



HAL
open science

The ecology and evolution of wildlife cancers: Applications for management and conservation

Rodrigo Hamede, Rachel Owen, Hannah Siddle, Sarah Peck, Menna Jones, Antoine M Dujon, Mathieu Giraudeau, Benjamin Roche, Beata Ujvari, Frédéric Thomas

► **To cite this version:**

Rodrigo Hamede, Rachel Owen, Hannah Siddle, Sarah Peck, Menna Jones, et al.. The ecology and evolution of wildlife cancers: Applications for management and conservation. *Evolutionary Applications*, 2020, 13 (7), pp.1719 - 1732. <10.1111/eva.12948>. <hal-03027879>

HAL Id: hal-03027879

<https://hal.umontpellier.fr/hal-03027879v1>

Submitted on 27 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons CC BY 4.0 - Attribution - International License



The ecology and evolution of wildlife cancers: Applications for management and conservation

Rodrigo Hamede^{1,2} | Rachel Owen³ | Hannah Siddle³ | Sarah Peck⁴ |
Menna Jones¹ | Antoine M. Dujon² | Mathieu Giraudeau⁵ | Benjamin Roche⁵ |
Beata Ujvari^{1,2} | Frédéric Thomas⁵

¹School of Natural Sciences, University of Tasmania, Hobart, Tas., Australia

²Centre for Integrative Ecology, School of Life and Environmental Sciences, Deakin University, Vic., Australia

³Centre for Biological Sciences, University of Southampton, Southampton, UK

⁴Wildlife Veterinarian, Veterinary Register of Tasmania, South Hobart, Tas., Australia

⁵Centre de Recherches Ecologiques et Evolutives sur le Cancer/Centre de Recherches en Ecologie et Evolution de la Santé, Unité Mixte de Recherches, Institut de Recherches pour le Développement 224-Centre National de la Recherche Scientifique 5290-Université de Montpellier, Montpellier, France

Correspondence

Rodrigo Hamede, School of Natural Sciences, University of Tasmania, Private Bag 55, Hobart, Tasmania 7001, Australia.
Email: rkhamede@utas.edu.au

Funding information

Agence Nationale de la Recherche, Grant/Award Number: ANR-18-CE35-0009; Australian Research Council, Grant/Award Number: DE170101116 and LP170101105; University of Tasmania Foundation; Rotary Club Les Sables d'Olonne; CNRS International Associated Laboratory

Abstract

Ecological and evolutionary concepts have been widely adopted to understand host-pathogen dynamics, and more recently, integrated into wildlife disease management. Cancer is a ubiquitous disease that affects most metazoan species; however, the role of oncogenic phenomena in eco-evolutionary processes and its implications for wildlife management and conservation remains undeveloped. Despite the pervasive nature of cancer across taxa, our ability to detect its occurrence, progression and prevalence in wildlife populations is constrained due to logistic and diagnostic limitations, which suggests that most cancers in the wild are unreported and understudied. Nevertheless, an increasing number of virus-associated and directly transmissible cancers in terrestrial and aquatic environments have been detected. Furthermore, anthropogenic activities and sudden environmental changes are increasingly associated with cancer incidence in wildlife. This highlights the need to upscale surveillance efforts, collection of critical data and developing novel approaches for studying the emergence and evolution of cancers in the wild. Here, we discuss the relevance of malignant cells as important agents of selection and offer a holistic framework to understand the interplay of ecological, epidemiological and evolutionary dynamics of cancer in wildlife. We use a directly transmissible cancer (devil facial tumour disease) as a model system to reveal the potential evolutionary dynamics and broader ecological effects of cancer epidemics in wildlife. We provide further examples of tumour-host interactions and trade-offs that may lead to changes in life histories, and epidemiological and population dynamics. Within this framework, we explore immunological strategies at the individual level as well as transgenerational adaptations at the population level. Then, we highlight the need to integrate multiple disciplines to undertake comparative cancer research at the human-domestic-wildlife interface and their environments. Finally, we suggest strategies for screening cancer incidence in wildlife and discuss how to integrate ecological and evolutionary concepts in the management of current and future cancer epizootics.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. *Evolutionary Applications* published by John Wiley & Sons Ltd

KEYWORDS

cancer, disease ecology, host–pathogen interactions, natural selection, transmissible tumour, wildlife management

1 | INTRODUCTION

Over the last two decades, significant efforts have been made to incorporate ecological and evolutionary principles to better understand the dynamics of wildlife diseases and their impact on wild populations (Galvani, 2003; Tompkins, Dunn, Smith, & Telfer, 2011; Vander Wal et al., 2014). The reciprocal interactions between host and pathogens are in many ways analogous to the interplay of ecological and evolutionary processes between species and their environment. Thus, the eco-evolutionary processes and feedbacks in emerging host–pathogen systems are currently considered key in epidemiology and disease management (Brosi, Delaplane, Boots, & Roode, 2017; Coen & Bishop, 2015; Grenfell et al., 2004). Cancer is a disease that evolved with the transition to multicellularity (Aktipis & Nesse, 2013) and therefore affects most metazoans on earth. It corresponds to a family of potentially lethal pathologies in which normal cells lose their typical cooperative behaviour, proliferate, spread and hence become malignant. Despite the ubiquitous nature of cancers in wildlife, the role of the oncobiota (i.e. oncogenic phenomena from precancerous lesions to metastatic cancer, Thomas et al., 2017) in ecological and evolutionary processes has been historically neglected (but see Thomas et al., 2017; Vittecoq et al., 2013) and its applications for wildlife management and conservation remain mostly in their infancy. Given that cancer is an evolving disease where the ecological context of tumour–host interactions is of paramount relevance for disease progression and immunological responses, evolutionary principles have recently been used in oncology as a novel approach for developing therapeutic treatments (Enriquez-Navas, Wojtkowiak, & Gatenby, 2015; Willyard, 2016; Zhang, Cunningham, Brown, & Gatenby, 2017).

Oncogenic phenomena can act as important agents of selection by having differential effects on the survival, life history, reproductive success and fitness of hosts (Thomas et al., 2017, 2018; Ujvari, Beckmann, et al., 2016). These processes can shape phenotypic, genetic and epigenetic variance across individuals, populations and species. Carcinogenesis is a complex process that depends on trade-offs at the cellular and organismal levels, and, in turn, these trade-offs interact with individuals and species, and hence ecosystems (Jacqueline et al., 2017; Pesavento, Agnew, Keel, & Woolard, 2018; Wu, Wang, Ling, & Lu, 2016). Thus, cancer should not be studied in isolation but as an interacting force of selection between species and their changing environments. Furthermore, in a century characterized by rapid environmental changes, species are increasingly facing additional ecological and immunological trade-offs that in turn may increase cancer risk (Jacqueline et al., 2017). Unravelling the synergistic effects of environmental degradation, ecological and evolutionary processes, and susceptibility to cancer is nonetheless a complex task. Recognizing these complexities using a multidisciplinary approach will permit the

understanding of important concepts underpinning cancer emergence and evolution and at the same time identify novel and integrative frameworks for managing cancers in wildlife.

The misleading assumption that cancers in wildlife are rare stems from the logistic difficulties in detecting their occurrence and monitoring their prevalence: in most cases, afflicted hosts are preyed upon or die unseen (Vittecoq et al., 2013). This suggests that most cancers in the wild are unreported and understudied. In addition, infectious agents are now well recognized as important drivers of cancer causation. For example, 15%–20% of all cancers in humans have been associated with a direct infectious origin (i.e. oncoviruses) (Alizon, Bravo, Farrell, & Roberts, 2019; Ewald & Swain Ewald, 2015). There is considerable evidence that environmental factors are a major contributor to cancer risk. Anthropogenic activities such as urbanization, chemical contamination and knock-on effects from rapid environmental changes have been associated with high cancer prevalence in wildlife and a lack of upregulation of anticancer defence mechanisms in these carcinogenic habitats (Giraudeau, Sepp, Ujvari, Ewald, & Thomas, 2018; Giraudeau et al., 2020; Pesavento et al., 2018; Sepp, Ujvari, Ewald, Thomas, & Giraudeau, 2019). However, only recently has cancer been considered a disease of conservation concern (McAloose & Newton, 2009) and transmissible cancers regarded as a new modality of infectious disease (Metzger & Goff, 2016). The increasing number of virus-associated and directly transmitted cancers detected in wildlife (Table 1), particularly for species already endangered (Gulland, Trupkiewicz, Spraker, & Lowenstine, 1996; James et al., 2019; McCallum et al., 2009; Williams et al., 1994; Woolford et al., 2008), demonstrates the urgent need for developing a holistic framework for studying oncogenic phenomena in the wild. Studying patterns of emergence, tumour–host interactions and evolutionary processes between hosts and malignant cells will also provide new insights into our understanding of how cancer defence mechanisms arise and evolve in nature (Nunney, 2013).

2 | IMMUNE RESPONSES TO INFECTIOUS CANCERS IN WILDLIFE

Infectious cancers can be broadly grouped into two categories: directly transmissible cancers, where the infectious agent is the cancer cell itself (Ostrander, Davis, & Ostrander, 2016), and indirectly transmissible cancers, where the infectious agent is a pathogen such as a virus that induces cancer formation (Ewald & Swain Ewald, 2015). Although there are similarities in terms of host immune responses, the interaction between these types of infectious cancers with the host immune system is multifaceted and in some cases cancer-specific, as described in detail below. The vertebrate immune system consists of two arms: the innate immune system, which functions

TABLE 1 Examples of infectious cancer in wildlife populations, their aetiology and prevalence

Taxa	Species	IUCN status	Cancer type	Aetiology	Prevalence	Reference
Amphibian	Leopard frog (<i>Rana pipiens</i>)	NL	Renal adenocarcinoma	Viral	10%	Granoff (1973)
	Tiger salamander (<i>Ambystoma tigrinum</i>)	LC	Pollution-related melanophoromas	Various pollutants	30%–50%	Rose and Harshbarger (1977)
	Japanese newt (<i>Cynops pyrrhogaster</i>)	LC	Cutaenous papillomas	Virus-like agent	Up to 5.45%	Asashima, Komazaki, Satou, and Oinuma (1982)
	Alpine newt (<i>Ichthyosaura alpestris</i>)	LC	Cutaenous papillomas	Unknown	20%–30%	Greven and Guex (2018)
	Montseny brook newt (<i>Calotriton arnoldi</i>)	CE	Skin tumour	Unknown	Up to 29%	Martínez-Silvestre, Amat, Bargalló, and Carranza (2011)
Avian	Barn swallow (<i>Hirundo rustica</i>)	LC	Cutaneous masses	Radiation	1.50%	Møller, Bonisoli-Alquati, and Mousseau (2013)
	White fronted goose (<i>Anser albifrons</i>)	LC	Lipoma/fibroma	Unknown	23%	Daoust, Wobeser, Rainnie, and Leighton (1991)
Fish	Walleye (<i>Sander vitreus</i>)	LC	Walleye dermal sarcoma	Viral	Unknown	Coffee, Casey, and Bowser (2013)
	Walleye (<i>Sander vitreus</i>)	LC	Discrete epidermal hyperplasia	Viral	Unknown	Coffee et al. (2013)
	Chinook salmon (<i>Oncorhynchus tshawytscha</i>)	NL	Plasmacytoid leukaemia	Viral	5%	Coffee et al. (2013)
	European smelt (<i>Osmerus eperlanus</i>)	LC	Spawning papillomatosis	Viral	5.50%	Coffee et al. (2013)
	Damselfish (<i>Stegastes partitus</i>)	LC	Neurofibromatosis	Virus-like agent	Up to 23%	Coffee et al. (2013)
	Brown bullhead (<i>Ameiurus nebulosus</i>)	LC	Cutaneous papillomas and squamous cell carcinoma	Pollutants	19%–58%	Baumann et al. (2008)
	English sole (<i>Parophrys vetulus</i>)	NL	Liver cancer	Pollutants	24%	Malins et al. (1988)
	Northern pike (<i>Esox lucius</i>)	LC	Lymphoma	Viral	21%	Papas, Dahlberg, and Sonstegard (1976)
	White sucker (<i>Catostomus commersonii</i>)	LC	Squamous cell carcinoma	Pollutants	33%–58%	Blazer et al. (2017)
	Dusky dolphins (<i>Lagenorhynchus obscurus</i>)	LC	Genital papilloma	Viral	57%–74%	Van Bresseem, Waerebeek, Piérard, and Desaintes (1996)
Mammal	Long beaked dolphin (<i>Delphinus capensis</i>)	DD	Genital papilloma	Viral	50%	Van Bresseem et al. (1996)
	Bottlenose dolphin (<i>Tursiops truncatus</i>)	LC	Genital papilloma	Viral	33%	Van Bresseem et al. (1996)
	Burmeister's porpoise (<i>Phocoena spinipinnis</i>)	NT	Genital papilloma	Viral	48.50%	Van Bresseem et al. (1996)
	Tasmanian devil (<i>Sarcophilus harrisii</i>)	E	Transmissible tumour	Transmissible cancer	40%–80%	Pearse and Swift (2006)
	Tasmanian devil (<i>Sarcophilus harrisii</i>)	E	Transmissible tumour	Transmissible cancer	20%–40%	Pye, Pemberton, et al. (2016)
	Beluga whale (<i>Delphinapterus leucas</i>)	LC	Intestinal adenocarcinoma	Pollutant	27%	Martineau et al. (2002)
	Sperm whale (<i>Physeter macrocephalus</i>)	V	Genital papilloma	Viral	10%	Lambertsen, Kohn, Sundberg, and Buergelt (1987)
	Island fox (<i>Urocyon littoralis catalinae</i>)	NT	Ceruminous gland hyperplasia/carcinoma	Ectoparasite	50%	Vickers et al. (2015)
	Raccoon (<i>Procyon lotor</i>)	LC	Brain tumours	Viral	15%	Cruz et al. (2013)
	Western bandicoot (<i>Perameles bougainville</i>)	V	Cutaneous hyperplasia/carcinoma	Viral	9%–18%	Woolford et al. (2008)
	California sea lion (<i>Zalophus californianus</i>)	LC	Urogenital carcinomas	Viral, pollutants, bacteria, genetic	18%	Deming et al. (2018)

(Continues)

TABLE 1 (Continued)

Taxa	Species	IUCN status	Cancer type	Aetiology	Prevalence	Reference
Mollusc	Soft shell clam (<i>Mya arenaria</i>)	NL	Transmissible cancer	Transmissible cancer	6.1%–95%	Brousseau (1987)
	Mussel (<i>Mytilus trossulus</i>)	NL	Transmissible cancer	Transmissible cancer	0%–29%	Mix (1983)
	Cockle (<i>Cerastoderma edule</i>)	NL	Transmissible cancer	Transmissible cancer	2%–46%	Poder and Auffret (1986)
	Carpenter shell clam (<i>Polittapes aureus</i>)	NL	Transmissible cancer	Transmissible cancer	42%	Metzger et al. (2016)
	Mussel (<i>Mytilus chilensis</i>)	NL	Transmissible cancer	Transmissible cancer	5%–13%	Yonemitsu et al. (2019)
	Mussel (<i>Mytilus edulis</i>)	NL	Transmissible cancer	Transmissible cancer	4%	Yonemitsu et al. (2019)
Reptile	Green sea turtles (<i>Chelonia mydas</i>)	E	Dermal fibropapillomas	Viral	60%	Jones, Ariel, Burgess, and Read (2016)
	Hawksbill turtle (<i>Eretmochelys imbricata</i>)	CE	Dermal fibropapillomas	Viral	Unknown	Poli, Lopez, Mesquita, Saska, and Mascarenhas (2014)
	Loggerhead turtle (<i>Caretta caretta</i>)	V	Dermal fibropapillomas	Viral	Unknown	Page et al. (2015)

Note: The conservation status of affected species according to the International Union for Conservation of Nature (IUCN) is shown: CE, Critically Endangered; DD, Data Deficient; E, Endangered; LC, Least Concern; NL, Not Listed; NT, Near Threatened; V, Vulnerable.

to induce systemic inflammation and nonspecific immune responses (Hato & Dagher, 2015), and the adaptive immune system, which exerts specific immune responses against pathogens or tumour-associated antigens (Cooper & Alder, 2006). Understanding the interaction between infectious cancers and the host immune system is key to developing effective disease management strategies.

2.1 | Directly transmissible cancers

There are nine known directly transmissible cancers: one in domestic dogs (CTVT; Murgia, Pritchard, Kim, Fassati, & Weiss, 2006), two independently evolved transmissible tumours in Tasmanian devils (*Sarcophilus harrisii*) (devil facial tumour disease [DFTD] and devil facial tumour 2 [DFT2]) (Pearse & Swift, 2006; Pye, Pemberton, et al., 2016) and six lineages of transmissible neoplasia circulating in six species of marine bivalves (Metzger et al., 2016; Yonemitsu et al., 2019). In CTVT and DFTD, immune evasion is at least partially achieved through downregulation of the major histocompatibility complex (MHC) proteins from the tumour cells' surface (Siddle et al., 2013; Yang, Chandler, & Dunne-Anway, 1987). MHC is a highly polymorphic group of proteins which label infected or cancerous cells for immune destruction by T cells (Wieczorek et al., 2017). Thus, removal of MHC from the cell surface hides the cancer from host immune cells and prevents clearance by the adaptive immune system. It has been demonstrated in both CTVT and DFTD that restoration of MHC to the cell surface can result in specific immune responses against the tumour cells (Hsiao et al., 2008; Tovar et al., 2017). In contrast, DFT2 expresses MHC on the cell surface (Caldwell et al., 2018); however, recent evidence suggests that DFT2 is currently losing its MHC-I expression from the cell surface (Ong, Lyons, Woods, & Flies, 2019), thereby enhancing its transmissibility potential.

Major histocompatibility complex polymorphism enables immune recognition of many pathogens, ensuring species survival in the face of epidemics (Savage & Zamudio, 2011; Sommer, 2005). Low polymorphism has been linked to reduced species fitness and a lower ability to recognize novel pathogens (Belasen, Bletz, Leite, Toledo, & James, 2019; Maibach & Vigilant, 2019), although this is not always the case (Castro-Prieto, Wachter, & Sommer, 2010). Low genetic diversity in Tasmanian devil populations, particularly in MHC genes (Cheng et al., 2012; Morris, Austin, & Belov, 2013; Siddle, Kreiss, et al., 2007), may have reduced the ability of the devil's immune system to distinguish self from non-self-malignant cells, facilitating the emergence of transmissible tumours (Caldwell et al., 2018). These mechanisms of emergence have been implicated in both DFTD and CTVT (Murchison et al., 2014; Siddle, Sanderson, Sanderson, & Belov, 2007), although the absence of MHC molecules from circulating tumours indicates that the host immune system has exerted pressure on the cancer cells during their evolution, as has been observed in single-organism cancer (McGranahan et al., 2017). The immune responses seen in DFTD, DFT2 and CTVT hosts are largely tumour-specific, indicating activation of the adaptive immune system against the cancer cells (Cohen, 1972; Hsiao et al., 2008; Pye,

Hamede, et al., 2016; Tovar et al., 2017). It is unclear whether marine bivalves raise any immune response against their transmissible neoplasia, although the lack of an adaptive immune system and MHC in invertebrates suggests that they may be more vulnerable to direct transmission of cancerous cell lines (Gestal et al., 2008; Metzger & Goff, 2016). At least until stronger anticancer defences (resistance) are selected for in these species, individuals could potentially achieve higher fitness by increasing their tolerance to cancer, that is surviving despite the presence of tumours (Thomas et al., 2019). Further studies would be necessary to test this hypothesis and to determine the extent to which the ecological and evolutionary drivers of tumour suppressor gene expression observed in certain vertebrates (i.e. elephants, see Abegglen et al., 2015) are also relevant in invertebrates. Currently, there is no empirical evidence for an exogenous initiator for any clonally transmissible cancers (Metzger et al., 2016; Murchison et al., 2012, 2014; Stammnitz et al., 2018). A promising direction worth to explore in light of the increasing number of transmissible cancers (Metzger & Goff, 2016; Ujvari, Beckmann, et al., 2016; Ujvari Gatenby, & Thomas, 2016bb) is to determine the contribution of the immune system complexity to the emergence of contagious malignant cell lines and whether transmissible tumours have an immune cell originator.

2.2 | Indirectly transmissible cancers

There are several examples of indirectly transmissible cancers in nature that induce variable host immune responses and are commonly associated with infection by oncogenic pathogens, though additional initiating factors are often implicated in tumorigenesis. In Atlantic bottlenose dolphins (*Tursiops truncatus*) suffering from papillomavirus, there is systemic inflammation and an activated innate immune response, with a partially activated adaptive immune response targeted against the virus rather than the tumour (Bossart et al., 2008; Rehtanz et al., 2010). Similarly, systemic inflammatory immune responses have been observed in green sea turtles (*Chelonia mydas*) suffering from virus-associated fibropapillomatosis alongside reduced lymphocyte proliferation, which may indicate immune exhaustion and a reduced capacity to raise an adaptive immune response to the tumour or pathogen (Cray, Varella, Bossart, & Lutz, 2001). A similar reduction of T-cell function has been demonstrated in Tasmanian devils following DFTD infection, suggesting that while the mode of avoiding T-cell recognition in DFTD is still not fully understood, there are similarities in certain immune evasion mechanisms (Cheng et al., 2019). In California sea lions (*Zalophus californianus*) suffering from Otarine herpes virus-1 (OthV-1)-associated urogenital carcinoma (King et al., 2002), there is a strong correlation between environmental organochlorine contamination and cancer incidence despite equivalent OthV-1 infection rates (Randhawa, Gulland, Ylitalo, DeLong, & Mazet, 2015). A link has also been demonstrated between MHC diversity and cancer risk (Bowen et al., 2005), indicating a genetic component to the disease that mirrors the emergence of directly transmissible tumours (Ujvari et al., 2018).

The ceruminous gland tumours affecting the Santa Catalina Island fox (*Urocyon littoralis catalinae*) are associated with ear mite infestations, and a generalized systemic inflammatory environment caused by bite wounds combined with a specific immune response to ear mite infection is thought to encourage tumour formation (Moriarty et al., 2015; Vickers et al., 2015). Similar mechanisms have been suggested in the emergence and transmission of facial tumours in the Tasmanian devil due to their aggressive social interactions (Hamede, McCallum, & Jones, 2013; Stammnitz et al., 2018).

The complex underlying causes of infectious cancers caused by pathogens often result in a systemic and nonspecific immune response that is not protective, causing chronic infection and tumour persistence (Browning, Gulland, Hammond, Colegrove, & Hall, 2015; Moriarty et al., 2015). One common feature that may underpin the emergence of directly and indirectly transmissible cancers is low genetic diversity, as evidenced by Tasmanian devils (Siddle, Kreiss, et al., 2007), Santa Catalina Island foxes (Hofman et al., 2015) and California sea lions (Acevedo-Whitehouse, Gulland, Greig, & Amos, 2003). However, many wild populations with extremely low genetic diversity thrive without increased cancer incidence (Weber, Stewart, Schienman, & Lehman, 2004), indicating that genetic diversity cannot alone be causative and that more complex interactions may be responsible for carcinogenesis. Although strong associations exist between pathogens and indirectly transmissible tumours, most infected individuals do not develop cancer, indicating that infection alone is not entirely the cause of tumour growth (Rehtanz et al., 2010; Vickers et al., 2015).

Infectious cancers are the result of complex combinations of genetic susceptibility, pathogenic infections, and abiotic and behavioural factors that allow the emergence and transmission of tumour cells or pathogens between individuals (i.e. the "perfect storm," see Ujvari, Beckmann, et al., 2016; Ujvari Gatenby, & Thomas, 2016aa). Understanding the interplay between these risk factors during the emergence and spread of cancers that are either caused by pathogens or by contagious cancer cell lines will not only help in managing current epidemics but also help to identify and manage emerging epidemics before they become widespread.

3 | ECOLOGICAL, EPIDEMIOLOGICAL AND EVOLUTIONARY DYNAMICS OF CANCERS IN WILDLIFE

Cancer emergence and progression do not occur in a vacuum, but rather in a complex suite of ecological and evolutionary interactions. In the same way that hosts can compensate for the fitness effects of parasitic infections (i.e. phenotypic plasticity of life-history traits), cancer is expected to trigger host responses to cope with the immunological and physiological demands of growing tumours. The diverse effects of cancer in host fitness (i.e. vulnerability to predation, susceptibility to coinfection with other pathogens, limited reproductive output, reduced ability to disperse) often result in host responses and adaptive processes early in cancer development.

For example, an experimental study demonstrated that *Drosophila* (*D. melanogaster*) with induced colorectal cancer are able to adjust their life-history traits by reaching the peak of oviposition significantly earlier than healthy ones (Arnal et al., 2017). Furthermore, there is evidence that the social environment of hosts can have a significant impact on cancer progression. *Drosophila* with induced colorectal cancer had faster tumour growth rates when kept in isolation than did flies in control groups (Dawson et al., 2018). These responses demonstrate the intricate and dynamic relationships between hosts and oncogenic processes and the ability of hosts to trade off fitness costs at different stages of disease. Likewise, host social structure, behaviour and sexual selection have the potential to affect contact rates and hence the transmission of infectious cancers (Vittecoq et al., 2015). Environmental factors driving the emergence of cancers, whether from anthropogenic sources (e.g. carcinogenic pollutants,) or natural sources (e.g. viral oncogenes), suggest a continuum of interactions and selection between host and oncogenic processes. Furthermore, increasing evidence from genomic studies suggests that certain oncogenes are capable of mutating and jumping hosts in species with disparate habitats and environmental attributes (Cortes-Hinojosa et al., 2019; Literak et al., 2010).

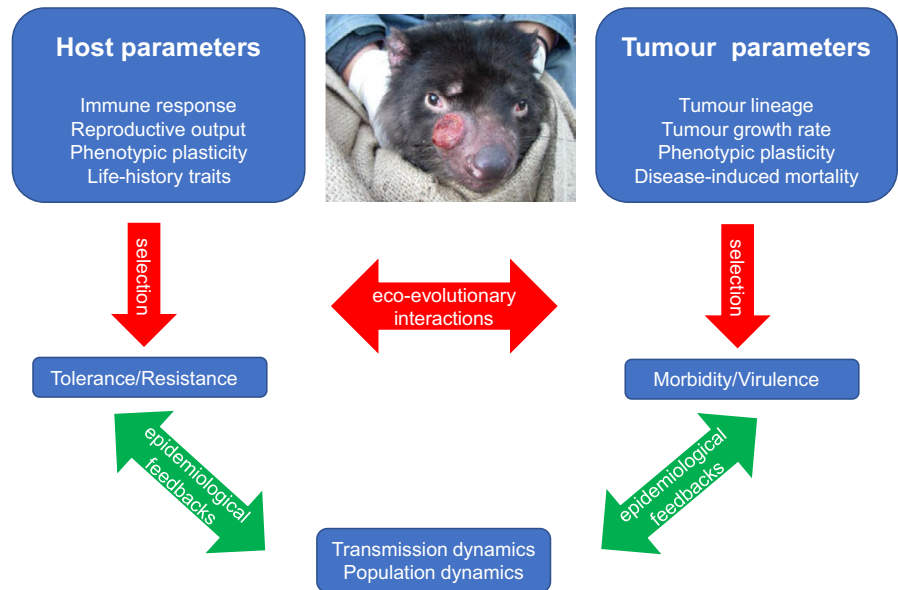
The vast majority of deaths caused by cancer are related to metastases, that is the development of secondary malignancies arising from the primary site of cancer in the host's body. However, metastatic cancer is the endpoint of a much more complex process with several stages, ranging from precancerous lesions to localized establishment and disseminated growths (Vittecoq et al., 2013). In some circumstances, cancerous lesions might never metastasize, either because the hosts die from other causes or due to the development of defence mechanisms such as tolerance and resistance. While tolerance (the ability to reduce disease costs in host fitness) and resistance (the ability to reduce disease burden or eliminate the disease) are mechanisms that have been mostly studied in host-parasite systems, there is now increasing evidence that these defence strategies are also applicable to oncogenic phenomena (Margres et al., 2018; Thomas et al., 2019). This is particularly relevant for transmissible cancers, where malignant cell lines persist beyond the host's life expectancy and selective processes favour the development of coping strategies across generations. For example, Tasmanian devils affected by DFTD have developed tolerance to the tumours, with females being able to maintain body condition at significantly larger tumour volumes than males (Ruiz-Aravena et al., 2018). Additionally, survival after DFTD infection has increased significantly in long-term diseased areas (Wells et al., 2017). An example of resistance to cancer occurs in CTVT, which originated in a wild canid between 6,000 and 10,000 years ago and currently affects domestic dogs (Baez-Ortega et al., 2019; Murgia et al., 2006). Infected dogs are able to develop immune responses, causing tumour regression and recovery (Das & Das, 2000). Although CTVT may have been highly lethal early in its evolutionary history (see also Leathlobhair et al., 2018), it now coexists with its hosts (Strakova & Murchison, 2015). Coexistence between dogs and CTVT might be the result of continuous selective processes between the cancer cell line and hosts over millennia.

However, the strong selective pressures of cancer can also operate on extremely short time scales. For example, a small proportion of Tasmanian devils have developed immune responses to DFTD resulting in natural tumour regressions in as little as 8–10 years (4–5 generations) after the cancer epidemic (Pye, Hamede, et al., 2016). The extremely high mortality of DFTD and the subsequent catastrophic population declines resulted in selection in regions of the genome that are associated with immune function and cancer risk (Epstein et al., 2016).

The high mortality caused by DFTD, where almost all individuals die within a year of attaining sexual maturity and their first mating event, would place strong selection pressure on life-history traits. In response to the cancer epidemic, a significant shift to younger populations and a 16-fold increase in the proportion of females able to breed in their first year (precocious sexual maturity) has been observed in several diseased sites (Jones et al., 2008; Lachish, McCallum, & Jones, 2009). Likewise, offspring sex ratios are more female-biased in diseased mothers compared to healthy mothers and litter size per female is significantly larger in populations where DFTD is present (Lachish et al., 2009; Lazenby et al., 2018). The rapid phenotypic and genotypic responses in the Tasmanian devil demonstrate that fast evolutionary processes in response to cancer can occur on ecologically relevant time scales. These processes can occur not just at the host level but also at the tumour level. Despite being a clonal cancer cell line, molecular studies have shown that DFTD is also subject to evolutionary plasticity (Murchison et al., 2012; Pearse et al., 2012; Ujvari, Beckmann, et al., 2016; Ujvari, Gatenby et al., 2016aa). More importantly, the evolutionary dynamics in the tumour can affect individuals and populations in different contexts. As the DFTD epidemic unfolded, a sudden local replacement of tumour karyotype (from tetraploid to diploid) resulted in a significant increase of infection rates and population decline (Hamede et al., 2015). Observed differential growth rates between tetraploid and diploid tumours (Hamede, Beeton, Carver, & Jones, 2017) may also select for polymorphism in tumour virulence. This may provide scope for an evolutionary arms race between cancer cells and hosts. At the host level, a broad range of eco-immunological dynamics such as seasonal dynamics of stress, demographic variation in immune expression profiles, reproductive hormones and immune senescence, as well as genetic and phenotypic variation, may interact with cancer susceptibility and tumour progression. At the tumour level, selection should favour lineages that reach optimal virulence, a trade-off between transmission rate and disease-induced mortality (Ebert & Bull, 2003).

The Tasmanian devil–DFTD system provides a unique opportunity to understand the interplay of ecological, evolutionary and epidemiological dynamics in response to cancer (Figure 1). Both tumours and devils have been consistently studied at multiple scales across the species' distributional range since the beginning of the epidemic. The observed selection and eco-evolutionary dynamics in DFTD should be used as a benchmark for studying and managing DFT2, for which limited information exists (James et al., 2019). More importantly, this knowledge could allow the use of several modelling approaches to predict the evolutionary trajectory of malignant

FIGURE 1 The Tasmanian devil and its transmissible tumour (DFTD), an ideal model system to understand how species adapt and evolve in response to infectious cancers and study the interplay of ecological, evolutionary and epidemiological processes. Blue boxes represent host and tumour parameters under selection through evolutionary interactions (red arrows). Host tolerance and resistance and tumour morbidity and virulence are under selection through ecological and evolutionary interactions. These interactions feed back into epidemiological and population dynamics (green arrows)



cells as well as evaluating critical epidemiological parameters such as tumour virulence, host susceptibility and tolerance/resistance to infection.

4 | FOLLOWING THE CANCER FOOTPRINT: FROM SPECIES CONSERVATION TO ECOSYSTEM FUNCTION

Cancer may be of particular concern in the small population paradigm in conservation, where stochastic causes of mortality can present a significant threat. Small populations or threatened species with low genetic diversity might be more susceptible to cancer (Ujvari et al., 2018). Population-level effects of cancer, such as reduction in population growth rate and cascading effects flowing through community and ecosystem levels, can be difficult to document. Establishing causal links between population decline and oncogenic processes in wildlife is fraught with the difficulties of long-term investigation and establishing the cause of mortalities in a sufficient proportion of the population. The clearest and best documented case of population decline caused by cancer is the Tasmanian devil-DFTD system (see Box 1).

Genetically isolated populations or those affected by other threatening processes can become more susceptible to cancer or mutagenic agents. For example, the critically endangered Santa Catalina Island fox neared extinction from hyperpredation by native eagles facilitated by abundant feral pigs (Roemer, Coonan, Garcelon, Bascompte, & Laughrin, 2001). Santa Catalina island foxes are also highly susceptible to exotic diseases (Crooks, Scott, & Vuren, 2001) and to ceruminous gland tumours, for which chronic inflammation from bacterial and mite infestation may promote tumorigenesis (Vickers et al., 2015). The genetic distinctiveness of this subspecies may predispose it to cancer: it has one of the highest rates of cancer observed in a wild population (Vickers et al., 2015). In further examples, debilitation by oncogenic viruses and tumour-associated

mortality limit population growth in small, isolated island populations of the western barred bandicoot (*Perameles bouganville*) and in a small population of Attwater's subspecies of the prairie chicken (*Tympanuchus cupido attwateri*) (McAloose & Newton, 2009). The prevalence of herpes virus-associated fibropapillomatosis is increasing in sea turtles, particularly in green sea turtles along the coasts of Florida and the Caribbean and Hawaiian Islands; it is considered to be a contributing factor to the ongoing population decline in these endangered species. Evidence of tumour regression offers a pathway for recovery (Guimarães, Gitirana, Wanderley, Monteiro-Neto, & Lobo-Hajdu, 2013; Tagliolatto, Guimarães, Lobo-Hajdu, & Monteiro-Neto, 2016) and potentially the evolution of resistance, as is occurring in Tasmanian devils (Epstein et al., 2016; Margres et al., 2018).

Documented evidence of trophic cascades triggered by cancer-induced population decline is rare and often not known (e.g. Santa Catalina Island fox; Vickers et al., 2015). Again, the Tasmanian devil-DFTD host-pathogen system provides the clearest and best documented case study. The progressive spatial and temporal patterns of devil population decline as DFTD has spread from east to west across the island state provide a rare natural experiment on the influential top-down role of this apex predator and primary scavenger in structuring Tasmanian ecosystems (Hollings, Jones, Mooney, & McCallum, 2014, 2016). The decline in devil populations has released invasive mesopredators from competition, with cascading effects on the decline in populations of small native mammals (Hollings, Jones, Mooney, & McCallum, 2016). Introduced pest species such as feral cats (*Felis catus*) and black rats (*Rattus rattus*) have increased in abundance (Cunningham et al., 2018). While the native mesopredator, the spotted-tailed quoll (*Dasyurus maculatus*), relax their temporal avoidance of devils when devils are at low density (Cunningham, Scoleri, Johnson, Barmuta, & Jones, 2019), it is possible that competition with the similar-sized feral cat, which has a higher fecundity (two rather than one litter per year), may counter the competitive release

from devils. Cats may be holding the smaller native mesopredator, the eastern quoll (*Dasyurus viverrinus*), in a “predator pit” where cats are at high density. Devil decline may also trigger a disease cascade, as the increased numbers of cats have been associated with a higher seroprevalence of *Toxoplasma gondii* in Tasmanian herbivores (Hollings, Jones, Mooney, & McCallum, 2013). These processes at the species and ecosystem levels highlight the broad-scale effects of cancer in wildlife and the vital need to document and study its implications for ecosystem functioning.

5 | APPLICATIONS FOR MANAGEMENT AND CONSERVATION

Studies of cancer in wildlife have been mostly accidental and reactive. There has been a historical lack of consistency in studying tumorigenesis across species and monitoring the broad-scale effects of cancer in wildlife. With few exceptions, such as Tasmanian devils and DFTD and CTVT in dogs, most studies are limited to a few postmortem examinations. In addition, many oncoviruses are asymptomatic and difficult to diagnose, making the epidemiological efforts needed to detect cancer in wildlife even more challenging. The discovery of eight transmissible cancers in the last 25 years (two in Tasmanian devils and six in marine

bivalves) suggests that (a) some species/environments might be particularly susceptible to these type of cancers, (b) there is a relationship between environmental change/disturbance and emergence of transmissible cancers or (c) transmissible cancers may have been more common throughout the evolutionary history of multicellular species, but our ability to detect them has only recently improved due to advances in biotechnology and multidisciplinary efforts in surveying populations. Furthermore, while multicellular organisms have been exposed to oncogenic phenomena throughout their evolutionary history, sudden changes in ecological and environmental conditions may result in a mismatch, promoting the development of cancers in the wild. Experimental studies in model systems can be a valuable tool to further understand mechanisms of selection and fitness costs associated with oncogenesis and tumour progression (i.e. Dawson et al., 2018).

Expanding the range and scope of studies on wildlife cancer is necessary to increase our ability to undertake comparative research at the human–wildlife–domestic interface and their environments. Robust data sets are key to further the development of fields such as comparative oncology and to understanding the prognosis, responses and survival in a broad range of malignancies across tissues, individuals, populations and species. Contrasting the biology and evolutionary ecology of tumours across species will provide a new perspective for understanding patterns of carcinogenesis and help

Box 1. The birth, spread and impact of transmissible cancers in Tasmanian devils

First detected in 1996 in northeast Tasmania, DFTD has spread across most of the distributional range of the devil in Tasmania (Figure 2; Hawkins et al., 2006; Lazenby et al., 2018). Clearly visible primary tumours (Loh et al., 2006), high recapture probability in trapping surveys (Lachish, Jones, & McCallum, 2007), and a concerted field monitoring and research effort have enabled clear causal links between DFTD spread, population decline and cascading effects at the ecosystem level to be established (Lazenby et al., 2018; McCallum et al., 2007).

Devil population decline accelerates 3 years after local disease emergence because the infection increases exponentially (McCallum et al., 2009), reaching a 60% decline after 5–6 years and up to a 90% decline in some areas (Lachish et al., 2007; McCallum et al., 2007). The rapid population decline led to the species being listed as *Endangered* at the international (IUCN Red List), national and state levels (Hawkins, McCallum, Mooney, Jones, & Holdsworth, 2009). Strong frequency-dependent transmission, likely caused by biting during the mating season, also contributed to concerns of extinction as a possible outcome of the epidemic (McCallum et al., 2009). However, to date, no local extinctions have been reported and long-term diseased populations persist despite high prevalence of tumours (Hamede et al., 2015; Lazenby et al., 2018). The rapid evolutionary response of devils to DFTD (within 4–6 generations) indicates that the adaptive shift is operating on the genetic variation present prior to the DFTD epidemic (Epstein et al., 2016), despite the low genetic diversity (Jones, Paetkau, Geffen, & Moritz, 2004; Siddle, Marzec, Cheng, Jones, & Belov, 2010) resulting from population bottlenecks during the last glacial maximum and during the Holocene (Brüniche-Olsen, Jones, Austin, Burridge, & Holland, 2014).

In 2014, a second and independently evolved transmissible cancer (DFT2) was discovered at the d'Entrecasteaux peninsula in south-eastern Tasmania. DFT2 and DFTD coexist in the same population, and a limited number cases of coinfection (both diseases in the same individual) have been reported (Kwon et al., 2018). Both tumours have been reported to be of neuroectodermal origin and most likely evolved from devils in north-eastern and south-eastern Tasmania (Stammnitz et al., 2018; Storfer et al., 2018). So far, DFT2 seems to be confined to the peninsula where it was first reported, although monitoring efforts outside of the peninsula have been limited (James et al., 2019). The population response to DFT2 and the epidemiological, ecological and evolutionary interactions between devils and DFTD are currently unknown; however, competition and selective processes are expected to occur at individual and population levels. In that sense, current and future research will be vital to predict epidemiological and evolutionary dynamics in the devil/DFTD/DFT2 study system.

mitigate risks of cancer emergence in the wild. We therefore suggest using a three-level approach to the study of wildlife cancer that will provide a solid link between fundamental research in cancer biology, eco-evolutionary processes and management and conservation.

First, increased networking and collaborative studies between disease ecologists and cancer biologists would maximize the capacity to diagnose cancer in the wild (Dujon et al. submitted to this Special Issue). A growing number of wildlife, human and environmental health surveys are undertaken globally at multiple scales, and initiatives such as *One Health* and *Conservation Medicine* are providing a practical link for disciplines that previously worked in isolation. This provides an ideal scenario for coordinating and expanding cancer surveillance at the ecosystem level. In parallel, new technologies and promising biomarkers for neoplasia are becoming valuable cost-effective diagnostic tools for monitoring cancer prevalence in captive and wild populations (Gourlan, Douay, & Telouk, 2019; Kourou, Exarchos, Exarchos, Karamouzis, & Fotiadis, 2015). The combination of technologies, coordination of surveys and diagnostic capacity offers a new platform to detect cancer in the wild, which will improve our ability to swiftly respond to epizootics and collect critical data.

Second, expertise should be drawn from different disciplines by integrating cancer research in humans, domestic species and wildlife populations to understand cellular, organismal and environmental mechanisms of carcinogenesis and their epidemiological and evolutionary patterns. While evolutionary biology and ecology are disciplines that have been recently integrated into oncology, the relevance of eco-evolutionary processes for recognizing cancer as an important agent of selection remains to be developed and integrated into wildlife management and conservation. For example,

evolutionary principles can be used for disentangling within- and between-host dynamics and trade-offs in response to cancer. Evaluating the role of infectious cancers as important agents of selection across populations provides a holistic and adaptive framework for understanding the adaptive capabilities of different species in response to oncogenic processes (Russell et al., 2018). The integration of these disciplines will also help to disentangle the biological/environmental mechanisms of cancer emergence and evaluate the diversity and lethality of tumours across taxa.

Finally, the knowledge generated from the cross-discipline framework should be used to develop adaptive management strategies and general guidelines in response to infectious cancers in wildlife. For example, understanding the long-term effects of the DFTD epidemic in Tasmanian devils—from its devastating population declines to the resulting functional changes in the genome—is critical for evaluating the extent to which management interventions are required. On the one hand, wildlife managers working with threatened wildlife are often focused on maximizing genetic diversity and reducing inbreeding (i.e. genetic rescue). On the other hand, modern genomic techniques have recently allowed the identification of adaptive genetic variation in response to drastic threatening processes such as disease epidemics or environmental degradations. The notion that rapid evolutionary changes in response to emerging infectious diseases can result in highly adapted genotypes and phenotypes by natural selection has given rise to the hope that populations in dire decline can be rescued through evolution (i.e. evolutionary rescue; Carlson, Cunningham, & Westley, 2014; DiRenzo et al., 2018). The adaptive capacity of Tasmanian devils in response to DFTD at different spatial and temporal scales (Epstein et al., 2016; Hamede, McCallum, & Jones, 2019) suggests that eco-evolutionary processes need to be thoroughly considered by wildlife managers and that rigorous evaluation of host–tumour interactions should be a priority to improve the conservation prospects of species in the face of epidemics (Hohenlohe et al., 2019).

The holistic vision proposed here is particularly relevant for most species affected by infectious cancers. The eradication of infectious cancers is not usually a plausible outcome; therefore, adaptations to these oncogenic processes are likely to evolve. Given the rapid environmental changes we are facing globally, the carcinogenic contaminants circulating in natural habitats and the increasing overlap among human, domestic and wildlife populations, greater attention should be given to screening for the development of neoplastic diseases across species and environments. In this sense, wildlife cancer can act a sentinel of environmental disturbance and species susceptibility to other threatening processes.

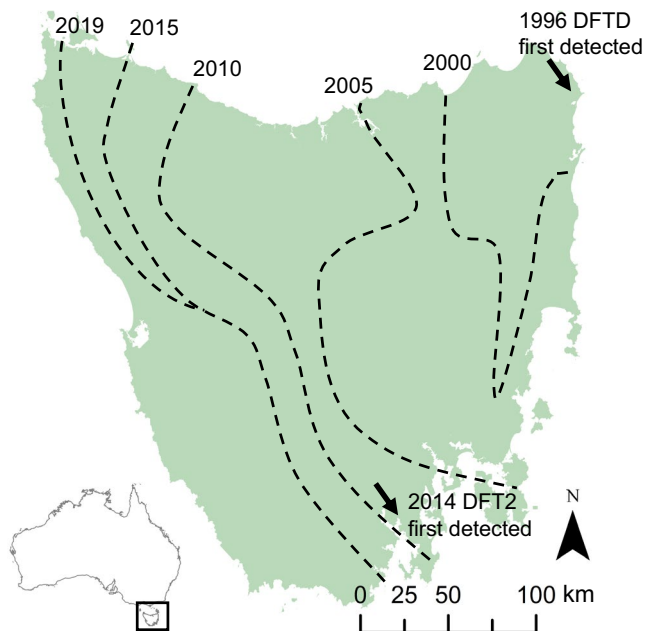


FIGURE 2 Map of Tasmanians showing the emergence and progression of two transmissible cancers in Tasmanian devils. While DFTD has spread from east to west over the last 25 years (dashed lines), DFT2 is still confined to the geographic area where it was first discovered

6 | CONCLUDING REMARKS

Much of the historical understanding of cancer has come from studies of human tumours and experimental research in laboratory mice. Because of this, cancer was until recently perceived as an evolutionary dead end. Studies in wildlife are now providing

novel perspectives for understanding eco-evolutionary processes at the cellular and organismal levels. As we look towards the future, there is a unique opportunity to integrate human, experimental and animal cancer research. The examples provided here highlight that cancer in wildlife is the result of a diverse range of mechanisms, including the emergence of novel cancer cell lines able to result in allograft transmission, an increasing number of virus-associated oncogenes, environmental change and carcinogenic pollutants. The pervasive nature of cancer in wildlife opens the field for studying the genesis of malignant cells, coping mechanisms at the individual level and transgenerational adaptations at the population level. Understanding how species respond and adapt to oncogenic processes and the trade-offs of suppressing malignant cell growth at the interface of environmental, ecological and evolutionary burdens should become a priority for oncologists, evolutionary biologists, disease ecologists and wildlife managers.

ACKNOWLEDGEMENTS

This work was supported by the Australian Research Council (DE170101116; LP170101105), the University of Tasmania Foundation (Save the Tasmanian Devil Appeal), Deakin SEBE_RGS_2019, ANR TRANSCAN (ANR-18-CE35-0009), the Rotary Club Les Sables d'Olonne and a CNRS International Associated Laboratory Grant.

CONFLICT OF INTEREST

None declared.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analysed in this study.

ORCID

Rodrigo Hamede  <https://orcid.org/0000-0003-1526-225X>

Beata Ujvari  <https://orcid.org/0000-0003-2391-2988>

Frédéric Thomas  <https://orcid.org/0000-0003-2238-1978>

REFERENCES

- Abegglen, L. M., Caulin, A. F., Chan, A., Lee, K., Robinson, R., Campbell, M. S., ... Schiffman, J. D. (2015). Potential mechanisms for cancer resistance in elephants and comparative cellular response to DNA damage in humans. *Jama-Journal of the American Medical Association*, 314, 1850–1860. <https://doi.org/10.1001/jama.2015.13134>
- Acevedo-Whitehouse, K., Gulland, F., Greig, D., & Amos, W. (2003). Disease susceptibility in California sea lions. *Nature*, 422, 35. <https://doi.org/10.1038/422035a>
- Aktipis, C. A., & Nesse, R. M. (2013). Evolutionary foundations for cancer biology. *Evolutionary Applications*, 6, 144–159. <https://doi.org/10.1111/eva.12034>
- Alizon, S., Bravo, I. G., Farrell, P. J., & Roberts, S. (2019). Towards a multi-level and a multi-disciplinary approach to DNA oncovirus virulence. *Philosophical Transactions of the Royal Society B-Biological Sciences*, 374, 5. <https://doi.org/10.1098/rstb.2019.0041>
- Arnal, A., Jacqueline, C., Ujvari, B., Leger, L., Moreno, C., Faugere, D., ... Thomas, F. (2017). Cancer brings forward oviposition in the fly *Drosophila melanogaster*. *Ecology and Evolution*, 7, 272–276. <https://doi.org/10.1002/ece3.2571>
- Asashima, M., Komazaki, S., Satou, C., & Oinuma, T. (1982). Seasonal and geographical changes of spontaneous skin papillomas in the Japanese newt *Cynops pyrrhogaster*. *Cancer Research*, 42, 3741–3746.
- Baez-Ortega, A., Gori, K., Strakova, A., Allen, J. L., Allum, K. M., Bansa-Issa, L., ... Murchison, E. P. (2019). Somatic evolution and global expansion of an ancient transmissible cancer lineage. *Science*, 365(6452), 464–+, eaa9923. <https://doi.org/10.1126/science.aau9923>
- Baumann, P. C., LeBlanc, D. R., Blazer, V. S., Meier, J. R., Hurley, S. T., & Kiryu, Y. (2008). Prevalence of tumors in brown bullhead from three lakes in Southeastern Massachusetts, 2002. <https://pubs.usgs.gov/sir/2008/5198/pdf/sir2008-5198.pdf>
- Belasen, A. M., Bletz, M. C., Leite, D. D. S., Toledo, L. F., & James, T. Y. (2019). Long-term habitat fragmentation is associated with reduced MHC IIB diversity and increased infections in amphibian hosts. *Frontiers in Ecology and Evolution*, 6, 236.
- Blazer, V. S., Walsh, H., Braham, R., Hahn, C., Mazik, P., & McIntyre, P. (2017). Tumours in white suckers from Lake Michigan tributaries: Pathology and prevalence. *Journal of Fish Diseases*, 40, 377–393.
- Bossart, G. D., Romano, T. A., Peden-Adams, M. M., Rice, C. D., Fair, P. A., Goldstein, J. D., ... Reif, J. S. (2008). Hematological, biochemical, and immunological findings in Atlantic Bottlenose Dolphins (*Tursiops truncatus*) with orogenital papillomas. *Aquatic Mammals*, 34, 166–177. <https://doi.org/10.1578/AM.34.2.2008.166>
- Bowen, L., Aldridge, B. M., DeLong, R., Melin, S., Buckles, E. L., Gulland, F., ... Johnson, M. L. (2005). An immunogenetic basis for the high prevalence of urogenital cancer in a free-ranging population of California sea lions (*Zalophus californianus*). *Immunogenetics*, 56, 846–848. <https://doi.org/10.1007/s00251-004-0757-z>
- Brosi, B. J., Delaplane, K. S., Boots, M., & de Roode, J. C. (2017). Ecological and evolutionary approaches to managing honeybee disease. *Nature Ecology & Evolution*, 1, 1250–1262. <https://doi.org/10.1038/s41559-017-0246-z>
- Brousseau, D. J. (1987). Seasonal aspects of sarcomatous neoplasia in *Mya arenaria* (soft-shell clam) from Long Island Sound. *Journal of Invertebrate Pathology*, 50, 269–276.
- Browning, H. M., Gulland, F. M. D., Hammond, J. A., Colegrove, K. M., & Hall, A. J. (2015). Common cancer in a wild animal: The California sea lion (*Zalophus californianus*) as an emerging model for carcinogenesis. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 370, 20140228. <https://doi.org/10.1098/rstb.2014.0228>
- Brüniche-Olsen, A., Jones, M. E., Austin, J. J., Burridge, C. P., & Holland, B. R. (2014). Extensive population decline in the Tasmanian devil predates European settlement and devil facial tumor disease. *Biology Letters*, 10, 20140619.
- Caldwell, A., Coleby, R., Tovar, C., Stammnitz, M. R., Kwon, Y. M., Owen, R. S., ... Siddle, H. V. T. (2018). The newly-arisen Devil facial tumour disease 2 (DFT2) reveals a mechanism for the emergence of a contagious cancer. *eLife*, 7, e35314. <https://doi.org/10.7554/eLife.35314>
- Carlson, S. M., Cunningham, C. J., & Westley, P. A. H. (2014). Evolutionary rescue in a changing world. *Trends in Ecology & Evolution*, 29, 521–530. <https://doi.org/10.1016/j.tree.2014.06.005>
- Castro-Prieto, A., Wachter, B., & Sommer, S. (2010). Cheetah paradigm revisited: MHC diversity in the world's largest free-ranging population. *Molecular Biology and Evolution*, 28, 1455–1468. <https://doi.org/10.1093/molbev/msq330>
- Cheng, Y., Makara, M., Peel, E., Fox, S., Papenfuss, A. T., & Belov, K. (2019). Tasmanian devils with contagious cancer exhibit a constricted T-cell repertoire diversity. *Communications Biology*, 2, 99. <https://doi.org/10.1038/s42003-019-0342-5>
- Cheng, Y., Stuart, A., Morris, K., Taylor, R., Siddle, H., Deakin, J., ... Belov, K. (2012). Antigen-presenting genes and genomic copy number

- variations in the Tasmanian devil MHC. *BMC Genomics*, 13, 1–14. <https://doi.org/10.1186/1471-2164-13-87>
- Coen, L. D., & Bishop, M. J. (2015). The ecology, evolution, impacts and management of host-parasite interactions of marine molluscs. *Journal of Invertebrate Pathology*, 131, 177–211. <https://doi.org/10.1016/j.jip.2015.08.005>
- Coffee, L., Casey, J., & Bowser, P. (2013). Pathology of tumors in fish associated with retroviruses: A review. *Veterinary Pathology*, 50, 390–403.
- Cohen, D. (1972). Detection of humoral antibody to the transmissible venereal tumour of the dog. *International Journal of Cancer*, 10, 207–212. <https://doi.org/10.1002/ijc.2910100126>
- Cooper, M. D., & Alder, M. N. (2006). The evolution of adaptive immune systems. *Cell*, 124, 815–822. <https://doi.org/10.1016/j.cell.2006.02.001>
- Cortés-Hinojosa, G., Subramaniam, K., Wellehan, J. F. X., Ng, T. F. F., Delwart, E., McCulloch, S. D., ... Waltzek, T. B. (2019). Genomic sequencing of a virus representing a novel type within the species Dyopipapillomavirus 1 in an Indian River Lagoon bottlenose dolphin. *Archives of Virology*, 164, 767–774. <https://doi.org/10.1007/s00705-018-04117-5>
- Cray, C., Varella, R., Bossart, G. D., & Lutz, P. (2001). Altered in vitro immune responses in Green Turtles (*Chelonia mydas*) with fibropapillomatosis. *Journal of Zoo and Wildlife Medicine*, 32, 436–440.
- Crooks, K. R., Scott, C. A., & Van Vuren, D. H. (2001). Exotic disease and an insular endemic carnivore, the island fox. *Biological Conservation*, 98, 55–60.
- Cruz, F. N. D. Jr, Giannitti, F., Li, L., Woods, L. W., Del Valle, L., Delwart, E., & Pesavento, P. A. (2013). Novel polyomavirus associated with brain tumors in free-ranging raccoons, western United States. *Emerging Infectious Diseases*, 19, 77.
- Cunningham, C. X., Johnson, C. N., Barmuta, L. A., Hollings, T., Woehler, E., & Jones, M. E. (2018). Top carnivore decline has cascading effects on scavengers and carrion persistence. *Proceedings of the Royal Society B: Biological Sciences*, 285, 20181582. <https://doi.org/10.1098/rspb.2018.1582>
- Cunningham, C. X., Scoleri, V., Johnson, C. N., Barmuta, L. A., & Jones, M. E. (2019). Temporal partitioning of activity: Rising and falling top-predator abundance triggers community-wide shifts in diel activity. *Ecography*, 42, 1–12. <https://doi.org/10.1111/ecog.04485>
- Daoust, P.-Y., Wobeser, G., Rainnie, D. J., & Leighton, F. A. (1991). Multicentric intramuscular lipomatosis/fibromatosis in free-flying white-fronted and Canada geese. *Journal of Wildlife Diseases*, 27, 135–139.
- Das, U., & Das, A. K. (2000). Review of canine transmissible venereal sarcoma. *Veterinary Research Communications*, 24, 545–556. <https://doi.org/10.1023/a:1006491918910>
- Dawson, E. H., Bailly, T. P. M., Dos Santos, J., Moreno, C., Devilliers, M., Maroni, B., ... Mery, F. (2018). Social environment mediates cancer progression in *Drosophila*. *Nature Communications*, 9, 7. <https://doi.org/10.1038/s41467-018-05737-w>
- Deming, A. C., Colegrove, K. M., Duignan, P. J., Hall, A. J., Wellehan, J. F., & Gulland, F. M. (2018). Prevalence of urogenital carcinoma in stranded California sea lions (*Zalophus californianus*) from 2005–15. *Journal of Wildlife Diseases*, 54, 581–586.
- DiRenzo, G. V., Zipkin, E. F., Grant, E. H. C., Royle, J. A., Longo, A. V., Zamudio, K. R., & Lips, K. R. (2018). Eco-evolutionary rescue promotes host-pathogen coexistence. *Ecological Applications*, 28, 1948–1962. <https://doi.org/10.1002/eap.1792>
- Ebert, D., & Bull, J. J. (2003). Challenging the trade-off model for the evolution of virulence: Is virulence management feasible? *Trends in Microbiology*, 11, 15–20. [https://doi.org/10.1016/s0966-842x\(02\)00003-3](https://doi.org/10.1016/s0966-842x(02)00003-3)
- Enriquez-Navas, P. M., Wojtkowiak, J. W., & Gatenby, R. A. (2015). Application of evolutionary principles to cancer therapy. *Cancer Research*, 75, 4675–4680. <https://doi.org/10.1158/0008-5472.Can-15-1337>
- Epstein, B., Jones, M., Hamede, R., Hendricks, S., McCallum, H., Murchison, E. P., ... Storfer, A. (2016). Rapid evolutionary response to a transmissible cancer in Tasmanian devils. *Nature Communications*, 7, 12684.
- Ewald, P. W., & Swain Ewald, H. A. (2015). Infection and cancer in multicellular organisms. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 370, 20140224. <https://doi.org/10.1098/rstb.2014.0224>
- Galvani, A. P. (2003). Epidemiology meets evolutionary ecology. *Trends in Ecology & Evolution*, 18, 132–139. [https://doi.org/10.1016/s0169-5347\(02\)00050-2](https://doi.org/10.1016/s0169-5347(02)00050-2)
- Gestal, C., Roch, P., Renault, T., Pallavicini, A., Paillard, C., Novoa, B., ... Figueras, A. (2008). Study of diseases and the immune system of bivalves using molecular biology and genomics. *Reviews in Fisheries Science*, 16, 133–156. <https://doi.org/10.1080/10641260802325518>
- Giraudeau, M., Sepp, T., Ujvari, B., Ewald, P. W., & Thomas, F. (2018). Human activities might influence oncogenic processes in wild animal populations. *Nature Ecology & Evolution*, 2, 1065–1070. <https://doi.org/10.1038/s41559-018-0558-7>
- Giraudeau, M., Watson, H., Powell, D., Vincze, O., Thomas, F., Sepp, T., ... Isaksson, C. (2020). Will urbanisation affect the expression level of genes related to cancer of wild great tits? *Science of the Total Environment*, 714, 135793. <https://doi.org/10.1016/j.scitotenv.2019.135793>
- Gourlan, A. T., Douay, G., & Telouk, P. (2019). Copper isotopes as possible neoplasia biomarkers in captive wild felids. *Zoo Biology*, 38, 371–383. <https://doi.org/10.1002/zoo.21504>
- Granoff, A. (1973). Herpesvirus and the Lucke tumor. *Cancer Research*, 33, 1431–1433.
- Grenfell, B. T., Pybus, O. G., Gog, J. R., Wood, J. L., Daly, J. M., Mumford, J. A., & Holmes, E. C. (2004). Unifying the epidemiological and evolutionary dynamics of pathogens. *Science*, 303, 327–332. <https://doi.org/10.1126/science.1090727>
- Greven, H., & Guex, G.-D. (2018). Histology and fine structure of epidermal papillomas in the Alpine newt *Ichthyosaura alpestris* (Urodela: Salamandridae). *Vertebrate Zoology*, 68, 5–19.
- Guimarães, S. M., Gitirana, H. M., Wanderley, A. V., Monteiro-Neto, C., & Lobo-Hajdu, G. (2013). Evidence of regression of fibropapillomas in juvenile green turtles *Chelonia mydas* caught in Niterói, south-east Brazil. *Diseases of Aquatic Organisms*, 102, 243–247. <https://doi.org/10.3354/dao02542>
- Gulland, F. M. D., Trupkiewicz, J. G., Spraker, T. R., & Lowenstine, L. J. (1996). Metastatic carcinoma of probable transitional cell origin in 66 free-living California sea lions (*Zalophus californianus*), 1979 to 1994. *Journal of Wildlife Diseases*, 32, 250–258. <https://doi.org/10.7589/0090-3558-32.2.250>
- Hamede, R. K., Beeton, N. J., Carver, S., & Jones, M. E. (2017). Untangling the model muddle: Empirical tumour growth in Tasmanian devil facial tumour disease. *Scientific Reports*, 7, 7. <https://doi.org/10.1038/s41598-017-06166-3>
- Hamede, R. K., McCallum, H., & Jones, M. E. (2013). Biting injuries and transmission of Tasmanian devil facial tumour disease. *Journal of Animal Ecology*, 82, 182–190. <https://doi.org/10.1111/j.1365-2656.2012.02025.x>
- Hamede, R. K., McCallum, H., & Jones, M. E. (2019). Learning to live with cancer – insights from local adaptations in Tasmanian devils and transmissible tumours in West Pencil Pine. In C. Hogg, S. Fox, D. Pemberton, & K. Belov (Eds.), *Saving the Tasmanian devil: Recovery using science-based management* (pp. 93–99). Canberra, Australia: CSIRO.
- Hamede, R. K., Pearse, A. M., Swift, K., Barmuta, L. A., Murchison, E. P., & Jones, M. E. (2015). Transmissible cancer in Tasmanian

- devils: Localized lineage replacement and host population response. *Proceedings of the Royal Society B: Biological Sciences*, 282, 122–128. <https://doi.org/10.1098/rspb.2015.1468>
- Hato, T., & Dagher, P. C. (2015). How the innate immune system senses trouble and causes trouble. *Clinical Journal of the American Society of Nephrology*, 10, 1459–1469. <https://doi.org/10.2215/CJN.04680514>
- Hawkins, C. E., Baars, C., Hesterman, H., Hocking, G. J., Jones, M. E., Lazenby, B., ... Wiersma, J. (2006). Emerging disease and population decline of an island endemic, the Tasmanian devil *Sarcophilus harrisii*. *Biological Conservation*, 131, 307–324.
- Hawkins, C. E., McCallum, H., Mooney, N., Jones, M. E., & Holdsworth, M. (2009). *Sarcophilus harrisii*. IUCN Red List of threatened species. Version 2009.1.
- Hofman, C. A., Rick, T. C., Hawkins, M. T. R., Funk, W. C., Ralls, K., Boser, C. L., ... Maldonado, J. E. (2015). Mitochondrial genomes suggest rapid evolution of dwarf California Channel Islands Foxes (*Urocyon littoralis*). *PLoS ONE*, 10, e0118240.
- Hohenlohe, P. A., McCallum, H. I., Jones, M. E., Lawrance, M. F., Hamede, R. K., & Storfer, A. (2019). Conserving adaptive potential: Lessons from Tasmanian devils and their transmissible cancer. *Conservation Genetics*, 20, 81–87. <https://doi.org/10.1007/s10592-019-01157-5>
- Hollings, T., Jones, M. E., Mooney, N., & McCallum, H. (2013). Wildlife disease ecology in changing landscapes: Mesopredator release and toxoplasmosis. *International Journal for Parasitology*, 2, 110–118. <https://doi.org/10.1016/j.ijppaw.2013.02.002>
- Hollings, T., Jones, M. E., Mooney, N., & McCallum, H. (2014). Trophic cascades following the disease-induced decline of an apex predator, the Tasmanian devil. *Conservation Biology*, 28, 36–75. <https://doi.org/10.1111/cobi.12152>
- Hollings, T., Jones, M. E., Mooney, N., & McCallum, H. (2016). Disease-induced decline of an apex predator drives invasive dominated states and threatens biodiversity. *Ecology*, 97, 394–405. <https://doi.org/10.1890/15-0204.1>
- Hsiao, Y. W., Liao, K. W., Chung, T. F., Liu, C. H., Hsu, C. D., & Chu, R. M. (2008). Interactions of host IL-6 and IFN-gamma and cancer-derived TGF-beta1 on MHC molecule expression during tumor spontaneous regression. *Cancer Immunology and Immunotherapy*, 57, 1091–1104. <https://doi.org/10.1007/s00262-007-0446-5>
- Jacqueline, C., Biro, P. A., Beckmann, C., Moller, A. P., Renaud, F., Sorci, G., ... Thomas, F. (2017). Cancer: A disease at the crossroads of trade-offs. *Evolutionary Applications*, 10, 215–225. <https://doi.org/10.1111/eva.12444>
- James, S., Jennings, G., Kwon, Y. M., Stammnitz, M., Fraik, A., Storfer, A., ... Hamede, R. (2019). Tracing the rise of malignant cell lines: Distribution, epidemiology and evolutionary interactions of two transmissible cancers in Tasmanian devils. *Evolutionary Applications*, <https://doi.org/10.1111/eva.12831>
- Jones, K., Ariel, E., Burgess, G., & Read, M. (2016). A review of fibropapillomatosis in green turtles (*Chelonia mydas*). *The Veterinary Journal*, 212, 48–57.
- Jones, M. E., Cockburn, A., Hamede, R., Hawkins, C., Hesterman, H., Lachish, S., ... Pemberton, D. (2008). Life-history change in disease-ravaged Tasmanian devil populations. *Proceedings of the National Academy of Sciences of the United States of America*, 105, 10023–10027. <https://doi.org/10.1073/pnas.0711236105>
- Jones, M. E., Paetkau, D., Geffen, E., & Moritz, C. (2004). Genetic diversity and population structure of Tasmanian devils, the largest marsupial carnivore. *Molecular Ecology*, 13, 2197–2209. <https://doi.org/10.1111/j.1365-294X.2004.02239.x>
- King, D. P., Hure, M. C., Goldstein, T., Aldridge, B. M., Gulland, F. M. D., Saliki, J. T., ... Stott, J. L. (2002). Otarine herpesvirus-1: A novel gammaherpesvirus associated with urogenital carcinoma in California sea lions (*Zalophus californianus*). *Veterinary Microbiology*, 86, 131–137. [https://doi.org/10.1016/S0378-1135\(01\)00497-7](https://doi.org/10.1016/S0378-1135(01)00497-7)
- Kourou, K., Exarchos, T. P., Exarchos, K. P., Karamouzis, M. V., & Fotiadis, D. I. (2015). Machine learning applications in cancer prognosis and prediction. *Computational and Structural Biotechnology Journal*, 13, 8–17. <https://doi.org/10.1016/j.csbj.2014.11.005>
- Kwon, Y. M., Stammnitz, M. R., Wang, J., Swift, K., Knowles, G. W., Pye, R. J., ... Murchison, E. P. (2018). Tasman-PCR: A genetic diagnostic assay for Tasmanian devil facial tumour diseases. *Royal Society Open Science*, 5, 10. <https://doi.org/10.1098/rsos.180870>
- Lachish, S., Jones, M. E., & McCallum, H. (2007). The impact of disease on the survival and population growth rate of the Tasmanian devil. *Journal of Animal Ecology*, 76, 926–936.
- Lachish, S., McCallum, H., & Jones, M. (2009). Demography, disease and the devil: Life-history changes in a disease-affected population of Tasmanian devils (*Sarcophilus harrisii*). *Journal of Animal Ecology*, 78, 427–436. <https://doi.org/10.1111/j.1365-2656.2008.01494.x>
- Lambertsen, R. H., Kohn, B. A., Sundberg, J. P., & Buergelt, C. D. (1987). Genital papillomatosis in sperm whale bulls. *Journal of Wildlife Diseases*, 23, 361–367.
- Lazenby, B. T., Tobler, M. W., Brown, W. E., Hawkins, C. E., Hocking, G. J., Hume, F., ... Pemberton, D. (2018). Density trends and demographic signals uncover the long-term impact of transmissible cancer in Tasmanian devils. *Journal of Applied Ecology*, 55, 1368–1379. <https://doi.org/10.1111/1365-2664.13088>
- Literak, L., Robesova, B., Majlathova, V., Majlath, I., Kulich, P., Fabian, P., & Roubalova, E. (2010). Herpesvirus-associated papillomatosis in a green lizard. *Journal of Wildlife Diseases*, 46, 257–261.
- Loh, R., Bergfeld, J., Hayes, D., O'Hara, A., Pyecroft, S., Raidal, S., & Sharpe, R. (2006). The pathology of devil facial tumor disease (DFTD) in Tasmanian devils (*Sarcophilus harrisii*). *Veterinary Pathology*, 43, 890–895.
- Maibach, V., & Vigilant, L. (2019). Reduced bonobo MHC class I diversity predicts a reduced viral peptide binding ability compared to chimpanzees. *BMC Evolutionary Biology*, 19, 14. <https://doi.org/10.1186/s12862-019-1352-0>
- Malins, D. C., McCain, B. B., Landahl, J. T., Myers, M. S., Krahn, M. M., Brown, D. W., ... Roubal, W. T. (1988). Neoplastic and other diseases in fish in relation to toxic chemicals: An overview. *Aquatic Toxicology*, 11, 43–67.
- Margres, M. J., Ruiz-Aravena, M., Hamede, R., Jones, M. E., Lawrance, M. F., Hendricks, S. A., ... Storfer, A. (2018). The genomic basis of tumor regression in Tasmanian devils (*Sarcophilus harrisii*). *Genome Biology and Evolution*, 10, 3012–3025. <https://doi.org/10.1093/gbe/evy229>
- Martineau, D., Lemberger, K., Dallaire, A., Labelle, P., Lipscomb, T. P., Michel, P., & Mikaelian, I. (2002). Cancer in wildlife, a case study: Beluga from the St. Lawrence estuary, Québec, Canada. *Environmental Health Perspectives*, 110, 285–292.
- Martinez-Silvestre, A., Amat, F., Bargalló, F., & Carranza, S. (2011). Incidence of pigmented skin tumors in a population of wild Montseny brook newt (*Calotriton arnoldi*). *Journal of Wildlife Diseases*, 47, 410–414.
- McAloose, D., & Newton, A. L. (2009). Wildlife cancer: A conservation perspective. *Nature Reviews Cancer*, 9, 517–526. <https://doi.org/10.1038/nrc2665>
- McCallum, H., Jones, M., Hawkins, C., Hamede, R., Lachish, S., Sinn, D. L., ... Lazenby, B. (2009). Transmission dynamics of Tasmanian devil facial tumor disease may lead to disease-induced extinction. *Ecology*, 90, 3379–3392. <https://doi.org/10.1890/08-1763.1>
- McCallum, H., Tompkins, D. M., Jones, M., Lachish, S., Marvanek, S., Lazenby, B., ... Hawkins, C. E. (2007). Distribution and impacts of Tasmanian devil facial tumor disease. *EcoHealth*, 4, 318–325. <https://doi.org/10.1007/s10393-007-0118-0>
- McGranahan, N., Rosenthal, R., Hiley, C. T., Rowan, A. J., Watkins, T. B. K., Wilson, G. A., ... Dessimoz, C. (2017). Allele-specific HLA loss and immune escape in lung cancer evolution. *Cell*, 171, 1259–1271. <https://doi.org/10.1016/j.cell.2017.10.001>

- Metzger, M. J., & Goff, S. P. (2016). A sixth modality of infectious disease: Contagious cancer from devils to clams and beyond. *PLoS Path.* 12, e1005904. <https://doi.org/10.1371/journal.ppat.1005904>
- Metzger, M. J., Villalba, A., Carballal, M. J., Iglesias, D., Sherry, J., Reinisch, C., ... Goff, S. P. (2016). Widespread transmission of independent cancer lineages within multiple bivalve species. *Nature*, 534, 705–709. <https://doi.org/10.1038/nature18599>
- Mix, M. (1983). Haemic neoplasms of bay mussels, *Mytilus edulis* L., from Oregon: Occurrence, prevalence, seasonality and histopathological progression. *Journal of Fish Diseases*, 6, 239–248.
- Møller, A., Bonisoli-Alquati, A., & Mousseau, T. (2013). High frequency of albinism and tumours in free-living birds around Chernobyl. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 757, 52–59.
- Moriarty, M. E., Vickers, T. W., Clifford, D. L., Garcelon, D. K., Gaffney, P. M., Lee, K. W., ... Boyce, W. M. (2015). Ear mite removal in the Santa Catalina Island fox (*Urocyon littoralis catalinae*): Controlling risk factors for cancer development. *PLoS ONE*, 10, e0144271. <https://doi.org/10.1371/journal.pone.0144271>
- Morris, K., Austin, J. J., & Belov, K. (2013). Low major histocompatibility complex diversity in the Tasmanian devil predates European settlement and may explain susceptibility to disease epidemics. *Biology Letters*, 9, 20120900. <https://doi.org/10.1098/rsbl.2012.0900>
- Murchison, E. P., Schulz-Trieglaff, O. B., Ning, Z., Alexandrov, L. B., Bauer, M. J., Fu, B., ... Stratton, M. R. (2012). Genome sequencing and analysis of the Tasmanian Devil and its transmissible cancer. *Cell*, 148, 780–791. <https://doi.org/10.1016/j.cell.2011.11.065>
- Murchison, E. P., Wedge, D. C., Alexandrov, L. B., Fu, B., Martincorena, I., Ning, Z., ... Stratton, M. R. (2014). Transmissible dog cancer genome reveals the origin and history of an ancient cell lineage. *Science*, 343, 437–440. <https://doi.org/10.1126/science.1247167>
- Murgia, C., Pritchard, J. K., Kim, S. Y., Fassati, A., & Weiss, R. A. (2006). Clonal origin and evolution of a transmissible cancer. *Cell*, 126, 477–487. <https://doi.org/10.1016/j.cell.2006.05.051>
- Ní Leathlobhair, M., Perri, A. R., Irving-Pease, E. K., Witt, K. E., Linderholm, A., Haile, J., ... Frantz, L. A. F. (2018). The evolutionary history of dogs in the Americas. *Science*, 361, 81–85. <https://doi.org/10.1126/science.aao4776>
- Nunney, L. (2013). The real war on cancer: The evolutionary dynamics of cancer suppression. *Evolutionary Applications*, 6, 11–19. <https://doi.org/10.1111/eva.12018>
- Ong, C. E. B., Lyons, A. B., Woods, G. M., & Flies, A. S. (2019). Inducible IFN-gamma expression for MHC-I upregulation in devil facial tumor cells. *Frontiers in Immunology*, 9, 9. <https://doi.org/10.3389/fimmu.2018.03117>
- Ostrander, E. A., Davis, B. W., & Ostrander, G. K. (2016). Transmissible tumors: Breaking the cancer paradigm. *Trends in Genetics*, 32, 1–15. <https://doi.org/10.1016/j.tig.2015.10.001>
- Page, A., Norton, T. M., Harms, C., Mader, D., Herbst, L. H., Stedman, N., & Gottdenker, N. L. (2015). Case descriptions of fibropapillomatosis in rehabilitating loggerhead sea turtles *Caretta caretta* in the southeastern USA. *Diseases of Aquatic Organisms*, 115, 185–191. <https://doi.org/10.3354/dao02878>
- Papas, T. S., Dahlberg, J. E., & Sonstegard, R. A. (1976). Type C virus in lymphosarcoma in northern pike (*Esox lucius*). *Nature*, 261, 506.
- Pearse, A. M., & Swift, K. (2006). Allograft theory: Transmission of devil facial-tumour disease. *Nature*, 439, 549.
- Pearse, A.-M., Swift, K., Hodson, P., Hua, B., McCallum, H., Pyecroft, S., ... Belov, K. (2012). Evolution in a transmissible cancer: A study of the chromosomal changes in devil facial tumor (DFT) as it spreads through the wild Tasmanian devil population. *Cancer Genetics*, 205, 101–112. <https://doi.org/10.1016/j.cancergen.2011.12.001>
- Pesavento, P. A., Agnew, D., Keel, M. K., & Woolard, K. D. (2018). Cancer in wildlife: Patterns of emergence. *Nature Reviews Cancer*, 18, 646–661. <https://doi.org/10.1038/s41568-018-0045-0>
- Poder, M., & Auffret, M. (1986). Sarcomatous lesion in the cockle *Cerastoderma edule*: I. Morphology and population survey in Brittany, France. *Aquaculture*, 58, 1–8.
- Poli, C., Lopez, L. C. S., Mesquita, D. O., Saska, C., & Mascarenhas, R. (2014). Patterns and inferred processes associated with sea turtle strandings in Paraíba State, Northeast Brazil. *Brazilian Journal of Biology*, 74, 283–289. <https://doi.org/10.1590/1519-6984.13112>
- Pye, R., Hamede, R., Siddle, H. V., Caldwell, A., Knowles, G. W., Swift, K., ... Woods, G. M. (2016). Demonstration of immune responses against devil facial tumour disease in wild Tasmanian devils. *Biology Letters*, 12, 0553.
- Pye, R. J., Pemberton, D., Tovar, C., Tubio, J. M. C., Dun, K. A., Fox, S., ... Woods, G. M. (2016). A second transmissible cancer in Tasmanian devils. *Proceedings of the National Academy of Sciences of the United States of America*, 113, 374–379. <https://doi.org/10.1073/pnas.1519691113>
- Randhawa, N., Gulland, F., Ylitalo, G. M., DeLong, R., & Mazet, J. A. K. (2015). Sentinel California sea lions provide insight into legacy organochlorine exposure trends and their association with cancer and infectious disease. *One Health*, 1, 37–43. <https://doi.org/10.1016/j.onehit.2015.08.003>
- Rehtanz, M., Ghim, S.-J., McFee, W., Doescher, B., Lacave, G., Fair, P. A., ... Jenson, A. B. (2010). Papillomavirus antibody prevalence in free-ranging and captive Bottlenose Dolphins (*Tursiops truncatus*). *Journal of Wildlife Diseases*, 46, 136–145. <https://doi.org/10.7589/0090-3558-46.1.136>
- Roemer, G. W., Coonan, T. J., Garcelon, D. K., Bascompte, J., & Laughrin, L. (2001). Feral pigs facilitate hyperpredation by golden eagles and indirectly cause the decline of the island fox. *Animal Conservation*, 4, 307–318.
- Rose, F. L., & Harshbarger, J. C. (1977). Neoplastic and possibly related skin lesions in neotenic tiger salamanders from a sewage lagoon. *Science*, 315–317.
- Ruiz-Aravena, M., Jones, M. E., Carver, S., Estay, S., Espejo, C., Storfer, A., & Hamede, R. K. (2018). Sex bias in ability to cope with cancer: Tasmanian devils and facial tumour disease. *Proceedings of the Royal Society B: Biological Sciences*, 285, 20182239. <https://doi.org/10.1098/rspb.2018.2239>
- Russell, T., Madsen, T., Thomas, F., Raven, N., Hamede, R., & Ujvari, B. (2018). Oncogenesis as a selective force: Adaptive evolution in the face of a transmissible cancer. *BioEssays*, 40, 9. <https://doi.org/10.1002/bies.201700146>
- Savage, A. E., & Zamudio, K. R. (2011). MHC genotypes associate with resistance to a frog-killing fungus. *Proceedings of the National Academy of Sciences of the United States of America*, 108, 16705–16710. <https://doi.org/10.1073/pnas.1106893108>
- Sepp, T., Ujvari, B., Ewald, P. W., Thomas, F., & Giraudeau, M. (2019). Urban environment and cancer in wildlife: Available evidence and future research avenues. *Proceedings of the Royal Society B: Biological Sciences*, 286, 8. <https://doi.org/10.1098/rspb.2018.2434>
- Siddle, H. V., Kreiss, A., Eldridge, M. D. B., Noonan, E., Clarke, C. J., Pyecroft, S., ... Belov, K. (2007). Transmission of a fatal clonal tumor by biting occurs due to depleted MHC diversity in a threatened carnivorous marsupial. *Proceedings of the National Academy of Sciences of the United States of America*, 104, 16221–16226. <https://doi.org/10.1073/pnas.0704580104>
- Siddle, H. V., Kreiss, A., Tovar, C., Yuen, C. K., Cheng, Y., Belov, K., ... Kaufman, J. (2013). Reversible epigenetic down-regulation of MHC molecules by devil facial tumour disease illustrates immune escape by a contagious cancer. *Proceedings of the National Academy of Sciences of the United States of America*, 110, 5103–5108. <https://doi.org/10.1073/pnas.1219920110>
- Siddle, H. V., Marzec, J., Cheng, Y., Jones, M. E., & Belov, K. (2010). MHC gene copy number variation in Tasmanian devils: Implications for the spread of a contagious cancer. *Proceedings of the Royal Society*

- B: *Biological Sciences*, 277, 2001–2006. <https://doi.org/10.1098/rspb.2009.2362>
- Siddle, H. V., Sanderson, C., & Belov, K. (2007). Characterization of a major histocompatibility complex class I and class II genes from the Tasmanian devil (*Sarcophilus harrisi*). *Immunogenetics*, 59, 753–760. <https://doi.org/10.1007/s00251-007-0238-2>
- Sommer, S. (2005). The importance of immune gene variability (MHC) in evolutionary ecology and conservation. *Frontiers in Zoology*, 2, 16. <https://doi.org/10.1186/1742-9994-2-16>
- Stammnitz, M. R., Coorens, T. H. H., Gori, K. C., Hayes, D., Fu, B., Wang, J., ... Murchison, E. P. (2018). The origins and vulnerabilities of two transmissible cancers in Tasmanian Devils. *Cancer Cell*, 33, 607–619. <https://doi.org/10.1016/j.ccell.2018.03.013>
- Storfer, A., Hohenlohe, P. A., Margres, M. J., Patton, A., Fraik, A. K., Lawrance, M., ... Jones, M. E. (2018). The devil is in the details: Genomics of transmissible cancers in Tasmanian devils. *PLoS Path*, 14, 7. <https://doi.org/10.1371/journal.ppat.1007098>
- Strakova, A., & Murchison, E. P. (2015). The cancer which survived: Insights from the genome of an 11 000 year-old cancer. *Current Opinion in Genetics & Development*, 30, 49–55. <https://doi.org/10.1016/j.gde.2015.03.005>
- Tagliolatto, A. B., Guimarães, S. M., Lobo-Hajdu, G., & Monteiro-Neto, C. (2016). Characterization of fibropapillomatosis in green turtles *Chelonia mydas* (Cheloniidae) captured in a foraging area in south-eastern Brazil. *Diseases of Aquatic Organisms*, 121, 233–240. <https://doi.org/10.3354/dao03020>
- Thomas, F., Giraudeau, M., Gouzerh, F., Boutry, J., Renaud, F., Pujol, P., ... Ujvari, B. (2019). The evolution of resistance and tolerance as cancer defences. *Parasitology International*. <https://doi.org/10.1017/S0031182019001501>
- Thomas, F., Jacqueline, C., Tissot, T., Henard, M., Blanchet, S., Loot, G., ... Ujvari, B. (2017). The importance of cancer cells for animal evolutionary ecology. *Nature Ecology & Evolution*, 1, 1592–1595. <https://doi.org/10.1038/s41559-017-0343-z>
- Thomas, F., Vavre, F., Tissot, T., Vittecoq, M., Giraudeau, M., Bernex, F., ... Ujvari, B. (2018). Cancer is not (only) a senescence problem. *Trends in Cancer*, 4, 169–172. <https://doi.org/10.1016/j.trecan.2018.01.002>
- Tompkins, D. M., Dunn, A. M., Smith, M. J., & Telfer, S. (2011). Wildlife diseases: From individuals to ecosystems. *Journal of Animal Ecology*, 80, 19–38. <https://doi.org/10.1111/j.1365-2656.2010.01742.x>
- Tovar, C., Pye, R. J., Kreiss, A., Cheng, Y., Brown, G. K., Darby, J., ... Woods, G. M. (2017). Regression of devil facial tumour disease following immunotherapy in immunised Tasmanian devils. *Scientific Reports*, 7, 43827.
- Ujvari, B., Beckmann, C., Biro, P. A., Arnal, A., Tasiemski, A., Massol, F., ... Thomas, F. (2016). Cancer and life-history traits: Lessons from host-parasite interactions. *Parasitology*, 143, 533–541. <https://doi.org/10.1017/s0031182016000147>
- Ujvari, B., Gatenby, R. A., & Thomas, F. (2016a). The evolutionary ecology of transmissible cancers. *Infection Genetics and Evolution*, 39, 293–303. <https://doi.org/10.1016/j.meegid.2016.02.005>
- Ujvari, B., Gatenby, R. A., & Thomas, F. (2016b). Transmissible cancers, are they more common than thought? *Evolutionary Applications*, 9, 633–634. <https://doi.org/10.1111/eva.12372>
- Ujvari, B., Klaassen, M., Raven, N., Russell, T., Vittecoq, M., Hamedé, R., ... Madsen, T. (2018). Genetic diversity, inbreeding and cancer. *Proceedings of the Royal Society B-Biological Sciences*, 285, 8. <https://doi.org/10.1098/rspb.2017.2589>
- Van Bresse, M., Van Waerebeek, K., Piérard, G. E., & Desaintes, C. (1996). Genital and lingual warts in small cetaceans from coastal Peru. *Diseases of Aquatic Organisms*, 26, 1–10.
- Vander Wal, E., Garant, D., Calmé, S., Chapman, C. A., Festa-Bianchet, M., Millien, V., ... Pelletier, F. (2014). Applying evolutionary concepts to wildlife disease ecology and management. *Evolutionary Applications*, 7, 856–868. <https://doi.org/10.1111/eva.12168>
- Vickers, T. W., Clifford, D. L., Garcelon, D. K., King, J. L., Duncan, C. L., Gaffney, P. M., & Boyce, W. M. (2015). Pathology and epidemiology of ceruminous gland tumors among endangered Santa Catalina Island foxes (*Urocyon littoralis catalinae*) in the Channel Islands, USA. *PLoS ONE*, 10, e0143211. <https://doi.org/10.1371/journal.pone.0143211>
- Vittecoq, M., Ducasse, H., Arnal, A., Møller, A. P., Ujvari, B., Jacqueline, C. B., ... Thomas, F. (2015). Animal behaviour and cancer. *Animal Behaviour*, 101, 19–26. <https://doi.org/10.1016/j.anbehav.2014.12.001>
- Vittecoq, M., Roche, B., Daoust, S. P., Ducasse, H., Missé, D., Abadie, J., ... Thomas, F. (2013). Cancer: A missing link in ecosystem functioning? *Trends in Ecology & Evolution*, 28, 628–635. <https://doi.org/10.1016/j.tree.2013.07.005>
- Weber, D. S., Stewart, B. S., Schienman, J., & Lehman, N. (2004). Major histocompatibility complex variation at three class II loci in the northern elephant seal. *Molecular Ecology*, 13, 711–718. <https://doi.org/10.1111/j.1365-294X.2004.02095.x>
- Wells, K., Hamedé, R. K., Kerlin, D. H., Storfer, A., Hohenlohe, P. A., Jones, M. E., & McCallum, H. I. (2017). Infection of the fittest: Devil facial tumour disease has greatest effect on individuals with highest reproductive output. *Ecology Letters*, 20, 770–778. <https://doi.org/10.1111/ele.12776>
- Wieczorek, M., Abualrous, E. T., Sticht, J., Álvaro-Benito, M., Stolzenberg, S., Noé, F., & Freund, C. (2017). Major Histocompatibility Complex (MHC) class I and MHC class II proteins: Conformational plasticity in antigen presentation. *Frontiers in Immunology*, 8, 292. <https://doi.org/10.3389/fimmu.2017.00292>
- Williams, E. H., Bunkley-Williams, L., Peters, E. C., Pinto-Rodriguez, B., Matos-Morales, R., Mignucci-Giannoni, A. A., ... Boulon, R. H. (1994). An epizootic of cutaneous fibropapillomas in green turtles *Chelonia mydas* of the Caribbean: Part of a panzootic? *Journal of Aquatic Animal Health*, 6, 70–78. [https://doi.org/10.1577/1548-8667\(1994\)06<0070:AEOCFI>2.3.CO;2](https://doi.org/10.1577/1548-8667(1994)06<0070:AEOCFI>2.3.CO;2)
- Willyard, C. (2016). Tumours are subject to the same rules of natural selection as any other living thing. Clinicians are now putting that knowledge to use. *Nature*, 532, 166–168. <https://doi.org/10.1038/532166a>
- Woolford, L., O'Hara, A. J., Bennett, M. D., Slaven, M., Swan, R., Friend, J. A., ... Warren, K. S. (2008). Cutaneous papillomatosis and carcinomatosis in the Western barred bandicoot (*Perameles bougainville*). *Veterinary Pathology*, 45, 95–103. <https://doi.org/10.1354/vp.45-1-95>
- Wu, C. I., Wang, H. Y., Ling, S. P., & Lu, X. M. (2016). The ecology and evolution of cancer: The ultra-microevolutionary process. In N. M. Bonini (Ed.), *Annual review of genetics* (Vol. 50, pp. 347–369). Palo Alto, CA: Annual Reviews.
- Yang, T. J., Chandler, J. P., & Dunne-Anway, S. (1987). Growth stage dependent expression of MHC antigens on the canine transmissible venereal sarcoma. *British Journal of Cancer*, 55, 131–134.
- Yonemitsu, M. A., Giersch, R. M., Polo-Prieto, M., Hammel, M., Simon, A., Cremonese, F., ... Metzger, M. J. (2019). A single clonal lineage of transmissible cancer identified in two marine mussel species in South America and Europe. *eLife*, 8, 21. <https://doi.org/10.7554/eLife.47788>
- Zhang, J. S., Cunningham, J. J., Brown, J. S., & Gatenby, R. A. (2017). Integrating evolutionary dynamics into treatment of metastatic castrate-resistant prostate cancer. *Nature Communications*, 8, 9. <https://doi.org/10.1038/s41467-017-01968-5>

How to cite this article: Hamedé R, Owen R, Siddle H, et al. The ecology and evolution of wildlife cancers: Applications for management and conservation. *Evol Appl*. 2020;13:1719–1732. <https://doi.org/10.1111/eva.12948>