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Temporal Patterns of Abundance of *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) and Mitochondrial DNA Analysis of *Ae. albopictus* in the Central African Republic

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Abstract

The invasive Asian tiger mosquito Aedes albopictus (Diptera: Culicidae) was first reported in central Africa in 2000, in Cameroon, with the indigenous mosquito species Ae. aegypti (Diptera: Culicidae). Today, this invasive species is present in almost all countries of the region, including the Central African Republic (CAR), where it was first recorded in 2009. As invasive species of mosquitoes can affect the distribution of native species, resulting in new patterns of vectors and concomitant risk for disease, we undertook a comparative study early and late in the wet season in the capital and the main cities of CAR to document infestation and the ecological preferences of the two species. In addition, we determined the probable geographical origin of invasive populations of Ae. albopictus with two mitochondrial DNA genes, COI and ND5. Analysis revealed that Ae. aegypti was more abundant earlier in the wet season and Ae. albopictus in the late wet season. Used tyres were the most heavily colonized productive larval habitats for both species in both seasons. The invasive species Ae. albopictus predominated over the resident species at all sites in which the two species were sympatric. Mitochondrial DNA analysis revealed broad low genetic diversity, confirming recent introduction of Ae. albopictus in CAR. Phylogeographical analysis based on COI polymorphism indicated that the Ae. albopictus haplotype in the CAR population segregated into two lineages, suggesting multiple sources of Ae. albopictus. These data may have important implications for vector control strategies in central Africa.

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Introduction

Aedes aegypti Linneaus 1762 and Ae. albopictus Skuse 1894, two mosquitoes belonging to the Stegomyia subgenus, are the main epidemic vectors of dengue and chikungunya viruses worldwide [1,2,3,4]. Both species are established in sub-Saharan Africa, where Ae. aegypti is native [5]. Ae. albopictus originated in Asia [6] and has invaded Europe, the Americas and Africa during the past three decades. This rapid global spread was favoured by international trade, especially of used tyres [7], and by the differing physiology and ecology of many populations, which allows the species to thrive in a wide range of climates and habitats [8]. Since 2000, Ae. albopictus has invaded several central African countries, including Cameroon [9], Gabon [10], Equatorial Guinea [11] and the Central African Republic (CAR) [12], where it occurs in human-dominated environments previously colonized by Ae. aegypti. Recently, the density of Ae. albopictus has reached levels compatible with arbovirus transmission. Ae. albopictus is

suspected to have played a major role in the transmission of chikungunya virus in Cameroon in 2006 [13] and was shown to be the main vector of both chikungunya and dengue virus in Gabon in 2007 and 2010 [1,14,15]. In addition, *Ae. albopictus* populations in Cameroon were shown to be orally susceptible to dengue-2 virus and highly competent for chikungunya virus [1]. It is therefore likely that this invasive mosquito also played a significant role in the chikungunya outbreak in the Republic of Congo in 2011 [16].

Coexistence of *Ae. aegypti* and *Ae. albopictus* has been documented in several regions in the world, where the larvae sometimes share common developmental sites [17,18,19,20]. In areas of South America and South-East Asia where the two species are sympatric, they segregate into different habitats on the basis of environmental factors [17,21,22]. *Ae. aegypti* usually dominates in densely crowded urban areas, whereas *Ae. albopictus* dominates in suburban or rural areas. Nevertheless, *Ae. albopictus* can also colonize urban habitats, especially when *Ae. aegypti* is absent [23]. Overlap in the spatial

Author Summary

Aedes aegypti and Ae. albopictus are the main vectors of human arboviral diseases such as dengue and chikungunya. Ae. aegypti is indigenous in the Central African Republic (CAR), whereas Ae. Albopictus, originating from Asian forests, was first reported in 2009. To determine the consequences of this invasion of Ae. albopictus for epidemiological transmission of arboviruses, we conducted a comparative study in the early and late wet season in the capital, Bangui, and in the other main cities of the country to document infestation by the two species and their ecological preferences. In addition, we explored the geographical origin of populations of Ae. albopictus with two mitochondrial DNA genes (COI and ND5). We demonstrate that Ae. aegypti predominates early and Ae. albopictus late in the wet season. Ae. albopictus was the most prevalent species in almost all the sites investigated, except Bouar, where only Ae. aegypti was found, suggesting that Ae. albopictus tends to supplant Ae. aegypti in sympatric areas. Mitochondrial DNA analysis revealed broad low genetic diversity, confirming recent introduction of Ae. albopictus. Phylogeographical analysis with MtDNA COI gene suggested that Ae. albopictus in CAR came from multiple invasions and from multiple population sources

distribution of the two species is thought to result in competitive interaction. Displacement of *Ae. aegypti* after invasion by *Ae. albopictus* was documented in south-eastern USA and Brazil [24,25,26] and was suspected in Réunion and Mayotte [27,28,29]. Conversely, in Asia, *Ae. aegypti* has an overall competitive advantage over *Ae. albopictus*, especially in urban areas [16,30,31]. Although the outcome of competitive interactions between these two species has not yet been studied in Africa, studies in Cameroon showed that invasion by *Ae. albopictus* led to replacement of the native species *Ae. aegypti* in cities in which both species are present [18,20].

Several phylogeographical studies have been undertaken to determine the origin of invasive populations of *Ae. albopictus* with isoenzymatic and mitochondrial markers. Recent studies with mitochondrial markers challenged the hypothesis of a common origin of North and South American populations [32], and it was suggested that the Brazilian populations were related to South-East Asian rather than temperate Asian populations [33]. Other analyses based on *COI* polymorphism indicate that *Ae. albopictus* populations in Cameroon are related to tropical rather than temperate or subtropical out-groups [34].

Invasion of central Africa by Ae. albopictus genetically competent for dengue or chikungunya virus [1] and subsequent modification of Aedes populations might affect the epidemiology of these two viruses and lead to major outbreaks. The control of such diseases is based on entomological surveillance and vector control and requires extensive background information on the biology of the mosquito vectors involved. In addition, as the biological traits of mosquitoes are genetically determined [35] and as these traits may influence virus transmission in newly colonized areas, it is important to determine the geographical origin of invading populations. We undertook a study to assess the extent of infestation by Ae. aegypti and Ae. albopictus in Bangui, the main urban area of CAR, and nearby localities, focusing on larval habitats and spatial distribution. We also explored the phylogenetic relations between the Ae. albopictus populations colonizing CAR and out-group populations sampled worldwide.

Materials and Methods

Ethics statement

Institutional clearance for this study, including the sampling of mosquitoes, was approved by the national ethical and scientific committees in charge of validating study designs in CAR. For entomological investigation performed on private land or in private residences, all owners or residents gave permission for the study to be conducted.

Study sites

Mosquitoes were collected between April and November 2012 at seven localities in southern CAR: Mbaïki (3°52N,17°59E), Batalimo (3°40N, 18°27E), Mongoumba (3°38N, 18°35E), Boda (4°18N, 17°27E), Berberati (4°15N, 15°47E), Bouar (5°56N, 15°35E) and Bangui (04°21N, 18°33E) (Figure 1). The surveys were limited to this part of the country as it was the only part that was safe and accessible. The larval ecology of Ae. aegypti and Ae. albopictus was characterized in Bangui, the capital, with a population of about 900 000. The city is located on the right bank of the Ubangi River, which forms the border between CAR and the Democratic Republic of the Congo. Bangui comprises two blocks: the centre is modern, with urban buildings from the preindependence period, while the suburbs are unplanned and sparsely populated. The climate is of the Guinean forest type, with alternation of two seasons: a rainy season from March to mid-December and a dry season from mid-December to February. The average annual rainfall is 1543 mm, and the minimum and maximum temperatures are around 15°C and 38°C, respectively.

Sampling and entomological surveys of immature stages

We undertook ecological characterization of Ae. aegypti and Ae. albopictus in Bangui and assessed the current spatial distributions of the two species in the southern part of the country. In Bangui, entomological surveys were carried out twice, in April and October 2012, corresponding to early and later in the wet season, respectively. Surveys were undertaken in clusters of houses sampled randomly, each cluster consisting of 10 houses per quarter in each of eight boroughs. In the field surveys, each selected house was geo-referenced with a GPS and visited to record all natural and artificial containers of water (potential containers) and those containing immature stages (larvae and pupae) of Ae. aegypti and Ae. albopictus (positive containers). Whenever they were present, immature stages were collected for further counting and identification in the insectarium at the Institut Pasteur of Bangui. Positive larval development sites were also geo-referenced, and the type of container, the container volume, the volume, source, use and quality (clear, tinted, organic matter) of water, the presence of plant debris inside the container, the presence of vegetation around the container and sun exposure were noted, with the number of inhabitants per house. On the basis of the nature, the source and the use of the water, potential containers were classified into domestic, peri-domestic and natural. Domestic containers were defined as human-filled receptacles, whereas peri-domestic (e.g. discarded containers) and natural receptacles (e.g. rock and tree holes, leaf axils, empty shells and nuts) were those filled by rain. Larvae and pupae were returned to the insectaries and isolated from predators such as Culex (Lutzia) tigripes larvae, counted (larvae L3-4 and pupae), reared to adults and then identified from morphological identification keys [36,37]. The number of immature stages of each species was estimated from the proportion of emerging adults of each species.



Figure 1. Location of mosquito sampling sites in the Central African Republic. doi:10.1371/journal.pntd.0002590.g001

At sites other than Bangui, the surveys were undertaken only in the late wet season. Entomological investigation consisted of a complete inventory of potential larval breeding sites (natural, peridomestic and domestic) and positive sites (with at least one *Aedes* larvae or pupae). Immature stages were collected from positive sites, recorded, transported in insectaries and reared to adult stage for identification.

Mosquitoes identified as *Ae. albopictus* were stored in individual tubes containing a desiccant at -20° C for further molecular analysis.

Entomological indexes

The level of *Ae. aegypti* and *Ae. albopictus* infestation was assessed from standard indexes based on immature stages, including the house index (percentage of houses positive for larvae and/or pupae) and the Breteau index (number of positive containers per 100 houses). Additional indexes based on the presence or absence and the number of larvae or pupae were also used, including the larvae index (number of pupae e per 100 houses) and the pupae index (number of pupae per 100 houses) [20]. The productivity of a container type was defined as the number of L3-4 or pupae in each divided by the total number of L3-4 or pupae in all container types [38]. The larvae (L3-4) per person index and the pupae per person index were also estimated [39].

Statistical analysis

All statistical analyses were performed in STATA version 11 (StataCorp College Station, Texas 77845). The distribution of

each variable was observed. The type of container, water turbidity, the presence of vegetal debris inside the container, the presence of vegetation around the container, sun exposure and the presence of any immature stage of Ae. albopictus and Ae. aegypti were defined as categorical variables and expressed as percentages. The effect of each variable on the presence of vectors was examined in the chi-square or Fisher exact test. Numerical variables (container volume, volume of water inside the container, number of L3-4 and pupae) were described as means and standard deviations and compared in the Student t test or the Kruskal-Wallis test when the Student t test was not appropriate. Contingency tables were generated and the relation between container characteristic and presence or absence of L3-4 and pupae (immature stage) of Ae. aegypti or Ae albopictus was analysed using chi-square (or Fisher exact test if appropriate). A p value <0.05 was considered significant. In a second step, the presence or absence of immature stages was analysed by binary logistic regression with a conditional backwards stepwise procedure. The potential predictors tested corresponded to the main larval habitat characteristics described above. A test of correlation was also performed to determine the relations between numbers of L3-4 and pupae of Ae. aegypti or Ae. albopictus and certain breeding site characteristics, such as the container volume, volume of water inside the container, distance to the nearest building and distance of the container to plants. The GPS coordinates of houses surveyed and positive larval habitats of the two species were projected onto maps with ArcGis software (ArcGis®9.2, ESRI).

		Early wet se	ason					Late wet sea	tson			
Species	House index	Breteau index	Larvae index	Pupae index	Larvae per person index	Pupae per person index	House index	Breteau index	Larvae index	Pupae index	Larvae per person index	Pupae per person index
Ae. aegypti	9.03	14.4	768.6	99.1	0.7	0.1	21.8	16.5	913.4	84.4	0.8	0.08
Ae. albopictus	7.3	12.7	437.8	52.2	0.4	0.05	21.8	16.2	1381.8	120.7	1.3	0.1
d	0.8	0.7	<10 ⁻³	<10 ⁻³	<10 ⁻³	<10 ⁻³	-	0.9	<10 ⁻³	$< 10^{-3}$	<10 ⁻³	<10 ⁻³

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Table

	Early wet season					Late wet season				
Type of container	n inspected	% positive	% Ae. aegypti only	% Ae. albopictus only	% mixed	n inspected	% positive	% Ae. aegypti only	% Ae. albopictus only	% mixed
	n=176	n=52	n = 7	n=1	n = 44	n = 209	n = 97	n=11	n=12	n= 74
Domestic	31	17.3	14.3	0.0	18.2	18	8.2	0.0	8.3	9.4
Watering place*	9	0.0	0.0	0.0	0.0	5	1.0	0.0	0.0	1.4
Water storage	11	0.0	0.0	0.0	0.0	S	1.0	0.0	0.0	1.4
Flower pots	14	17.3	14.3	0.0	18.2	8	6.2	0.0	8.3	6.7
Peri-domestic	143	80.8	71.4	100.0	81.8	189	90.8	100.0	91.7	89.2
Used tyres	49	51.9	42.9	100.0	50.0	79	46.4	45.5	25.0	50.0
Discarded tanks	63	28.8	28.6	0.0	31.8	82	30.0	45.5	50.0	24.3
Miscellaneous	31	0.0	0.0	0.0	0.0	28	14.4	9.0	16.6	14.9
Natural	2	1.9	14.3	0.0	0.0	2	1.0	0.0	0.0	1.4
n inspected, number of poter containers containing only Ae *Container used to give drinki doi:10.1371/journal.pritd.0002:	ntial containers inspecte <i>: albopictus;</i> mixed, cor ing-water to pets. 590.t002	ed; % positive, per ntainers infested w	centage of contair <i>i</i> th at least one lar	ners infested with at le va or pupa of each sp	east one larva or becies;	r pupa of one species;	Ae. aegypti only, c	ontainers containii	ıg only Ae. aegypti; A	e. albopictus only,

Mitochondrial DNA analysis for Ae. albopictus

Sequence polymorphisms in the mitochondrial genes encoding for the nicotinamide adenine dinucleotide dehydrogenase subunit 5 (ND5) and for cytochrome oxidase I (COI) were explored in 95 individual Ae. albopictus mosquitoes from CAR. We included in the analysis 22 specimens of Ae. albopictus from Franceville (1°37'S, 13°34'E) and Dienga (1°52'S, 12°40'E) in Gabon, collected at the larval stage in June 2013. DNA extraction and PCR amplification were done as described previously [34]. Mosquito DNA extracts were used as templates to amplify a 400-bp fragment of ND5 and a 550-bp fragment of COI. PCR products were purified and sent to GATC Biotech (Konstanz, Germany) for sequencing. Sequences were cleaned, when necessary, with SEQSCAPE software 2.5 (Applied Biosystems) and aligned with Clustal W [40]. ND5 and COI sequences were numbered according to the reference sequences GeneBank ID JF309321 and JF309317, respectively. Basic sequence statistics, including the number of haplotypes per sample, the number of segregating sites (S), haplotype diversity, nucleotide diversity (π) and the average number of nucleotide differences, were computed with DnaSP 4.10.9 [41]. The statistical tests of Tajima [42], Fu and Li [43] and Fu [44] were used with DnaSP to test non-neutral evolution and deviation from mutation-drift equilibrium. The phylogenetic relations between COI and ND5 haplotypes recorded in CAR and previously published sequences (Table S1) of Ae. albopictus from Asia, the Americas, the Indian Ocean, Europe and central Africa were explored by Bayesian inference analysis. MrModeltest v2.2 [45] was first used to select the model that best fit the ND5 and COI nucleotide sequence data (under Akaike's information criterion). Analyses were performed with MrBayes 3.1.2 [46], and four Markov chains were run for 200 000 generations (sampling every 10 generations) to allow adequate time for convergence. The first 50 000 resulting trees were discarded as burn-in, and the remaining 150 000 sampled trees were used to estimate the 50% majority rule consensus tree and the Bayesian posterior probabilities. All Markov chain Monte Carlo runs were repeated twice to confirm consistent approximation of the posterior parameter distributions.

Results

Pre-imaginal infestation

We investigated 354 houses in 34 clusters or quarters in Bangui, with 3855 inhabitants. Of 176 potential larval development sites investigated early in the wet season, 52 (29.5%) contained immature stages of *Ae. aegypti* and/or *Ae. albopictus*. Late in the wet season, 97 of 209 potential larval habitats surveyed (46.4%) were positive. Several other mosquito species were found with *Ae. aegypti* and *Ae. albopictus* at both surveys: *Anopheles gambiae* s.l. Giles 1902, *Culex quinquefasciatus* Say 1823, *Culex perfuscus* Edwards 1914, *Culex tigripes* De Grandpré & De Charmoy 1900.

Early in the wet season, all the larval infestation indexes calculated for *Ae. aegypti* were significantly higher $(p < 10^{-3}, \text{ chi-square test})$ than those for *Ae. albopictus*, except the house and Breteau indexes, for which no significant difference was found. In contrast, higher infestation with *Ae. albopictus* was observed late in



Figure 2. Total abundance of immature stages of *Aedes aegypti* **and** *Ae. albopictus* **per container.** Each two-letter abbreviation on the x-axis corresponds to a type of container as follows: WS, water storage; FP, flower pot; WP, watering place; UT, used tyres; DT, discarded tanks; MI, miscellaneous; NA, natural. doi:10.1371/journal.pntd.0002590.q002

Table 3. Container characteristics associated with the presence of immature stages of Ae. albopictus and Ae. aegypti in Bangui.

	Early we	et season						Late wet	season					
		Aedes albo	pictus		Aedes a	egypti			Aedes albu	opictus		Aedes ae	egypti	
Category	Number	%	Univariate, OR (CI 95%)	Multivariate, OR (CI 95%)	%	Univariate, OR (CI 95%)	Multivariate, OR (CI 95%)	Number	%	Univariate, OR (CI 95%)	Multivariate, OR (CI 95%)	%	Univariate, OR (CI 95%)	Multivariate, OR (CI 95%)
Type of cont	niner													
Water storage	11	0	Reference	Reference	0	Reference	Reference	Ŋ	20	Reference	Reference	20	Reference	NA
Flower pot	14	57.1	5.8 (2.6–12.7)*	* 6.1 (1.6–23.6)*	64.3	6.3 (2.9–13.6)*	9.1 (2.4–34.9)*	80	75	2.1 (1.1–3.7)*	0.5 (0.07–3.8)	62.5	3.8 (0.8–16.8)	NA
Watering place	9	0	Reference	Reference	0	Reference	Reference	5	20	Reference	Reference	20	Reference	Reference
Used tires	49	44.9	9.4 (2.8–31.2)*	* 3.9 (1.6–9.8)*	51	10.9 (3.2–36.7)	* 4.7 (1.9–11.6)*	79	49.4	6.4(1.2-33.1)*	0.3 (0.09–0.8)	48.1	2.1 (1.2–3.8)*	0.3 (0.1–1.0)
Discarded tank	s 63	22.2	Reference	Reference	23.8	Reference	Reference	85	29.4	Reference	Reference	28.2	Reference	Reference
Miscellaneous	31	0	Reference	Reference	0	Reference	Reference	25	44	Reference	Reference	40	Reference	Reference
Natural	2	0	Reference	Reference	50	Reference	Reference	2	50	Reference	Reference	50	Reference	Reference
Sun exposure	A :													
Yes	82	13.4	Reference	Reference	14.6	Reference	Reference	44	45.5	1.3 (0.9–2.8)	NA	45.5	1.4 (0.7–2.8)	NA
No	94	35.1	3.5 (1.6–7.5)*	0.5 (0.2–1.3)	40.4	3.9 (1.9–8.3)*	0.3 (0.1–0.9)	165	38.8	Reference	NA	36.4	Reference	NA
Nature of wa	ter													
Clear	133	22.6	Reference	NA	24.1	Reference	Reference	170	40.6	Reference	NA	36.5	Reference	NA
Turbid	44	31.8	0.6 (0.3–1.3)	NA	40.9	2.2 (1.1–4.5)*	1.4 (0.5–3.4)	37	37.8	0.9 (0.4–1.8)	NA	45.9	1.5 (0.7–3.0)	NA
Polluted	6	0	NA	NA	0	NA	NA	11	45.5	1.2 (0.3–4.1)	NA	45.5	NA	NA
Plant debris	inside the (container												
Yes	87	46	18.1 (6.1–53.7)* 8.9 (2.7–29.5)*	49.4	11.4 (4.7–27.6)	* 4.1 (1.5–11.3)*	117	45.3	1.6 (0.9–2.8)	NA	45.3	2.0 (1.1–3.5)	1.5 (0.8–3.0)
No	89	4.5	Reference	Reference	7.9	Reference	Reference	92	33.7	Reference	NA	29.3	Reference	Reference
Vegetation a	round the	container												
Yes	64	40.6	3.6 (1.7–7.3)*	2.0 (0.8–4.8)	48.4	4.6 (2.3–9.2)*	3.3 (1.4–8.0)*	80	67.5	6.8 (3.7–12.7)*	16.7 (5.5–51.0)*	63.8	6.0 (3.3-11.2)*	12.7 (4.5–36.0)*
No	112	16.1	Reference	Reference	17	Reference	Reference	129	23.3	Reference	Reference	22.5	Reference	Reference
OR, odds ratio; *significant ass NA, not applic; doi:10.1371/jou	Cl, confider ociation; ible; Referen rnal.pntd.00	nce interval; nce, 'comparat 102590.t003	or group' for es	timating the OR.										



Legend= • Inspected houses, Positive breeding sites, Proportion of Ae. aegypti, Proportion of Ae. albopictus.

Figure 3. Spatial distribution of surveyed houses and positive larval habitats of *Aedes* spp. in Bangui. The surveys were conducted during the early wet season (A) and the late wet season (B). doi:10.1371/journal.pntd.0002590.q003

the wet season for all indexes except the house and Breteau indexes (Table 1). The proportion of containers infested by *Ae. albopictus* only was significantly higher late rather than early in the wet season (p<0.05, Fisher exact test), whereas no significant difference was found in the proportion of containers infested by *Ae. aegypti* only early and late in the wet season (Table 2). The proportion of containers infested by *Ae. albopictus* and by *Ae. albopictus* with or without *Ae. albopictus* and by *Ae. albopictus* with or without *Ae. aegypti* were 98.5% and 86.5% in the early wet season and 87.6% and 88.6% in the late wet season, respectively (data not shown). No statistically significant difference in the proportions of containers infested by *Ae. albopictus* with or without *Ae. albopictus* with or without *Ae. albopictus* with or without *Ae. aegypti* with or without *Ae. albopictus* with or without *Ae. aegypti* with or without *Ae. aegypti* was found in any collection period, suggesting that infestation of containers by these species is comparable, irrespective of the season.

Container occupancy by Ae. aegypti and Ae. albopictus

During entomological surveys in both periods, all three defined categories of container were found: domestic (watering place, water storage and flower pots), peri-domestic (used tyres, discarded tanks, miscellaneous) and natural containers (leaf axils of *Colocasia* spp. taro plants). Peri-domestic containers represented the main infested container type in both periods, with a prevalence of infestation of 80.8% and 90.8%, respectively (Table 2). The most productive containers for both species during the two periods of investigation were used tyres, although the distribution of larvae (L3–4) was over-dispersed early in the wet season (Figure 2). The domestic containers were more likely to contain larvae early (17.3%) than late in the wet season

(8.2%) ($p{<}0.05,$ chi-square test), flower pots being the most productive domestic containers.

We used a binary logistic regression model to test the association between container characteristics and the presence of immature stages of *Ae. aegypti* and *Ae. albopictus*. Multivariate analyses showed that early in the wet season the presence of the two species was significantly associated with the type of container (used tyres or flower pots), the presence of plant debris inside the container and the presence of vegetation in the vicinity of containers (for *Ae. aegypti* only), whereas late in the wet season, only the presence of vegetation around the potential containers was significantly associated with the presence of the two species (Table 3).

We also explored the correlation between numbers of larvae and pupae of *Ae. aegypti* and *Ae. albopictus* and breeding site characteristics, such as distance of a container from a building and from plants, container volume and water volume. Early in the wet season, the distance of a container from plants was significantly inversely correlated with the number of larvae of both species (correlation coefficient (r) = -0.15, p<0.05 for *Ae. aegypti*; r = -0.18, p<0.02 for *Ae. albopictus*) and the number of pupae (r = -0.20, p<0.01 for *Ae. aegypti*; r = -0.20, p<0.01 for *Ae. albopictus*), whereas late in the wet season no significant correlation was found between container characteristics and productivity.

Spatial distribution of immature stage of *Aedes* spp.

In the 52 positive larval habitats identified early in the wet season, 3556 specimens of immature stages of *Aedes* spp. were

Table 4. Immature	mosquito sampli	ng in southern Ce	entral African Republ	lic.					
Locality	Period of sampling	Type of container inspected	Containers inspected	Positive containers	Container with <i>Ae.</i> <i>aegypti</i> only	Container with <i>Ae.</i> <i>albopictus</i> only	Mixed	Number of <i>Ae.</i> <i>albopictus</i> identified (%)	Number of <i>Ae.</i> <i>aegypti</i> identified (%)
Mbaïki	October 2012	Used tyres, discardec tanks	1 18	7	0	9	-	256 (96.6)	9 (3.4)
Batalimo	October 2012	Used tyres, discardec tanks, tree holes	1 8	9	-	m	2	156 (98.1)	3 (1.9)
Mongoumba	October 2012	Watering place, discarded tanks	13	4	0	0	2	25 (89.3)	3 (10.7)
Boda	November 2012	Used tyres, earthen j	ar11	5	0	4	, -	86 (98.9)	1 (1.1)
Berberati	November 2012	Car wrecks, discarded tanks, tin cans	d 23	9	0	0	9	266 (66.8)	132 (33.2)
Bouar	November 2012	Used tyres	11	6	6	0	0	0	511 (100)
doi:10.1371/journal.pntd.0	002590.t004								

Table 5. MtDNA *COI* and *ND5* haplotypes recorded in *Ae. albopictus* in the Central African Republic.

	ND5			<i>COI</i>	
		223			333
		068			2046
Haplotype*	Frequency	885	Haplotype	Frequency	4656
Ref. [JF309321]		ΑΤΤ	Ref. [JF309317]		Т G T C
H1 [KC979137]	103		H1 [KC979140]	86	
H2 [KC979138]	5	G . A	H2 [KC979141]	4	С
H3 [KC979139]	5	. C A	H3 [KC979142]	1	. A
			H4 [KC979143]	1	. A A T

Only polymorphic positions are shown and are numbered with reference (Ref) to the published *Ae. albopictus* sequences for *ND5* (JF309321; Cameroon) and *COI* (JF309317; Cameroon). Dots represent identity with respect to the reference

*GenBank accession number in brackets.

Frequency, number of times the haplotype was found in the total sample. doi:10.1371/journal.pntd.0002590.t005

identified, 60% of which were *Ae. aegypti* and 40% *Ae. albopictus*. In contrast, late in the wet season, 4250 specimens of *Aedes* spp. were recorded in 97 positive containers, of which 36% corresponded to *Ae. aegypti* and 64% to *Ae. albopictus*. These data suggest that *Ae. aegypti* is more prevalent early in the wet season and *Ae. albopictus* late in the wet season. The spatial distribution (Figure 3) of the two species showed that *Ae. aegypti* mosquitoes occur throughout Bangui early in the wet season, whereas *Ae. albopictus* is present in almost all environments late in the wet season, suggesting efficient expansion of this species, which appeared to be most prevalent in suburban areas. No trend in segregation of the species according to unplanned or planned environment was found (p>0.05, data not shown).

Distribution of *Ae. aegypti* and *Ae. albopictus* in southern CAR

Surveys in six additional locations in southern CAR during the late wet season showed that the two species coexisted and often shared the same larval habitats, except at Bouar, where *Ae. aegypti* was found alone. *Ae. albopictus* was the more prevalent in all localities in which both species were found (Table 4).

Mitochondrial DNA analysis of Ae. albopictus

Nucleotide sequences of the mtDNA *ND5* gene were retrieved from 91 specimens originating from six localities in CAR. Complete overlap of all the fragments spanned 399 nucleotides, of which two were polymorphic, defining three haplotypes, resulting in low haplotype and nucleotide indexes. The most frequent haplotype, H1 (89%), was detected in all geographical samples (Tables 5 and 6).

Sequences of the mtDNA *COI* gene were obtained from 70 specimens. Complete overlap of all fragments spanned 426 nucleotides, of which four were polymorphic (overall nucleotide diversity, $\pi = 0.0005$), defining four distinct haplotypes. Haplotype I dominated (91%) and was encountered in all localities (Tables 5 and 6). Sequences of mtDNA from the *COI* and *ND5* genes were also retrieved from 22 *Ae. albopictus* specimens from two localities in Gabon. The analysis revealed the existence of only one haplotype

Table 6. Summary statistics for mtDNA gene polymorphism in Ae. albopictus in the Central African Republic.

Locality	N	Mt gene	Нр	s	HpD	π	к	D	D*	F*	Fs
Berberati	12	ND5	H1, H2, H3	2	0.53	0.0014	0.57	-0.38	-0.37	-4.42	-0.36
	10	COI	H1, H2	2	0.35	0.0008	0.35	0.01	0.80	0.68	0.41
Boda	6	ND5	H1	0	0.00	0.0000	NC	NC	NC	NC	NC
	6	COI	H1	0	0.00	0.0000	NC	NC	NC	NC	NC
Mongoumba	8	ND5	H1	0	0.00	0.0000	NC	NC	NC	NC	NC
	6	COI	H1	0	0.00	0.0000	NC	NC	NC	NC	NC
Batalimo	22	ND5	H1, H3	1	0.24	0.0008	0.24	-0.17	0.63	0.47	0.30
	10	COI	H1, H2, H3	2	0.37	0.0009	0.40	-1.40	-1.58	-1.71	-1.16
Mbaïki	10	ND5	H1, H2	1	0.35	0.0008	0.35	0.01	0.80	0.68	0.417
	9	COI	H1, H2	1	0.22	0.0005	0.22	-1.08	-1.18	-1.28	-0.26
Bangui	33	ND5	H1, H3	1	0.06	0.0000	0.06	-1.14	-1.71	-1.78	-1.29
	29	COI	H1, H4	3	0.07	0.0005	0.26	-1.73	-2.66*	-2.77*	0.16
Overall	91	ND5	H1, H2, H3	2	0.20	0.0005	0.21	-0.73	0.69	0.29	-1.01
	70	COI	H1, H2, H3, H4	4	0.16	0.0005	0.22	-1.54	-1.33	-1.64	-2.51

N, number of sequences analysed; Hp, number of haplotypes; S, number of segregating sites; HpD, haplotype diversity; π , nucleotide diversity; K, average number of nucleotide differences; D, Tajima statistic; D* and F*, Fu and Li statistics; Fs, Fu statistic; NC, not computed; *p < 0.05.

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(H1) for each gene (Table 5). The sequence of the main CAR and Gabon *ND5* (H1) and *COI* (H1) haplotypes perfectly matched the dominant haplotypes found in Cameroonian *Ae. albopictus* samples from a database (Table 5). When all the sequences were analysed as a unique sample, the Tajima D, Fu and Li F* and D*, and Fu Fs statistics for the *COI* gene were negative but not statistically significant so (Table 6). Negative values for these indexes indicate an excess of rare polymorphisms in a population and suggest either population expansion or background selection [43].

In order to determine the geographical origin of the *Ae. albopictus* populations that are invading CAR, the phylogenetic relations between *COI* and *ND5* sequences recorded in CAR and previously published sequences were assessed by Bayesian inference. The *COI* sequences segregated into two lineages (Figure 4). The first encompassed specimens from tropical areas (Brazil, Cambodia, India, Thailand and Viet Nam), including all the Cameroonian haplotypes and two haplotypes from CAR (H1-CAR and H2-CAR). The second lineage encompassed temperate and subtropical areas (France, Greece, Madagascar, Reunion and the USA) and, surprisingly, two haplotypes from CAR (H3-CAR and H4-CAR). All the sequences were monophyletic at *ND5*.

Discussion

This detailed study suggests that *Ae. aegypti* is most prevalent in the early wet season and *Ae. albopictus* in the late wet season. Used tyres were the most productive container for both species, independently of season. In the survey across southern CAR in the late wet season, *Ae. albopictus* was the dominant species at all sites except Bouar.

In Bangui, we found significant differences in infestation rates by larval *Ae. aegypti* and *Ae. albopictus* according to season. The *Ae. aegypti* indexes were significantly higher than those for *Ae. albopictus* in the early wet season, with the opposite situation in the late wet season, suggesting lower abundance of *Ae. albopictus* in the early and higher abundance in the late wet season. These findings are consistent with those of studies in southern Florida, USA [47]. Although both species have desiccant-resistant eggs, Juliano et al. [48] showed that *Ae. aegypti* eggs are more tolerant to high temperatures than those of *Ae. albopictus*. This would explain why resident *Ae. aegypti* is more prevalent than invasive *Ae. albopictus* in the early wet season (i.e. the warmer season) in locations where the two species are sympatric.

The larvae of both species preferentially colonized peridomestic containers, especially used tyres and discarded tanks, irrespective of the collection period. In agreement with observations made in Cameroon [9,18,20], peri-domestic containers represented the bulk of the containers infested by Ae. aegypti or Ae. albopictus, thus differing from the situation in other parts of the world, particularly in Asia, where domestic containers such as water storage tanks were most commonly infested with Ae. aegypti [38,49]. In many sub-Saharan towns, unplanned urbanization and lack of waste management lead to widespread water collection, thus favouring the proliferation of *Aedes* spp. The two species studied here breed in the same type of container, with a preference for used tyres, flower pots, containers with plant debris and vegetation surrounding the container early in the wet season; late in the wet season, only larval habitats surrounded by vegetation were significantly associated with the presence of immature stages of Aedes spp. Micro-environmental factors therefore affect the presence of larval stages in breeding sites. In addition, used tyres were found to be the most productive containers for larvae and pupae in both sampling periods. Both species are native to the forest and breed mainly in natural tree holes, which share the characteristics of tyres, as the dark colour and the dark interior provides an attractive resting or oviposition site for Aedes spp. The presence of plant debris inside a larval habitat can serve as a food source or a micro-habitat to hide and avoid predators [50]. Surrounding vegetation can provide shade to reduce the water temperature of the larval habitat [51]. The association of the two species with the same micro-environmental conditions suggests that the invasive species, Ae. albopictus, shares the ecological niche of the resident species, Ae. aegypti. Competition for resources will, however, lead to segregation of habitats according to macroenvironmental variations such as urban environmental gradients,





0.03

Figure 4. Bayesian inference hypothesis of *Ae. albopictus* **phylogeny based on** *COI* (**A**) **and** *ND5* (**B**) **sequence data.** The phylogeny was constructed with MrBayes 3.1.2, ngen = 2 000 000. Best-fitting models selected with the MR model test (under AIC) were HKY for *COI* and HKY+I+G for the *ND5* nucleotide datasets. Branch support is indicated by the posterior probability values. Accession numbers of *COI* and *ND5* out-group sequences are given in supporting information file Table S1. doi:10.1371/journal.pntd.0002590.g004

as shown by other authors [7,21,51], or reduction of abundance of the indigenous species [25,26]. In addition, recent work shows that the two species are able to mate in nature and that *Ae. albopictus* males effectively sterilize *Ae. aegypti* females [52,53]. The authors suggest that this form of mating interference, called satyrization, could explain the competitive displacement of resident *Ae. aegypti* by the invasive *Ae. albopictus* where they co-occur.

The invasive species *Ae. albopitus* was more prevalent in all the sites investigated, except in Bouar, where only *Ae. aegypti* was found. This suggests rapid spread and good adaptation of *Ae. albopictus* in CAR. Previous surveys reported the presence of this species only in Bangui and Bayanga [12,54], at lower proportions (container index below 5) than observed in this study. The low density of *Ae. albopictus* reported in 2010 prompted Diallo et al. [12] to propose that its introduction is recent, probably through migratory flow and trade between the CAR and neighbouring countries, especially Cameroon, where the species was recorded for the first time in central Africa in 2000 [9]. Bouar is located near Cameroon at 6°N latitude, beyond which *Ae. albopictus* has not been found. This observation is consistent with studies in Cameroon, which suggest that the northern limit of *Ae. albopictus* invasion in Africa is around 6°N [18,20].

The higher prevalence of *Ae. albopictus* at all the sites investigated is in agreement with the findings of studies in other central African countries (Cameroon and Gabon), which suggest a dominance of the invasive species over the indigenous species in sites where the two species co-exist [1,14,18,20]. A decrease in indigenous *Ae. aegypti* after invasion by *Ae. albopictus* was also suspected in several localities in the Indian Ocean, such as Mayotte and Reunion [27,28,29]. In addition, invasive species have a competitive advantage over native species or first established invasive species, as observed in Brazil and south-eastern USA, where established invasive *Ae. aegypti* were displaced by recently invading *Ae. albopictus* [25,26], and in Asia, where *Ae. aegypti* has an overall competitive advantage over *Ae. albopictus*, especially in urban areas [31,55].

MtDNA markers have been used extensively to assess the genetic diversity of *Ae. albopictus* populations across most of its geographical range. The degree of polymorphism found in *ND5* and *COI* sequences in this study was low (three haplotypes for *ND5* and four for *COI*), consistent with previous studies of populations sampled in newly invaded areas [32,33,34,56,57], in which the number of haplotypes per country never exceeded five, regardless of the mtDNA marker used (*ND5, COI* or *Cytb*). In CAR, the low overall mtDNA diversity is consistent with recent introduction of a few founder females, as suggested by Diallo et al. [12], or may be related to ubiquitous *Wolbachia* infection in populations of this species, as suggested by Armbruster et al. [58]. Analyses of *COI* sequences revealed that central African *Ae. albopictus* are partly related to a tropical lineage (H1-CAR and H2-CAR) and partly to a temperate or subtropical lineage (H3-CAR and H4-CAR),

References

- Paupy C, Ollomo B, Kamgang B, Moutailler S, Rousset D, et al. (2010) Comparative role of *Aedes albopictus* and *Aedes aegypti* in the emergence of dengue and chikungunya in central Africa. Vector Borne Zoonotic Dis 10: 259–266.
- Kow CY, Koon LL, Yin PF (2001) Detection of dengue viruses in field caught male *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) in Singapore by typespecific PCR. J Med Entomol 38: 475–479.
- Gubler DJ (2002) The global emergence/resurgence of arboviral diseases as public health problems. Arch Med Res 33: 330–342.
- Reiter P (2010) Yellow fever and dengue: a threat to Europe? Euro Surveill 15: 19509.
- Mattingly PF (1967) Taxonomy of *Aedes aegypti* and related species. Bull World Health Organ 36: 552–554.
- Gratz NG (2004) Critical review of the vector status of *Aedes albopictus*. Med Vet Entomol 18: 215–227.

although H3 and H4 for COI are represented by only one specimen each. These results suggest that the populations present in CAR are derived from multiple invasions and multiple population sources. It is likely that Cameroon, which shares a border with CAR, was the main source of the invasion. Nevertheless, our previous study in Cameroon indicated that *Ae. albopictus* is related only to a tropical lineage, such as H1-CAR and H2-CAR haplotypes, suggesting that haplotypes H3-CAR and H4-CAR were introduced independently, from a temperate or a subtropical source, and make a minor contribution to the invasion in CAR. As CAR is landlocked, with no direct access to the sea, introduction of this species could have been by air with the transport of logistical equipment by foreign armed forces or the ubiquitous nongovernmental organizations.

The high infestation indexes of both species (particularly of *Ae. albopictus*) suggest an imminent risk for large outbreaks of arbovirus infections such as dengue and chikungunya in CAR, as in Cameroon in 2006 [13] and Gabon in 2007 [59] and 2010 [14], where the species was identified as or suspected to be the main vector. *Ae. albopictus* was also suspected of being responsible for transmission of chikungunya virus during the large outbreak in the Republic of Congo in 2011 [16].

As the dynamics of epidemics are correlated with the seasonal dynamics of vector populations [60], additional sampling, covering additional locations and spanning several seasons, would be beneficial. Nevertheless, our data on the spatial distribution, container type and productivity of larval development sites provide a useful basis for planning vector control programmes.

Supporting Information

Table S1 Phylogenetic relations between *COI* and *ND5* haplotypes recorded in the Central African Republic and previously published sequences of *Ae. albopictus* from Asia, the Americas, the Indian Ocean, Europe and central Africa. (DOC)

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Author Contributions

Conceived and designed the experiments: BK CN MK. Performed the experiments: BK CN. Analyzed the data: BK CN AM CP. Contributed reagents/materials/analysis tools: BK CN MK. Wrote the paper: CN EN CP MK.

- Reiter P (1998) Aedes albopictus and the world trade in used tires, 1988–1995: the shape of things to come? J Am Mosq Control Assoc 14: 83–94.
- Paupy C, Delatte H, Bagny L, Corbel V, Fontenille D (2009) Aedes albopictus, an arbovirus vector: from the darkness to the light. Microbes Infect 11: 1177–1185.
- Fontenille D, Toto JC (2001) Aedes (Stegonyia) albopictus (Skuse), a potential new dengue vector in southern Cameroon. Emerg Infect Dis 7: 1066–1067.
- Coffinet T, Mourou JR, Pradines B, Toto JC, Jarjaval F, et al. (2007) First record of *Aedes albopictus* in Gabon. J Am Mosq Control Assoc 23: 471–472.
- Toto JC, Abaga S, Carnevale P, Simard F (2003) First report of the oriental mosquito *Aedes albopictus* on the West African island of Bioko, Equatorial Guinea. Med Vet Entomol 17: 343–346.
- Diallo M, Laganier R, Nangouma A (2010) First record of Aedes albopictus (Skuse 1894), in Central African Republic. Trop Med Int Health 15: 1185–1189.
- Peyrefitte CN, Rousset D, Pastorino BA, Pouillot R, Bessaud M, et al. (2007) Chikungunya virus, Cameroon, 2006. Emerg Infect Dis 13: 768–771.

- Paupy C, Kassa Kassa F, Caron M, Nkoghe D, Leroy EM (2012) A chikungunya outbreak associated with the vector *Aedes albopictus* in remote villages of Gabon. Vector Borne Zoonotic Dis 12: 167–169.
- Pages F, Peyrefitte CN, Mve MT, Jarjaval F, Brisse S, et al. (2009) Aedes albopictus mosquito: the main vector of the 2007 chikungunya outbreak in Gabon. PLoS One 4: e4691.
- Kelvin AA (2011) Outbreak of chikungunya in the Republic of Congo and the global picture. J Infect Dev Ctries 5: 441–444.
- Braks MA, Honorio NA, Lourencqo-De-Oliveira R, Juliano SA, Lounibos LP (2003) Convergent habitat segregation of *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) in southeastern Brazil and Florida. J Med Entomol 40: 785– 794.
- Simard F, Nchoutpouen E, Toto JC, Fontenille D (2005) Geographic distribution and breeding site preference of *Aedes albopictus* and *Aedes aegypti* (Diptera: Culicidae) in Cameroon, Central Africa. J Med Entomol 42: 726–731.
- Chen CD, Nazni WA, Lee HL, Seleena B, Mohd Masri S, et al. (2006) Mixed breeding of *Aedes aegypti* (L.) and *Aedes albopictus* Skuse in four dengue endemic areas in Kuala Lumpur and Selangor, Malaysia. Trop Biomed 23: 224–227.
- Kamgang B, Happi JY, Boisier P, Njiokou F, Herve JP, et al. (2010) Geographic and ecological distribution of the dengue and chikungunya virus vectors *Aedes* aegypti and *Aedes albopictus* in three major Cameroonian towns. Med Vet Entomol 24: 132–141.
- Rey JR, Nishimura N, Wagner B, Braks MA, O'Connell SM, et al. (2006) Habitat segregation of mosquito arbovirus vectors in south Florida. J Med Entomol 43: 1134–1141.
- 22. Tsuda Y, Suwonkerd W, Chawprom S, Prajakwong S, Takagi M (2006) Different spatial distribution of *Aedes aegypti* and *Aedes albopictus* along an urbanrural gradient and the relating environmental factors examined in three villages in northern Thailand. J Am Mosq Control Assoc 22: 222–228.
- Delatte H, Dehecq JS, Thiria J, Domerg C, Paupy C, et al. (2008) Geographic distribution and developmental sites of *Aedes albopictus* (Diptera: Culicidae) during a chikungunya epidemic event. Vector Borne Zoonotic Dis 8: 25–34.
- O'Meara GF, Evans LF Jr, Gettman AD, Cuda JP (1995) Spread of Aedes albopictus and decline of Aedes aegypti (Diptera: Culicidae) in Florida. J Med Entomol 32: 554–562.
- Lounibos LP (2002) Invasions by insect vectors of human disease. Annu Rev Entomol 47: 233–266.
- Juliano SA, Lounibos LP (2005) Ecology of invasive mosquitoes: effects on resident species and on human health. Ecol Lett 8: 558–574.
- Bagny L, Delatte H, Elissa N, Quilici S, Fontenille D (2009) Acdes (Diptera: Culicidae) vectors of arboviruses in Mayotte (Indian Ocean): distribution area and larval habitats. J Med Entomol 46: 198–207.
- Bagny L, Delatte H, Quilici S, Fontenille D (2009) Progressive decrease in Aedes aegypti distribution in Reunion Island since the 1900s. J Med Entomol 46: 1541– 1545.
- Bagny L, Arnoux S, Delatte H, Lajoie G, Fontenille D (2012) Spread of invasive *Aedes albopictus* and decline of resident *Aedes aegypti* in urban areas of Mayotte 2007–2010. Biol Invasions 14: 1623–1633.
- Rudnick A, Chan YC (1965) Dengue type 2 virus in naturally infected Aedes albopictus mosquitoes in Singapore. Science 149: 638–639.
- Gilotra SK, Rozeboom LE, Bhattacharya NC (1967) Observations on possible competitive displacement between populations of *Aedes aegypti* Linnaeus and *Aedes albopictus* Skuse in Calcutta. Bull World Health Organ 37: 437–446.
- Birungi J, Munstermann LE (2002) Genetic structure of Aedes albopictus (Diptera: Culicidae) populations based on mitochondrial ND5 sequences: evidence for an independent invasion into Brazil and United States. Ann Entomol Soc Am 95: 125–132.
- Mousson L, Dauga C, Garrigues T, Schaffner F, Vazeille M, et al. (2005) Phylogeography of Aedes (Stegonyia) aegypti (L.) and Aedes (Stegonyia) albopictus (Skuse) (Diptera: Culicidae) based on mitochondrial DNA variations. Genet Res 86: 1–11.
- Kamgang B, Brengues C, Fontenille D, Njiokou F, Simard F, et al. (2011) Genetic structure of the tiger mosquito, *Aedes albopictus*, in Cameroon (Central Africa). PLoS One 6: e20257.
- Tabachnick WJ (1994) Genetics of insect vector competence for arboviruses. Adv Dis Vector Res 10: 93–108.
- Jupp PG (1996) Mosquitoes of southern Africa. Culicinae and Toxorhynchitinae. Hartebeesporte: Ekogilde Publishers.

- Invasion of Aedes albopictus in Central Africa
- Edwards FW (1941) Mosquitoes of the Ethiopian region. III Culicine adults and pupae. Oxford: Oxford University Press.
- Hammond SN, Gordon AL, Lugo Edel C, Moreno G, Kuan GM, et al. (2007) Characterization of *Aedes aegypti* (Diptera: Culcidae) production sites in urban Nicaragua. J Med Entomol 44: 851–860.
- Barrera R (2009) Simplified pupal surveys of Aedes aegypti (L.) for entomologic surveillance and dengue control. Am J Trop Med Hyg 81: 100–107.
- Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. Nucleic Acids Res 22: 4673–4680.
- Rozas J, Sanchez-DelBarrio JC, Messeguer X, Rozas R (2003) DnaSP, DNA polymorphism analyses by the coalescent and other methods. Bioinformatics 19: 2496–2497.
- Tajima F (1989) The effect of change in population size on DNA polymorphism. Genetics 123: 597–601.
- Fu YX, Li WH (1993) Statistical tests of neutrality of mutations. Genetics 133: 693–709.
- Fu YX (1997) Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. Genetics 147: 915–925.
- Nylander JA, Ronquist F, Huelsenbeck JP, Nieves-Aldrey JL (2004) Bayesian phylogenetic analysis of combined data. Syst Biol 53: 47–67.
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19: 1572–1574.
- Reiskind MH, Lounibos LP (2012) Spatial and temporal patterns of abundance of *Aedes aegypti* L. and *Aedes albopictus* (Skuse) in southern Florida. Med Vet Entomol. doi: 10.1111/mve.12000.
- Juliano SA, O'Meara GF, Morrill JR, Cutwa MM (2002) Desiccation and thermal tolerance of eggs and the coexistence of competing mosquitoes. Oecologia 130: 458–469.
- Kittayapong P, Strickman D (1993) Distribution of container-inhabiting Aedes larvae (Diptera: Culicidae) at a dengue focus in Thailand. J Med Entomol 30: 601–606.
- Barrera R, Amador M, Clark GG (2006) Ecological factors influencing *Aedes aegypti* (Diptera: Culicidae) productivity in artificial containers in Salinas, Puerto Rico. J Med Entomol 43: 484–492.
- Cox J, Grillet ME, Ramos OM, Amador M, Barrera R (2007) Habitat segregation of dengue vectors along an urban environmental gradient. Am J Trop Med Hyg 76: 820–826.
- Tripet F, Lounibos LP, Robbins D, Moran J, Nishimura N, et al. (2011) Competitive reduction by satyrization? Evidence for interspecific mating in nature and asymmetric reproductive competition between invasive mosquito vectors. Am J Trop Med Hyg 85: 265–270.
- Bargielowski IE, Lounibos LP, Carrasquilla MC (2013) Evolution of resistance to satyrization through reproductive character displacement in populations of invasive dengue vectors. Proc Natl Acad Sci U S A 110: 2888–2892.
- Ngoagouni Č, Kamgang B, Manirakiza A, Nangouma A, Paupy C, et al. (2012) Entomological profile of yellow fever epidemics in the Central African Republic, 2006–2010. Parasit Vectors 5: 175.
- Rudnick A (1965) Studies of the ecology of dengue in Malaysia: a preliminary report. J Med Entomol 2: 203–208.
- Maia RT, Scarpassa VM, Maciel-Litaiff LH, Tadei WP (2009) Reduced levels of genetic variation in *Aedes albopictus* (Diptera: Culicidae) from Manaus, Amazonas State, Brazil, based on analysis of the mitochondrial DNA ND5 gene. Genet Mol Res 8: 998–1007.
- Usmani-Brown S, Cohnstaedt L, Munstermann LE (2009) Population genetics of *Aedes albopictus* (Diptera: Culicidae) invading populations, using mitochondrial nicotinamide adenine dinucleotide dehydrogenase subunit 5 sequences. Ann Entomol Soc Am 102: 144–150.
- Armbruster P, Damsky WE Jr, Giordano R, Birungi J, Munstermann LE, et al. (2003) Infection of new- and old-World *Aedes albopictus* (Diptera: Culicidae) by the intracellular parasite Wolbachia: implications for host mitochondrial DNA evolution. J Med Entomol 40: 356–360.
- Leroy EM, Nkoghe D, Ollomo B, Nze-Nkogue C, Becquart P, et al. (2009) Concurrent chikungunya and dengue virus infections during simultaneous outbreaks, Gabon, 2007. Emerg Infect Dis 15: 591–593.
- Thaikruea L, Charearnsook O, Reanphumkarnkit S, Dissomboon P, Phonjan R, et al. (1997) Chikungunya in Thailand: a re-emerging disease? Southeast Asian J Trop Med Public Health 28: 359–364.