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Order within chaos: Harnessing *Plasmodium falciparum var* gene extreme polymorphism for malaria epidemiology

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**Genotyping methods**

The characterization of *Plasmodium falciparum* genetic diversity is key to understand evolutionary pressure, measure the impact of elimination campaigns, monitor drug resistance, etc. A common genotyping method is to PCR amplify microsatellite markers or polymorphic genes, such as the *msp* family, the size of the resulting amplicon(s) being variable between different isolates [1]. Amplicon ultra-deep sequencing now allows multiplexing samples and are particularly useful to determine the proportion of each strain in infections with multiple genotypes [2]. Alternatively, genotyping a few dozen single nucleotide polymorphisms (SNPs) generates a molecular barcode specific to each isolate. Whole genome sequencing (WGS) is the most comprehensive approach, but the cost remains high for large-scale studies.

Over the last decade, the Day lab has pioneered a PCR approach based on *P. falciparum*’s most polymorphic gene family: the *var* genes [3]. The roughly 60 *var* genes have a similar organization consisting in a succession of Duffy binding-like (DBL) and cysteine-rich interdomain region (CIDR) domains with the near ubiquitous presence of DBLα subtype (Fig 1A) [4]. Through this approach, the DBLα domain is amplified using degenerate primers that match two short conserved motifs on either side of the domain. The approximately 450 bp-long sequence in between is extremely variable, making the total number of unique DBLα domains worldwide virtually infinite [5]. This cost-effective method is used as a surveillance tool across a variety of epidemiological settings. Unlike microsatellites that are presumably neutral markers, the DBLα typing offers the added advantage of characterizing the parasite’s most immunogenic surface protein family [6].

**DBLα sequences differentiate *Plasmodium falciparum* populations**

To test the suitability of DBLα sequences for population genetics analysis, Tonkin-Hill and colleagues processed 32,682 sequences from 1,248 *P. falciparum* isolates collected in 10 different countries located in South America, Africa, Asia, and Oceania [7]. To account for the frequent recombination events between *var* genes, they developed a novel computationally intensive method known as jumping hidden Markov Model (JHMM) which is able to infer the posterior probability that each location in an isolate’s DBLα type amino acid sequence is most closely related to every other DBLα type [8,9]. Through this approach, the DBLα domain is amplified using degenerate primers that match two short conserved motifs on either side of the domain. The approximately 450 bp-long sequence in between is extremely variable, making the total number of unique DBLα domains worldwide virtually infinite [5]. This cost-effective method is used as a surveillance tool across a variety of epidemiological settings. Unlike microsatellites that are presumably neutral markers, the DBLα typing offers the added advantage of characterizing the parasite’s most immunogenic surface protein family [6].

DBLα sequences provide an estimate of the expected proportion of relatedness between isolates. These proportions were then aggregated to provide estimates of an isolate’s DBLα repertoire that most closely matched each country, hence attributing a geographical origin to each isolate (Fig 1B). Country-specific clustering was observed, even in Africa where *P. falciparum*
Fig 1. Relevance of using DBLα sequences for malaria epidemiology. (A) Schematic view of DBLα domain in PfEMP1, the protein encoded by var genes. The amino acid sequence of DBLα domain, with letter size proportional to the prevalence in the P. falciparum population, shows two conserved motifs on either side. Universal primers were designed to match the corresponding DNA sequence. (B) Each isolate’s geographical origin is estimated by comparing DBLα types found in the isolate with DBLα types found in a specific country. (C) World map with each country from South America (blue), Africa (green), and Asia or Oceania (red) highlighted. Rectangle areas are proportional to the weight of each country in the overall estimated geographical origin of isolates, based on DBLα matching proportions. For example, isolates from Uganda share a relatively high number of DBLα with Ghana and Gabon, while PNG is more isolated. Each country is preferentially self-matching. Data from Tonkin-Hill and colleagues [7]. Made with Natural Earth.

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genetic diversity is particularly high [10], indicating that the majority of DBLα types are country specific. Excluding intracountry comparisons, countries located in the same continent show greater matching proportions of DBLα sequences, especially for Africa and South America. Regarding intercontinent comparisons, DBLα types coming from South America show greater relatedness to DBLα types found in Africa compared to Asia or Oceania (Fig 1C).

These results are in line with conclusions drawn from previous works, indicating that *P. falciparum* was introduced from Africa to South America during the slave trade [11]. Also, the JHMM proportions suggest that the *var* populations in Asia/Oceania more closely resemble African populations than those seen in South America, consistent with the expansion of *P. falciparum* out of Africa toward Asia [12]. Finally, the DBLα method is also able to distinguish between *Laverania* species.

**The exception to the rule: Conserved var genes**

While most DBLα in the Tonkin-Hill dataset were detected only once, some appeared to be conserved at the global scale. The top 100 most prevalent DBLα, present in at least 50 isolates, were found to be distributed all over the countries sampled. This result was confirmed with a BLAST search against the NCBI and MalariaGEN-derived databases [13]. Although the in vitro recombination rate of group A *var* genes is lower than group B or C [9], the DBLα1 (specific to group A *var* genes) was proportionally represented in the Top 100. The overall mind-boggling genetic diversity of DBLα sequences make these specific “conserved” DBLα even more interesting.

What is driving this selection? Some DBLα could be in linkage disequilibrium with drug resistance alleles [14]. However, the authors showed that only 10% of conserved DBLα were associated with drug resistance markers, indicating that other selection forces are at play and remained to be discovered. Interestingly, they identified conserved DBLα on chromosome 6, in which long-range haplotypes have been reported in African *P. falciparum* genomes [15].

**Plasmodium diversity: A matter of scale**

In summary, Tonkin-Hill and colleagues produced the first global scale DBLα types analysis. Their JHMM approach is promising when dealing with relatedness between sets of sequences, even though its computational cost remains to be lowered. Comparing DBLα sequences led to a clustering similar to what was observed using WGS in Africa, for a fraction of the cost. To further investigate the ability of DBLα comparisons to track *P. falciparum* populations, more samples are needed, especially from Ethiopia and the Democratic Republic of The Congo, where divergent parasites have recently been identified [10]. Follow-up studies should also include countries where malaria is highly seasonal, to measure the impact of transmission (or lack thereof) on parasite population genetic diversity.

To conclude, Tonkin-Hill and colleagues provide compelling evidence for further DBLα-based genotyping studies, not just for “varologists” but most epidemiological studies.

**References**


