



HAL
open science

Do malignant cells sleep at night?

Luis Enrique Cortés-Hernández, Zahra Eslami-S, Antoine Dujon, Mathieu Giraudeau, Beata Ujvari, Frédéric Thomas, Catherine Alix-Panabières

► **To cite this version:**

Luis Enrique Cortés-Hernández, Zahra Eslami-S, Antoine Dujon, Mathieu Giraudeau, Beata Ujvari, et al.. Do malignant cells sleep at night?. *Genome Biology*, 2020, 21 (1), 10.1186/s13059-020-02179-w . hal-03025763

HAL Id: hal-03025763

<https://hal.umontpellier.fr/hal-03025763>

Submitted on 26 Nov 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.




Distributed under a Creative Commons Attribution 4.0 International License

REVIEW

Open Access

Do malignant cells sleep at night?



Luis Enrique Cortés-Hernández^{1†}, Zahra Eslami-S^{1†}, Antoine M. Dujon^{2,3}, Mathieu Giraudeau², Beata Ujvari^{3,4}, Frédéric Thomas^{2†} and Catherine Alix-Panabières^{1,2,5*†} 

* Correspondence: c-panabieres@chu-montpellier.fr

[†]Luis Enrique Cortés-Hernández, Zahra Eslami-S, Frédéric Thomas and Catherine Alix-Panabières contributed equally to this work.

¹Laboratory of Rare Human Circulating Cells (LCCRH), University Medical Centre of Montpellier, Montpellier, France

²CREEC (CREES), Unité Mixte de Recherches, IRD 224–CNRS 5290–Université de Montpellier, Montpellier, France

Full list of author information is available at the end of the article

Abstract

Biological rhythms regulate the biology of most, if not all living creatures, from whole organisms to their constitutive cells, their microbiota, and also parasites. Here, we present the hypothesis that internal and external ecological variations induced by biological cycles also influence or are exploited by cancer cells, especially by circulating tumor cells, the key players in the metastatic cascade. We then discuss the possible clinical implications of the effect of biological cycles on cancer progression, and how they could be exploited to improve and standardize methods used in the liquid biopsy field.

Keywords: Circadian cycle, Tumor dissemination, Circulating tumor cells, Chronobiology, Phenology, Disease ecology

Introduction

Since the early 1940s, multicellular organisms are no longer considered autonomous entities, but rather “holobionts,” i.e., assemblages composed of the host and its associated commensal and mutualistic microorganisms and parasitic taxa [1, 2]. Recently, Thomas et al. [3] proposed that multicellular organisms have a long evolutionary history with a third category of living entities inside their bodies: cancer cell communities (oncobiota). From precancerous lesions to metastatic cancers, oncogenic processes are very frequent in humans and animals [4, 5], and not just during aging as often thought in the past [6]. From an evolutionary perspective, the transformation of normal cells into malignant cells that will form a cancer is equivalent to a speciation event inside the body [7, 8], preceded by a set of mutations that allow normal cells to acquire self-defined fitness functions and on which natural selection can act [9].

As the holobiont components are usually prisoners of their host, their environment is composed of at least two very different ecological dimensions: the host (i.e., the immediate environment) and the host habitat (i.e., the ecosystem). The first type of environmental variables that holobiont members experience are the host physiological, genetic, and phenotypic characteristics, such as sex, age, and immunocompetence. The second level of environmental variability is due to the biotic and abiotic factors that characterize the ecosystem in which the host lives (resource level, abundance of predators, seasons). These intra- and inter-individual



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

variables and their interactions contribute to the selective landscape in which holobiont members survive and maximize their lifetime reproductive success (i.e., their fitness). Examples of host-parasite interactions show that several parasite species have adapted to perceive, and then to respond in a state-dependent manner, to various signals linked to immediate and/or external fitness-related environmental parameters [10].

An interesting direction in this research area concerns the influence of the biological rhythms that coordinate organismal activities with the environment circadian rhythms. Importantly, holobiont members are not only influenced by the rhythm of the external biotic and abiotic environments, but also of the host biotic environment. Several studies have demonstrated the relevance of these biological cycles in host-parasite relationships. For example, parasites manifest periodic modulations of traits, such as virulence, development, and transmission, in function of their host behavioral or physiological rhythms, such as periodicity in the immune responses and/or host feeding rhythms [11–14]. Hosts may use biological rhythms to defend themselves against parasites (e.g., circadian clocks in immune cells can modulate the magnitude of *Leishmania* infection) [15], but parasites also may manipulate the host clocks to their own advantage. For instance, baculoviruses called *Spodoptera exigua* multiple nucleopolyhedroviruses (SeMNPV) can capture their host phototaxis pathway, thus forcing the host to climb to elevated positions that are more favorable for parasite transmission [16, 17]. Similar reciprocal interactions also exist between the host and its microbiota. Indeed, the microbiota rhythms and consequently their community structure and metabolic activity are regulated by their host diet and feeding time [18]. On the other hand, intestinal microbiota can, sometimes, program the host diurnal metabolic rhythms, thus influencing diurnal fluctuations of the host physiology and susceptibility to pathologies [19].

The connections between chronobiology, disease ecology, and evolutionary biology can help to understand interactions between holobiont members. Conversely, little is known on whether and how malignant cells are influenced by and/or exploit biological rhythms. Like many parasitic organisms, malignant cells depend on their host for sustenance, proliferation, and dispersal, and thus exploit their host for energy and resources. Therefore, although cancer cells are issued from the self (i.e., are not an exogenous organism), with an evolution that last only few decades at most (thus probably preventing the fine tuning of many adaptive responses, unlike true parasitic species where selection occurs during tens of thousands to millions of years), some adaptations that have evolved in the context of host-parasite interactions could also be relevant to cancer progression and dissemination (circulating tumor cells [CTCs]). Such adaptations might be relevant also for clinical applications, for instance in the *liquid biopsy* field (Fig. 1). Here, we combined mechanistic and evolutionary knowledge to investigate and discuss to which extent biological rhythms, especially the circadian rhythm, should be considered when studying the complexity of host-tumor cell interactions during tumorigenesis. We first provide an overview of the geological, physiological, cellular, and molecular factors implicated in circadian rhythms, then describe the impact of biological cycles on cancer development. Finally, we discuss how these principles could be implemented in novel therapeutic and preventive strategies related to the *liquid biopsy* field.

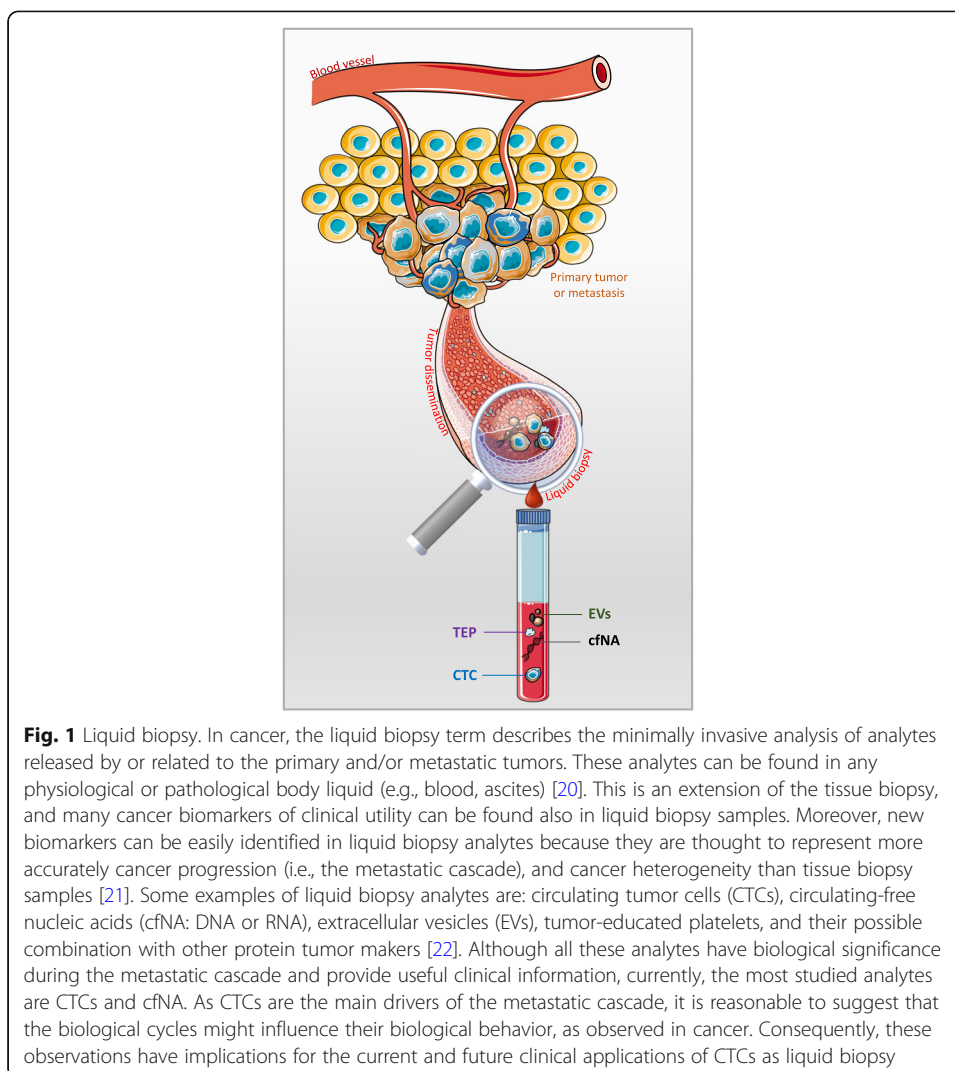


Fig. 1 Liquid biopsy. In cancer, the liquid biopsy term describes the minimally invasive analysis of analytes released by or related to the primary and/or metastatic tumors. These analytes can be found in any physiological or pathological body liquid (e.g., blood, ascites) [20]. This is an extension of the tissue biopsy, and many cancer biomarkers of clinical utility can be found also in liquid biopsy samples. Moreover, new biomarkers can be easily identified in liquid biopsy analytes because they are thought to represent more accurately cancer progression (i.e., the metastatic cascade), and cancer heterogeneity than tissue biopsy samples [21]. Some examples of liquid biopsy analytes are: circulating tumor cells (CTCs), circulating-free nucleic acids (cfNA: DNA or RNA), extracellular vesicles (EVs), tumor-educated platelets, and their possible combination with other protein tumor makers [22]. Although all these analytes have biological significance during the metastatic cascade and provide useful clinical information, currently, the most studied analytes are CTCs and cfNA. As CTCs are the main drivers of the metastatic cascade, it is reasonable to suggest that the biological cycles might influence their biological behavior, as observed in cancer. Consequently, these observations have implications for the current and future clinical applications of CTCs as liquid biopsy

Geological, physiological, cellular, and molecular factors of biological cycles

The movement of the Earth around the Sun and around its own axis are the main factors contributing to geophysical rhythms that result in the biological cycles observed in nature and have been a major evolutionary force. The alteration of any of these cycles may directly or indirectly cause a higher vulnerability to various infectious and/or chronic pathologies, including cancer [23–25]. All biological cycles and rhythms share similar physiological mechanisms. Nevertheless, it is possible to divide them according to the environmental factors they are associated with (e.g., sunlight, temperature, humidity, gravity, exercise, social cues, eating patterns). These cues are called “zeitgebers” or synchronizing cues, and the most predominant are those dictated by geophysical signals, such as the tidal, diel, lunar, and annual rhythms [11].

One of the most studied “zeitgebers” is the diel cycle, which is the fluctuation between day and night. The diel cycle dictates the biology of the circadian rhythm, with a duration of 24 to 26 h. Its main cue is exposure to sunlight that guides the central biological clock in vertebrates. This biological clock works as a network that is called the *vertebrate circadian axis* and includes the suprachiasmatic nucleus (SCN) in the

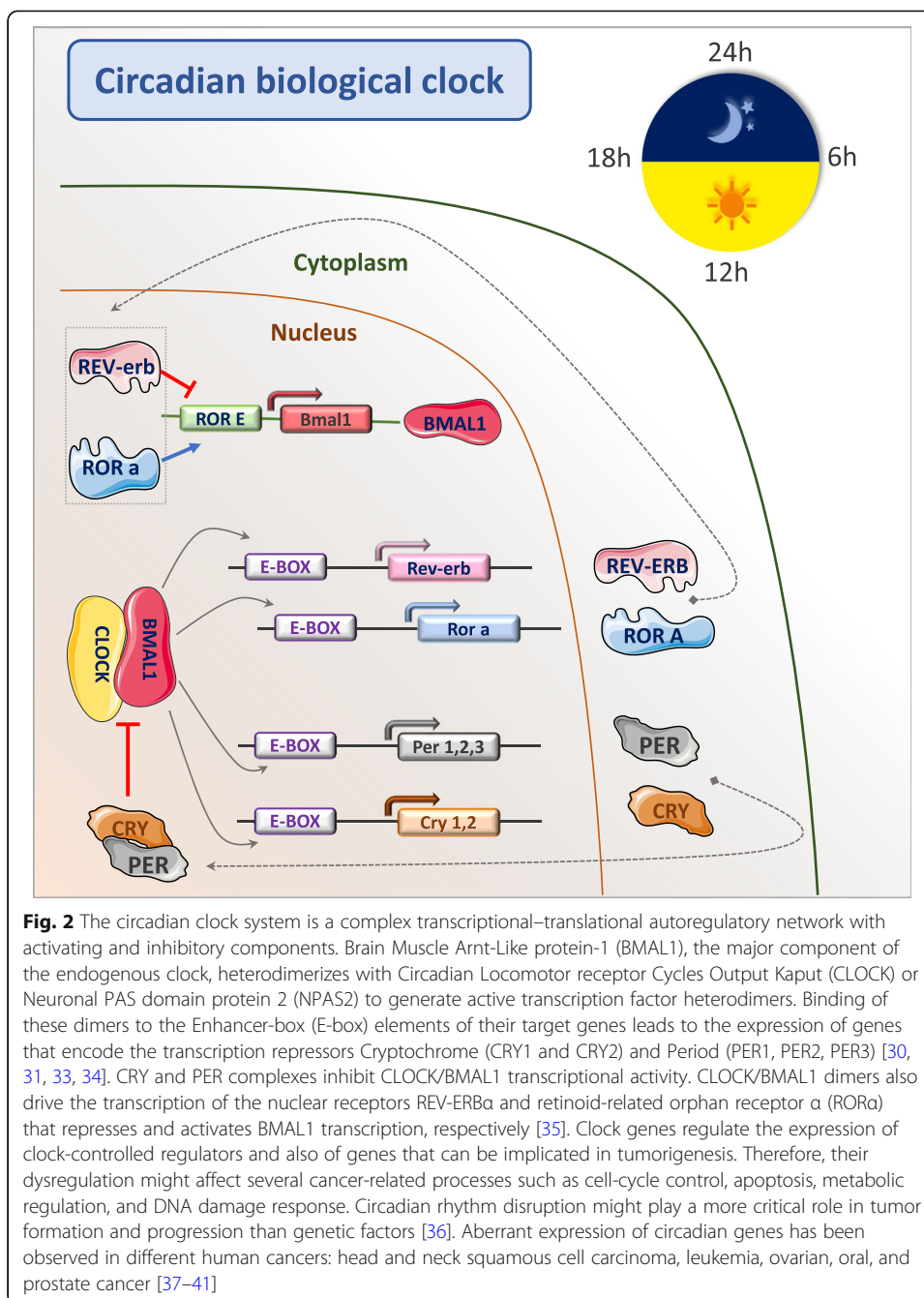
hypothalamus, the eyes, and the pineal complex [26–28]. The relative importance of each axis component varies in different species, depending on the selective pressures in the photic niche they occupy [29]. In mammals, the SCN has the main role, and the whole cycle is abolished upon removal of this organ.

In the SCN, the circadian rhythm is regulated by molecular mechanisms that set a transcription-translation feedback loop. Indeed, cells in the SCN are synchronized and use the proteins Brain Muscle Arnt-Like protein-1 (BMAL1) and Circadian Locomotor receptor Cycles Output Kaput (CLOCK) as transcription factors to regulate the expression of several genes related to the circadian cycle [30]. BMAL1 and CLOCK also induce the expression of members of the Period (PER) and Cryptochrome (CRY) families [31, 32] that accumulate and repress the function of these two transcription factors (Fig. 2).

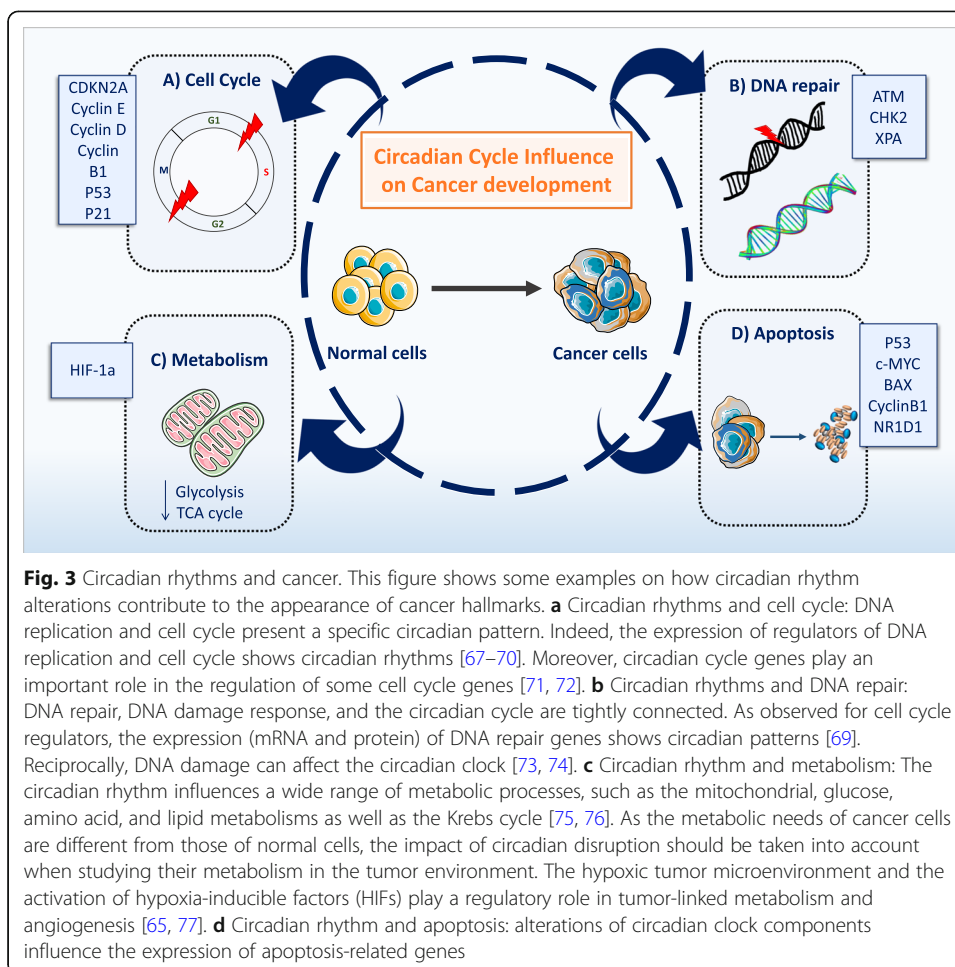
Biological cycles and cancer

Biological cycles influence nearly all physiological and biological aspects of an organism, and acute and chronic alterations of the circadian rhythm have been linked to various health issues and diseases [35, 42]. Humans evolved to be active during the day and to sleep at night. Recent society changes, especially in developed countries, have important consequences on the circadian rhythm. For example, studies on sleep deprivation have shown that the disruption of the sleep pattern produces cognitive alterations and behavioral changes in the short term [43, 44], and mood disorders (e.g., depression) in the long term [45]. On the other hand, sleep deprivation has been used as antidepressant therapy [46]. Other studies found that circadian system disturbance by exogenous factors (night shift work, physiologic perturbation, and exposure to light at night) is associated with higher cancer incidence and poor prognosis. For instance, shift work is correlated with higher risk of breast, prostate, lung, and colorectal cancer [47–50]. A recent cohort study with almost 10 years of follow-up found a significant association between increasing breast cancer risk and mean hours of night work per week [51]. Moreover, the cancer risk is increased by the number of years that an individual has spent working during the night [52, 53]. However, a meta-analysis concluded that night shift, including long-term shift work, has little or no effect on breast cancer incidence [54]. These discrepancies are probably due to the many factors involved in cancer development and progression.

Different studies have also investigated the effect of sleep duration on cancer risk. Short sleep duration has been correlated with higher cancer risk and the development of more aggressive tumors [55–57]. Conversely, longer sleep duration reduces the risk of breast cancer [58]. Moreover, cancer recurrence has been associated with the mean hours of sleep per night before cancer diagnosis, and shorter sleep duration was correlated with higher recurrence score [59]. A study with 5 years of follow-up before and after the diagnosis of prostate cancer showed that the risk of prostate cancer is higher in men with sleep disruption [60]. However, other studies did not find any association between sleep duration or sleep disruption and cancer risk [61, 62]. Finally, circadian rhythm alterations might be an independent prognostic risk factor of poor survival in patients with cancer and of treatment side effects [63, 64]. There is strong evidence about the existence of reciprocal interactions between cancer and the circadian clock components. Indeed, alterations of the circadian regulation and homeostatic balance



may facilitate the transformation of normal cells into malignant cells [35, 65]. Cancer initiation and progression are influenced by circadian cycle components through (i) direct or indirect regulation of different genes; (ii) interaction of circadian cycle proteins with other proteins; (iii) changes in redox state, cofactors, and post-translational modifications; and (iv) regulation of secreted factors with paracrine or endocrine function. These regulations have an effect on cellular processes, including nutrient metabolism, cell cycle, DNA repair, redox regulation, cellular secretion, protein folding, and autophagy [35, 66]. Figure 3 summarizes the molecular mechanisms related to the circadian cycle that are also involved in cancer development.



The circadian cycle is not the only rhythm that influences the organismal physiology. Another example in humans is the menstrual cycle that usually lasts between 24 and 38 days and is controlled by different hormones produced by the hypothalamus, pituitary, and ovaries. The risk of developing breast cancer is increased in women with higher number of cycles during their life [78, 79]. Furthermore, hormone replacement therapy and hormonal oral contraceptives promote abnormal mammary epithelial cell proliferation, resulting in higher breast cancer risk [80]. From a clinical point of view, this means that in premenopausal women, screening by mammography should be performed during the first week of the menstrual cycle [81], although this is not fully implemented in screening guidelines [82].

In addition, several studies have shown the importance of annual cycles in tumor development. For example, the seasonality of sunlight exposure and vitamin D synthesis might directly influence the risk of cancer [83–87]. Moreover, the analysis of long-term survival time series in function of the season of cancer diagnosis found a reduction in death rates among patients in whom breast cancer was diagnosed in fall [88]. Another study on skin cancer in Norway showed no significant variation of incidence and mortality rates in relation with seasons. However, a significant seasonal variation of cancer prognosis was observed [89]. Other factors might influence cancer development during

the year. Indeed, skin cancers might result from excessive exposure to ultraviolet light in tropical regions [90], and warmer air temperatures have been associated with lower cancer death rates [91]. As annual cycles are hard to study and many confounding factors might influence cancer outcome, there is still no clear evidence of their direct role in cancer.

The influence of biological cycles on the neuroimmune-endocrine system and its clinical implications for cancer progression and dissemination

In addition to the central biological clock in the SCN, all cells and tissues have some molecular clocks. These peripheral biological clocks strongly modulate the neuroimmune-endocrine system.

The immune system is significantly influenced by the circadian cycle through various hormonal and molecular pathways, particularly the hypothalamus-pituitary-adrenal axis (HPAA), where the secretion of the glucocorticoid hormone cortisol (hydrocortisone) from the suprarenal cortex peaks early in the day [92–94]. Glucocorticoid receptors are found in the whole organism, and their main effects concern (but are not limited to) the glucose metabolism and immune system [95]. In rheumatic diseases, the use of hydrocortisone as anti-inflammatory drug increases the risk of infections [96], and the cortisol circadian peak might predispose to infections at specific times of the day. Moreover, some infectious diseases show clearly circadian patterns of symptoms. For instance, in patients with tuberculosis, fever is often exacerbated at night [97]. In infectious diseases, this difference in fever patterns might be an adaptive trait to limit the symptoms of infection to the night, thus promoting viability during the day. Additionally, it has been shown that the expression of interleukin-6 (IL-6), the main mediator of fever, can be regulated by *Mycobacterium tuberculosis* in infected macrophages [98]. This facilitates infection progression because IL-6 also stimulates the secretion of cortisol during the immune system activation [99]. Moreover, in humans, IL-6 secretion follows a circadian rhythm [100]. These circadian mechanisms might be extrapolated to cancer as well. For instance, in patients with lymphoma, cytokines are strongly released by malignant leukocytes during the night, leading to nocturnal fever peaks [101]. Moreover, in pancreatic and colorectal cancer, IL-6 has been shown to support the formation of a pro-metastatic niche in the liver [102]. Currently, it is not fully understood how cancer-induced nighttime fever may be beneficial to cancer cells from a fitness perspective; however, its presence and high IL-6 concentrations are often considered bad prognostic factors [103–106].

In a different manner, the central biological clock can influence the peripheral biological clocks not only via the HPAA, but also via the sympathetic nervous system (SNS) by using epinephrine and norepinephrine as signaling molecules. The SNS is mainly active during daytime and induces an anti-inflammatory environment, in a similar manner to glucocorticoids. The SNS might also have a role in cancer progression. Indeed, in prostate cancer, the formation of sympathetic nerve fibers contributes to cancer development and dissemination [107], and sensory nerves are induced to trans-differentiate into sympathetic nerves directly by cancer cells [108].

Both HPAA and SNS are downregulated in the early sleep period when different hormones, such as leptin, melatonin, growth hormone, and prolactin, reach their peak. These hormones have synergic pro-inflammatory actions [95]. Additionally, melatonin

effectively inhibits epithelial to mesenchymal transition (EMT), which is considered one of the main mechanisms for cancer dissemination, via different pathways, such as reduction of IL-1 β /NF- κ B/MMP2/MMP9 signaling [109], and inhibition of Twist/Twist1 expression [110]. High melatonin levels also significantly suppress cell proliferation and induce apoptosis by inhibition of cyclooxygenase-2 and p300 [111]. Melatonin can also suppress cell invasion/migration through MMP-9-mediated ECM remodeling [112]. Melatonin also exerts inhibitory effects on metastatic HER2/neu-negative breast cancer cell migration and invasion of by repressing a panel of mesenchymal genes that regulate EMT [113]. Moreover, during the melatonin peak, hematopoietic stem cell self-renewal is enhanced through CD150 upregulation [114]. CD150 family members are key regulators of leukocyte activation and differentiation [115]. The daily cycle and secretion of different hormones are essential for the synchronized maturation of blood cells [114].

Furthermore, the immune performance of an organism is reduced by circadian cycle alterations. For example, sleep deprivation for just 24 h can drastically decrease the efficacy of the hepatitis A vaccine, an effect that is also associated with the lower cortisol release during the night [116]. This effect can be explained by the presence of two different “immune environments” in the day and in the night. Dimitrov et al. found that the number of effectors CD8⁽⁺⁾ T lymphocytes reaches its maximum peak during daytime. This outcome is driven by β -adrenergic and fractalkine (CX3CR1) receptors that are strongly expressed in these cells and increase the influence of catecholamines, related to the SNS [117]. In contrast, the number of naive T lymphocytes is lower during the day (and increases at night) when they are redistributed to the bone marrow by chemoattraction to the C-X-C motif chemokine 12 (CXCL12), the ligand of C-X-C chemokine receptor type 4 (CXCR4) that is strongly expressed on these cells. Likewise, He et al. demonstrated that CXCR4 governs the circadian fluctuation of leukocytes and myeloid lineages, such as neutrophils. Moreover, they showed that *BMALI* gene ablation in leukocyte subsets leads to a reduction of these circadian fluctuations [118].

These observations can be interpreted as different immune reaction mechanisms during the circadian cycle, in which effector CD8⁽⁺⁾ T lymphocytes represent the immediate immune reaction during the day, and naive T lymphocytes are the main actors of the adaptive (lasting) immune reaction during the night [117]. From an evolutionary perspective, this means that during cancer cell dissemination, CTCs must adapt and elude the immediate immune response that is most active during daytime, and evade the adaptive immune response during the night. Interestingly, CXCR4 is strongly expressed in several cancer types [119, 120] and in CTCs is associated with poor prognosis [121]. Therefore, we could hypothesize that similarly to naive T lymphocytes, CTCs might migrate to the bone marrow at specific times of the day to escape the immune system [122]. Additionally, a recent study found that CTCs are associated with neutrophils, and proposed that this might enhance their metastatic potential [123]. It might be possible that due to the neutrophil fluctuations associated with the circadian rhythms, the formation of CTC clusters is facilitated at specific times of the day, or upon disruption of the circadian cycle. Moreover, in an in vivo mice model, chronic circadian rhythm disruption enhances the CXCL12-CXCR4 axis activity and chemoattraction to component of the C-X-C motif chemokine 5-C-X-C chemokine receptor type 2 (CXCL5-CXCR2) axis. These two axes recruit myeloid-derived suppressor cells into the

tumor micro-environment, promoting cancer cell dissemination and metastasis formation. In agreement, inhibition of the CXCL5-CXCR2 axis limits the number of metastasis in mice [124]. The higher expression of components of these two axes upon circadian disruption might increase CTC fitness.

On the other hand, expression of specific markers can give some advantages to CTC subsets that will be selected. One example is the acquisition by CTCs of the “perfect” phenotype to escape the immune system. Indeed, expression of PD-L1 (*a camouflage to trap the immune system*) and/or CD47 (*the “do not eat me” signal*) by CTCs certainly help these disseminating tumor cells to survive the attack by immune cells. A recent study in *Bmal1*^{-/-} mice showed that *Bmal1* (one of the two main regulators of the circadian cycle) counter-balances *Pd-l1* expression in macrophages and plays a role in the immune response [125]. This kind of mechanism could, in theory, also be involved in CTC clearance from blood during the day, with possible implications for the sampling time and evaluation of this marker in CTCs.

Another interesting observation, with potential consequences for tumor dissemination, is that circadian variations can predispose to higher platelet activation and coagulation, thus potentially promoting CTC survival. For instance, cardiovascular events have day/night patterns with peaks in the morning, suggesting a potential link with the endogenous circadian clock that controls platelet activation. This hypothesis is supported by the observation that the expression peaks of platelet surface activated glycoprotein (GP) IIb-IIIa, GPIb, and P-selectin also occur at that time [126]. These molecules are involved in the reciprocal interaction of platelets and CTCs. Indeed, platelet P-selectin interacts with tumor cell CD44, and the fibrinogen receptors GPIIb-IIIa are involved in platelet adherence on CTCs and in the formation of platelet-CTC emboli [127]. Moreover, platelets can protect CTCs by forming a protective cloak and by conferring a “pseudo-normal” phenotype against natural killer cells [128]. As platelet-coated CTCs have been detected in the blood of patients with metastatic cancer [129], the circadian peaks of platelet activation and of surface marker expression suggest possible roles for the circadian clock in CTC escape from the immune system. However, the role of the circadian cycle in CTCs and the potential consequences on CTC clone fitness have not been elucidated yet.

These observations might have implications for CTC use as *liquid biopsy*. Most studies on CTCs do not report the sampling time [130, 131]. Only one group tried to determine the circadian cycle role in tumor cell dissemination. To evaluate a possible circadian variation in CTC number in patients with metastatic breast cancer, they carried out two studies ($n = 51$ and $n = 23$ samples) in which CTCs were quantified 12 h apart (at 8:00 a.m. and at 8:00 p.m. of the same day) [130, 131]. The authors concluded that CTC count, determined with the CellSearch® system, does not significantly fluctuate depending on the sampling time. These data suggest that the circadian rhythm does not influence tumor cell dissemination; however, they need to be confirmed in a bigger cohort of patients in a multi-center trial involving independent research groups. Moreover, blood samples were not collected during the night. Additionally, the Nyquist theorem states that to detect a cycle, sampling rate must be at least twice the signal frequency [132, 133]. Two samples may theoretically be enough to show crude day/night differences, but the cycle amplitude (e.g., expressed in number of CTCs) may be difficult to estimate, if samples are not collected when CTC numbers

are highest and lowest. Therefore, at least four samples equidistant in time should be required to clearly define a cycle. Moreover, only two data points for 24 h might not allow highlighting possible scenarios in which CTC number increases during a short time window. These statements need to be considered in the design of future studies.

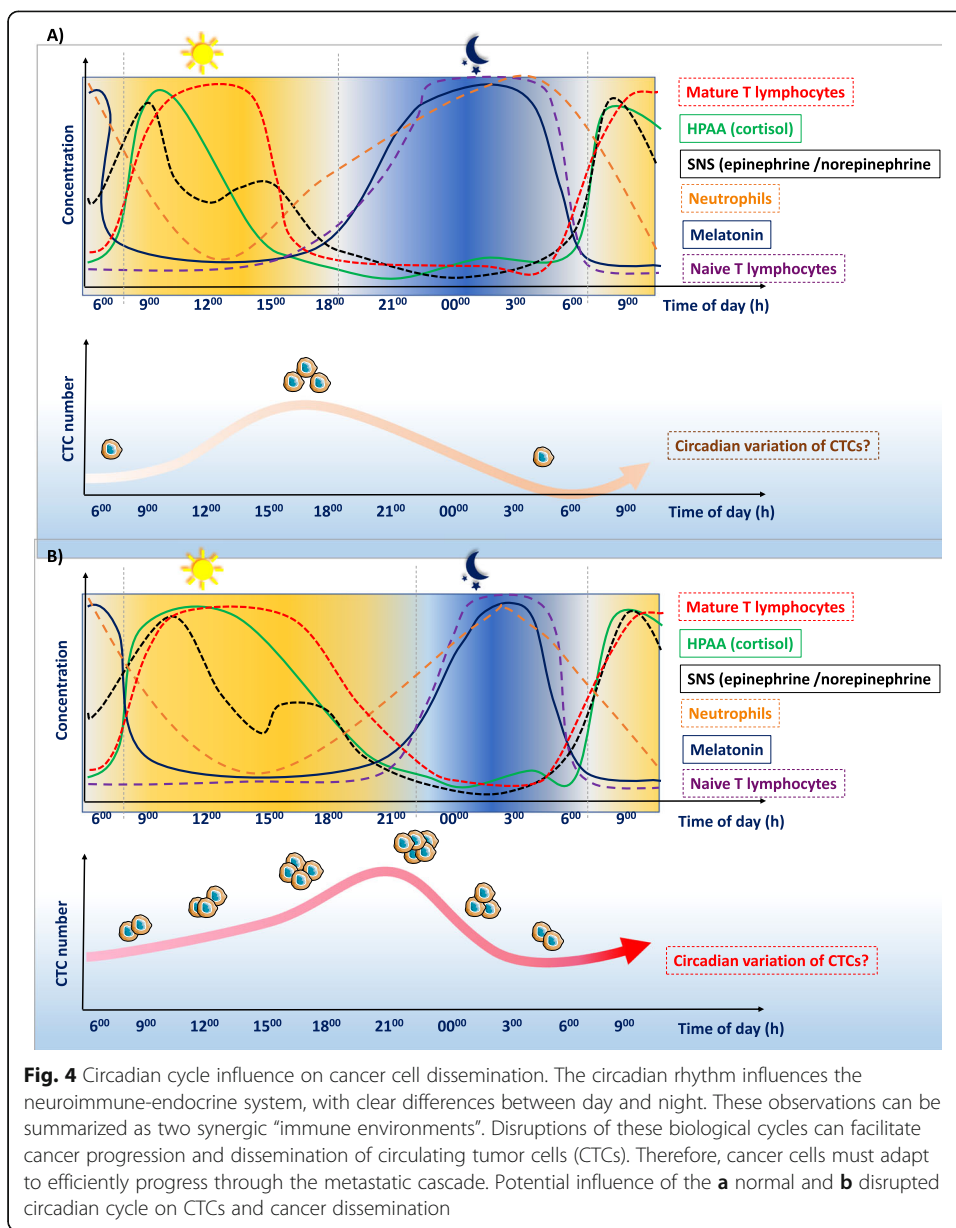
Furthermore, independently of CTC number in blood, the biology of each CTC can be different according to its neuroimmune-endocrine environment. For instance, expression of CXCR4 might fluctuate in CTCs during the day, and this might have important clinical implications. Indeed, these tumor cells can migrate and hide in specific niches in the bone marrow, which is considered a “reservoir” for disseminated cells where they can enter dormancy after having left the primary tumor and survived in the circulation [20]. In addition, CTCs might take advantage of the anti-inflammatory effects of cortisol (the secretion of which is strongly increased upon sleep disruptions) to escape the immune surveillance and survive. Moreover, alterations in the circadian cycle will modify the influence of this cycle on cancer cell dissemination (Fig. 4). Altogether, the possible fluctuations related to the biological cycles may play a crucial role in defining the best moment for blood sampling to increase the chance of CTC detection.

On the other hand, the knowledge on circadian biology has been successfully applied to maximize the balance between chemotherapy efficacy and toxicity. For instance, the early clinical trials on oxaliplatin, which is one of the main therapies against colorectal cancer, showed high toxicity, and therefore, its further development was stopped [134]. However, a chronopharmacology approach allowed identifying the best time for oxaliplatin administration, strongly improving patient survival [135]. In the *liquid biopsy* field, the circadian cycle-based schedules have been rarely implemented, although it would be important to include this strategy in future clinical trials on *liquid biopsy* and in cancer research in general [136].

Evolutionary considerations

The data described in the previous sections indicate that biological rhythms, especially the circadian cycle, should not be neglected when studying cancer. The circadian rhythm influences the host defense efficiency against cancer. Moreover, malignant cells, like any other cell in the holobiont, are directly or indirectly influenced by the host internal rhythms.

In addition, it should be important to determine whether during tumorigenesis, malignant cells can acquire some features, like parasites and microbiota, that allow them to exploit their host circadian rhythm, and/or even to manipulate it to improve their proliferation and dispersal. One major difference between parasitic organisms/bacteria and malignant cells is that in most cases, cancer cells are not transmitted between hosts (but see [137]). Thus, although cancer is an ancestral disease [138], each cancer must “reinvent the wheel” because their evolutionary adaptations disappear when their host dies. As the “lifestyle” of malignant cells is under selective pressure for only few decades, highly sophisticated adaptive responses, as observed in organisms exposed to natural selection for millions of years, may be unlikely to appear [139]. On the other hand, in only few years, natural selection can act on the extreme diversity of cells typically generated by tumors, leading to the emergence of clones that show the best features for malignant progression, metastasis development, and resistance to immune



attack and to therapies. In such “successful” tumors, adaptations in relationship with the host circadian rhythm may be detected.

Assuming that the conditions allowing the selection of these adaptive traits are present, two hypotheses concerning the behavior of malignant cells, especially CTCs, relative to the host circadian rhythm can be proposed. First, oncogenic selection might lead to the emergence of malignant cells that avoid, or limit their activities during the part of the circadian rhythm that is not favorable or dangerous for them, and amplify them during advantageous time windows (see for instance [140, 141]). Second, malignant cells could acquire the ability to alter their host circadian cycle to promote directly or indirectly their proliferation or dispersal (see for instance [142]).

Concerning the first hypothesis, the number of circulating immune cells in humans varies during the circadian cycle [118, 143]. Moreover, some studies highlighted the

asynchrony in cell proliferation and metabolism between host and malignant tissues [144]. Finally, there is experimental evidence that some periods of the day are better than others for cancer cell proliferation and spread. For instance, using two genetically distinct mouse strains and two different tumor cell models (i.e., fibrosarcoma and melanoma), Hrushesky et al. [145] determined the tumor-take frequency after subcutaneous tumor cell inoculation, and the number of pulmonary tumor nodules after intravenous tumor cell injection at six equidistant times of the day. With fibrosarcoma cells, they found that tumor-take frequency was lower after injection close to the daily sleep/wake boundary. Similarly, with melanoma cells, the daily sleep/wake boundary was the time associated with the greatest resistance to metastatic spread. In this context, it is not surprising that therapies altering the tumor biological rhythms are increasingly considered as efficient to prevent cancer proliferation and spread, in addition to their limited toxicity for the surrounding healthy tissue [146].

Concerning the second hypothesis, few studies found that in patients with cancer (notably breast and ovarian cancer [147, 148]), the distinction between nighttime and daytime activities is reduced, suggesting a possible circadian rhythm disruption. Insomnia is also frequently reported by patients with breast, gynecological, and lung cancer [149]. Although these circadian rhythm disorders may have a variety of causes, a direct effect of the tumor on sleep is possible [149], for instance by affecting the secretion of the cytokines that modulate the sleep-wake cycle. Interestingly, several studies have suggested that the aberrant clock gene expression observed in many tumors might promote cancer cell survival, and have identified the mechanisms through which malignant cells induce sleep disruption (see [150] for a recent review). However, they did not consider their findings in the framework of a potential adaptive context of host manipulation. We strongly believe that the hypothesis of circadian rhythm manipulation by cancer is an additional reason to adopt a Darwinian approach in cancer research. More studies are needed to determine whether cancer cells with the ability to disrupt circadian rhythms are directly favored by selection, or whether this is only a side effect with positive effects on carcinogenesis. In any case, in accordance with the idea that fighting cancer adaptation might improve therapies, pharmacological modulation of clock-related proteins (REV-ERB receptor activity) is increasingly considered as an effective anticancer strategy [151].

Concluding remarks

While the circadian timing of cancer treatments is progressively acknowledged, e.g., [152–154], few studies have tested the hypothesis that malignant cells could exploit and/or manipulate the host biological rhythms. Yet, it is important to determine whether, when, and how malignant cells can acquire such adaptive traits in order to prevent their selection and to limit their spreading. This is particularly true for CTCs because this could strongly influence the speed and rate of expansion of invasive cancers. As several non-genetic variables are involved in tumor formation and progression, it is also essential to determine whether the acquisition of the ability to exploit/manipulate circadian rhythms by malignant cells has to be considered within the framework of phenotypic plasticity or just as a genetic adaptation. It is also important to consider the whole holobiont in which these malignant adaptations could occur, by considering the microbiota and the parasitic organisms present in the host, because they may have

common or conflicting interests concerning the host exploitation, e.g., [155]. Only such an integrative approach will provide a correct assessment of the context in which malignant cells that can exploit/manipulate the host biological rhythms are selected. As one single discipline or biological model cannot correctly describe all these phenomena, scientists from different fields must engage in exchanges and collaborations. Demonstrating that malignant cells can exploit/manipulate their host biological rhythms is only the first step. Indeed, this knowledge will have to be integrated in the design of preventive or curative strategies, and for improving the identification of life periods when the risk of invasive cancer initiation is highest. It will also be necessary to determine whether these adaptations rely on a more or less constant/obligatory sequence of events, with the same proximate factors (i.e., evolutionary convergence) that could potentially be targeted by specific therapies. Another promising research direction is to explore the proximate mechanisms used by parasites that also manipulate the host circadian rhythms to favor their own fitness, e.g., [156]. If such strategies rely on similar biochemical precursors, it would be important to test whether therapies that target manipulative activities in parasites might be equally effective against manipulation by cancer cells (see for instance [157]).

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13059-020-02179-w>.

Additional file 1. Review history.

Acknowledgements

The authors thank Dr. Elisabetta Andermarcher for assistance with her comments and proofreading that greatly improved the manuscript. The figures were designed with the assistance of and modification to the images provided by the Servier Medical Art database <http://smart.servier.com/>. Further information pertaining to the license and disclaimer notices can be found here: <https://creativecommons.org/licenses/by/3.0/>.

Review history

The review history is available as Additional file 1.

Peer review information

Anahita Bishop was the primary editor of this article and managed its editorial process and peer review in collaboration with the rest of the editorial team.

Authors' contributions

All authors contributed to the conception of the work, drafting the article, critical revision of the article, and final approval of the version to be published. LECH and ZES contributed equally to this work.

Funding

ZES, LECH, and CAP are supported by the ELBA project that has received funding from the European Union Horizon 2020 Research and Innovation program under the Marie Skłodowska-Curie grant agreement No 765492. CAP is also supported by The National Institute of Cancer (INCa, <http://www.e-cancer.fr>), SIRIC Montpellier Cancer Grant INCa_Inserm_DGOS_12553, and the ERA-NET TRANSCAN 2 JTC 2016 PROLIPSY. FT, MG, BU are supported by an ANR TRANSCAN (ANR-18-CE35-0009), the Rotary Club Les Sables d'Olonne, and a CNRS International Associated Laboratory Grant. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Availability of data and materials

Not applicable.

Competing interests

CAP is one of the patent holders (US Patent Number 16,093,934) for detecting and/or characterizing circulating tumor cells. The remaining authors declare no conflict of interest.

Author details

¹Laboratory of Rare Human Circulating Cells (LCCRH), University Medical Centre of Montpellier, Montpellier, France.

²CREEC (CREES), Unité Mixte de Recherches, IRD 224–CNRS 5290–Université de Montpellier, Montpellier, France.

³Centre for Integrative Ecology, School of Life and Environmental Sciences, Deakin University, Waurn Ponds, Victoria,

Australia. ⁴School of Natural Sciences, University of Tasmania, Hobart, Tasmania, Australia. ⁵Institut Universitaire de Recherche Clinique (IURC), 641, avenue du Doyen Gaston Giraud, 34093 Montpellier Cedex 5, France.

Received: 30 May 2020 Accepted: 13 October 2020

Published online: 12 November 2020

References

- Baedke J, Fábregas-Tejeda A, Nieves DA. The holobiont concept before Margulis. *J Exp Zool Part B Mol Dev Evol*. 2020; 334(3):149–55.
- Cavalier-Smith T. Symbiosis as a source of evolutionary innovation: speciation and morphogenesis. Margulis L, Fester R, editors. *Trends Ecol Evol* 1992;7(12):422–423.
- Thomas F, Jacqueline C, Tissot T, Henard M, Blanchet S, Loot G, et al. The importance of cancer cells for animal evolutionary ecology. *Nat Ecol Evol*. 2017;1(11):1592–5.
- Bissell MJ, Hines WC. Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression. *Nat Med*. 2011;17(3):320–9.
- Folkman J, Kalluri R. Cancer without disease. *Nature*. 2004;427(6977):787.
- Thomas F, Vavre F, Tissot T, Vittecoq M, Giraudeau M, Bernex F, et al. Cancer is not (only) a senescence problem. *Trends in Cancer*. 2018;4(3):169–72.
- Duesberg P, Mandrioli D, McCormack A, Nicholson JM. Is carcinogenesis a form of speciation? *Cell Cycle*. 2011;10(13): 2100–14.
- Vincent MD. The animal within: carcinogenesis and the clonal evolution of cancer cells are speciation events sensu stricto. *Evolution (N Y)*. 2010;64(4):1173–83.
- Gatenby RA, Avdieiev S, Tsai KY, Brown JS. Integrating genetic and non-genetic drivers of somatic evolution during carcinogenesis: the biplane model. *Evol Appl*. 2020;13:1651–9.
- Thomas F, Brown SP, Sukhdeo M, Renaud F. Understanding parasite strategies: a state-dependent approach? *Trends Parasitol*. 2002;18(9):387–90.
- Westwood ML, O'Donnell AJ, de Bekker C, Lively CM, Zuk M, Reece SE. The evolutionary ecology of circadian rhythms in infection. *Nat Ecol Evol*. 2019;3(4):552–60.
- Martinez-Bakker M, Helm B. The influence of biological rhythms on host-parasite interactions. *Trends Ecol Evol*. 2015; 30(6):314–26.
- Reece SE, Prior KF, Mideo N. The life and times of parasites: rhythms in strategies for within-host survival and between-host transmission. *J Biol Rhythm*. 2017;32(6):516–33.
- O'Donnell AJ, Prior KF, Reece SE. Host circadian clocks do not set the schedule for the within-host replication of malaria parasites. *Proceedings Biol Sci*. 2020;287(1932):20200347.
- Kiessling S, Dubeau-Laramée G, Ohm H, Labrecque N, Olivier M, Cermakian N. The circadian clock in immune cells controls the magnitude of Leishmania parasite infection. *Sci Rep*. 2017;7(1):10892.
- Van Houte S, Van Oers MM, Han Y, Vlak JM, Ros VID. Baculovirus infection triggers a positive phototactic response in caterpillars to induce "tree-top" disease. *Biol Lett*. 2014;10(12):20140680.
- De Bekker C, Will I, Das B, Adams RMM. The ants (Hymenoptera: Formicidae) and their parasites: effects of parasitic manipulations and host responses on ant behavioral ecology. *Myrmecological News*. 2018;28:1–24.
- Liang X, Bushman FD, FitzGerald GA. Rhythmicity of the intestinal microbiota is regulated by gender and the host circadian clock. *Proc Natl Acad Sci U S A*. 2015;112(33):10479–84.
- Kuang Z, Wang Y, Li Y, Ye C, Ruhn KA, Behrendt CL, et al. The intestinal microbiota programs diurnal rhythms in host metabolism through histone deacetylase 3. *Science*. 2019;365(6460):1428–34.
- Pantel K, Alix-Panabières C. Liquid biopsy and minimal residual disease — latest advances and implications for cure. *Nat Rev Clin Oncol*. 2019;16(7):409–24.
- Eslami-S Z, Cortés-Hernández LE, Alix-Panabières C. The metastatic cascade as the basis for liquid biopsy development. *Front Oncol*. 2020;10:1055.
- Eslami-S Z, Cortes-Hernandez LE, Cayrefourcq L, Alix-Panabieres C. The different facets of liquid biopsy: a kaleidoscopic view. *Cold Spring Harb Perspect Med*. 2019;10(6):a037333.
- Klerman EB. Clinical aspects of human circadian rhythms. *J Biol Rhythm*. 2005;20(4):375–86.
- Evans JA, Davidson AJ. Health consequences of circadian disruption in humans and animal models. *Prog Mol Biol Transl Sci*. 2013;119:283–323.
- Rana S, Mahmood S. Circadian rhythm and its role in malignancy. *J Circadian Rhythms*. 2010;8.
- Reppert SM, Weaver DR. Coordination of circadian timing in mammals. *Nature*. 2002 Aug;418(6901):935–41.
- Tosini G, Bertolucci C, Foà A. The circadian system of reptiles: a multioscillatory and multiphotoreceptive system. *Physiol Behav*. 2001 Mar;72(4):461–71.
- Borjigin J, Zhang LS, Calinescu A-A. Circadian regulation of pineal gland rhythmicity. *Mol Cell Endocrinol*. 2012;349(1): 13–9.
- Menaker M, Moreira LF, Tosini G. Evolution of circadian organization in vertebrates. *Brazilian J Med Biol Res*. 1997;30(3): 305–13.
- Vitaterna MH, King DP, Chang AM, Kornhauser JM, Lowrey PL, McDonald JD, et al. Mutagenesis and mapping of a mouse gene, clock, essential for circadian behavior. *Science*. 1994;264(5159):719–25.
- Bae K, Jin X, Maywood ES, Hastings MH, Reppert SM, Weaver DR. Differential functions of mPer1, mPer2, and mPer3 in the SCN circadian clock. *Neuron*. 2001;30(2):525–36.
- van der Horst GT, Muijtjens M, Kobayashi K, Takano R, Kanno S, Takao M, et al. Mammalian Cry1 and Cry2 are essential for maintenance of circadian rhythms. *Nature*. 1999;398(6728):627–30.
- Farshadi E, Yan J, Leclere P, Goldbeter A, Chaves I, van der Horst GTJ. The positive circadian regulators CLOCK and BMAL1 control G2/M cell cycle transition through cyclin B1. *Cell Cycle*. 2019;18(1):16–33.
- Li H-X. The role of circadian clock genes in tumors. *Onco Targets Ther*. 2019;12:3645–60.

35. Sulli G, Lam MTY, Panda S. Interplay between circadian clock and cancer: new frontiers for cancer treatment. *Trends in cancer*. 2019;5(8):475–94.
36. Keith LG, Oleszczuk JJ, Laguens M. Circadian rhythm chaos: a new breast cancer marker. *Int J Fertil Womens Med*. 2001; 46(5):238–47.
37. Yang M-Y, Chang J-G, Lin P-M, Tang K-P, Chen Y-H, Lin HY-H, et al. Downregulation of circadian clock genes in chronic myeloid leukemia: alternative methylation pattern of hPER3. *Cancer Sci*. 2006;97(12):1298–307.
38. Tokunaga H, Takebayashi Y, Utsunomiya H, Akahira J-I, Higashimoto M, Mashiko M, et al. Clinicopathological significance of circadian rhythm-related gene expression levels in patients with epithelial ovarian cancer. *Acta Obstet Gynecol Scand*. 2008;87(10):1060–70.
39. Hsu C-M, Lin S-F, Lu C-T, Lin P-M, Yang M-Y. Altered expression of circadian clock genes in head and neck squamous cell carcinoma. *Tumour Biol*. 2012;33(1):149–55.
40. Relles D, Sendeci J, Chipitsyna G, Hyslop T, Yeo CJ, Arafat HA. Circadian gene expression and clinicopathologic correlates in pancreatic cancer. *J Gastrointest Surg*. 2013;17(3):443–50.
41. Xiong H, Yang Y, Yang K, Zhao D, Tang H, Ran X. Loss of the clock gene PER2 is associated with cancer development and altered expression of important tumor-related genes in oral cancer. *Int J Oncol*. 2018;52(1):279–87.
42. Masri S, Sassone-Corsi P. The emerging link between cancer, metabolism, and circadian rhythms. *Nat Med*. 2018;24(12): 1795–803.
43. Killgore WDS. Effects of sleep deprivation on cognition. *Prog Brain Res*. 2010;185:105–29.
44. Saghir Z, Syeda JN, Muhammad AS, Balla Abdalla TH. The amygdala, sleep debt, sleep deprivation, and the emotion of anger: a possible connection? *Cureus*. 2018;10(7):e2912.
45. Watson NF, Harden KP, Buchwald D, Vitiello MV, Pack AI, Strachan E, et al. Sleep duration and depressive symptoms: a gene-environment interaction. *Sleep*. 2014;37(2):351–8.
46. Dallaspesza S, Benedetti F. Sleep deprivation therapy for depression. *Curr Top Behav Neurosci*. 2015;25:483–502.
47. Papanitiou K, Devore EE, Massa J, Strohmaier S, Vetter C, Yang L, et al. Rotating night shift work and colorectal cancer risk in the nurses' health studies. *Int J Cancer*. 2018;143(11):2709–17.
48. Salamanca-Fernández E, Rodríguez-Barranco M, Guevara M, Ardanaz E, Olry de Labry Lima A, Sánchez MJ. Night-shift work and breast and prostate cancer risk: updating the evidence from epidemiological studies. *An Sist Sanit Navar* 2018; 41(2):211–226.
49. Yang W, Shi Y, Ke X, Sun H, Guo J, Wang X. Long-term sleep habits and the risk of breast cancer among Chinese women: a case-control study. *Eur J Cancer Prev*. 2019;28(4):323–9.
50. Kloog I, Haim A, Stevens RG, Portnov BA. Global co-distribution of light at night (LAN) and cancers of prostate, colon, and lung in men. *Chronobiol Int*. 2009;26(1):108–25.
51. Jones ME, Schoemaker MJ, McFadden EC, Wright LB, Johns LE, Swerdlow AJ. Night shift work and risk of breast cancer in women: the Generations Study cohort. *Br J Cancer*. 2019;121(2):172–9.
52. Schernhammer ES, Laden F, Speizer FE, Willett WC, Hunter DJ, Kawachi I, et al. Night-shift work and risk of colorectal cancer in the nurses' health study. *J Natl Cancer Inst*. 2003;95(11):825–8.
53. Lee S, Donehower LA, Herron AJ, Moore DD, Fu L. Disrupting circadian homeostasis of sympathetic signaling promotes tumor development in mice. *PLoS One* 2010;7(5):e10995.
54. Travis RC, Balkwill A, Fensom GK, Appleby PN, Reeves GK, Wang X-S, et al. Night shift work and breast cancer incidence: three prospective studies and meta-analysis of published studies. *J Natl Cancer Inst*. 2016;108(12):djw169.
55. Kakizaki M, Kuriyama S, Sone T, Ohmori-Matsuda K, Hozawa A, Nakaya N, et al. Sleep duration and the risk of breast cancer: the Ohsaki Cohort Study. *Br J Cancer*. 2008;99(9):1502–5.
56. Xiao Q, Signorello LB, Brinton LA, Cohen SS, Blot WJ, Matthews CE. Sleep duration and breast cancer risk among black and white women. *Sleep Med*. 2016;20:25–9.
57. Soucise A, Vaughn C, Thompson CL, Millen AE, Freudenheim JL, Wactawski-Wende J, et al. Sleep quality, duration, and breast cancer aggressiveness. *Breast Cancer Res Treat*. 2017;164(1):169–78.
58. Wu AH, Wang R, Koh W-P, Stanczyk FZ, Lee H-P, Yu MC. Sleep duration, melatonin and breast cancer among Chinese women in Singapore. *Carcinogenesis*. 2008;29(6):1244–8.
59. Thompson CL, Li L. Association of sleep duration and breast cancer OncotypeDX recurrence score. *Breast Cancer Res Treat*. 2012;134(3):1291–5.
60. Sigurdardottir LG, Valdimarsdottir UA, Mucci LA, Fall K, Rider JR, Schernhammer E, et al. Sleep disruption among older men and risk of prostate Cancer. *Cancer Epidemiol biomarkers Prev a Publ Am Assoc Cancer Res cosponsored by Am Soc Prev Oncol* 2013;22(5):872–879.
61. Kakizaki M, Inoue K, Kuriyama S, Sone T, Matsuda-Ohmori K, Nakaya N, et al. Sleep duration and the risk of prostate cancer: the Ohsaki Cohort Study. *Br J Cancer*. 2008;99(1):176–8.
62. Vogtmann E, Levitan EB, Hale L, Shikany JM, Shah NA, Endeshaw Y, et al. Association between sleep and breast cancer incidence among postmenopausal women in the Women's Health Initiative. *Sleep*. 2013;36(10):1437–44.
63. Palesh O, Aldridge-Gerry A, Zeitzer JM, Koopman C, Neri E, Giese-Davis J, et al. Actigraphy-measured sleep disruption as a predictor of survival among women with advanced breast cancer. *Sleep*. 2014;37(5):837–42.
64. Li W, Kwok CC-H, Chan DC-W, Wang F, Tse LA. Weak circadian rhythm increases neutropenia risk among breast cancer patients undergoing adjuvant chemotherapy. *Breast Cancer Res Treat*. 2018;168(2):483–93.
65. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646–74.
66. Rijo-Ferreira F, Takahashi JS. Genomics of circadian rhythms in health and disease. *Genome Med*. 2019;11(1):82.
67. Smaaland R. Circadian rhythm of cell division. *Prog Cell Cycle Res*. 1996;2:241–66.
68. Matsuo T, Yamaguchi S, Mitsui S, Emi A, Shimoda F, Okamura H. Control mechanism of the circadian clock for timing of cell division in vivo. *Science*. 2003;302(5643):255–9.
69. Wang J, Mauvoisin D, Martin E, Atger F, Galindo AN, Dayon L, et al. Nuclear proteomics uncovers diurnal regulatory landscapes in mouse liver. *Cell Metab*. 2017;25(1):102–17.
70. Soták M, Sumová A, Pácha J. Cross-talk between the circadian clock and the cell cycle in cancer. *Ann Med*. 2014;46(4):221–32.
71. Fu L, Pelicano H, Liu J, Huang P, Lee C. The circadian gene *Period2* plays an important role in tumor suppression and DNA damage response in vivo. *Cell*. 2002;111(1):41–50.

72. Gréchez-Cassiau A, Rayet B, Guillaumond F, Teboul M, Delaunay F. The circadian clock component BMAL1 is a critical regulator of p21WAF1/CIP1 expression and hepatocyte proliferation. *J Biol Chem*. 2008;283(8):4535–42.
73. Oklejewicz M, Destici E, Tamanini F, Hut RA, Janssens R, van der Horst GTJ. Phase resetting of the mammalian circadian clock by DNA damage. *Curr Biol*. 2008;18(4):286–91.
74. Papp SJ, Huber A-L, Jordan SD, Kriebs A, Nguyen M, Moresco JJ, et al. DNA damage shifts circadian clock time via Hausp-dependent Cry1 stabilization. *Elife*. 2015;4:e04883.
75. Verlande A, Masri S. Circadian clocks and cancer: timekeeping governs cellular metabolism. *Trends Endocrinol Metab*. 2019;30(7):445–58.
76. Reinke H, Asher G. Crosstalk between metabolism and circadian clocks. *Nat Rev Mol Cell Biol*. 2019;20(4):227–41.
77. Petrova V, Annicchiarico-Petruzzelli M, Melino G, Amelio I. The hypoxic tumour microenvironment. *Oncogenesis*. 2018; 7(1):10.
78. Atashgaran V, Wrin J, Barry SC, Dasari P, Ingman WW. Dissecting the biology of menstrual cycle-associated breast cancer risk. *Front Oncol*. 2016;6:267.
79. Olsson HL, Olsson ML. The menstrual cycle and risk of breast cancer: a review. *Front Oncol*. 2020;10:21.
80. Henderson BE, Feigelson HS. Hormonal carcinogenesis. *Carcinogenesis*. 2000;21(3):427–33.
81. Miglioretti DL, Walker R, Weaver DL, Buist DSM, Taplin SH, Carney PA, et al. Accuracy of screening mammography varies by week of menstrual cycle. *Radiology*. 2011;258(2):372–9.
82. Siu AL. Screening for breast cancer: U.S. preventive services task force recommendation statement. *Ann Intern Med*. 2016;164(4):279–96.
83. Chen P, Hu P, Xie D, Qin Y, Wang F, Wang H. Meta-analysis of vitamin D, calcium and the prevention of breast cancer. *Breast Cancer Res Treat*. 2010;121(2):469–77.
84. Ferraroni M, La Vecchia C, D'Avanzo B, Negri E, Franceschi S, Decarli A. Selected micronutrient intake and the risk of colorectal cancer. *Br J Cancer*. 1994;70(6):1150–5.
85. Lappe JM, Travers-Gustafson D, Davies KM, Recker RR, Heaney RP. Vitamin D and calcium supplementation reduces cancer risk: results of a randomized trial. *Am J Clin Nutr*. 2007;85(6):1586–91.
86. Lipworth L, Rossi M, McLaughlin JK, Negri E, Talamini R, Levi F, et al. Dietary vitamin D and cancers of the oral cavity and esophagus. *Ann Oncol*. 2009;20(9):1576–81.
87. Ahn J, Peters U, Albanes D, Purdue MP, Abnet CC, Chatterjee N, et al. Serum vitamin D concentration and prostate cancer risk: a nested case-control study. *J Natl Cancer Inst*. 2008;100(11):796–804.
88. Roychoudhuri R, Robinson D, Coupland V, Holmberg L, Møller H. Season of cancer diagnosis exerts distinct effects upon short- and long-term survival. *Int J Cancer*. 2009;124(10):2436–41.
89. Moan J, Lagunova Z, Bruland Ø, Juzeniene A. Seasonal variations of cancer incidence and prognosis. *Dermatoendocrinol*. 2010;2(2):55–7.
90. Roué T, Nacher M. Epidemiology of cancer in the tropical areas. In: Droz JP, Carme B, Couppié P, Nacher M. TC (eds), editor. *Tropical Hemato-Oncology*. Springer, Cham; 2015. p. 17–23.
91. Dockery DW, Pope CA, Xu X, Spengler JD, Ware JH, Fay ME, et al. An association between air pollution and mortality in six U.S. cities. *N Engl J Med*. 1993;329(24):1753–9.
92. Chan S, Debono M. Review: Replication of cortisol circadian rhythm: new advances in hydrocortisone replacement therapy. *Ther Adv Endocrinol Metab*. 2010;1(3):129–38.
93. Krieger DT, Allen W, Rizzo F, Krieger HP. Characterization of the normal temporal pattern of plasma corticosteroid levels. *J Clin Endocrinol Metab*. 1971;32(2):266–84.
94. Weitzman ED, Fukushima D, Nogeire C, Roffwarg H, Gallagher TF, Hellman L. Twenty-four hour pattern of the episodic secretion of cortisol in normal subjects. *J Clin Endocrinol Metab*. 1971;33(1):14–22.
95. Besedovsky L, Lange T, Born J. Sleep and immune function. *Pflugers Arch*. 2012;463(1):121–37.
96. Youssef J, Novosad SA, Winthrop KL. Infection risk and safety of corticosteroid use. *Rheum Dis Clin N Am*. 2016;42(1): 157–76.
97. Tsao TC, Tsai YH, Lan RS, Shieh WB, Lee CH. Fever characteristics in tuberculosis—clinical observation in 597 cases. *Chang yi xue za zhi*. 1989;12(2):81–8.
98. Martinez AN, Mehra S, Kaushal D. Role of interleukin 6 in innate immunity to mycobacterium tuberculosis infection. *J Infect Dis*. 2013;207(8):1253–61.
99. Bethin KE, Vogt SK, Muglia LJ. Interleukin-6 is an essential, corticotropin-releasing hormone-independent stimulator of the adrenal axis during immune system activation. *Proc Natl Acad Sci U S A*. 2000;97(16):9317–22.
100. Nilsson G, Lekander M, Åkerstedt T, Axelsson J, Ingre M. Diurnal variation of circulating Interleukin-6 in humans: a meta-analysis. *PLoS One*. 2016;11(11):e0165799.
101. Foggo V, Cavenagh J. Malignant causes of fever of unknown origin. *Clin Med*. 2015;15(3):292–4.
102. Lee JW, Stone ML, Porrett PM, Thomas SK, Komar CA, Li JH, et al. Hepatocytes direct the formation of a pro-metastatic niche in the liver. *Nature*. 2019;567(7747):249–52.
103. Preti HA, Cabanillas F, Talpaz M, Tucker SL, Seymour JF, Kurzrock R. Prognostic value of serum interleukin-6 in diffuse large-cell lymphoma. *Ann Intern Med*. 1997;127(3):186–94.
104. Cortes JE, Talpaz M, Cabanillas F, Seymour JF, Kurzrock R. Serum levels of interleukin-10 in patients with diffuse large cell lymphoma: lack of correlation with prognosis. *Blood*. 1995;85(9):2516–20.
105. Blay JY, Negrier S, Combaret V, Attali S, Goillot E, Merrouche Y, et al. Serum level of interleukin 6 as a prognosis factor in metastatic renal cell carcinoma. *Cancer Res*. 1992;52(12):3317–22.
106. Zell JA, Chang JC. Neoplastic fever: a neglected paraneoplastic syndrome. *Support Care Cancer Off J Multinatl Assoc Support Care Cancer*. 2005;13(11):870–7.
107. Magnon C, Hall SJ, Lin J, Xue X, Gerber L, Freedland SJ, et al. Autonomic nerve development contributes to prostate cancer progression. *Science*. 2013;341(6142):1236361.
108. Amit M, Takahashi H, Dragomir MP, Lindemann A, Gleber-Netto FO, Pickering CR, et al. Loss of p53 drives neuron reprogramming in head and neck cancer. *Nature*. 2020;578(7795):449–54.
109. Wang X, Wang B, Xie J, Hou D, Zhang H, Huang H. Melatonin inhibits epithelial-to-mesenchymal transition in gastric cancer cells via attenuation of IL-1 β /NF- κ B/MMP2/MMP9 signaling. *Int J Mol Med*. 2018;42(4):2221–8.

110. Chao C-C, Chen P-C, Chiou P-C, Hsu C-J, Liu P-I, Yang Y-C, et al. Melatonin suppresses lung cancer metastasis by inhibition of epithelial-mesenchymal transition through targeting to Twist. *Clin Sci (Lond)*. 2019;133(5):709–22.
111. Wang J, Xiao X, Zhang Y, Shi D, Chen W, Fu L, et al. Simultaneous modulation of COX-2, p300, Akt, and Apaf-1 signaling by melatonin to inhibit proliferation and induce apoptosis in breast cancer cells. *J Pineal Res*. 2012;53(1):77–90.
112. Reiter RJ, Rosales-Corral SA, Tan D-X, Acuna-Castroviejo D, Qin L, Yang S-F, et al. Melatonin, a full service anti-cancer agent: inhibition of initiation, progression and metastasis. *Int J Mol Sci*. 2017;18(4):843.
113. Mao L, Summers W, Xiang S, Yuan L, Dauchy RT, Reynolds A, et al. Melatonin represses metastasis in Her2-positive human breast cancer cells by suppressing RSK2 expression. *Mol Cancer Res*. 2016;14(11):1159–69.
114. Golan K, Kumari A, Kollet O, Khatib-Massalha E, Subramaniam MD, Ferreira ZS, et al. Daily onset of light and darkness differentially controls hematopoietic stem cell differentiation and maintenance. *Cell Stem Cell*. 2018;23(4):572–585.e7.
115. Veillette A. Immune regulation by SLAM family receptors and SAP-related adaptors. *Nat Rev Immunol*. 2006;6(1):56–66.
116. Lange T, Dimitrov S, Bollinger T, Diekelmann S, Born J. Sleep after vaccination boosts immunological memory. *J Immunol*. 2011;187(1):283–90.
117. Dimitrov S, Benedict C, Heutling D, Westermann J, Born J, Lange T. Cortisol and epinephrine control opposing circadian rhythms in T cell subsets. *Blood*. 2009;113(21):5134–43.
118. He W, Holtkamp S, Hergenhan SM, Kraus K, de Juan A, Weber J, et al. Circadian expression of migratory factors establishes lineage-specific signatures that guide the homing of leukocyte subsets to tissues. *Immunity*. 2018;49(6):1175–1190.e7.
119. Müller A, Homey B, Soto H, Ge N, Catron D, Buchanan ME, et al. Involvement of chemokine receptors in breast cancer metastasis. *Nature*. 2001;410(6824):50–6.
120. Chatterjee S, Behnam Azad B, Nimmagadda S. The intricate role of CXCR4 in cancer. *Adv Cancer Res*. 2014;124:31–82.
121. Salgia R, Weaver RW, McCleod M, Stille JR, Yan SB, Roberson S, et al. Prognostic and predictive value of circulating tumor cells and CXCR4 expression as biomarkers for a CXCR4 peptide antagonist in combination with carboplatin-etoposide in small cell lung cancer: exploratory analysis of a phase II study. *Investig New Drugs*. 2017;35(3):334–44.
122. Alix-Panabières C, Brouillet J-P, Fabbro M, Yssel H, Rousset T, Maudelonde T, et al. Characterization and enumeration of cells secreting tumor markers in the peripheral blood of breast cancer patients. *J Immunol Methods*. 2005;299(1):177–88.
123. Szczerba BM, Castro-Giner F, Vetter M, Krol I, Gkoutela S, Landin J, et al. Neutrophils escort circulating tumour cells to enable cell cycle progression. *Nature*. 2019;566(7745):553–7.
124. Hadadi E, Taylor W, Li X-M, Aslan Y, Villote M, Rivière J, et al. Chronic circadian disruption modulates breast cancer stemness and immune microenvironment to drive metastasis in mice. *Nat Commun*. 2020;11(1):3193.
125. Deng W, Zhu S, Zeng L, Liu J, Kang R, Yang M, et al. The circadian clock controls immune checkpoint pathway in sepsis. *Cell Rep*. 2018;24(2):366–78.
126. Scheer FAJL, Michelson AD, Frelinger AL, Evoniuk H, Kelly EE, McCarthy M, et al. The human endogenous circadian system causes greatest platelet activation during the biological morning independent of behaviors. *PLoS One*. 2011;6(9):e24549.
127. Camerer E, Qazi AA, Duong DN, Cornelissen I, Advincula R, Coughlin SR. Platelets, protease-activated receptors, and fibrinogen in hematogenous metastasis. *Blood*. 2004;104(2):397–401.
128. Nieswandt B, Hafner M, Echtenacher B, Mannel DN. Lysis of tumor cells by natural killer cells in mice is impeded by platelets. *Cancer Res*. 1999;59(6):1295–300.
129. Jiang X, Wong KHK, Khankhel AH, Zeinali M, Reategui E, Phillips MJ, et al. Microfluidic isolation of platelet-covered circulating tumor cells. *Lab Chip*. 2017;17(20):3498–503.
130. Martín M, García-Sáenz JÁ, De Las Casas MLM, Vidaurreta M, Puente J, Veganzones S, et al. Circulating tumor cells in metastatic breast cancer: timing of blood extraction for analysis. *Anticancer Res*. 2009;29(10):4185–7.
131. García-Sáenz JÁ, Martín M, Maestro M, Vidaurreta M, Veganzones S, Villalobos L, et al. Circulating tumoral cells lack circadian-rhythm in hospitalized metastatic breast cancer patients. *Clin Transl Oncol*. 2006;8(11):826–9.
132. Nyquist H. Certain topics in telegraph transmission theory. *Trans Am Inst Electr Eng*. 1928;47(2):617–44.
133. Shannon CE. Communication in the presence of noise. *Proc IRE*. 1949;37(1):10–21.
134. Extra JM, Espie M, Calvo F, Ferme C, Mignot L, Marty M. Phase I study of oxaliplatin in patients with advanced cancer. *Cancer Chemother Pharmacol*. 1990;25(4):299–303.
135. Lévi F. Circadian chronotherapy for human cancers. *Lancet Oncol*. 2001;2(5):307–15.
136. Cederroth CR, Albrecht U, Bass J, Brown SA, Dyhrfeld-Johnsen J, Gachon F, et al. Medicine in the fourth dimension. *Cell Metab*. 2019;30(2):238–50.
137. Ujvari B, Gatenby RA, Thomas F. The evolutionary ecology of transmissible cancers. *Infect Genet Evol*. 2016;39:293–303.
138. Domazet-Lošo T, Tautz D. Phylostratigraphic tracking of cancer genes suggests a link to the emergence of multicellularity in metazoa. *BMC Biol*. 2010;8:66.
139. Arnal A, Ujvari B, Crespi B, Gatenby RA, Tissot T, Vittecoq M, et al. Evolutionary perspective of cancer: myth, metaphors, and reality. *Evol Appl*. 2015;8(6):541–4.
140. Tissot T, Massol F, Ujvari B, Alix-Panabières C, Loeuille N, Thomas F. Metastasis and the evolution of dispersal. *Proceedings Biol Sci*. 2019;286(1916):20192186.
141. Lloyd MC, Gatenby RA, Brown JS. Ecology of the metastatic process. In: *Ecology and evolution of Cancer*; 2017. p. 153–65.
142. Tissot T, Arnal A, Jacqueline C, Poulin R, Lefèvre T, Mery F, et al. Host manipulation by cancer cells: expectations, facts, and therapeutic implications. *BioEssays*. 2016;38(3):276–85.
143. Born J, Lange T, Hansen K, Mölle M, Fehm HL. Effects of sleep and circadian rhythm on human circulating immune cells. *J Immunol*. 1997;158(9):4454–64.
144. Fu L, Kettner NM. The circadian clock in cancer development and therapy. In: *Progress in molecular biology and translational science*; 2013. p. 221–82.
145. Hrushesky WJM, Lester B, Lannin D. Circadian coordination of cancer growth and metastatic spread. *Int J Cancer*. 1999; 83(3):365–73.
146. Proscenc ZU. Circadian rhythms and new options for novel anticancer therapies. *ChronoPhysiology Ther*. 2015;5:1–13.

147. Fodde R, Smits R, Clevers H. APC, signal transduction and genetic instability in colorectal cancer. *Nat Rev Cancer*. 2001; 1(1):55–67.
148. Yang C, Tang D. Patient-specific carotid plaque progression simulation. *C Model Eng Sci*. 2000;1(2):119–31.
149. Lowery A. Sleep and cancer. In: Press OU, editor. *Psycho-Oncology*. 2015. p. 231–8.
150. Walker WH, Borniger JC. Molecular mechanisms of cancer-induced sleep disruption. *Int J Mol Sci*. 2019;20(11):2780.
151. Ercolani L, Ferrari A, De Mei C, Parodi C, Wade M, Grimaldi B. Circadian clock: time for novel anticancer strategies? *Pharmacol Res*. 2015;100:288–95.
152. Lévi F, Okyar A, Dulong S, Innominato PF, Clairambault J. Circadian timing in cancer treatments. *Annu Rev Pharmacol Toxicol*. 2010;50(1):377–421.
153. Innominato PF, Roche VP, Palesh OG, Ulusakarya A, Spiegel D, Lévi FA. The circadian timing system in clinical oncology. *Ann Med*. 2014;46(4):191–207.
154. Hrushesky WJM. Circadian timing of cancer chemotherapy. *Science*. 1985;228(4695):73–5.
155. Dheilly NM. Holobiont-holobiont interactions: redefining host-parasite interactions. *PLoS Pathog*. 2014;10(7):1–4.
156. Prior KF, O'Donnell AJ, Rund SSC, Savill NJ, van der Veen DR, Reece SE. Host circadian rhythms are disrupted during malaria infection in parasite genotype-specific manners. *Sci Rep*. 2019;9(1):10905.
157. Patra G, Sarkar S. Nurse cell biology of *Trichinella spiralis*. *Int J Adv Res Technol*. 2014;3(4):133–8.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

