{org}}$ (‰) - organics	$\delta^{13}\text{C}_{\text{nod}}$ (‰) - nodules	Section	Sample	Height (m)	CaCO ₃ (%)	$\delta^{13}\text{C}_{\text{org}}$ (‰) - organics	$\delta^{13}\text{C}_{\text{nod}}$ (‰) - nodules
Western	18TP99	115.5	11	-24.5	-11	Western	18TP35	51.5	18	-24	-6.9
Western	18TP98	113.5	8	-26.5	-6.9	Western	18TP34	50.1	64	-25.2	-8.3
Western	18TP97	112	6	TOC too low	-	Western	18TP33	49	18	-24.1	-8.9
Western	18TP96	110.5	13	TOC too low	-9.8	Western	18TP32	46.5	25	-23.8	-6.8
Western	18TP95	109	16	TOC too low	-10.7	Western	18TP31	45.2	24	-25.9	-7.4
Western	18TP94	108	18	-23.9	-9.2	Western	18TP30	43.8	15	-23.5	-10.1
Western	18TP93	107.9	49	TOC too low	-	Western	18TP29	42.5	27	TOC too low	-7
Western	18TP92	107.5	17	-24.5	-11.5	Western	18TP28	41.4	29	-25	-6.8
Western	18TP91	106.4	16	TOC too low	-	Western	18TP27	40.8	19	-25.8	-
Western	18TP90	106	8	-25.4	-9.5	Western	18TP26B	39.5	8	-24	-9.3
Western	18TP89	105.6	8	-25	-9.2	Western	18TP26	38.1	18	-24.5	-10.6
Western	18TP88	105.1	8	-25.7	-10	Western	18TP25	37.5	20	-24.6	-
Western	18TP87	104	13	-25.5	-11.2	Western	18TP24	36.4	14	-23.5	-
Western	18TP86	102.8	16	-26.1	-10	Western	18TP23	34.5	19	-24.6	-
Western	18TP85	101.1	17	-24.9	-10.3	Western	18TP22B	33.3	15	-25.1	-
Western	18TP84B	100.3	10	-24.4	-10.1	Western	18TP22A	33.6	13	-25	-
Western	18TP84	99.8	9	-24	-10.1	Western	18TP21	30.8	53	-25.2	-10.8
Western	18TP83	98.8	18	-26	-10.4	Western	18TP20	27.3	32	TOC too low	-
Western	18TP82	98.3	8	-24.1	-11.3	Western	18TP19	26.3	21	-24.5	-
Western	18TP81	97.6	66	TOC too low	-9.3	Western	18TP18	24	30	TOC too low	-
Western	18TP80	96.2	8	-25.4	-9.3	Western	18TP17	21.8	27	-23.8	-
Western	18TP79	94.5	24	-26	-10	Western	18TP16	20.4	15	-24.6	-
Western	18TP78	93.5	25	TOC too low	-9.6	Western	18TP15	18.2	12	-26	-
Western	18TP77	92.2	21	-24.4	-8.3	Western	18TP14	17.5	10	-25.5	-
Western	18TP76	91.2	22	-25.5	-8	Western	18TP13	16.9	14	-25.3	-
Western	18TP75	88.8	12	-22.4	-7.7	Western	18TP12	14.4	34	-24.8	-10.7
Western	18TP74	87.5	23	-24.5	-6.7	Western	18TP11	13	65	-24.1	-11.2
Western	18TP73	86.1	28	-24.9	-7.3	Western	18TP10	12	21	-25.1	-
Western	18TP72	85.3	28	-25	-6.4	Western	18TP09	11.2	19	-24.2	-
Western	18TP71	84.3	26	-25.2	-	Western	18TP08	10.8	10	-25.2	-
Western	18TP70	83.3	27	-25.2	-	Western	18TP07	9.8	32	-25.8	-8.8
Western	18TP69	82.3	26	-24.8	-	Western	18TP06	8.9	27	-25.8	-9.8
Western	18TP68	81.2	23	-25.8	-	Western	18TP05	6.5	10	-25.2	-10.2
Western	18TP67C	81.2	27	-25.5	-5.8	Western	18TP04	5	8	-24	-10.6
Western	18TP67B	80.75	24	-26	-5.7	Western	18TP03	2.9	15	-24.2	-10.3
Western	18TP67NEW	80.3	28	-24.3	-6.2	Western	18TP02	1.5	10	-24.6	-8.7
Western	18TP67	80.3	33	-23.2	-6.5	Western	18TP01	0	5	-24.3	-7.7
Western	18TP66	80	25	-24.6	-6.8	Eastern	18TP131	48.9	24	-26	-11.1
Western	18TP65	79.4	24	-23.4	-6.5	Eastern	18TP130	47.5	13	TOC too low	-11.8
Western	18TP64	78.8	24	-22.1	-6.1	Eastern	18TP129	46	25	TOC too low	-10.8
Western	18TP63	78.3	22	-24.6	-6.6	Eastern	18TP128	43.9	14	-25.2	-10.2
Western	18TP62	77.9	23	-24.1	-6.1	Eastern	18TP127	40.5	9	-25.8	-
Western	18TP61	77.5	21	-24.1	-6	Eastern	18TP126	39.9	11	-25.8	-11.8
Western	18TP60	77	20	-24.6	-6.7	Eastern	18TP125	38.1	8	-25.5	-
Western	18TP59	76.3	24	-23.6	-6.1	Eastern	18TP124	34.9	11	-24.7	-
Western	18TP58	75.3	22	-23.3	-6.1	Eastern	18TP123	32	9	-25.2	-
Western	18TP57	74.3	20	-23.5	-	Eastern	18TP122	30.5	10	-25.7	-
Western	18TP56	73	19	-22.3	-6.2	Eastern	18TP121	28.8	12	-25.1	-
Western	18TP55	72.5	31	-25.9	-8.7	Eastern	18TP120	28.3	13	-26	-10.8
Western	18TP54	72.1	13	-24.5	-9.3	Eastern	18TP119	27.9	32	-26.9	-
Western	18TP53	71.1	15	-24.9	-9.4	Eastern	18TP118	26.6	30	-25.7	-
Western	18TP52	70.2	17	-24.7	-9.2	Eastern	18TP117	23.5	24	TOC too low	-
Western	18TP51	69.2	20	-23	-7.4	Eastern	18TP116	21.7	9	-25.7	-9.2
Western	18TP50	68.2	23	-24.5	-6.6	Eastern	18TP115	19.8	10	-24.5	-
Western	18TP49	67.3	23	-24.6	-6.6	Eastern	18TP114	17.2	10	-24.7	-9.9
Western	18TP48	66.3	21	-24.2	-6.6	Eastern	18TP113	15.4	5	-25	-
Western	18TP47	65.1	20	-21	-6.4	Eastern	18TP112	13.5	5	TOC too low	-
Western	18TP46	63.8	15	-21.6	-	Eastern	18TP111	11.5	6	-24	-
Western	18TP45	63	16	-21.2	-5.6	Eastern	18TP110	10.5	18	-22.7	-
Western	18TP44	61.8	21	-21.1	-5.5	Eastern	18TP109	9.5	13	-24.4	-9.6
Western	18TP43	60.2	22	-21.9	-5.8	Eastern	18TP108	8.9	11	-23.8	-7.5
Western	18TP42	59.5	20	-21.9	-	Eastern	18TP107	8.1	10	-23.5	-8
Western	18TP41	58.8	20	-24.4	-	Eastern	18TP106	6.7	13	-23.2	-
Western	18TP40	58.1	24	-22.7	-5.8	Eastern	18TP105	5.2	11	-25.2	-
Western	18TP39	55.9	9	-25.7	-6.7	Eastern	18TP104	4.5	22	-24.3	-10.1
Western	18TP38	54.5	12	TOC too low	-7.3	Eastern	18TP103	4	12	-24.6	-10.2
Western	18TP37	53.2	16	-26.3	-8	Eastern	18TP102	1.8	19	-24.2	-9.5
Western	18TP36	52.6	20	-25.6	-8.2	Eastern	18TP101	0.2	13	-25.2	-9.6

277
278
279

Table 1. Chemostratigraphical data of the Shapaja area, including section, sample labels, heights (m), CaCO₃ content (%), δ¹³C_{org} values (‰, VPDB) and δ¹³C_{nod} values (‰, VPDB).



280
281
282
283
284
285
286
287
288
289
290
291
292
293
294
295
296

Fig. 2. Stratigraphic sections encompassing the late Eocene–early Oligocene interval at Shapaja, with location of chemostratigraphical samples, resulting δ¹³C curves (on organic matter [δ¹³C_{org}] and pedogenic carbonate nodules [δ¹³C_{nod}], in ‰), and chronological interpretations (right) based on δ¹³C curves around the Eocene-Oligocene transition in ODP Site 1218 (Erhardt et al., 2013) and reference δ¹³C composite curve (Cramer et al., 2009 modified by Vandenberghe et al., 2012). Top = western section; bottom = eastern section. Ideograms denote fossil content (charophytes, wood, leaf imprints, seeds/fruits, mollusks, crustaceans, and vertebrates, respectively; see Fig. 1). C.N., Calcareous Nannoplankton. EOT, Eocene–Oligocene Transition. GPTS, geomagnetic polarity timescale.

4.2 Plant remains

4.2.1 Leaf and fruit impressions (Fig. 3 A-H)

Ten magnoliopsid dicot morphotypes were recognized in the Shapaja section, through leaf and fruit impressions (nine morphotypes and one [PZ4], respectively), recovered from TAR-20 (late

297 Eocene), TAR-21sup (EOT), and TAR-01 (early Oligocene). Most morphotypes are documented by
298 several specimens. These morphotypes are fully described, compared, and illustrated in the SI.
299 TAR-20 yielded a single leaf morphotype (PZ5; Fig. S1M) and TAR-21sup three (PZ1-3; Fig. 3A-C), all
300 of uncertain affinities. TAR-01 provided by far the most diversified macroflora, with a fruit
301 morphotype (PZ4: legume; Fig. 3D) and seven leaf morphotypes (PZ1, PZ3, PZ6-10; Fig. 3E-G).
302 Three of them document Fabaceae Fabales (PZ4, 8, and 9). Leaf morphotypes PZ1 and PZ3 were
303 recognized both at TAR-21sup and TAR-01.

304 All leaves from the Shapaja section have entire margins and pinnate primary venation. Most
305 have nanophyll and/or microphyll leaflet laminar sizes (dimorphic leaflets in PZ10; Fig. 3F), with
306 petiolate leaf and marginal blade attachments. PZ1 and PZ2 have larger leaflets (mesophyll and
307 notophyll, respectively; Fig. 3A,C). Some leaves are compound pinnae or bipinnate, imparipinnate
308 with opposite (PZ8; Fig. 3F) or alternate (PZ9; Fig. 3G), non-overlapping pinnules. The
309 leaves/leaflets are either oblong, obovate, ovate, or elliptic in shape (PZ9), mostly with an acute
310 apex. Major secondaries are brochidodromous in most morphotypes, except for PZ6 and PZ8
311 (craspedodromous). PZ2 has affinities with the late Miocene morphotype IP2 from Shumanza
312 ([Feussom-Tcheumeleu et al., 2019](#)) and with *Machaerium* (Fabaceae). All four fruit specimens
313 available at TAR-01 are straight and symmetric legumes, with placental and non-placental margins
314 (well-developed, but very narrow and non-winged), seven symmetrical seminal chambers
315 separated by a septum, and a convex apex (Fig. 3D).



316
 317 **Fig. 3.** Plants (leaf and fruit impressions; permineralized seeds and fruit) from the late Eocene-
 318 early Oligocene Shapaja section, San Martín, Peruvian Amazonia. **A**, *PZ1*, non-monocot of
 319 uncertain affinities (PVMUSM-60; TAR-21sup). **B**, *PZ2*, non-monocot of uncertain affinities,
 320 resembling Fabaceae (PVMUSM-66; TAR-21sup). **C**, *PZ3*, non-monocot of uncertain affinities
 321 (PVMUSM-68; TAR-21sup). **D**, *PZ4*, Fabaceae fruit (PVMUSM-71; TAR-01). **E**, *PZ7*, non-monocot of
 322 uncertain affinities (PVMUSM-78; TAR-01). **F**, *PZ8*, Fabaceae compound leaf (PVMUSM-80; TAR-
 323 01). **G**, *PZ9*, compound leaf of uncertain affinities (PVMUSM-82; TAR-01). **H**, *PZ10*, non-monocot
 324 of uncertain affinity (PVMUSM-83; TAR-01). **I**, cf. *Elatine* seeds showing reticulate seed coats, left
 325 (TAR-74), right (TAR-01) photographed under epifluorescence. **J**, Two Passifloroideae seed types
 326 showing conspicuously pitted surfaces (TAR-21). **K**, Araceae (cf. *Monstera*) seed; top in lateral
 327 view, bottom in ventral view, note depressed hilum (TAR-21). **L**, Loculicidal fruit of euphorbiaceous
 328 affinity. Scale bars = 5 mm (A–H), 2 mm (J left, L), 1 mm (J right, K), and 500 µm (I).

329
 330 4.2.2 Seeds

331 Approximately forty seeds were identified throughout the Shapaja section. However, most of the
 332 specimens came from late Eocene samples, particularly from locality TAR-21. Seeds that were
 333 taxonomically identified are briefly described here.

- 334 - cf. *Elatine* (TAR-74, TAR-01): two small seeds (~600-700 µm long), slightly curved, operculate,
335 with reticulate coat (Fig. 3I).
- 336 - Passifloroideae (TAR-21): two different seed morphotypes of the passion fruit family were
337 identified from TAR-21. Both seeds are bilateral, ellipsoidal to ovoid with a rounded base and a
338 pointed apex, chalaza apical, and a conspicuously pitted surface (>20 depressions) (Fig. 3J).
- 339 - Araceae (cf. *Monstera*): The most common seed morphotype from the Shapaja section, with ~30
340 specimens. The seeds are ~2 mm high and 2.5 mm wide, flattened ventrally and with a dorsal
341 ridge, the micropyle is on the opposite side of the hilum from the chalaza. The hilum appears as a
342 depressed channel (Fig. 3K).
- 343 Three other seeds are less understood due to their fragmentary preservation and will require
344 additional study to confirm their affinity, but they can be preliminarily assigned to the families
345 Vitaceae (TAR-20), Poaceae (TAR-20), and Nymphaeaceae (TAR-73). A single loculicidal fruit of
346 euphorbiaceous affinity is also reported from sample TAR-21 (Fig. 3L).

348 4.2.3 Charophytes and wood.

349 All localities but TAR-72, TAR-21sup, and TAR-13 have yielded charophyte oogonia and wood
350 chunks (millimetric to centimetric, either silicified or oxidized). They were not identified thus far.

352 4.2.4 Pollen.

353 Several samples were processed but they were all sterile with the exception of a sample at TAR-
354 72, i.e., 12 stratigraphic meters below TAR-21 (latest Eocene). The TAR-72 sample had a poor
355 recovery of organic matter mostly dominated by coaly matter and a few grains that include the
356 magnoliopsid *Psilatricolporites* sp., Fungi monocellate, and the fern spores *Psilatriteles* sp. and
357 *Baculatriteles* sp. The high level of organic matter degradation suggests a seasonal water-table
358 that oxidized the organic matter.

360 4.3 Mollusks and decapods

361 **Mollusks** are mostly represented by freshwater gastropod inner casts, with two distinct species of
362 small pachychilids (TAR-01; tropical distribution; Fig. 5A), three specimens of a small planorbid
363 (*Helisoma* sp.; one in TAR-20, two in TAR-21), several ampullariids either large or small (TAR-22;
364 Fig. 4C), and small unidentified turritiform snails (TAR-20, TAR-72, and TAR-73), and a bulliform snail
365 of unknown affinities (two specimens in TAR-21; Fig. 4D). An external cast of a pluricentimetric
366 bivalve with paired valves, referred to as a probable corbiculid of freshwater-brackish affinities,
367 was unearthed at TAR-22 (Fig. 4A-B).

368 **Decapod** claw and carapace fragments were found by hundreds in most localities (TAR-74, TAR-20,
369 TAR-21, TAR-13, TAR-22, TAR-73, and TAR-01). All of them are assigned to Trichodactylidae, a
370 South and Central American family of freshwater crabs of tropical affinities, abundant in
371 Amazonian lowland streams today (Yeo et al., 2008), but with a scarce fossil record ranging the
372 middle Eocene–late Miocene in Western Amazonia (Klaus et al., 2017). None of the specimens
373 from the Shapaja decapod sample, partly studied by Klaus et al. (2017) for the TAR-01 locality, is
374 identifiable at the genus or species level.

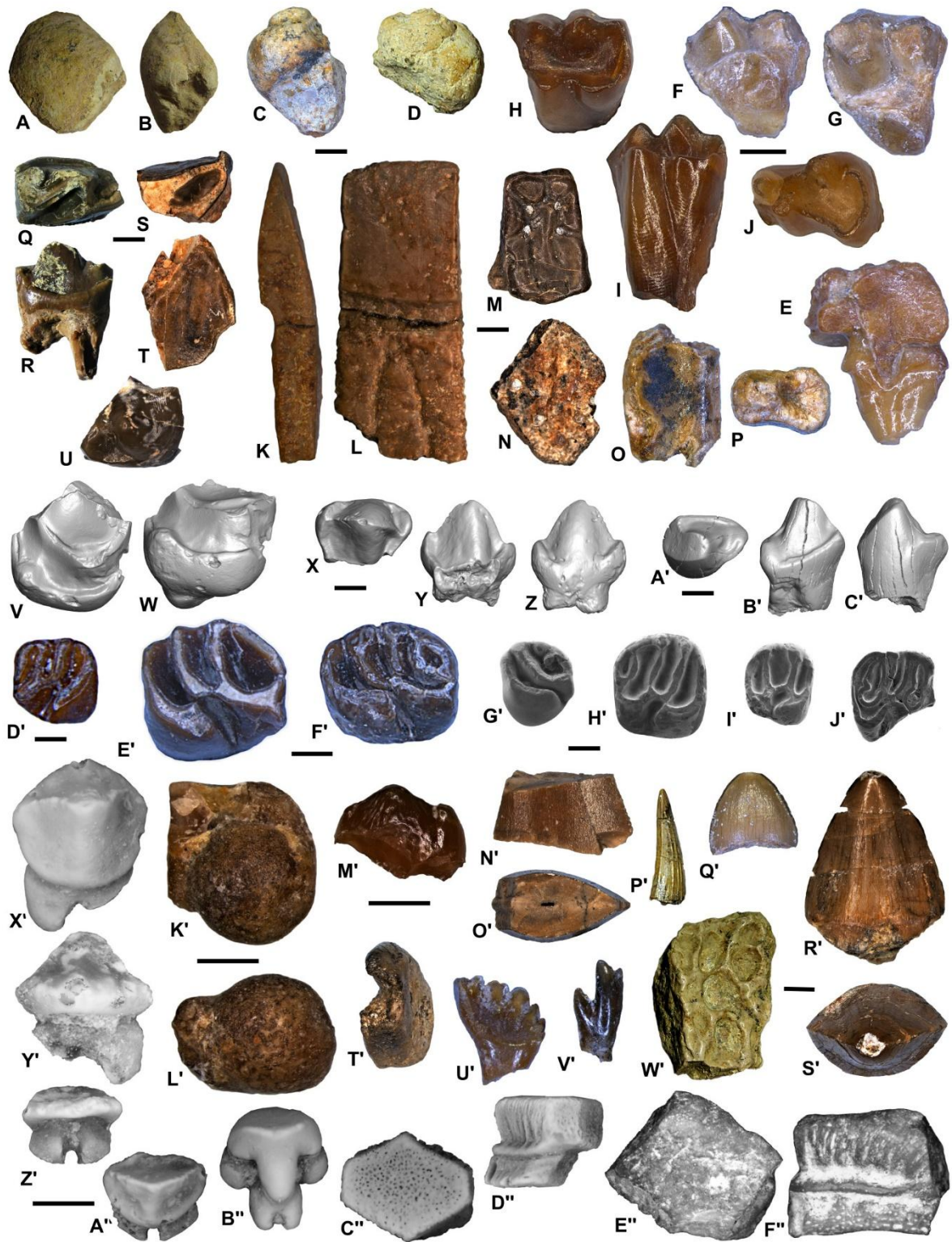
376 4.4 Vertebrates:

377 4.4.1 Metatherian mammals

378 Two fragmentary dental remains from TAR-74 and TAR-01, preventing precise taxonomic
379 identification (Fig. 4E), and a 1-mm wide astragalus (Fig. 5B-D) are referred to as **Marsupialiformes**
380 **incertae sedis**.

381 The extinct order **Polydolopimorphia** is represented in Shapaja by five taxa assigned to
382 Bonapartheriiformes. Aside from a new prepidolopid bonapartherioid documented at TAR-74 (Fig.

383 4F-G), at least two taxa are assigned to argyrolagid argyrolagids. The affinities of argyrolagids
384 (gerbil-like marsupials) are still under debate. There is evidence supporting Argyrolagidae as
385 representatives of polydolopimorphians (Goin et al., 2009; Chornogubsky & Goin, 2015) and of
386 paucituberculatans (Sánchez-Villagra, 2001; Beck, 2017); in addition, a recent phylogeny indicates
387 that argyrolagids are not paucituberculatans, and that they are phylogenetically close to
388 peramelemorphs (Abello and Candela, 2020). Nonetheless, this discussion is out of the scope of
389 this study, so we will assign here argyrolagids to ?Polydolopimorphia. Fragmentary and heavily-
390 worn molars from TAR-20 and TAR-72 (late Eocene) are identified as cf. *Proargyrolagus*. In
391 contrast, specimens from TAR-21 (latest Eocene) and TAR-01 (early Oligocene) are well preserved
392 and abundant (upper and lower cheek teeth). They document two distinct new species of
393 *Proargyrolagus*: a small species in TAR-21 (*P. nov. sp. A*; Fig. 4H-J) and a larger one in TAR-01 (*P.*
394 *nov. sp. B*; Fig. 5E-F). Accordingly, the Shapaja section attests to the earliest occurrence of i)
395 *Proargyrolagus*, previously from upper Oligocene Salla Beds, Bolivia (Wolff, 1984; Sánchez-Villagra
396 & Kay, 1997), and ii) the earliest and northernmost occurrences of Argyrolagidae, further pointing
397 to the unsuspected and long-termed presence of these unique metatherians throughout the
398 Eocene–Miocene interval in Proto-Amazonia (see Antoine et al., 2016). These teeth are “proto-
399 hypsodont” (i.e., high-crowned) and complete specimens retain closed roots.
400 Unidentifiable plagiaulacoid premolars are also recognized in TAR-20, TAR-72 (late Eocene) and
401 TAR-01 (early Oligocene), and assigned to polydolopimorphians of uncertain affinities.



402
 403
 404
 405
 406
 407
 408
 409
 410
 411

Fig. 4. Representative faunal content of late Eocene–earliest Oligocene paleocommunities near Shapaja, San Martín, Peruvian Amazonia (Assemblages 1 and 2; see Tables 2–4). **A–B**, MUSM 3955, Corbiculidae indet., natural inner cast with joined valves, in lateral (A) and dorsal views (B), TAR-22. **C**, MUSM 3956, Gastropoda indet., natural inner cast in front view, TAR-22. **D**, MUSM 3957, Gastropoda indet., bulliform, natural inner cast in front view, TAR-22. **E**, MUSM 3958, Marsupialiformes *incertae sedis*, left P3 in labial view, TAR-74. **F–G**, MUSM 3959, Prepidolopidae gen. et sp. nov., right Mx in lingual (F) and occlusal views (G), TAR-74. **H–J**, *Proargyrolagus* nov. sp. A, TAR-21: MUSM 3960, left M2 in occlusal (H) and lingual views (I); MUSM 3961, left m1 in occlusal view (J). **K–M**, *Parastegosimpsonia peruana*: MUSM 3962, mobile osteoderm in cross (K)

412 and lateral views (L), TAR-21; MUSM 3963, fixed osteoderm in external view (M), TAR-22. **N**,
 413 MUSM 3964, Peltephilidae indet., fixed osteoderm, TAR-21. **O-P**, MUSM 3965, Mylodontidae
 414 indet., molariform in lateral (O) and vertical views (P), TAR-22. **Q-R**, MUSM 3966, Leontiniidae
 415 indet., right p3 in occlusal (Q) and labial views (R), TAR-74. **S-T**, MUSM 3967, Adianthidae indet.,
 416 fragmentary right M2-3 in occlusal (S) and lingual views (T), TAR-21. **U**, MUSM 3968,
 417 Didolodontidae indet., fragment of a left ?m1 in occlusal view, TAR-22. **V-W**, MUSM 3969,
 418 Anthropeoidea indet. 1, right upper molar fragment in occlusal (V) and lingual views (W), TAR-21. **X-**
 419 **C'**, Anthropeoidea indet. 2 (small) TAR-21: MUSM 3970, right upper premolar (labial part) in
 420 occlusal (X), lingual (Y), and labial views (Z); MUSM 3971, left p2 in occlusal (A'), lingual (B'), and
 421 labial views (C'). **D'**, MUSM 3972, *Pozomys* nov. sp., left m1 in occlusal view, TAR-74; **E'-F'**,
 422 *Eopululo* cf. *wigmorei*, TAR-74: E', MUSM 3973, left M1/2 in occlusal view; F', MUSM 3974, left
 423 dP4 in occlusal view. **G'**, MUSM 2965, *Eoincamys parvus*, left M3 in occlusal view, TAR-22. **H'**,
 424 MUSM 2925, *Kichkasteiromys raimondii*, right M1/2 (holotype) in occlusal view, TAR-21. **I'**, MUSM
 425 2960, *Selvamys paulus*, right M2 in occlusal view, TAR-22; **J'**, MUSM 3332, *Tarapotomys*
 426 *subandinus*, fragmentary right M1 in occlusal view, TAR-20. **K'-L'**, MUSM 3975, Anura indet., distal
 427 humerus in anterior (K') and distal views (L'), TAR-20. **M'**, MUSM 3976, unidentified ?teiid
 428 squamate tooth in lateral view, TAR-21. **N'-O'**, MUSM 3977, Sebecidae indet., fragmentary tooth
 429 in lingual (N') and apical views (O'), TAR-74. **P'**, MUSM 3978, Gavioloidea indet., tooth in sagittal
 430 view, TAR-22. **Q'**, MUSM 3979, Caimaninae indet., globular tooth in lingual view, TAR-22. **R'-S'**,
 431 MUSM 3980, Caimaninae indet., leaf-shaped tooth in lingual (R') and apical views (S'). **T'**, MUSM
 432 3981, Lepidosirenidae indet., large dental plate in apical view, TAR-72. **U'**, MUSM 3982,
 433 Anostomidae, cf. *Leporinus* sp., hexacuspoid tooth, TAR-21. **V'**, MUSM 3983, Loricariidae indet.,
 434 muffle-shaped tooth, TAR-21. **W'**, MUSM 3984, cf. *Phractocephalus* sp., fragmentary cranial bone,
 435 TAR-22. **X'-Y'**, MUSM 3985, *Paratrygon* nov. sp., female anterior tooth in lingual (X') and labial
 436 views (Y'), TAR-22. **Z'-A''**, MUSM 3986, *Potamotrygon* nov. sp., female anterior tooth in lingual (Z')
 437 and labial views (A''), TAR-21. **B''**, MUSM 3987, *Pristis* sp., oral tooth in lingual view, TAR-22. **C''-D''**,
 438 MUSM 3988, *Myliobatis* sp., tooth of a neonate/young individual in occlusal (C'') and lateral views
 439 (D''), TAR-21. **E''-F''**, MUSM 3989, ?*Myliobatis* sp., tooth in occlusal (E'') and lateral views (F''), TAR-
 440 21. V-C' are 3D surface renderings. Scale bars = 500 µm (E-J, V-D', G'-L', U'-V', E''-F''), 1 mm (E'-F',
 441 M'-O', X'-D''), 2 mm (C, K-P, S-U, Q'-T'), 5 mm (D, Q-R, P'), and 10 mm (A-B).

442
 443 **Paucituberculata** (shrew opossums) are the most diverse order in Shapaja, with at least 12 taxa,
 444 all belonging to the extinct superfamily Palaeothentoidea (Fig. 5G-I). The records from Shapaja
 445 encompass two primitive species of Non-Pichipilidae palaeothentoids (NPP) in uppermost Eocene
 446 levels (TAR-72 and TAR-21), one of them being close to *Perulestes*, previously described at Santa
 447 Rosa (Goin & Candela, 2004) with close allies in early Contamana faunas (Antoine et al., 2016).
 448 Representatives of Palaeothentidae and Abderitidae are much more diversified in TAR-01 (early
 449 Oligocene; Fig. 5I). In addition, fragmentary remains from TAR-72 and TAR-21 and referred to
 450 Palaeothentinae might document the earliest occurrence of the subfamily, previously recorded in
 451 the late Oligocene of Argentina and Bolivia (Abello, 2007; Rincón et al., 2015).
 452 Contrary to all other localities, TAR-13 and TAR-22 have yielded a single marsupialiform specimen
 453 (a tiny astragalus of an unidentified marsupialiform [Fig. 5B-D] and a palaeothentoid molar,
 454 respectively; Table 2). This low abundance is particularly striking with respect to the volume of
 455 sediment treated at TAR-22 (214 kg; see SI). No metatherian specimen was recognized at TAR-73.

456
 457 4.4.2 Eutherian mammals.

458 **Cingulates** (armadillos). Two osteoderms (one movable and one fixed) from TAR-21 and TAR-22
 459 (Fig. 4K-M), are referable to *Parastegosimpsonia peruana*, an astegotheriine dasypodid originally
 460 described at Santa Rosa (Ciancio et al., 2013). Two other osteoderms are too eroded/fragmentary

461 for a precise identification but attributable to the Peltephilidae (TAR-21; Fig. 4N) and Dasypodidae
462 (TAR-01) families.

463 **Pilosans** (sloths). A molariform belonging to a small mylodontid (under study by FP) was recovered
464 from TAR-22 (Fig. 4O-P). Smaller in size than the smallest known representative of the family
465 (*Brievabradys laventensis*, middle Miocene of Colombia; [Villarreal, 2000](#)), this “8”-shaped tooth is
466 somewhat reminiscent to last upper molariforms of the late Oligocene *Octodontotherium* and
467 *Paroctodontotherium* (Deseadan of Argentina and Bolivia, respectively; [Hoffstetter, 1956](#); [Shockey
et al., 2011](#)). Excluding *Pseudoglyptodon* (an Oligocene sloth-like xenarthran of dubious affinities;
468 [McKenna et al., 2006](#)), this fossil from TAR-22 likely stands as the oldest record of a true sloth (for
469 a review, see [Pujos et al., 2017](#)).

471
472 As for **Astrapotheria** (elephant-like native ungulates), the occurrence of unidentified
473 Astrapotheriidae is suspected in TAR-21 and TAR-01, through small fragments of large-sized cheek
474 teeth with apparent vertical and horizontal decussation on enamel ([Koenigswald, 1997](#)).

475 Among **Notoungulata**, representatives of the suborders Toxodontia (horse- and rhino-like native
476 ungulates) and Typotheria (rabbit-like native ungulates) were recognized throughout the section,
477 on the basis of isolated teeth and tooth fragments. Unfortunately, most remains are fragmentary
478 and not assignable at the family level or below. Toxodontia indet. are documented by fragments of
479 large-sized teeth (TAR-74, TAR-20, and TAR-01). An early-diverging leontiniid occurs in TAR-74, as
480 recorded by a mesodont lower premolar (p3; length = 16 mm; width = 11 mm; Fig. 4Q-R) with a
481 strong distolabial extension of the protolophid, a bunoid entoconid united to the hypolophid, a
482 short entolophid, an un-isolated fossettid, and a talonid longer than the trigonid. Somewhat
483 reminiscent of p3-4 of *Elmerriglesia* from Deseadan beds of Patagonia (e.g., [Shockey et al., 2012](#)), it
484 is much closer morphologically to the p3 of *Scarrittia barranquensis*, from La Cantera (early
485 Oligocene, Patagonia; [Ribeiro et al., 2010](#)), from which it only differs in being 30% smaller and 40%
486 narrower. Typotheria may have been more diversified than toxodonts, with two distinct bispecific
487 faunas, in TAR-74 and TAR-22. More precisely, TAR-74 (late Eocene) yields a small hypsodont
488 cheek tooth fragment, with cement, pointing to an unidentified intertheriid or hegetotheriid and
489 low-crowned tooth fragments of a small unidentified typothere (morph 1). This morph 1 also likely
490 occurs in TAR-21. In TAR-22 (earliest Oligocene), a medium-sized molar fragment, hypsodont, with
491 a closed fossette/fossettid and a wide enamel band, is assignable to a late “archaeohyracid”
492 (*Archaeohyrax*, *Archaeotypotherium*, or *Protarchaeohyrax*), while smaller-sized low-crowned
493 fragmentary teeth (decidua teeth?) may document another typothere referred to as
494 “Archaeohyracidae” indet. A 14mm-long lower tooth from TAR-73, eroded and heavily worn, has
495 diverging roots. It is assigned to Notoungulata indet.

496 **Litopterna** (horse- and camel-like native ungulates) are not abundant in the Shapaja section. At
497 TAR-21, the mesiolabial tip of a right upper molar documents an adianthine adianthid (Fig. 4S-T).
498 The corresponding tooth is low-crowned, with a sharp and labially-protruding parastyle, a shallow
499 oblique fossette, a smooth labial cingulum, thin enamel, and a saddle-shaped neck both lingually
500 and labially. Its morphological pattern closely resembles that of M2-3s of *Tricoelodus* from
501 Deseadan beds of Argentina and Bolivia ([Cifelli & Soria, 1983](#)), but with a size 50% larger than in
502 both species assigned to *Tricoelodus*. This specimen likely fills a stratigraphic gap between the
503 Eocene indalecines and the late Oligocene adianthines, only occurring at mid- and high latitudes
504 ([Cifelli & Soria, 1983](#)).

505 We refer to a tooth fragment as an unidentified didolodontid (TAR-22; Fig. 4U). This pristine
506 specimen preserves the distolingual part of a left lower molar (probably m1), of brachydont and
507 bunodont condition. It has a prominent and conical entoconid, a much thinner and lower
508 hypoconulid with a low postentocristid joining them, and a marked postentocingulid between
509 both cuspids. Such morphological features closely resemble those of the m1 of *Didolodus* from the

510 late middle Eocene of Patagonia (e.g., [Simpson, 1967](#)), the latter being 15% larger than the
511 Shapaja specimen. It is much larger than *Ernestokokenia* and *Saltaodus* ([Gelfo et al., 2019](#)). Direct
512 comparison with *Sallalodus deutherotherioides* is not possible, as its original hypodigm is restricted
513 to two associated upper molars, larger sized than the specimen from TAR-22 ([Soria & Hoffstetter,
514 1983](#)).

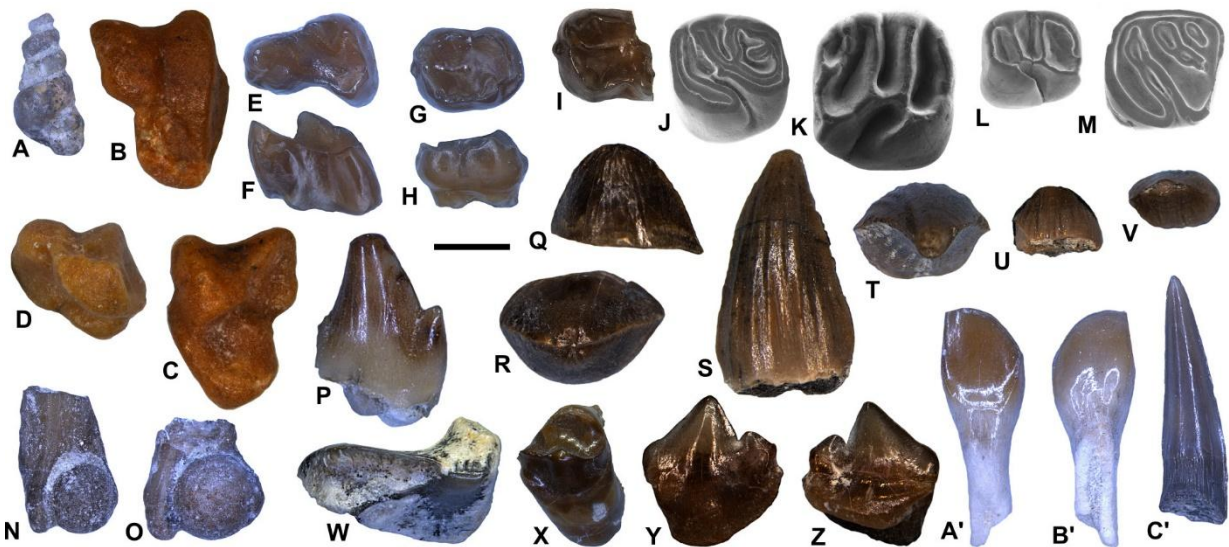
515 **Pyrotheria** (mastodon-like native ungulates) are documented at Shapaja by tooth fragments with
516 a lophodont Bauplan, thick enamel with vertically-oriented bands ([Von Koenigswald et al., 2014](#)),
517 and a typical sagittally-oblique wear pattern. They are tentatively assigned either to *Pyrotheria*
518 indet. (TAR-20 and TAR-21; latest Eocene) or to the pyrotheriid cf. *Griphodon* sp. (TAR-13, TAR-22,
519 and TAR-01; earliest Oligocene) when their features and dimensions match closely those of
520 *Griphodon*. *Griphodon peruvianus* was originally described in Paleogene deposits from the vicinity
521 of Shapaja (probably Eocene in age; [Anthony & Richards, 1924](#); [Patterson, 1942](#)).
522 Other tooth fragments were assigned to unidentified native ungulates (TAR-72).

523
524 **Chiropterans** (bats). Distolabial fragments of two hyperdilambodont upper molars from TAR-21
525 and TAR-01 (estimated length < 2mm) are assignable to unidentified chiropterans (Chiroptera
526 indet.).

527
528 **Primates** (monkeys). In the Shapaja section, primates have so far been found only at TAR-21.
529 Despite the large amount of sediment sampled and treated by wet screening, the fossil material
530 documenting primates is particularly rare compared to that of metatherians or rodents from the
531 same locality (see specific paragraphs). It consists of five dental remains, most of which are
532 fragmentary (a well-preserved trigon region of an upper molar [Fig. 4V-W], an upper molar
533 fragment, the buccal part of an upper premolar [Fig. 4X-Z], a complete but worn lower second
534 premolar [Fig. 4A'-C'], and a worn lower molar). These fossils attest to the co-occurrence of at
535 least two tiny but size-distinct species. Pending the discovery of new specimens, the paucity and
536 fragmentary nature of the currently available material does not allow evaluating the phylogenetic
537 affinities of these two species. However, despite the scarce morphological information, the lingual
538 part of a half upper molar displays a complete and quite strong lingual cingulum, with a minute
539 but well-defined hypocone, without a pre-hypocrista (Fig. 4W). Such a simple pattern illustrates, to
540 some extent, the degree of primitiveness of that primate, which rather matches that of some basal
541 anthropoids from the Paleogene of Africa and *Perupithecus* from Santa Rosa (early Oligocene;
542 [Bond et al., 2015](#); [Seiffert et al., 2020](#)) or even *Parvimico* from Madre de Dios (early Miocene; [Kay
543 et al., 2019](#)) than that of most subsequent stem and/or crown platyrrhines of South America (i.e.,
544 Homunculidae and extant families; for details, see [Marivaux et al., 2016](#)).

545
546 **Rodents** are by far the most conspicuous and species-rich mammalian group in the Shapaja
547 section, with teeth uncovered in all localities, and ca. 500 specimens identified taxonomically.
548 TAR-21 and TAR-01 yielded most rodent material. This material was extensively described in [Boivin
549 et al. \(2018, 2019a, 2019b\)](#) and we only provide here an updated overview, including additional
550 specimens from the last field campaigns (2018-2019). Twenty-two distinct caviomorph taxa
551 document stem Caviomorpha (sensu [Boivin et al., 2019a](#)), representatives of three extant
552 superfamilies (i.e., Chinchilloidea [chinchillas], Erethizontoidea [New World porcupines], and
553 Octodontoidea [spiny rats]; Table 3), and caviomorphs of uncertain affinities. Caviomorphs from
554 the Shapaja localities notably display a well-marked disparity regarding crown height, as
555 brachydont, mesodont, and subprotohypsodont / protohypsodont morphs are recognized ([Boivin
556 et al., 2018](#)). Although small-sized rodents widely predominate in the Shapaja section, specimens
557 attesting to the presence of larger-sized taxa were found in late Eocene localities such as TAR-74
558 (*Eopululo* cf. *wigmorei*; Fig. 4E'-F') and TAR-21 (Caviomorpha gen. et sp. indet. 5), and in the

559 earliest Oligocene TAR-22 locality (Caviomorpha gen. et sp. indet. 2). In terms of taxonomic
 560 richness, the most diversified rodent fauna from Shapaja is recorded at TAR-21 (with eight co-
 561 occurring species), followed by TAR-22 (six, including *Selvamys paulus* [Fig. 4I']), TAR-01 and TAR-
 562 20 (five), TAR-74 and TAR-13 (four), and lastly TAR-72 (two). The earliest rodent fauna (TAR-74:
 563 Assemblage 1) is fully distinct from all younger ones. It includes a new representative of *Pozomys*
 564 (Fig. 4D'), a genus originally restricted to late middle Eocene localities of Contamana (Boivin et al.,
 565 2017) and recognized at Santa Rosa (Arnal et al., 2020). It also yields *Eopululo* cf. *wigmorei*, i.e. a
 566 close ally of a species defined at Santa Rosa (Frailey & Campbell, 2004), a taxon tentatively
 567 approximated to *Eoincamys* (a stem chinchilloid, distinct from that of TAR-13), and an unidentified
 568 caviomorph. Overlying localities yield quite homogenous rodent faunas over the TAR-20 to TAR-22
 569 interval, with genera and/or species in common, especially among *Eoincamys* and *Tarapotomys*
 570 (Assemblage 2; Table 3): *Eoincamys valverdei* occurs in all localities from this interval; *Eoincamys*
 571 *parvus* is recognized at TAR-20, TAR-21, and TAR-22 (Fig. 4G'); *Tarapotomys subandinus* is
 572 documented in all localities, except in TAR-13 (Fig. 4J'); Caviomorpha nov. gen. et sp. ranges from
 573 TAR-20 to TAR-13, and an unidentified caviomorph (Caviomorpha gen. et sp. indet. 1) occurs at
 574 TAR-20 and TAR-21. The early erethizontoid *Kichkasteiromys raimondii* is restricted to TAR-21 (Fig.
 575 4H'). Conversely, the uppermost locality of the section (TAR-01) provides a very distinctive fauna,
 576 with *Tarapotomys mayoensis*, *Shapajamys labocensis*, *Mayomys confluens*, *Eoincamys* cf. *pascuali*,
 577 and an unidentified chinchilloid (Assemblage 3; Table 3; Fig. 5J-M). Moreover, only five rodent
 578 species are recognized at TAR-01, which is a strikingly low taxonomic diversity with respect to the
 579 number of specimens uncovered (406 cheek teeth) and to the volume of sediment treated (470
 580 kg; see SI).
 581



582
 583 **Fig. 5.** Representative faunal content of the early Oligocene paleocommunities near Shapaja, San
 584 Martín, Peruvian Amazonia (Assemblage 3; see Tables 2-4). All specimens originate from TAR-01,
 585 except for MUSM 4018 (B-D: TAR-13). **A**, MUSM 3990, Pachyichilidae indet. sp. 2, natural inner cast
 586 in front view. **B-D**, MUSM 4018, Marsupialiformes *incertae sedis*, right astragalus in dorsal (**B**),
 587 plantar (**C**) and distal views (**D**). **E-F**, MUSM 3991, *Proargyrolagus* nov. sp. B, left m1 in occlusal (**B**)
 588 and labial views (**C**). **G-H**, MUSM 3992, Palaeothentoidea indet. 2, right ?m4 in occlusal (**D**) and
 589 lingual views (**E**). **I**, MUSM 3993, Abderitidae indet., fragmentary right m2, in occlusal view. **J**,
 590 MUSM 3496, *Tarapotomys mayoensis*, left M2 (holotype) in occlusal view. **K**, MUSM 2995,
 591 *Shapajamys labocensis*, right M2 (holotype) in occlusal view. **L**, MUSM 3159, *Mayomys confluens*,
 592 left M1 in occlusal view. **M**, MUSM 3492, *Eoincamys* cf. *pascuali*, right m1 in occlusal view. **N**,
 593 MUSM 3994, Anura indet., distal humerus fragment in anterior view. **O**, MUSM 3995, Anura
 594 indet., non-Pipidae, distal humerus fragment in anterior view. **P**, MUSM 3996, Unidentified

595 squamate tricuspid tooth in lingual view. **Q-R**, MUSM 3997, Caimaninae indet., broken tooth, with
596 irregular enamel in labial (N) and apical views (O). **S-T**, MUSM 3998, Caimaninae indet., tooth with
597 grooved enamel in labial (P) and apical views (Q). **U-V**, MUSM 3999, Caimaninae indet.,
598 durophagous tooth, in labial (R) and apical views (S). **W**, MUSM 4000, Lepidosirenidae indet., large
599 dental plate. **X**, MUSM 4001, cf. *Colossoma* sp., tooth in apical view. **Y-Z**, MUSM 4002, cf.
600 *Serrasalmus* sp., 4th or 5th tooth of the dentary, in labial (V) and lingual views (W). **A'-B'**, MUSM
601 4003, *Leporinus* sp., tooth in palatine (X) and ab-palatine views (Y). **C'**, MUSM 4004, *Hydrolycus*
602 sp., tooth in lingual view. Scale bar = 500 µm (B-D), 1 mm (E-P, U-V, Y-Z) and 2 mm (A, Q-T, W-X,
603 A'-C').
604

605 4.4.3 Anurans (frogs)

606 A dozen of postcranial elements of very small dimensions were recovered in four localities over
607 the studied section. All of them are of uncertain taxonomic affinities. They consist of fragmentary
608 humeri (TAR-74, TAR-20 [Fig. 4K'-L'], and TAR-01 [two morphs, including non-Pipidae; Fig. 5N-O]),
609 radio-ulnae (TAR-20, TAR-21, and TAR-01), and ilium (TAR-01).
610

611 4.4.4 Squamates (lizards)

612 Isolated millimetric tricuspid teeth document three distinct squamate morphotypes at TAR-21
613 (morph 1: corrugated enamel; Fig. 4M') and TAR-01 (morph 2: high-crowned and sharp-edged [Fig.
614 5P]; morph 3: bulbous crown), probably assignable to scincomorphan lizards (teiids?). Neither
615 snake remains nor lacertilian osteoderms were unearthed in the concerned localities.
616

617 4.4.5 Chelonians (turtles and tortoises)

618 Chelonian remains occur throughout the section (TAR-20, TAR-21, TAR-22, and TAR-01). The
619 concerned fossil specimens consist of isolated smooth-surfaced scutes, referable to unidentified
620 pelomedusoid pleurodiran turtles, and documenting a wide size range. In particular, they are
621 substantially large sized at TAR-22.
622

623 4.4.6 Crocodylomorphs (crocodylians and their stem relatives)

624 At Shapaja, crocodylomorphs are documented from most localities by basically isolated, small-
625 sized teeth. The Shapaja communities include two (TAR-74, TAR-72, and TAR-22) up to four (TAR-
626 21), and even five co-occurring crocodylomorph taxa (TAR-01).
627 Sebecosuchians (extinct terrestrial crocs) occur in TAR-74, TAR-72, TAR-22, and TAR-01 (Fig. 4N'-
628 O'). Teeth, easily recognizable thanks to their triangular profile, almond-shaped cross section and
629 serrated edges, are referred to as Sebecidae indet. (sensu [Pol & Powell, 2011](#)). The fragmentary
630 state of the available sample and the sparse Paleogene fossil record of sebecosuchians in tropical-
631 equatorial areas of South America discards to refine further their taxonomic assignment or to
632 consider the co-occurrence of several taxa.
633 Elongated teeth with fluted enamel, typical of longirostrine fish-eating crocodylians are recognized
634 throughout the section (from TAR-74 up to TAR-01). They are either assignable to unidentified
635 crocodyloids or gavialoids (Fig. 4P') in all localities. Only at TAR-20 a large tooth with a clear S-
636 shaped profile can be referred to as a gryposuchine gavialoid.
637 The record of caimanine teeth far exceeds that of other crocodylomorph components at Shapaja
638 (TAR-20 to TAR-01; Fig. 4Q'-S', 5Q-V). A complete skull (under study by RSG) and an osteoderm
639 referable to this group were also recovered at TAR-20 and TAR-21, respectively. Teeth may
640 document several morphs related to caimans, including globular, blunt with a short crown, leaf-
641 shaped (MUSM 3980; Fig. 4R'-S') and conical with strongly-fluted enamel morphs, aside from the
642 more "generalized" caiman dental shapes (Fig. 5Q-V; for a review, see [Salas-Gismondi et al., 2015](#)).
643

644 4.4.7 Ichthyofauna

645 **Dipnoi** (lungfish)

646 Dipnoan dental plates of a single morph and dentine/bone structure, but of two distinct sizes,
647 were uncovered in localities of the Shapaja section. Small ones occur throughout the considered
648 section (from TAR-74 to TAR-01). They are mostly documented through tiny fragments (Fig. 3),
649 sometimes as conspicuous elements of the ichthyofaunal community (up to 21 and 24 specimens
650 at TAR-21 and TAR-01, respectively; Fig. 5W). The largest ones only occur in late Eocene localities
651 (TAR-20, TAR-72, TAR-21; Fig. 4T'), which is not primarily linked to stream energy, as a similar grain
652 size is recorded at TAR-22 and TAR-73 (earliest Oligocene). They are far smaller than tooth plates
653 of *Lepidosiren* recorded in Miocene deposits of Brazil, Colombia, and Peru (Lundberg, 1997; Toledo
654 & Bertini, 2005). All these remains are referred to as Lepidosirenidae indet.

656 **Actinopterygians** (ray-finned fish)

657 Characiforms (characins and kin) dominate the actinopterygian record (see SI), with a constant
658 composition throughout the Shapaja section (from TAR-74 to TAR-01). Most if not all localities
659 have yielded isolated teeth referable to serrasalmids (pacu, cf. *Colossoma* sp.; 2,230 specimens at
660 TAR-01; Fig. 5X), anostomids (cf. *Leporinus* sp., incisor-like and hexacuspids [rare; Fig. 4U'];
661 *Leporinus* sp., incisor-like and paddle-shaped teeth [629 specimens at TAR-01; Fig. 5A'-B'], and
662 cynodontids (cf. *Hydrolycus* sp., monocuspid, sharp and dagger-like teeth; 359 specimens at TAR-
663 01; Fig. 5C'). Unidentified hook-like, pad-like, and acuminate unicuspid pharyngeal teeth add to
664 the characiform record. TAR-01 yielded three millimetric razor-sharp bicuspid teeth with an
665 interlock furrow, unquestionably referable to as a crown piranha, and closely resembling the 4th or
666 5th tooth of the dentary of *Serrasalmus* (cf. *Serrasalmus* sp., Fig. 5Y-Z; Shellis & Berkovitz, 1976).
667 In contrast, siluriforms (catfish) are mostly documented by pectoral and/or dorsal spines at TAR-
668 20, TAR-21, and TAR-01 (Siluriformes indet.). A small muffle-shaped tooth from TAR-21 and a small
669 spine with a typical punctuated ornamentation from TAR-01 are assigned to an unidentified
670 loricariid (armored catfish; Loricariidae indet.). The tooth has a small accessory cusp at the tip of
671 the crown (Fig. 4V'), contrary to the loricariid teeth from Miocene deposits of Contamana (Antoine
672 et al., 2016). A cranial fragment of a large-sized goliath catfish recovered at TAR-22 is recognizable
673 thanks to the ornamented external aspect, with thick ridges and elongated sulci (Fig. 4W'). It is
674 referred to as a pimelodid, cf. *Phractocephalus* sp. (for comparison, see Lundberg & Aguilera,
675 2003; Aguilera et al., 2008).

677 **Selachians** (sharks, skates, and rays)

678 Both the oldest (TAR-74) and the youngest Shapaja localities (TAR-73 and TAR-01) have not yielded
679 a single selachian specimen, whereas all interbedded localities gather one to four co-occurring
680 selachian species. Myliobatiforms are fully dominant over pristiforms (see SI). The most
681 conspicuous elements by far are freshwater stingrays (Potamotrygonidae), with a new species of
682 *Potamotrygon* documented at TAR-20, TAR-21, and TAR-22 (one to 40 teeth per locality; Fig. 4Z'-
683 A''), *Potamotrygon canaanorum* at TAR-13 (previously restricted to upper Oligocene-upper
684 Miocene deposits from Contamana area; Chabain et al., 2017), and a representative of *Paratrygon*
685 (*Paratrygon* sp.) at TAR-21 and TAR-22 (Fig. 4X'-Y'). To our knowledge, these are the first fossil
686 occurrences of this Recent monotypic genus. Myliobatids occur at TAR-20, TAR-72, and TAR-13
687 (with a neonate tooth in each locality), and TAR-21 (20 teeth of *Myliobatis* sp. [Fig. 4E''-F''] and a
688 tooth assigned to the bat ray ?*Rhinoptera*). Some *Myliobatis* specimens are particularly large (30
689 mm-long) at TAR-21, whereas unusually small batoid specimens in TAR-20, TAR-72, and TAR-13 are
690 cautiously interpreted as documenting neonate/young specimens of bat rays (?*Myliobatis*; Fig.
691 4C''-D''). In addition, a single oral tooth of the sawfish *Pristis* sp. was recognized at TAR-22 (Fig.
692 4B''). To sum up, the Shapaja selachian sample likely attests to the presence of a single and

693 consistent community between TAR-20 and TAR-22, with brackish/coastal plain affinities, as
694 indicated by bat rays and *Pristis*.

695 696 5 Discussion

697 698 5.1 Age

699 Here we performed, for the first time, chemostratigraphical investigation using carbon isotopes on
700 dispersed organic matter ($\delta^{13}\text{C}_{\text{org}}$) and pedogenic nodules ($\delta^{13}\text{C}_{\text{nod}}$) at Shapaja (Western and
701 Eastern sections), in order to i) provide a stratigraphic framework of the upper part of the Pozo
702 Fm. in the Tarapoto area (Peruvian Amazonia), ii) refine the position of nine fossil-bearing levels in
703 this stratigraphic framework. Unravelling the age of fossil-bearing levels in terrestrial to proximal
704 marine sections is challenging mainly due to the lack of marine biostratigraphical reference fossils.
705 Moreover, usage of certain terms has changed through time regarding the Eocene/Oligocene
706 boundary (EOB; Premoli-Silva & Jenkins, 1993; Hutchison et al., 2020). In common practice, the
707 EOB is linked to the Oi-1 event, based on $\delta^{18}\text{O}$ perturbations (Van Mourik and Brinkhuis, 2005).
708 The late Eocene–early Oligocene interval comprised several isotopic events initially defined by
709 Miller et al. (1991). The oldest of these events are major $\delta^{18}\text{O}$ and $\delta^{13}\text{C}$ positive shifts, starting in
710 the latest Eocene and ending in the earliest Oligocene (e.g., Coxall et al., 2005; Katz et al., 2008;
711 Lear et al., 2008; Vandenberghe et al., 2021; Fig. 2). This isotopic event, called Eocene-Oligocene
712 (climate) transition (EOT), lasting ~500 kyr (Coxall and Pearson, 2007), then encompasses the
713 Eocene-Oligocene boundary. The EOB occurs approximately two thirds of the way through the EOT
714 (Hutchison et al., 2020). Using a high-resolution carbon isotope study of the ODP site 1218,
715 Erhardt et al. (2013) showed that the EOT positive shift is followed by two positive $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$
716 excursions called Oi-1 and Oi-1a, both earliest Oligocene in age. This isotopic pattern was also
717 observed by Zhifei et al. (2004) in ODP Leg 208 Site 1262, 1265 and 522. These trends are followed
718 by another positive excursion (Oi-1b), clearly showing lighter maximal values than Oi-1 and Oi-1a
719 excursions. While diagenesis increasingly alters the oxygen-isotope signature of sediments, C-
720 isotope geochemistry is less influenced by diagenesis (Weissert et al., 2008). Oxygen isotopes
721 ($\delta^{18}\text{O}$) on organic matter do not provide primary signals and are therefore not useful for
722 stratigraphy. Conversely, variations in terrestrial organic carbon-isotope composition ($\delta^{13}\text{C}_{\text{org}}$)
723 allow for using C-isotope stratigraphy as tool for correlating marine and terrestrial records, as
724 demonstrated in a wide array of sections and time intervals (e.g. Gröcke et al., 1999). These
725 variations have been satisfactorily used for allocating the position of Paleogene fossil-bearing
726 levels in terrestrial sections (e.g. Yans et al., 2014a,b; Noiret et al., 2016), including the EOB and
727 around (Benammi et al., 2019).

728 The Shapaja section was previously assigned an early Oligocene age (Klaus et al., 2017; Boivin et
729 al., 2018, 2019a, 2019b), then a late Eocene–early Oligocene age range, on the basis of
730 paleontological content and lithological correlations (Assemat et al., 2019). In the Western section
731 of Shapaja, our new isotopic data on both organics ($\delta^{13}\text{C}_{\text{org}}$) and pedogenic nodules ($\delta^{13}\text{C}_{\text{nod}}$)
732 clearly show (Fig. 2): i) a positive shift attributed to the EOT event, ii) followed by quite positive
733 values considered as Oi-1, iii) overlaid by a positive excursion interpreted as Oi-1a. These isotope
734 trends perfectly match those observed at the global cooling of the Eocene–Oligocene transition
735 described above (Erhardt et al., 2013). During Paleogene to Miocene times, similar C-isotope
736 patterns, showing two successive high-amplitude positive $\delta^{13}\text{C}$ excursions, are only recorded after
737 the Mid-Eocene Climatic Optimum (MECO, Bartonian) and around the Mid-Miocene Climatic
738 Optimum (MMCO, Langhian; Zachos et al., 2001; Luciani et al., 2010; Vandenberghe et al., 2012).
739 As mentioned above, the MECO and MMCO time intervals are widely at odds with the inferred
740 biostratigraphic age range of Shapaja localities studied here. Moreover, middle Miocene fossils
741 have been collected ~1,800 m above TAR-01 (at TAR-31 locality; Marivaux et al., 2020) whereas

742 late Middle Eocene rodents were recognized at TAR-67, i.e., 280 m below the lowermost locality
743 (TAR-74; Fig. 1). The uppermost positive peak at Shapaja (-6.9 ‰ at 113.5m) may be interpreted as
744 Oi-1b but this remains tentative since i) this peak is based on a single point and ii) $\delta^{13}\text{C}_{\text{org}}$ does not
745 show the same trend for this sample. The main part of the Eastern section shows a negative trend,
746 interpreted as global negative trend observed below the EOT (Fig. 2), which further matches local
747 lithological sequences and taxonomic affinities of the concerned paleocommunities.
748 In Shapaja, the C-isotope values on organics and pedogenic nodules are globally consistent,
749 showing similar trends (Fig. 2). However, two minor discrepancies are observed in the following
750 intervals: i) hypothetical base of the EOT located in the $\delta^{13}\text{C}_{\text{org}}$ curve ~20 m above its location in
751 the $\delta^{13}\text{C}_{\text{nod}}$ curve, and ii) negative trend of the Oi-1a excursion, well-marked in the $\delta^{13}\text{C}_{\text{nod}}$ curve
752 (from -5.8 ‰ at 81.2 m to -11.3 ‰ at 98.3 m) and less marked in the $\delta^{13}\text{C}_{\text{org}}$ curve. These local
753 inconsistencies may be explained by the poor carbon content of organics (usually < 0.05 %) in all
754 studied samples. In Shapaja, organic matter most probably experienced intense recent
755 oxidation/weathering, leading to potential biases in the resulting $\delta^{13}\text{C}_{\text{org}}$ values/trends. We
756 therefore consider the data on pedogenic nodules as more robust (meteoric water-sourced),
757 allowing us to use the $\delta^{13}\text{C}_{\text{nod}}$ values as the best primary signals where data on both the studied
758 materials (organics vs nodules) are not consistent.
759 Our data in the Western section suggest that the EOT is observed between 53 and 59 m using
760 $\delta^{13}\text{C}_{\text{org}}$ data, and between 32 and 59 m, using $\delta^{13}\text{C}_{\text{nod}}$ data. Thus, the EOB is located somewhere in
761 this latter interval (Fig. 6). It is not possible, however, to refine the location of the EOB, at our
762 current stage of knowledge. Based on these new chemostratigraphical data, we conclude that i)
763 TAR-20 and TAR-72 (and TAR-74, even if this underlying level was not sampled in our study) are
764 late Eocene in age, ii) TAR-21 is located at the earliest EOT (i.e., most probably latest Eocene in
765 age), iii) TAR-21sup is within the EOT (latest Eocene? – earliest Oligocene?), and iv) TAR-22, TAR-
766 13, TAR-73, and TAR-01 are early Oligocene in age.
767 Among metatherians, most new records of palaeothenitids and abderitids at Shapaja are
768 consistent with previously-estimated divergence times (i.e., earliest Oligocene; [Abello et al., 2018](#)).
769 However, the presence of palaeothenitines by the EOT at Shapaja implies an unexpected ghost
770 lineage of the group, and as such, a much earlier divergence, not only of the palaeothenitids and
771 abderitids, but also of the major palaeothenitid lineages (i.e. Palaeothenitinae and Acdestinae).
772 Further studies (in progress) will allow to test the current taxonomic assignments and to refine
773 diversification timings of non-Pichipilidae Palaeothenitoidea.

774 775 5.2 Environment

776 The abundance of pedogenic nodules (calcretes) throughout the latest Eocene–early Oligocene
777 interval points to contrasted seasonal rainfall and water-table oscillation. The poor palynomorph
778 recovery at TAR-72 is compatible with understory vegetation in a tropical rainforest (Fungi
779 monocellate and fern spores) and with seasonal water-table fluctuation.
780 Quantitative paleoclimatic inferences were not performed due to the limited number of leaf and
781 seed fossil material that could be collected in the field. The Shapaja record, however, consists
782 mostly of nanophyll leaflets of compound leaves or microphyll leaves. In addition, most of the
783 fossil leaves recovered came from the early Oligocene section (i.e., TAR-01), which could indicate
784 that the forests at this time likely adapted to longer periods of drought (e.g., strong seasonal
785 precipitation pattern) by becoming deciduous and reducing the leaf lamina (see [Peppe et al., 2011](#)).
786 Nevertheless, larger collections are necessary to confirm this hypothesis. Interestingly,
787 recent studies documenting macrofloras throughout the late Eocene to early Oligocene from
788 Neotropical localities also provide evidence to support an increment in seasonal conditions in this
789 region ([Calvillo-Canadell and Cevallos-Ferriz, 2005](#); [Woodcock et al., 2017](#); [Martínez et al., this issue](#)),
790 a pattern that has been previously proposed for northern high-latitude localities ([Eldrett et](#)

791 [al., 2009](#)). Most of the taxonomically identified fossil seeds came from the latest Eocene locality
792 (TAR-21), which includes at least two taxa of Passifloroideae (passion fruit subfamily) and one
793 taxon of the Araceae (cf. *Monstera*). These plants are common vines or epiphytes in extant
794 tropical rainforests in the region. The presence of these plants in the latest part of the Eocene also
795 coincides with the occurrence of primates, indicative of tropical rainforest environment (Fig 4V-
796 C'). The persistence of vine/epiphytic plants suggests the presence of high-canopy trees in this
797 part of the section near the vicinity of streams (TAR-74 to TAR-73) and ponds (TAR-01) throughout
798 the section. Seeds of Nymphaeaceae (TAR-73) and cf. *Elatine* (Elatinaceae; TAR-74 and TAR-01; Fig.
799 3I), a common cosmopolitan plant of aquatic environments (e.g., ponds, freshwater lakes), also
800 indicate the presence of freshwater settings. Moreover, the oldest and youngest localities (TAR-74
801 and TAR-73 + TAR-01, respectively) testify to the presence of obligate freshwater settings, as
802 revealed by the mollusk, decapod, and fish communities. Conversely, a marine/brackish influence
803 can be hypothesized in the middle part of the section (between TAR-20 and TAR-22) thanks to the
804 conspicuous presence of myliobatid and pristid selachians (bat rays and sawfish, respectively).
805 These taxa co-occur with a freshwater ichthyofauna dominated by characiform actinopterygians
806 and dipnoans, thus pointing to an estuarine environment. The presence of neonate/young batoid
807 specimens at TAR-20, TAR-13, and TAR-72 might further support brackish conditions for the
808 concerned interval, as many *Myliobatis* females enter estuaries and coastal plains to give birth
809 today ([Mianzan et al., 2001](#)). Accordingly, we can hypothesize that the concerned settings were
810 close to such protected environments playing a nursery role for bat rays (even if the occurrence of
811 a minute euryhaline/freshwater myliobatid species of unknown affinities cannot be fully
812 discarded). In the uppermost levels of the section (TAR-73 and TAR-01), the fish community
813 characterizes again a freshwater environment without any marine influence (potamotrygonid
814 selachians, dipnoans, and characiform actinopterygians). Nodule-rich blue clays of TAR-01 may
815 have deposited in an oxbow lake, which would further explain the presence of piranhas (see CTA-
816 32 near Contamana; [Antoine et al., 2016](#)).

817

818 5.3 Community turnovers

819 In this section, we provide a short overview of the most prominent taxonomic groups for which
820 changes have been recognized in terms of community composition over the studied section. The
821 faunal components of the different localities were informally grouped into three assemblages
822 based on taxonomic composition of relevant groups. Assemblages 1 and 3 refer to the late Eocene
823 (TAR-74) and early Oligocene (TAR-01) metatherian, rodent, and fish assemblages, respectively,
824 while Assemblage 2 refers to the latest Eocene–earliest Oligocene transitional taxonomic
825 composition (TAR-20, TAR-72, TAR-21, TAR-22, and TAR-73).

826

827 Charophyte and plant remains (leaves, silicified wood, and seeds) are documented throughout the
828 complete studied section. Changes in the diversity of the leaf macroflora along the section are
829 hard to evaluate as most of the material collected came from only one Oligocene locality (TAR-01).
830 Nevertheless, we found two leaf morphotypes (PZ1 and PZ3, shared between TAR-21sup (EOT)
831 and TAR-01), and also one seed taxon, cf. *Elatine* (shared between TAR-74 and TAR-01), implying
832 that these plants persisted to the EOT and were tolerant of the new climatic conditions of the
833 early Oligocene. Although additional plant fossil material will be required to fully reconstruct the
834 forest types of the Shapaja section, our preliminary data suggests the presence of multistratified
835 rainforests during the latest part of the Eocene (TAR-21) and more open, deciduous forests in the
836 earliest Oligocene (TAR-21sup to TAR-01).

837

Taxa / Locality - Age	late Eocene				early Oligocene		
	TAR-74	TAR-20	TAR-72	TAR-21	TAR-13	TAR-22	TAR-01
Marsupialiformes incertae sedis	X				X		X
Polydolopimorphia	X						
Prepidolopidae gen. et sp. nov.	X						
?Prepidolopidae				X			
Polydolopimorphia indet.		X	X				X
?Polydolopimorphia							
Argyrolagidae							
cf. <i>Proargyrolagus</i>		X	X				
<i>Proargyrolagus</i> nov. sp. A (small)				X			
<i>Proargyrolagus</i> nov. sp. B (large)							X
Paucituberculata							
Palaeothentinae indet. 1			X				
Palaeothentinae indet. 2				X			
Palaeothentidae sp. nov. C							X
Palaeothentidae sp. nov. D							X
Palaeothentidae indet.							X
Abderitidae indet.							X
cf. <i>Perulestes</i>				X			
Palaeothentoidea, Abderitidae-Palaeothentidae indet. 1						X	
Palaeothentoidea, Abderitidae-Palaeothentidae indet. 2							X
Palaeothentoidea, non-Pichipilidae indet. 1 (basal)			X				
Palaeothentoidea, non-Pichipilidae indet. 2							X
Palaeothentoidea indet.							X
Co-occurring species	2	2	4	4	1	1	10
Weight of sediment sampled (kg)	220	185	119	762	20	214	470
	Ass 1	Assemblage 2			Transitional?		Ass 3

838

839

840

841

842

843

844

845

846

847

848

849

850

851

852

853

854

855

856

857

858

859

860

861

862

863

864

865

Table 2. Taxonomic composition of metatherian communities from the late Eocene–early Oligocene Shapaja section, San Martín, Peruvian Amazonia, suggesting the presence of two successive turnovers (Assemblages 1–3), as for rodents (Table 3) and fish assemblies (Table 4, Fig. 6). Ass, Assemblage.

As for mammals, only metatherians and rodents are sufficiently documented in terms of specimen numbers and taxonomic richness to provide evidence related to faunal turnovers (Tables 2–4). Among metatherians, the Assemblage 1 (TAR-74, late Eocene) yields a new prepidolopid and an unidentified marsupialiaform, but neither argyrolagids nor palaeothentoids, which fully dominate latest Eocene assemblages (from TAR-20 to TAR-21; Assemblage 2). Another obvious turnover is observed between TAR-21 and TAR-01 (Assemblages 2 vs. 3: no species in common). Indeed, latest Eocene localities (Assemblage 2) are fully dominated by small argyrolagids, very abundant in TAR-21 and co-occurring with another possible prepidolopid and some early palaeothentoids. By the early Oligocene, however, argyrolagids are much less conspicuous while there is a profusion of derived non-Pichipilidae palaeothentoids, with the presence of new taxa among Palaeothentidae and Abderitidae (TAR-01: Assemblage 3). Unfortunately, the timing of this turnover cannot be refined, due to the lack of marsupial remains in interbedded localities (TAR-13, TAR-22, and TAR-73; Table 2). Even though a sampling bias cannot be fully discarded, the differences between Shapaja Assemblages 2 and 3 might well illustrate a real change in taxonomic composition and diversity. A sharp decline of metatherian diversity by the EOT has already been observed as part of the Patagonian hinge ("*Bisagra patagónica*"; Goin et al., 2010, 2016): Patagonian faunas reveal the EOT as a major ecologically-induced turnover in the history of Metatheria, with the disappearance of some lineages and the diversification of other ones (Goin et al., 2016). With respect to other Eocene–Oligocene South American metatherian faunas, Shapaja communities are fully distinct from the Tinguiririca fauna, Chile (Flynn et al., 2003) and very dissimilar to those of Argentinean Patagonia (La Cancha and La Cantera) and Southeastern Brazil (Guabirota), except for the presence of basal NPPs and argyrolagoids (Goin et al., 2010; Sedor et al., 2017). Strikingly, except

866 for yielding a prepidolopid and basal NPPs (*Perulestes* and *Sasawatsu*), the Santa Rosa fauna does
 867 not have strong affinities with Shapaja metatherian assemblages: sparassodontans,
 868 microbiotherians, and other Polydolopimorphia (e.g., *Wamradolops* and *Rosendolops*) do not
 869 occur at Shapaja; Argyrolagidae, Palaeothentidae (including Palaeothentinae), and Abderitidae
 870 have not been found at Santa Rosa. Shapaja and coeval mid- and low-latitude metatherian
 871 paleocommunities need to be thoroughly revised and compared with their higher latitude
 872 counterparts, in order to unravel their evolutionary dynamics at the South American scale.
 873

Taxa / Locality - Age	late Eocene				early Oligocene		
	TAR-74	TAR-20	TAR-72	TAR-21	TAR-13	TAR-22	TAR-01
Caviomorpha							
Stem Caviomorpha							
<i>Pozomys</i> sp. nov.	X						
<i>Tarapotomys subandinus</i>		X	X	X		X	
<i>Tarapotomys mayoensis</i>							X
aff. <i>Tarapotomys</i> sp.				X			
Caviomorpha gen. et sp. nov.		X		X	X		
<i>Shapajamys labocensis</i>							X
Incertae sedis							
Caviomorpha gen. et sp. indet. 1		X		X			
Caviomorpha gen. et sp. indet. 2						X	
Caviomorpha gen. et sp. indet. 3						X	
Caviomorpha gen. et sp. indet. 4	X						
Caviomorpha gen. et sp. indet. 5				X			
Erethicavioi							
Erethizontoidea							
<i>Eopululo</i> cf. <i>wigmorei</i>	X						
<i>Kichkasteiromys raimondii</i>				X			
Octochinchilloi							
<i>Mayomys confluens</i>							X
Octodontoidea							
Adelphomyidae gen. et sp. indet.					X		
<i>Selvamys paulus</i>						X	
Chinchilloidea							
<i>Eoincamys valverdei</i>		X	X	X	X	X	
<i>Eoincamys parvus</i>		X		X		X	
<i>Eoincamys</i> cf. <i>pascuali</i>							X
? <i>Eoincamys</i> sp. 1 (TAR-74)	X						
? <i>Eoincamys</i> sp. 2 (TAR-13)					X		
Chinchilloidea gen. et sp. indet.							X
Co-occurring species	4	5	2	8	4	6	5
Weight of sediment sampled (kg)	220	185	119	762	20	214	470
	Ass 1	Assemblage 2					Ass 3

874
 875 **Table 3.** Taxonomic composition of caviomorph rodent communities from the late Eocene–early
 876 Oligocene Shapaja section, San Martín, Peruvian Amazonia, suggesting the presence of two
 877 successive turnovers (Assemblages 1–3), as for metatherians (Table 2) and fish assemblies (Table
 878 4, Fig. 6). Ass, Assemblage.
 879

880 According to our results, three distinct rodent communities are recognized in the studied Shapaja
881 section (Assemblages 1–3; Table 3). The Assemblage 1 and 3 are late Eocene and early Oligocene
882 in age, respectively, whilst the Assemblage 2 is well documented in both latest Eocene and earliest
883 Oligocene localities. Four species of this Assemblage 2 survive the EOT locally (*Tarapotomys*
884 *subandinus*, *Caviomorpha* gen. et sp. nov., *Eoincamys valverdei*, and *E. parvus*). More generally,
885 chinchilloids and erethizontoids first occur prior to the EOT, whereas octodontoids (as represented
886 by Adelphomyidae indet. gen. et sp. at TAR-13 and *Selvamys* at TAR-22) appear after the EOT, at
887 least locally. The most prominent turnover occurs between TAR-22 (Assemblage 2) and TAR-01
888 (Assemblage 3), with species replacements among *Tarapotomys* (*T. subandinus* versus *T.*
889 *mayoensis*) and *Eoincamys* (*E. valverdei* + *E. parvus* versus *E. cf. pascuali*), and the first appearance
890 of both *Shapajamys* and of *Mayomys* at TAR-01. As illustrated by distinct depositional settings, this
891 turnover likely concurs with environmental changes locally, potentially related to the EOT.
892 Nevertheless, the corresponding turnover does not coincide temporally with this event but occurs
893 somewhat later (by Oi-1a times; Fig. 2, 6). Strikingly, all stratigraphically-constrained Shapaja
894 rodent assemblages (1–3), spanning the late Eocene–early Oligocene time interval (ca. 36–32.5
895 Ma), are equally homotaxic with the Santa Rosa rodent fauna, notably in sharing a stem
896 caviomorph (*Pozomys*; TAR-74), erethizontoids (from TAR-74 upward), chinchilloids (with a strong
897 domination of *Eoincamys* at TAR-20 and younger localities), and adelphomyine octodontoids (at
898 TAR-13; [Boivin et al., 2018, 2019a, 2019b](#)). This raises questions about Santa Rosa i) consisting of
899 several successive fossil-yielding levels of distinct ages or ii) being a time-averaged level with
900 reworked specimens, as further suggested by its extreme rodent specific richness (17 co-occurring
901 species; [Arnal et al., 2020](#)), recalling that of upper Oligocene Salla Beds, Bolivia (multiple fossil-
902 bearing levels, 29–25 Ma; e.g., [Pérez et al., 2019](#)).

903 So far, no primate has been recorded in the early Oligocene TAR-01 locality, which has been
904 sampled as extensively as TAR-21 and has yielded a specimen-rich rodent fauna (868 specimens).
905 Most tiny primates are dependent on dense forest habitats, and can be highly sensitive to
906 environmental changes. Although a taphonomic bias cannot be ruled out for explaining this
907 contrast between TAR-21 and the well-sampled overlying early Oligocene localities (i.e., TAR-22
908 and TAR-01), the apparent rarefaction of primates in post-EOT deposits could also be linked to
909 paleoenvironmental changes, at least locally, thereby corresponding to the onset of drier/open
910 terrestrial environments by the EOT or afterwards.

911 Even if it is likely to predate it, the bispecific primate community at TAR-21 strongly recalls that of
912 Santa Rosa ([Bond et al., 2015; Seiffert et al., 2020](#)). Conversely, poorly-documented Shapaja
913 ungulate assemblages seem to be highly distinct, at least for northern South America.

Taxa / Locality - Age	late Eocene				early Oligocene			
	TAR-74	TAR-20	TAR-72	TAR-21	TAR-13	TAR-22	TAR-73	TAR-01
Selachii								
Myliobatiformes								
Myliobatidae								
<i>Myliobatis</i> sp.		X	X	X	X			
? <i>Rhinoptera</i> sp.				X				
Potamotrygonidae								
<i>Potamotrygon</i> nov. sp.		X		X		X	X	
<i>Potamotrygon canaanorum</i>					X			
<i>Paratrygon</i> sp.				X		X		
Pristiformes								
Pristidae								
<i>Pristis</i> sp.						X		
Actinopterygii								
Characiformes								
Serrasalminidae								
cf. <i>Colossoma</i> sp.	X	X	X	X	X	X	X	X
cf. <i>Serrasalmus</i> sp.								X
Anostomidae								
cf. <i>Leporinus</i> sp.	X	X		X	X	X	X	X
<i>Leporinus</i> sp.	X	X	X	X	X	X	X	X
Cynodontidae								
cf. <i>Hydrolycus</i> sp.	X	X	X	X	X	X	X	X
Characiformes indet. 1	X	X		X				X
Characiformes indet. 2		X						X
Characiformes indet. 3		X		X		X	X	X
Siluriformes								
Siluriformes indet.		X		X				X
Loricariidae indet.				X				X
Pimelodidae								
cf. <i>Phractocephalus</i> sp.						X		
Sarcopterygii								
Lepidosireniformes								
? <i>Lepidosiren</i> sp.	X	X	X	X		X	X	X
Co-occurring species	6	11	5	13	6	10	7	11
Weight of sediment sampled (kg)	220	185	119	762	20	214	12	470
	Ass 1	Assemblage 2						Ass 3

914
915
916
917
918
919
920
921
922
923
924
925
926
927
928

Table 4. Taxonomic composition of fish communities from the late Eocene–early Oligocene Shapaja section, San Martín, Peruvian Amazonia, suggesting a turnover pattern similar to that of metatherian and rodent paleocommunities (see Tables 2-3, Fig. 6). Blue-typed occurrences and locality names refer to the marine affinities of Assemblage 2 (mixohaline coastal plain), as hypothesized by the presence of myliobatiform and pristiform selachians. Ass, Assemblage.

The Shapaja fish faunas are strongly reminiscent of the modern Amazonian lowland ichthyofauna, with characiform and siluriform actinopterygians, potamotrygonid selachians, and dipnoans (Brito and Deynat, 2004). They also recall the middle Eocene–late Miocene freshwater-dominated fish communities from Contamana (Adnet et al., 2014; Antoine et al., 2016; Chabain et al., 2017). In more detail, the Shapaja fish communities have distinctive features (Table 4), such as the conspicuous presence of a new species of *Potamotrygon* (in TAR-20, TAR-21, and TAR-22), the first fossil occurrence of the discus ray *Paratrygon* (TAR-21 and TAR-22), and the earliest occurrence of loricariid siluriforms (TAR-21), of *Potamotrygon canaanorum* (TAR-13; previously restricted to

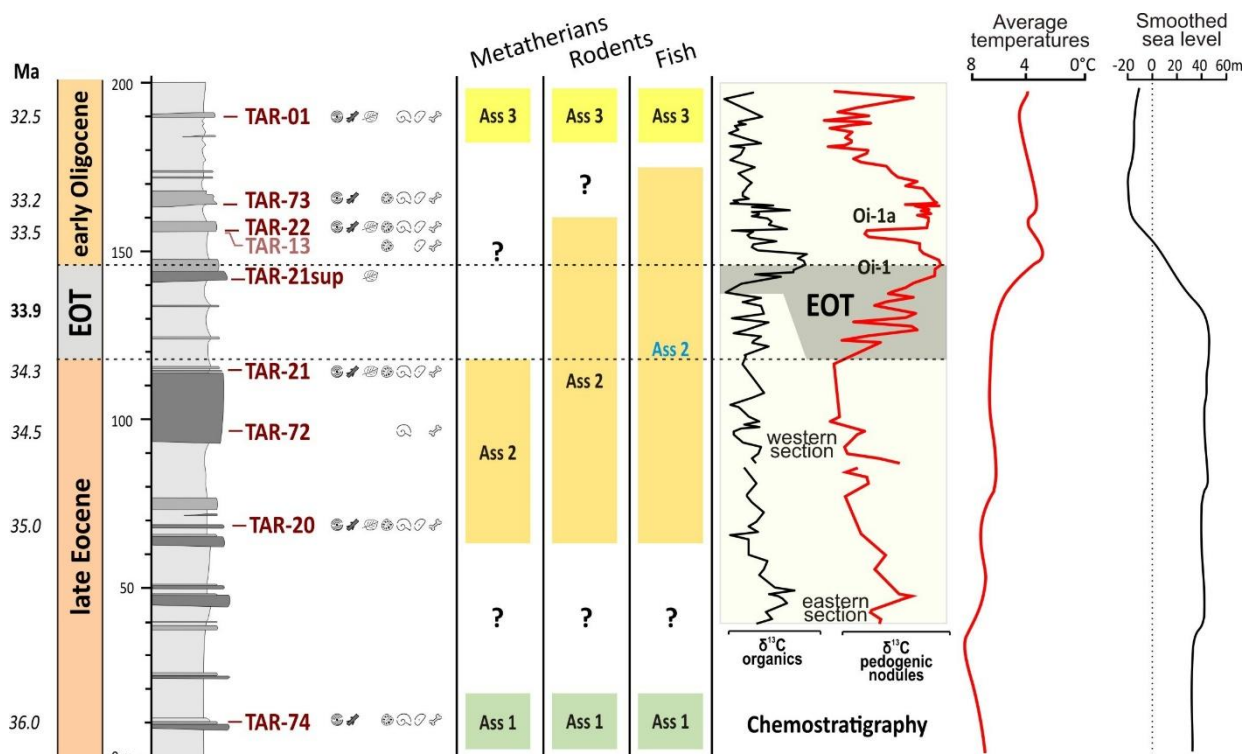
929 upper Oligocene settings; [Chabain et al., 2017](#)), and of piranhas closely allied to *Serrasalmus* (TAR-
930 01; previously recorded from upper Oligocene settings onward; [Antoine et al., 2016](#)). TAR-74 and
931 TAR-01 did not yield any selachian specimens, in spite of having been extensively sampled (Table
932 4; see SI). Despite the large amount of sediment sampled (220 kg), the fish community at TAR-74
933 only consists of characiforms and lepidosirenid dipnoans, typical of running-water settings, and
934 consistent with those of overlying localities (TAR-20 to TAR-73). However, the localities TAR-20 to
935 TAR-22 yielded selachians of marine/brackish affinities, such as *Myliobatis*, *Rhinoptera*, and/or
936 *Pristis*. The presence of *Myliobatis* neonates (at TAR-20, TAR-72, and TAR-13) may further point to
937 a mixohaline coastal plain environment (bat ray nurseries). In contrast, the distinctiveness of TAR-
938 01 in terms of ichthyofaunal content, without any selachians and further illustrated by the
939 presence of piranhas, may be related to the depositional setting (oxbow lake), contrasting with
940 that of all underlying localities, although the role of other EOT-related drivers cannot be discarded
941 (Table 4). In other words, the discrepancies between Shapaja fish Assemblages 1–3 appear to be
942 mainly related to environmental conditions, with a strict freshwater setting (TAR-74: Assemblage
943 1), a transgressive lag (brackish interval from TAR-20 to TAR-22: Assemblage 2), and a subsequent
944 regression (TAR-01: Assemblage 3). TAR-73 may correspond to a transitional phase between the
945 Assemblages 2 and 3 (no marine component, but a potamotrygonid).

946 947 5.4 Regional and/or global drivers

948 Assessing which drivers have provoked past biotic changes is quite challenging, especially
949 in areas as widely under-investigated paleontologically as tropical-equatorial regions of South
950 America. In the Shapaja section recording and bracketing the EOT, we will nonetheless try to
951 disentangle the potential roles of main regional and global drivers (i.e., vertical movements vs.
952 climatic and sea-level changes, respectively), likely to explain the biotic turnovers observed during
953 the late Eocene–earliest Oligocene interval.

954 From a tectono-sedimentary perspective, the Shapaja area is located in the Sub-Andean
955 Zone today. Recent regional syntheses for the Huallaga and Bagua basins in the northern Central
956 Andes consistently state that the concerned area only experienced Andean thrust-related
957 deformation much later, i.e., from late Miocene times onward ([Hermoza et al., 2005](#); [Roddaz et
958 al., 2010](#); [Eude et al., 2015](#); [Moreno et al., 2020](#)). During the late Eocene–Oligocene interval, this
959 region was steadily part of the foredeep depozone of the foreland basin. This time period
960 coincides with the onset of a tectonic loading stage, leading locally to flexural subsidence ([Roddaz
961 et al., 2010](#)). Such downward settling likely forced the late Eocene shift from freshwater to
962 brackish/marine-influenced depositional environments, as recorded locally between TAR-74 and
963 TAR-20 and further illustrated by the first ichthyofaunal turnover (Fig. 6: Assemblage 1 vs.
964 Assemblage 2). In addition, this shift occurred during a time period with no noticeable global sea-
965 level change (Fig. 6; [Miller et al., 2020](#)). Thereby, the concerned late Eocene biotic changes might
966 have been fully locally-driven.

967



968
 969 **Fig. 6.** Synthetic chrono-biostratigraphical chart of late Eocene–early Oligocene paleocommunities
 970 from Shapaja, San Martín, Peruvian Amazonia. Timeframe based on Vandenberghe et al. (2012)
 971 and current chemostratigraphical analyses. Ages in italics are approximate and tentative.
 972 Assemblages 1–3 derive from the present work, based on metatherian, rodent, and fish
 973 assemblages (selachians + actinopterygians) (see Tables 2–4). Blue-typed fish component refers to
 974 the marine affinities of Assemblage 2 (mixohaline coastal plain). Paleotemperature curve
 975 (Westerhold et al., 2020) and smoothed sea level curve (Miller et al., 2020) are adapted to match
 976 the timeframe of the section.
 977

978 In contrast, the time interval spanning the EOT and the earliest Oligocene records unstable
 979 and deteriorating global conditions: a drastic cooling led to a *ca.* 80 m drop in sea level between
 980 *ca.* 34 and 33.2 Ma (Fig. 6), provoked a sustained drop in precipitation, and increased seasonality,
 981 with a worldwide onset of drier climatic regimes (Hutchinson et al., 2018; Miller et al., 2020;
 982 Westerhold et al., 2020). Both the abundance of pedogenic nodules in the concerned interval of
 983 the Shapaja section and the concurrent shift from multistratified rainforest to more open
 984 deciduous forest consistently point to the strengthening of seasonal contrast, especially regarding
 985 rainfall (see previous sections). This global event temporally coincides with the decay of the
 986 Assemblage 2, as perceived on metatherians and rodents (Fig. 6). The apparently-delayed
 987 response of the ichthyofaunal community (turnover starting *ca.* 33–32.5 Ma) with respect to that
 988 of terrestrial components (plants and mammals: *ca.* 33.7 Ma) is perhaps related to the persistence
 989 of regional flexural subsidence accommodating locally the effect of global sea-level drop. In other
 990 words, during the EOT and the earliest Oligocene, Shapaja biotic community turnovers have
 991 probably been primarily globally-driven, particularly for their terrestrial components (plants and
 992 mammals).

993 TAR-01 probably records new stable environmental conditions, under a drier and more
 994 seasonally-contrasted climate, but its highly-distinctive metatherian and rodent assemblages
 995 prevent from discussing further their sustainability at any spatiotemporal scale (Assemblage 3; Fig.
 996 6).
 997

998 6 Conclusion

1000 The Shapaja section ranges the late Eocene–early Oligocene interval (*ca.* 36–32.5 Ma). As
 1001 suggested by chemostratigraphical results, this section records both the EOT and positive carbon
 1002 isotopic incursions interpreted as Oi-1 and Oi1a events. The latest Eocene–earliest Oligocene
 1003 interval evidences a marine/brackish influence, as indicated by the co-occurrence of several
 1004 selachians, likely forced by regional flexural subsidence. Obligate freshwater depositional
 1005 environments of the uppermost part of the section are compatible with the earliest Oligocene
 1006 global sea level drop. Leaf physiognomy (mainly nanophyll leaflets of compound leaves or
 1007 microphyll leaves with entire margins) tentatively suggests a warm seasonal climate during the
 1008 EOT interval, while the seeds of vine/epiphytic plants during the latest Eocene suggests the
 1009 presence of high-canopy trees near the vicinity of streams and/or ponds. Two biotic turnovers are
 1010 retrieved (metatherians, rodents, and fish): one during the late Eocene (*ca.* 35–36 Ma) and another
 1011 one several thousand years after the EOT (*ca.* 33 Ma). TAR-21 (latest Eocene) records the first
 1012 South American primates unquestionably predating the EOT. More broadly, this section
 1013 documents the earliest occurrences of several genera, families, and/or superfamilies: *Eopululo* and
 1014 *Erethizontoidea* (late Eocene); *Paratrygon*, *Proargyrolagus* and *Argyrolagidae*, *Palaeothentidae*,
 1015 *Parastegosimpsonia*, *Chinchilloidea*, *Eoincamys*, and *Loricariidae* (latest Eocene); *Adelphomyinae*,
 1016 *Mylodontidae*, *Abderitidae*, and true piranhas (earliest Oligocene).
 1017 The paleocommunities from Shapaja exemplify a local response of terrestrial and aquatic
 1018 organisms to drastic regionally- then globally-driven changes over the late Eocene–early Oligocene
 1019 interval, at low latitudes in South America.

1020

1021 **Declaration of competing interest**

1022 All authors declare that they do not have any conflict of interest.

1023

1024 **Acknowledgments:** We are indebted to Adriana Albino (Museo de La Plata, Argentina) for her
 1025 taxonomic assignment of squamate remains. Fieldwork and post-field analyses in Peru were
 1026 carried out thanks to the support from the National Geographic Society (grant n° 9679-15) and the
 1027 Campus France program of the French Ministry of Foreign Affairs to POA, from the Doctoral School
 1028 SIBAGHE/Gaia of the Montpellier University to MB, the Oak Spring Garden Foundation to F.
 1029 Herrera and from the Institut des Sciences de l'Evolution de Montpellier, and The Leakey
 1030 Foundation to LM. This work was further supported by an "Investissements d'Avenir" grant
 1031 managed by the Agence Nationale de la Recherche (CEBA, ANR-10-LABX-25-01) and by the
 1032 cooperative programs ECOS-FonCyT (A14-U01) and CoopIntEER CNRS-CONICET (n°252540), in the
 1033 frame of the ongoing cooperation agreement between the Museo de Historia Natural de la
 1034 Universidad Nacional Mayor San Marcos (Lima, Peru) and the Institut des Sciences de l'Evolution-
 1035 Université de Montpellier. We are particularly indebted to the invited editor, Carina Hoorn, to
 1036 Robin M. D. Beck and to an anonymous reviewer for their useful comments and helpful
 1037 suggestions on a previous version of the manuscript. This is the article ISEM-2021-039 SUD.

1038

1039

1040 **References**

1041

- 1042 Abello, M. A., 2007. Sistemática y bioestratigrafía de los Paucituberculata (Mammalia: Marsupialia) del Cenozoico de
 1043 América del Sur (Doctoral dissertation, Universidad Nacional de La Plata).
 1044 Abello, M.A., Candela, A.M. 2020. Paleobiology of *Argyrolagus* (Marsupialia, Argyrolagidae): an astonishing case of
 1045 bipedalism among South American mammals. *J. Mammal. Evol.* 27, 419–444.
 1046 Abello, M.A., Toledo, N., Ortiz-Jaureguizar, O., 2018. Evolution of South American Paucituberculata (Metatheria:
 1047 Marsupialia): adaptive radiation and climate changes at the Eocene-Oligocene boundary, *Hist. Biol.*, doi:
 1048 10.1080/08912963.2018.1502286

- 1049 Adnet, S., Salas Gismondi, R., Antoine, P.-O., 2014. Comparisons of dental morphology in river stingrays
1050 (Chondrichthyes: Potamotrygonidae) with new fossils from the middle Eocene of Peruvian Amazonia rekindle
1051 debate on their evolution. *Naturwissenschaften* 101 (1), 33–45.
- 1052 Aguilera, O.A., Bocquentin, J., Lundberg, J.G., Maciente, A., 2008. A new cajaro catfish (Siluriformes: Pimelodidae:
1053 *Phractocephalus*) from the Late Miocene of southwestern Amazonia and its relationship to †*Phractocephalus nassi*
1054 of the Urumaco Formation. *Paläont Z*, 82 (2), 231–245.
- 1055 Anthony, H.E., Richards, J.G., 1924. A new fossil perissodactyl from Peru. *Am. Mus. Novitates* 111.
- 1056 Antoine, P.-O. et al., 2016. A 60-million-year Cenozoic history of western Amazonian ecosystems in Contamana,
1057 eastern Peru. *Gondwana Res.* 31, 30–59.
- 1058 Arnal, M., Kramarz, A.G., Vucetich, M.G., Frailey, C.D., Campbell, K.E., 2020. New Palaeogene caviomorphs (Rodentia,
1059 Hystricognathi) from Santa Rosa, Peru: systematics, biochronology, biogeography and early evolutionary trends.
1060 *Pap. Palaeontol.* 6 (2), 193–216.
- 1061 Assemat, A. et al., 2019. Restes inédits de rongeurs caviomorphes du Paléogène de la région de Juanjui (Amazonie
1062 péruvienne) : systématique, implications macro-évolutives et biostratigraphiques. *Geodiversitas* 41 (20), 699–730.
- 1063 Beard, K.C., Coster, M.C., Salem, M.J., Chaimanee, Y., Jaeger, J.-J., 2017. Biogeographic provincialism shown by Afro-
1064 Arabian mammals during the middle Cenozoic: climate change, Red Sea rifting and global eustasy. In: Agius, D.A.,
1065 Khalil, E., Scerri, E.M.L., Williams, A. (Eds.), *Human Interaction with the Environment in the Red Sea - Selected*
1066 *Papers of Red Sea Project VI.* Koninklijke Brill NV, Leiden, pp. 48–68.
- 1067 Beck, R.M.D. 2018. The Skull of *Epidolops ameghinoi* from the Early Eocene Itaboraí Fauna, Southeastern Brazil, and
1068 the Affinities of the Extinct Marsupialiform Order Polydolopimorphia. *J. Mammal. Evol.* 24, 373–414.
- 1069 Benammi, M. et al., 2019. Geology, biostratigraphy and carbon isotope chemostratigraphy of the Paleogene fossil-
1070 bearing Dakhla sections, Southwestern Moroccan Sahara. *Geol. Mag.* 156 (1), 117–132.
- 1071 Berggren, W.A., Prothero, D.R. (Eds.), 1992. *Eocene-Oligocene climatic and biotic evolution: an overview.* Princeton
1072 University Press, Princeton, 568 pp.
- 1073 Boivin, M., Marivaux, L., Antoine, P.-O., 2019a. L'apport du registre paléogène d'Amazonie sur la diversification initiale
1074 des Caviomorpha (Hystricognathi, Rodentia) : implications phylogénétiques, macroévolutives et
1075 paléobiogéographiques. *Geodiversitas* 41 (4), 143–245.
- 1076 Boivin, M. et al., 2017. Late Oligocene caviomorph rodents from Contamana, Peruvian Amazonia. *Pap. Palaeontol.* 3
1077 (1), 69–109.
- 1078 Boivin, M. et al., 2018. Early Oligocene caviomorph rodents from Shapaja, Peruvian Amazonia. *Palaeontographica Abt.*
1079 *A* 311 (1-6), 87–156.
- 1080 Boivin, M., Marivaux, L., Salas-Gismondi, R., Vieytes, E.C., Antoine, P.-O., 2019b. Incisor enamel microstructure of
1081 Paleogene caviomorph rodents from Contamana and Shapaja (Peruvian Amazonia). *J. Mammal. Evol.* 26 (3),
1082 389–406.
- 1083 Bond, M. et al., 2015. Eocene primates of South America and the African origins of New World monkeys. *Nature* 520,
1084 538–541.
- 1085 Brito, P.M., Deynat, P.P., 2004. Freshwater stingrays from the Miocene of South America with comments on the rise of
1086 potamotrygonids (Batoidea, Myliobatiformes). In Arratia, G., M.V H. Wilson & R. Cloutier, *Recent advances in the*
1087 *origin and early radiation of vertebrates.* Dr. Friedrich Pfeil, Munich, pp. 575-582.
- 1088 Calvillo-Canadell, L., Cevallos-Ferriz, S.R.S., 2005. Diverse assemblage of Eocene and Oligocene Leguminosae from
1089 Mexico. *Int. J. Plant Sci.* 166, 671–692.
- 1090 Chabain, J. et al., 2017. Cenozoic batoids from Contamana (Peruvian Amazonia) with special focus on freshwater
1091 potamotrygonins (Chondrichthyes, Myliobatiformes) and their paleoenvironmental significance. *Geobios* 50,
1092 389–400.
- 1093 Chornogubsky, L., Goin, F.J., 2015. A review of the molar morphology and phylogenetic affinities of *Sillustania*
1094 *quechuense* (Metatheria, Polydolopimorphia, Sillustaniidae), from the early Paleogene of Laguna Umayo,
1095 southeastern Peru. *J. Vertebr. Paleontol.* 35(6), e983238.
- 1096 Ciancio, M.R., Carlini, A.A., Campbell, K.E., Scillato-Yané, G.J., 2013. New Palaeogene cingulates (Mammalia,
1097 Xenarthra) from Santa Rosa, Perú and their importance in the context of South American faunas. *J. Syst. Palaeont.*
1098 11(6), 727–741.
- 1099 Cifelli, R., Soria, M.F., 1983. Systematics of the Adiantidae (Liptoptera, Mammalia). *Am. Mus. Novitates*, 2771, 1-25.
- 1100 Coxall, H.K., Wilson, P., Palike, H., Lear, C., Backman, J., 2005. Rapid stepwise onset of Antarctic glaciation and deeper
1101 calcite compensation in the Pacific Ocean. *Nature* 433, 53–57.
- 1102 Coxall, H.K., Pearson, P.N., 2007. The Eocene–Oligocene Transition. In: Williams, M., Haywood, A.M., Gregory, F.J.,
1103 Schmidt, D.N. (Eds.), *Deep-Time Perspectives on Climate Change: Marrying the Signal from Computer Models and*
1104 *Biological Proxies.* The Micropalaeont. Soc., Spec. Pub. The Geol. Soc., London, pp. 351–387.
- 1105 DeConto, R.M., Pollard, D., 2003. Rapid Cenozoic glaciation of Antarctica induced by declining atmospheric CO₂.
1106 *Nature* 421, 245–249.

1107 Dunn, R.E., Strömberg, C.A.E., Madden, R.H., Kohn, M.J., Carlini, A.A., 2015. Linked canopy, climate, and faunal change
1108 in the Cenozoic of Patagonia. *Science* 347 (6219), 258–261.

1109 Dupont-Nivet, G. et al., 2007. Tibetan plateau aridification linked to global cooling at the Eocene-Oligocene
1110 transition. *Nature* 445, 635–638.

1111 Eldrett, J. S., Greenwood, D. R., Harding, I. C., Huber, M., 2009. Increased seasonality through the Eocene to Oligocene
1112 transition in northern high latitudes. *Nature* 459 (7249), 969–973.

1113 Erhardt, A. M, Pälke, H., Paytan, A., 2013. High-resolution record of export production in the eastern equatorial Pacific
1114 across the Eocene-Oligocene transition and relationships to global climatic records. *Paleoceanography* 28, 130–
1115 142.

1116 Eude, A., Roddaz, M., Bricchau, S., Brusset, S., Calderon, Y., Baby, P., Soula, J.-C., 2015. Controls on timing of
1117 exhumation and deformation in the northern Peruvian eastern Andean wedge as inferred from low-temperature
1118 thermochronology and balanced cross section. *Tectonics* 34 (4), 715–730.

1119 Feussom-Tcheumeleu et al., 2019. Upper Miocene depositional environments and landscapes, Shumanza section,
1120 Subandean Zone, Northern Peru. *Palaeobiodiversity and Palaeoenvironments* 100 (3), 719–735.

1121 Flynn, J.J., Wyss, A.R., Croft, D.A., Charrier, R., 2003. The Tinguiririca Fauna, Chile: biochronology, paleoecology,
1122 biogeography, and a new earliest Oligocene South American Land Mammal “Age”. *Pal. Pal. Pal.* 195, 229–259.

1123 Frailey, C.D., Campbell, K.E., 2004. Paleogene rodents from Amazonian Peru: the Santa Rosa local fauna. In: Campbell,
1124 K.E. (Ed.), *The Paleogene Mammalian Fauna of Santa Rosa, Amazonian Peru*. Nat. Hist. Mus. Los Angeles County,
1125 Los Angeles, pp. 71–130.

1126 Gelfo, J.N., Alonso, R.N., Madden, R.H., Carlini, A.A., 2019. An Eocene bunodont South American native ungulate
1127 (*Didolodontidae*) from the Lumbraera Formation, Salta Province, Argentina. *Ameghiniana* 57 (2), 132–145.

1128 Godfrey, L.R. et al., 2020. Mid-Cenozoic climate change, extinction, and faunal turnover in Madagascar, and their
1129 bearing on the evolution of lemurs. *BMC Evol. Biol.* 20, 97 (1–18).

1130 Goin, F.J., Candela, A.M., 2004. New Paleogene marsupials from the Amazon Basin of eastern Peru. In: Campbell, K.E.
1131 (Ed.), *The Paleogene Mammalian Fauna of Santa Rosa, Amazonian Peru*. Nat. Hist. Mus. Los Angeles County, Los
1132 Angeles, pp. 15–60.

1133 Goin, F.J., Abello, M.A., Chornogubsky, L., 2010. Middle Tertiary marsupials from central Patagonia (early Oligocene of
1134 Gran Barranca): understanding South America's *Grande Coupure*. In: Madden, R.H., Carlini, A.A., Vucetich, M.G.,
1135 Kay, R.F. (Eds.), *The Paleontology of Gran Barranca. Evolution and Environmental Change through the Middle*
1136 *Cenozoic of Patagonia*. Cambridge Univ. Press, Cambridge, pp. 69–105.

1137 Goin, F.J., Candela, A.M., Abello, M.A., Oliveira, E.V., 2009. Earliest South American paucituberculatans and their
1138 significance in the understanding of ‘pseudodiprotodont’ marsupial radiations. *Zool. J. Linnean Soc.* 155, 867–884.

1139 Goin F.J., Woodburne, M.O., Zimic, A.N., Martin, G.M., Chornogubsky, L., 2016. A brief history of South American
1140 metatherians. *Springer Earth System Sciences*, New York, 237 pp.

1141 Gröcke, D.R., Hesselbo, S., Jenkyns, H.C., 1999. Carbon-isotope composition of Lower Cretaceous fossil wood: ocean
1142 atmosphere chemistry and relation to sea-level change. *Geology*, 27, 155–158.

1143 Hermoza, W. et al., 2005. The Huallaga foreland basin evolution: thrust propagation in a deltaic environment,
1144 northern Peruvian Andes. *J. South Am. Earth Sc.* 19, 21–34.

1145 Hoorn, C. et al., 2010. Amazonia through time: Andean uplift, climate change, landscape evolution, and biodiversity.
1146 *Science* 330, 927–931.

1147 Hutchinson, D.K. et al., 2018. The Eocene-Oligocene transition: a review of marine and terrestrial proxy data, models
1148 and model-data comparisons. *Climate of the Past*, doi.org/10.5194/cp-2020-68.

1149 Janis, C.M., 1993. Tertiary mammal evolution in the context of changing climates, vegetation, and tectonic events.
1150 *Annu. Rev. Ecol. Syst.* 24, 467–500.

1151 Jaramillo, C., Rueda, M.J., Mora, G., 2006. Cenozoic plant diversity in the Neotropics. *Science* 311, 1893–1896.

1152 Katz, M.E. et al., 2008. Stepwise transition from the Eocene greenhouse to the Oligocene icehouse. *Nature Geosci.* 1,
1153 329–334.

1154 Kay, R.F. et al., 2019. *Parvimico materdei* gen. et sp. nov.: a new platyrrhine from the Early Miocene of the Amazon
1155 Basin, Peru. *J. Hum. Evol.* 134, 102628.

1156 Klaus, S., Magalhães, C., Salas-Gismondi, R., Gross, M., Antoine, P.-O., 2017. Palaeogene and Neogene brachyurans of
1157 the Amazon Basin: a revised first appearance date for primary freshwater crabs (*Brachyura*, *Trichodactylidae*).
1158 *Crustaceana* 90 (7-10), 953–967.

1159 Koenigswald, W.v., 1997. Brief survey of the enamel diversity at the schmelzmuster level in Cenozoic placental
1160 mammals. In: Koenigswald, W.v. & Sander, P.M. (Eds.), *Tooth enamel microstructure*. Rotterdam: Balkema, pp.
1161 137–161.

1162 Koenigswald, W.v., Martin, T., Billet, G., 2015. Enamel microstructure and mastication in *Pyrotherium romeroi*
1163 (*Pyrotheria*, *Mammalia*). *Paläontol. Z.*, 89(3), 593–609.

1164 Kohn, M.J. et al., 2015. Quasi-static Eocene-Oligocene climate in Patagonia promotes slow faunal evolution and mid-
1165 Cenozoic global cooling. *Pal. Pal. Pal.* 435, 24–37.

1166 Lear, C.H., Bailey, T.R., Pearson, P.N., Coxall, H.K., Rosenthal, Y., 2008. Cooling and ice growth across the Eocene-
1167 Oligocene transition. *Geology* 36, 251–254.

1168 Liu, Z. et al., 2009. Global cooling during the Eocene-Oligocene climate transition. *Science* 323, 1187–1190.

1169 Luciani, V. et al., 2010. Ecological and evolutionary response of Tethyan planktonic foraminifera to the middle Eocene
1170 climatic optimum (MECO) from the Alano section (NE Italy), *Pal. Pal. Pal.* 292, 82–95.

1171 Lundberg, J.G., 1997. Fishes of the La Venta fauna: additional taxa, biotic and paleoenvironmental implications. In:
1172 Kay, R.F., Madden, R.H., Cifelli, R.L. & Flynn, J.J. (Eds), *Vertebrate paleontology in the Neotropics: The Miocene
1173 fauna of La Venta, Colombia*. Smithsonian Institution Press, Washington, DC, pp. 67–91.

1174 Lundberg, J.G., Aguilera, O., 2003. The late Miocene *Phractocephalus* catfish (Siluriformes: Pimelodidae) from
1175 Urumaco, Venezuela: additional specimens and reinterpretation as a distinct species. *Neotrop. Ichthyol.* 1(2),
1176 97–109.

1177 Magioncalda, R., Dupuis, C., Smith, T., Steurbaut, E. and Gingerich, P. D., 2004. Paleocene–Eocene carbon isotope
1178 excursion in organic carbon and pedogenic carbonate: Direct comparison in a continental stratigraphic section.
1179 *Geology* 32(7), 553–556.

1180 Marivaux, L. et al., 2016. Neotropics provide insights into the emergence of New World monkeys: new dental evidence
1181 from the late Oligocene of Peruvian Amazonia. *J. Hum. Evol.* 97, 159–175.

1182 Marivaux, L., Adnet, S., Benammi, M., Tabuce, R., Benammi, M., 2017. Anomaluroid rodents from the earliest
1183 Oligocene of Dakhla, Morocco, reveal the long-lived and morphologically conservative pattern of the
1184 Anomaluridae and Nonanomaluridae during the Tertiary in Africa. *J. Syst. Palaeontol.* 15 (7), 539–569.

1185 Marivaux, L. et al., 2017. Earliest Oligocene hystricognathous rodents from the Atlantic margin of Northwestern
1186 Saharan Africa (Dakhla, Morocco): systematic, paleobiogeographical and paleoenvironmental implications. *J.
1187 Vertebr. Paleontol.* 37 (4), 1357567.

1188 Marivaux, L. et al., 2005. Anthropoid primates from the Oligocene of Pakistan (Bugti Hills): data on early anthropoid
1189 evolution and biogeography. *Proc. Natl. Acad. Sci. USA* 102 (24), 8436–8441.

1190 Marivaux, L., Welcomme, J.-L., Ducrocq, S., Jaeger, J.-J., 2002. Oligocene sivaladapid primate from the Bugti Hills
1191 (Balochistan, Pakistan) bridges the gap between Eocene and Miocene adapiform communities in southern Asia. *J.
1192 Hum. Evol.* 42 (4), 379–388.

1193 Martínez, C. et al., submitted. Paleoclimatic and paleoecological reconstruction of a late Eocene South American
1194 tropical dry forest. *Global and Planetary Change* (this issue).

1195 McKenna, M.C., Wyss, A.R., Flynn, J.J., 2006. Paleogene pseudoglyptodont xenarthrans from central Chile and
1196 Argentine Patagonia. *Am. Mus. Novitates* 3536, 1–18.

1197 Meng, J., McKenna, M.C., 1998. Faunal turnovers of Palaeogene mammals from the Mongolian Plateau. *Nature* 394,
1198 364–367.

1199 Mianzan, H. et al., 2001. The Río de la Plata Estuary, Argentina-Uruguay. In: Seeliger, U., Kjerfve, B. (Eds.), *Coastal
1200 Marine Ecosystems of Latin America*. Springer, Berlin, Heidelberg, pp. 185–204.

1201 Miller, K.G. et al., 2020. Cenozoic sea-level and cryospheric evolution from deep-sea geochemical and continental
1202 margin records. *Science Advances* 6, eaaz1346.

1203 Miller, K.G., Wright, J., Fairbanks, R., 1991. Unlocking the icehouse: Oligocene-Miocene oxygen isotopes, eustasy and
1204 margin erosion. *J. Geophys. Res.* 96, 6829–6848.

1205 Moreno, F. et al., 2020. Coupled Andean growth and foreland basin evolution, Campanian–Cenozoic Bagua Basin,
1206 northern Peru. *Tectonics* 39 (7), e2019TC005967.

1207 Mudelsee, M., Bickert, T., Lear, C.H., Lohmann, G., 2014. Cenozoic climate changes: A review based on time series
1208 analysis of marine benthic $\delta^{18}\text{O}$ records. *Rev. Geophys.* 52 (3), 333–374.

1209 Ni, X., Li, Q., Li, L., Beard, K.C., 2016. Oligocene primates from China reveal divergence between African and Asian
1210 primate evolution. *Science* 352 (6286), 673–677.

1211 Noiret, C. et al., 2016. New bio-chemostratigraphic dating of a unique early Eocene sequence from southern Europe
1212 results in precise mammalian biochronological tie-points. *Newslett. Strat.* 49 (3), 469–480.

1213 Pagani, M., Zachos, J.C., Freeman, K.H., Tipple, B., Bohaty, S., 2005. Marked decline in atmospheric carbon dioxide
1214 concentrations during the Paleogene. *Science* 309, 600–603.

1215 Patterson, B., Roper, S.H., Richards, J.G., 1942. Two Tertiary mammals from northern South America. *Am. Mus.
1216 Novitates* 1173, 1–7.

1217 Peppe, D.J., Hickey, L.J., Miller, I.M., Green, W.A., 2008. A Morphotype Catalogue, Floristic Analysis and Stratigraphic
1218 Description of the Aspen Shale Flora (Cretaceous–Albian) of Southwestern Wyoming. *Bull. Peabody Mus. Nat. Hist.*,
1219 49, 181–208.

1220 Peppe, D.J., Royer, D.L., Cariglino, B., Oliver, S.Y., Newman, S., Leight, E., Enikolopov, G., Fernandez-Burgos, M.,
1221 Herrera, F., Adams, J.M., Correa, E., Currano, E.D., Erickson, J.M., Hinojosa, L.F., Hoganson, J.W., Iglesias, A.,

- 1222 Jaramillo, C., Johnson, K.R., Jordan, G.J., Kraft, N.L.B., Lovelock, E.C., Lusk, C.H., Niinemets, U., Penuelas, J., Rapson,
1223 G., Wing, S.L., Wright, I.J., 2011. Sensitivity of leaf size and shape to climate: global patterns and paleoclimatic
1224 applications. *New Phytol.* 190, 724–739.
- 1225 Pérez, M.E. et al., 2019. New caviomorph rodents from the late Oligocene of Salla, Bolivia: taxonomic, chronological,
1226 and biogeographic implications for the Deseadan faunas of South America. *J. Syst. Palaeont.* 17 (10), 821–847.
- 1227 Pol, D., Powell, J.E., 2011. A new sebecid mesoeucrocodylian from the Rio Loro Formation (Palaeocene) of north-
1228 western Argentina. *Zool. J. Linn. Soc.* 163, 7–36.
- 1229 Pound, M.J., Salzmann, U., 2017. Heterogeneity in global vegetation and terrestrial climate change during the late
1230 Eocene to early Oligocene transition. *Sci. Rep.* 7:43386, 1–12.
- 1231 Premoli-Silva, I., Jenkins, D. G., 1993. Decision on the Eocene-Oligocene boundary stratotype. *Episodes*, 16 (3),
1232 379–382.
- 1233 Pujos, F., De Iuliis, G., Cartelle, C., 2017. A paleogeographic overview of tropical fossil sloths: towards an
1234 understanding of the origin of extant suspensory sloths?. *J. Mammal. Evol.* 24 (1), 19–38.
- 1235 Ribeiro, A.M., López, G.M., Bond, M., 2010. The Leontiniidae (Mammalia, Notoungulata) from the Sarmiento
1236 Formation at Gran Barranca, Chubut Province, Argentina. In: Madden, R.H., Carlini, A.A., Vucetich, M.G., Kay, R.F.
1237 (Eds.), *The paleontology of Gran Barranca: evolution and environmental change through the Middle Cenozoic of*
1238 *Patagonia* Cambridge University Press, New York, pp. 170–181.
- 1239 Rincón, A.D., Shockey, B.J., Anaya, F., Solórzano, A., 2015. Palaeoentid marsupials of the Salla Beds of Bolivia (late
1240 Oligocene): two new species and insights into the post-Eocene radiation of palaeoentoids. *J. Mammal. Evol.* 22
1241 (4), 455–471.
- 1242 Roddaz, M. et al., 2010. Cenozoic sedimentary evolution of the Amazonian foreland basin system. In: Hoorn, C.,
1243 Wesselingh, F.P. (Eds.), *Amazonia, Landscape and Species Evolution: A Look into the Past.* Blackwell-Wiley,
1244 Hoboken, pp. 61–88.
- 1245 Salas-Gismondi, R. et al., 2015. A Miocene hyperdiverse crocodylian community reveals peculiar trophic dynamics in
1246 proto-Amazonian mega-wetlands. *Proc. Roy. Soc. B* 282, 20142490.
- 1247 Sánchez Fernández, A.W., Chira Fernández, J.E., Valencia Muñoz, M.M., 1997. *Geología de los cuadrángulos de*
1248 *Tarapoto, Papa Playa, Utcucarca y Yanayacu 13-k, 13-l, 14-k, 14-l.* INGEMMET Boletín A, 94, Lima, 237 pp.
- 1249 Sánchez-Villagra, M.R., Kay, R. F., 1997. A skull of *Proargyrolagus*, the oldest argyrolagid (Late Oligocene Salla Beds,
1250 Bolivia), with brief comments concerning its paleobiology. *J. Vertebr. Paleontol.*, 17(4), 717–724.
- 1251 Sánchez-Villagra, M.R. 2001. The phylogenetic relationships of argyrolagid marsupials. *Zool. J. Linnean Soc.* 131(4),
1252 481–496.
- 1253 Sarkar, A., Sarangib, S., Ebiharac, M., Bhattacharyad, S.K., Raye, A.K., 2003. Carbonate geochemistry across the
1254 Eocene/Oligocene boundary of Kutch, western India: implications to oceanic O₂-poor condition and foraminiferal
1255 extinction. *Chem. Geol.* 201 (3–4), 281–293.
- 1256 Savage, D.E., Russell, D.E., 1983. *Mammalian Paleofaunas of the World.* Addison-Wesley, Reading, MA, 432 pp.
- 1257 Sedor, F. A., Oliveira, É.V., Silva, D.D., Fernandes, L.A., Cunha, R.F., Ribeiro, A.M., & Dias, E.V., 2017. A new South
1258 American Paleogene land mammal fauna, Guabirotuba Formation (southern Brazil). *J. Mammal. Evol.* 24 (1),
1259 39–55.
- 1260 Seiffert, E.R., 2007. Evolution and extinction of Afro-Arabian Primates near the Eocene-Oligocene boundary. *Folia*
1261 *Primatol.* 78, 314–327.
- 1262 Seiffert, E.R. et al., 2020. A parapithecoid stem anthropoid of African origin in the Paleogene of South America. *Science*
1263 368, 194–197.
- 1264 Shellis, R.P., Berkovitz, B.K.B., 1976. Observations on the dental anatomy of piranhas (Characidae) with special
1265 reference to tooth structure. *J. Zoology* 180 (1), 69–84.
- 1266 Shockey, B.J., Anaya, F., 2011. Grazing in a new late Oligocene mylodontid sloth and a mylodontid radiation as a
1267 component of the Eocene-Oligocene faunal turnover and the early spread of grasslands/savannas in South
1268 America. *J. Mammal. Evol.* 18 (2), 101–115.
- 1269 Shockey, B.J., Flynn, J.J., Croft, D.A., Gans, P., Wyss, A.R., 2012. New leontiniid Notoungulata (Mammalia) from Chile
1270 and Argentina: comparative anatomy, character analysis, and phylogenetic hypotheses. *Am. Mus. Novitates* 3737,
1271 1–64.
- 1272 Simpson, G.G., 1967. The beginning of the Age of Mammals in South America. *Bull. Am. Mus. Nat. Hist.* 137, 1–259.
- 1273 Soria, M.F., Hoffstetter, R., 1983. Présence d'un Condylarthre (*Salladolodus deuterotherioides* gen. et sp. nov.) dans le
1274 Déséadien (Oligocène inférieur) de Salla, Bolivie. *CR Acad. Sci.* 2, 297 (6), 549–552.
- 1275 Stehlin, H.G., 1909. Remarques sur les faunules de mammifères des couches éocènes et oligocènes du Bassin de Paris.
1276 *Bull. Soc. Geol. France* 9 (4), 488–520.
- 1277 Storme, J.-Y. et al., 2012. Paleocene/Eocene boundary section at Zumaia (Basque-Catatic Basin) revisited: new
1278 insights from high resolution magnetic susceptibility and carbon isotope chemostratigraphy on organic matter
1279 ($\delta^{13}\text{C}_{\text{org}}$): *Terra Nova* 24, 310–317.

1280 Toledo, C.E.V., Bertini, R.J., 2005). Occurrences of the fossil dipnoiformes in Brazil and its stratigraphic and
1281 chronological distributions. *Rev. Brasil. Paleontol.* 8 (1), 47–57
1282 Toumoulin, A. et al., 2020. Quantifying the effect of the Drake Passage opening on the Eocene Ocean. *Pal. Pal. Pal.* 35,
1283 e2020PA003889.
1284 Tramoy, R. et al., 2016. Stepwise paleoclimate change across the Eocene-Oligocene transition recorded in continental
1285 NW Europe by mineralogical assemblages and $\delta^{15}\text{N}_{\text{org}}$ (Rennes Basin, France). *Terra Nova* 28, 212–220.
1286 Traverse, A., 2007. *Paleopalynology: Second Edition*, 2nd ed, Topics in Geobiology. Springer Netherlands.
1287 Vandenberghe, N., Hilgen, F.J., Speijer, R.P., 2012. Chapter 28: The Paleogene period. In: Gradstein, F.M., Ogg, J.G.,
1288 Schmitz, M., Ogg, G. (Eds.), *The Geological Time Scale 2012*. Elsevier Science Ltd, Oxford, pp. 855–921.
1289 Van Mourik, C., Brinkhuis, H., 2005. The Massignano Eocene-Oligocene golden spike section revisited. *Stratigraphy* 2
1290 (1), 13–30.
1291 Villarroel, C., 2000. Un nuevo Mylodontinae (Xenarthra, Tardigrada) en la fauna de La Venta, Mioceno de Colombia: el
1292 estado actual de la familia Orophodontidae. *Rev. Acad. Colomb. Cie. Ex., Físicas y Naturales* 24 (90), 117–128.
1293 Wade, B. S., Pälike, H., 2004. Oligocene climate dynamics. *Paleoceanography* 19 (4), PA4019.
1294 Westerhold, T. et al., 2020. An astronomically dated record of Earth’s climate and its predictability over the last 66
1295 million years. *Science* 369, 1383–1387.
1296 Woodcock, D.W., Meyer, H.W., Prado, Y., 2017. The Piedra Chamana fossil woods (Eocene, Peru). *IAWA Journal* 38,
1297 313–365.
1298 Wolff, R.G., 1984. A new early Oligocene argyrolagid (Mammalia: Marsupialia) from Salla, Bolivia. *J. Vertebr. Paleontol.*
1299 4 (1), 108–113.
1300 Yans, J. et al., 2010. Carbon-isotope of fossil wood and dispersed organic matter from the terrestrial Wealden facies of
1301 Hautrage (Mons basin, Belgium). *Pal., Pal., Pal.* 291, 85–105.
1302 Yans, J. et al., 2014a. First carbon chemostratigraphy of the Ouled Abdoun phosphate Basin, Morocco: implications for
1303 dating and evolution of earliest African placental mammals. *Gondw. Res.* 25, 257–269.
1304 Yans, J. et al., 2014b. Refined bio- (benthic foraminifera, dinoflagellate cysts) and chemostratigraphy ($\delta^{13}\text{C}_{\text{org}}$) of the
1305 earliest Eocene at Albas-Le Clot (Corbières, France): implications for mammalian biochronology in Western Europe.
1306 *Newslett. Stratigr.* 47(3), 331–353.
1307 Yeo, D.C., Ng, P.K., Cumberlidge, N., Magalhaes, C., Daniels, S.R., Campos, M.R., 2007. Global diversity of crabs
1308 (Crustacea: Decapoda: Brachyura) in freshwater. In: Balian, E.V., Lévêque, C., Segers, H., Martens, K. (Eds.),
1309 *Freshwater animal diversity assessment*. Springer, Dordrecht, pp. 275–286.
1310 Zachos, J.C., Dickens, G.R., Zeebe, R.E., 2008. An early Cenozoic perspective on greenhouse warming and carbon-cycle
1311 dynamics. *Nature* 451, 279–283.
1312 Zachos, J.C., Pagani, M., Sloan, L., Thomas, E., Billups, K., 2001. Trends, rhythms, and aberrations in global climate 65
1313 Ma to Present. *Science* 292, 686–693.
1314 Zanazzi, A., Kohn, M.J., MacFadden, B.J., Terry, D.O., 2007. Large temperature drop across the Eocene-Oligocene
1315 transition in Central North America. *Nature* 445, 639–642.
1316 Zhifei, L., Shouting, T., Quanhong Z., Xinrong, C., Wei, H., 2004. Deep-water Earliest Oligocene Glacial Maximum
1317 (EOGM) in South Atlantic: *Chin. Sci. Bull.*, 49 (20), 2190–2197.
1318