



HAL
open science

Treatment May Be Harmful: Mechanisms/Prediction/Prevention of Drug-Induced DNA Damage and Repair in Multiple Myeloma

Claire Gourzones, Caroline Bret, Jérôme Moreaux

► **To cite this version:**

Claire Gourzones, Caroline Bret, Jérôme Moreaux. Treatment May Be Harmful: Mechanisms/Prediction/Prevention of Drug-Induced DNA Damage and Repair in Multiple Myeloma. *Frontiers in Genetics*, 2019, 10, pp.861. 10.3389/fgene.2019.00861 . hal-02552817

HAL Id: hal-02552817

<https://hal.umontpellier.fr/hal-02552817>

Submitted on 27 May 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Treatment May Be Harmful: Mechanisms/Prediction/Prevention of Drug-Induced DNA Damage and Repair in Multiple Myeloma

Claire Gourzones¹, Caroline Bret^{1,2,3} and Jerome Moreaux^{1,2,3,4*}

¹IGH, CNRS, Univ Montpellier, France, ²Department of Biological Hematology, CHU Montpellier, Montpellier, France, ³Univ Montpellier, UFR de Médecine, Montpellier, France, ⁴Institut Universitaire de France, Paris, France

OPEN ACCESS

Edited by:

Meral Beksac,
Ankara University, Turkey

Reviewed by:

Anders Waage,
Norwegian University of Science
and Technology, Norway
Sergio Roa,
University of Navarra, Spain

*Correspondence:

Jerome Moreaux
jerome.moreaux@igh.cnrs.fr

Specialty section:

This article was submitted to
Cancer Genetics,
a section of the journal
Frontiers in Genetics

Received: 26 April 2019

Accepted: 19 August 2019

Published: 18 September 2019

Citation:

Gourzones C, Bret C and Moreaux J
(2019) Treatment May Be Harmful:
Mechanisms/Prediction/Prevention
of Drug-Induced DNA Damage and
Repair in Multiple Myeloma.
Front. Genet. 10:861.
doi: 10.3389/fgene.2019.00861

Multiple myeloma (MM) is a malignancy characterized by accumulation of malignant plasma cells within the bone marrow (BM). MM is considered mostly without definitive treatment because of the inability of standard of care therapies to overcome drug-resistant relapse. Genotoxic agents are used in the treatment of MM and exploit the fact that DNA double-strand breaks are highly cytotoxic for cancer cells. However, their mutagenic effects are well-established and described. According to these effects, chemotherapy could cause harmful DNA damage associated with new driver genomic abnormalities providing selective advantage, drug resistance, and higher relapse risk. Several mechanisms associated with MM cell (MMC) resistance to genotoxic agents have been described, underlining MM heterogeneity. The understanding of these mechanisms provides several therapeutic strategies to overcome drug resistance and limit mutagenic effects of treatment in MM. According to this heterogeneity, adopting precision medicine into clinical practice, with the development of biomarkers, has the potential to improve MM disease management and treatment.

Keywords: multiple myeloma, DNA damage, genomic instability, drug resistance, genotoxic agents

INTRODUCTION

Multiple myeloma (MM) is the second most common hematological malignancy, with an incidence ranging from 4 to 6 per 100,000/year in the United States (Braggio et al., 2015). Malignant plasma cells display prominent genomic abnormalities arising during tumorigenesis and accumulating during disease progression. This series of complex molecular events involved in MM development includes chromosomal abnormalities, oncogene activation, and cellular communication signals dysregulation. MM treatment strategies have significantly evolved during the last decades with an expanding arsenal of anti-MM therapies. This was associated with a three-fold improvement in median survival. According to these significant advances, long-term side effects and harmful effects of treatment are an important issue with the development of drug-resistant subclones and formation of therapy-related cancers caused by lesions they create in normal cells. Thus, understanding the DNA damages induced by anti-MM therapies and DNA damage response is important to improve patient survival and reduce harmful effects related to treatment.

GENOMIC INSTABILITY IN MULTIPLE MYELOMA

Genome integrity is constantly assailed by diverse arrays of insults. These include genotoxic agents of exogenous origin and endogenous sources of DNA damage, such as reactive oxygen species, collisions between DNA replication and transcription, and programmed genomic rearrangements (Aguilera and Gomez-Gonzalez, 2008). B cells are particularly challenged in this regard. During their maturation, B lymphocytes are subjected different to genetic alterations, including V, D and J immunoglobulin genes recombination, Ig class switch recombination (CSR), and somatic hypermutation (SHM). To prevent immune deficiency, autoimmunity, or cancer, these biological processes should be tightly regulated (Gennery et al., 2000). MM cells are characterized by genomic instability, including chromosomal instability, mutations, and microsatellite instability. Recent genome sequencing studies of MM cells have uncovered the major genomic instability and molecular and subclonal heterogeneity of the disease (Lohr et al., 2014; Vikova et al., 2019).

DNA-DAMAGING AGENTS USED IN MULTIPLE MYELOMA TREATMENT

Melphalan DNA-damaging agent was introduced for the treatment of MM in 1962. Several combinations have been developed to the improve outcome of patients, including other DNA-damaging agents (Table 1). The high-dose melphalan regimen combined with autologous hematopoietic stem cell transplant (ASCT) has long been considered frontline therapy for newly diagnosed patients. Melphalan is a nitrogen mustard known to induce mono-alkylation of adenine and guanine together with interstrand DNA crosslinks (Osborne et al., 1995). Monoadducts represent 95% of the lesions, whereas 5% are interstrand DNA crosslinks (ICLs) (Muniandy et al., 2010). In MM cells, ICL number is correlated with the melphalan concentration used (Spanswick et al., 2002). ICLs are highly toxic for MM cells.

Results from studies with a long-term follow-up of patients treated with high-dose chemotherapy and ASCT demonstrated that 10%–15% of the patients remain alive after more than 10 years without relapse (Barlogie et al., 2006; van de Velde et al., 2007; Martinez-Lopez et al., 2011), demonstrating the efficacy of this

strategy in MM (van Rhee et al., 2014). The novel drugs such as immunomodulatory agents (IMiDs) and proteasome inhibitors (PIs) used during induction, consolidation, and maintenance stages have significantly improved the outcome of the patients (Barlogie et al., 2006; Cavo et al., 2010; Ladetto et al., 2010; Moreau et al., 2011; Neben et al., 2012; Sonneveld et al., 2012).

Other genotoxic drugs are used to treat patients with MM, including cyclophosphamide, doxorubicin, busulfan, vincristine, etoposide, cisplatin, bendamustine, liposomal doxorubicin, and melphalan flufenamide hydrochloride. Use of chemotherapeutic agents without cross-resistance could enhance peripheral blood stem cell collection and improve patient outcome related to better antitumor efficacy before ASCT. Cyclophosphamide is an alkylating agent inducing ICLs. Doxorubicin induces DNA double-strand DNA breaks (DSBs) related to intercalation into DNA and inhibition of topoisomerase II (TopII). Doxorubicin can also induce DNA adducts, free radicals release, and formaldehyde-dependent ICL formation (Bret et al., 2013). Etoposide is a DNA-damaging agent that induces DNA damage and inhibits DNA replication by suppressing the relaxation activity of TopIIA topoisomerase (Dobbelstein and Sorensen, 2015). Busulfan is a bifunctional alkylating agent. Vincristine is a microtubule-targeting agent that was demonstrated to increase DNA damage induced by DNA-damaging agents (Poruchynsky et al., 2015). Cisplatin is a platinum compound that modifies DNA, leading to the formation of intrastrand or interstrand crosslinks between bases. Cisplatin is used in DCEP regimen in combination with dexamethasone, etoposide, and cyclophosphamide (Dadacaridou et al., 2007; Park et al., 2014). Pegylated liposomal doxorubicin is a formulation of doxorubicin in liposomes with a prolonged circulation time (Voorhees et al., 2015). Bendamustine combines bifunctional alkylating nitrogen mustard group and a purine-like benzamidazol nucleus and was shown to overcome melphalan resistance in human myeloma cell lines (Cives et al., 2013). Melphalan flufenamide hydrochloride is a lipophilic alkylator characterized by intracellular hydrolysis after cellular uptake and release of active melphalan (Ray et al., 2016).

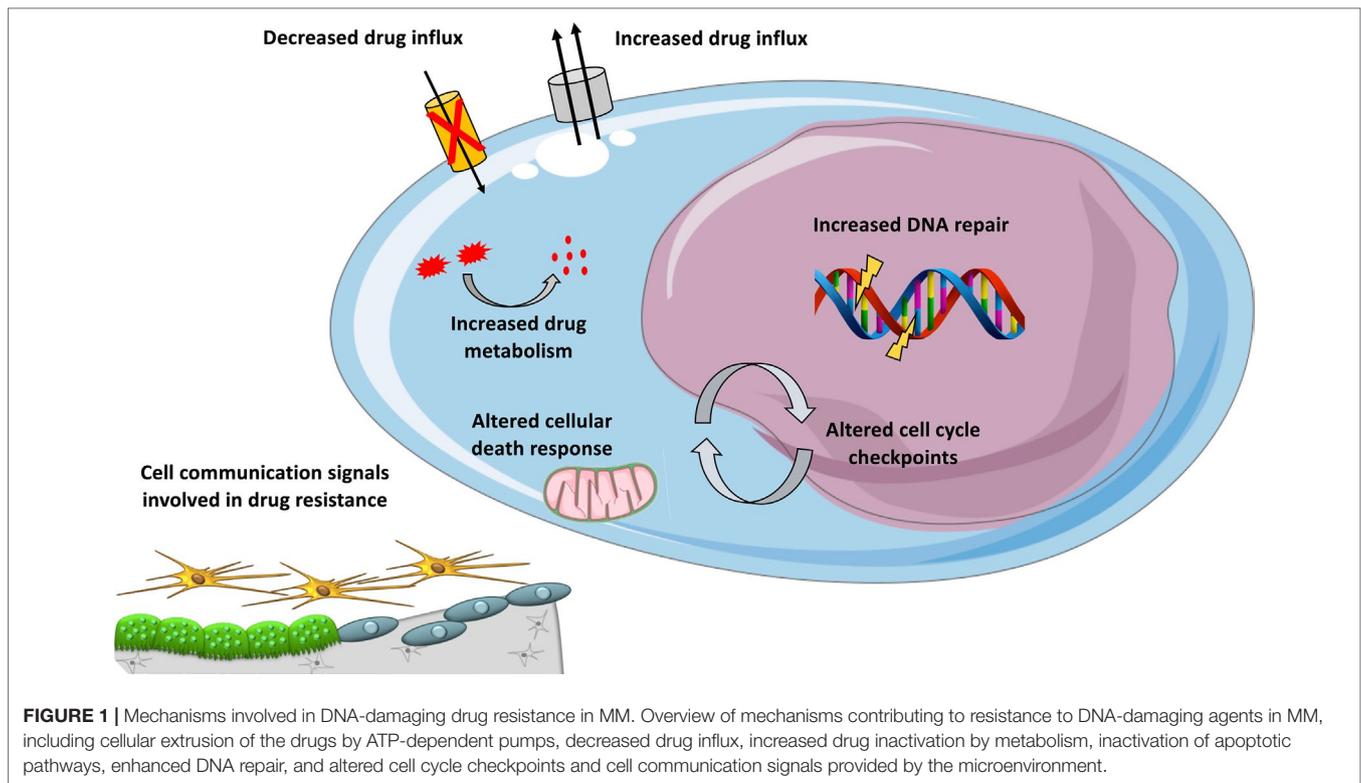
DRUG RESISTANCE MECHANISMS TO DNA-DAMAGING AGENTS

Drug Efflux, Cell Communication Signals, and Drug Metabolism

An important mechanism of multidrug resistance in cancer is the overexpression of ATP-binding cassette (ABC) transporters (Kathawala et al., 2015) (Figure 1). This family of transmembrane proteins use the energy generated by ATP hydrolysis to efflux cytotoxic compounds. The most studied ABC is dependent efflux pump P-glycoprotein (P-gp) coded by the *MDR1/ABCB1* gene. In MM, no significant expression of P-gp was detected in newly diagnosed MM and in patients treated with melphalan (Grogan et al., 1993). P-gp overexpression was demonstrated to be associated with resistance to glucocorticoid, etoposide, doxorubicin, and vincristine (Dalton, 1997). VAD treatment (vincristine, doxorubicin, and dexamethasone) was associated with P-gp overexpression in MM patients (Sonneveld et al., 2001; Yang et al., 2003). However, a clinical trial with ABCB1 inhibitor (Zosuquidar) did not show

TABLE 1 | DNA-damaging chemotherapeutic drugs used in MM.

| Agents |
|--|
| Melphalan (Alkeran®) |
| Cyclophosphamide (Cytoxan®) |
| Doxorubicin (Adriamycin®) |
| Busulfan (Myleran®) |
| Vincristine (Oncovin®) |
| VP-16 (Etoposide®) |
| Bendamustine (Treanda®) |
| Pegylated liposomal doxorubicin (Doxil®) |
| Melphalan flufenamide hydrochloride (Melflufen®) |
| Cisplatin (Platinol®) |



any benefit in progression-free or overall survival in refractory MM patients when combined with vincristine, doxorubicin, and dexamethasone (Friedenberg et al., 2006).

The behavior of MM cells is determined not only by their genetic or epigenetic background but also by their BM microenvironment. The majority of myeloma growth factors (MGFs) is secreted by the BM environment compared to autocrine MGFs (Mahtouk et al., 2010). Several studies have provided a comprehensive overview of MGF expression in the different BM cell subpopulations of MM patients (Podar et al., 2009; Mahtouk et al., 2010). Interactions between MM cells and bone marrow microenvironment could also play a role in DNA-damaging agents drug resistance (Figure 1). We have documented the rise of large concentrations of IL-6 9 days after high-dose melphalan in patients (Condomines et al., 2010). This large concentration of IL-6 will facilitate melphalan-resistant MMCs to survive within the BM. Patients treated with high-dose melphalan, stem cell transplantation, and anti-IL-6 antibody had a survival advantage when mixed with a large cohort of matched patients treated with melphalan and stem cell transplantation alone (Rossi et al., 2005). Cell adhesion-mediated drug resistance to doxorubicin, vincristine, and melphalan was described using human myeloma cell lines and primary MM cells from patients (Damiano et al., 1999; Noborio-Hatano et al., 2009; Neri et al., 2011a; Di Marzo et al., 2016). Bortezomib could overcome cell adhesion-mediated drug resistance through VLA-4 downregulation and inhibition of MM cell adhesion to stroma (Noborio-Hatano et al., 2009; Neri et al., 2011a). Cell adhesion-mediated drug resistance could also protect MM cells from etoposide toxicity (Hazlehurst et al., 2000). Targeting cell-to-cell communication between MM

cells and BM microenvironment could improve current therapeutic strategies using DNA-damaging agents.

In vivo, drugs could be metabolized (Figure 1) through a number of reactions occurring through two distinct consecutive phases: phase I and phase II drug metabolism. Phase I drug metabolizing enzymes consist primarily of oxidases, reductases, and dehydrogenases, and phase II enzymes play an important role in biotransformation and inactivation of drugs. A study reported that MMCs of patients with a favorable outcome after treatment with high-dose therapy and ASCT are characterized by an overexpression of genes coding for xenobiotic receptors and their downstream targets, including phase I and phase II drug metabolism enzymes and transporters (Hassen et al., 2014). At the opposite, high-risk patients were characterized by overexpression of genes involved in *Nrf2* and *ARNT* pathways (Hassen et al., 2014). These data underline a role of drug metabolism in chemotherapy resistance in MM and suggest that inhibitors targeting these pathways could open new perspectives to alleviate or overcome drug resistance.

DNA-Damaging Agents and DNA Repair Pathways

The fact that DNA double-strand breaks are highly cytotoxic is exploited by DNA-damaging agents used in the treatment of MM. According to the type of DNA damage, specific DNA repair pathways will be used to cope with DNA insults. For nucleotide lesions occurring on single strands, base excision repair (BER), nucleotide excision repair (NER), and mismatch repair (MMR) will be involved. For DSBs, there are two major pathways, including nonhomologous end-joining (NHEJ) and homologous

recombination (HR) DNA repair. The DNA damage response (DDR) sensor proteins will be involved in the detection of damaged DNA, leading to cellular response activation, including one or more DNA repair pathways. For DSBs, Ku proteins and MRN complex are the predominant sensors. Fanconi anemia proteins, Poly (ADP -ribose) polymerase (PARP), mismatch repair proteins (including MSH2, MSH3, MSH6, PMS2, and MLH1), and NER proteins (including XPC, CSA, and DDB2) are other DNA-damage sensors (Brown et al., 2017).

Single-Strand Damage DNA Repair

Nucleotide Excision Repair

Nucleotide excision repair (NER) removes helix-distorting adducts on DNA that could be caused by UV or radiation and participates in the repair of ICLs44 (Figures 2 and 3A). NER can be coupled to transcription [transcription-coupled nucleotide excision repair (TC-NER)] opposed to global genome nucleotide excision repair (GG-NER) (Friedberg, 2001; Hanawalt and Spivak, 2008). In cancer cells exposed to genotoxic agents, NER plays a key role in removal and repair of the DNA damage (Kirschner and Melton, 2010). It has been demonstrated that NER is a major DNA repair mechanism that removes cisplatin-induced DNA damage. In solid tumors, high expression of ERCC1, involved in NER, is associated with resistance to platinum-based therapy. NER removes helix-distorting adducts participating in the ICL repair. ERCC1 and XP proteins play major roles in the repair of DNA adducts and ICLs (Figure 3A). Furthermore, deficiency in NER was associated with higher sensitivity to platinum agents and a reduced capacity to repair ICLs (Kirschner and Melton, 2010).

Base Excision Repair

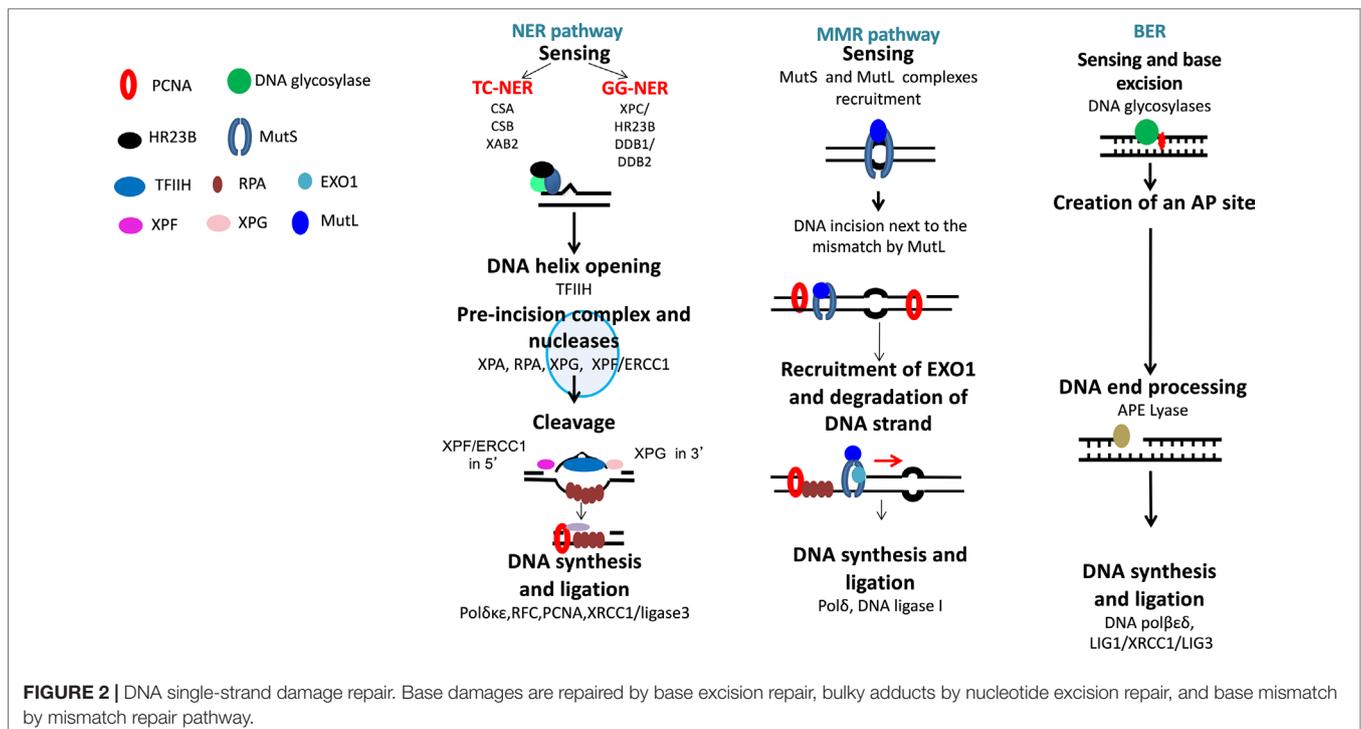
The base excision repair (BER) pathway repairs DNA damaged bases (Lindahl, 1993) (Figure 2). Conventional BER is initiated by specific DNA glycosylases that will hydrolyze N-glycosylic bond between the damaged base and the sugar phosphate backbone. This process is followed by end processing mechanism, repair synthesis, and ligation (Caldecott, 2008). However, different mechanisms could participate in BER pathways, depending on the physiological state of cancer cells and the type of DNA glycosylase involved (Caldecott, 2008).

Mismatch DNA Repair

Mismatch DNA repair (MMR) is involved in the repair of replication errors associated with nucleotide deletion or insertion or causing incorporation of a wrong nucleotide (Figure 2). MSH2-MSH6 defects in MMR are associated with drug resistance to temozolomide, nucleoside analogs, and platinum agents in solid cancers and Acute myeloid leukemia (AML) (Karran and Bignami, 1994; Fordham et al., 2011).

DSBs DNA Repair

DSBs can be generated during ICLs repair, by ROS, by DNA damaging used in MM treatment and during DNA replication when cells progress in S phase before completing single-strand DNA damage repair (Ciccia and Elledge, 2010; Deans and West, 2011). Several partially independent sensors can detect DSBs, including Ku70/Ku80, PARP, MRN, and RPA. Homologous recombination operates in the S and G2 phases of the cell cycle. Homologous recombination pathway is conservative and requires a homologous DNA template (Figure 4). At the opposite, NHEJ is nonconservative with direct ligation of DSB ends (Figure 4).



Homologous Recombination DNA Repair

The homologous recombination DNA repair (HRR) pathway is complex and occurred during S and G2 phases of the cell cycle (Ciccia and Elledge, 2010; Deans and West, 2011; Gourzones-Dmitriev et al., 2013). The DNA ends of the lesion are resected to allow invasion of the single strand into the sister chromatid used as a template for precise resynthesis of damaged DNA part. HRR plays an important role in genomic stability (Curtin, 2012; Curtin, 2013). The MRN complex (MRE11-RAD50-NBS1) recognizes DSBs, and CtIP recruits RPA (Williams et al., 2007; Jasin and Rothstein, 2013). BRCA1 interacts also with MRN and CtIP and promotes HRR and single-strand annealing (SSA). The resulting 3' ssDNA tails are bound by RPA, which is replaced with Rad51. The reaction is mediated by Rad52, Rad55, and Rad57. The RAD51 nucleoprotein filament promotes homology search. This is followed by exchange strand between the intact sister chromatid and the broken duplex and strand extension by DNA polymerase (Figure 4). HRR repairs DSBs, which occurs through exposure to topoisomerase inhibitors and DNA crosslinking agents used in MM treatment, including etoposide and doxorubicin. HRR plays a major role in stalled replication fork restart and ICL repair in association with the Fanconi anemia (FA) pathway (Figure 3B). HRR is more active in MM cells compared to normal plasma cells (Shammas et al., 2009; Roddam et al., 2010). Higher expressions of RAD50 and RAD51 proteins involved in HRR have been reported in primary MM cells and MM cell lines compared to

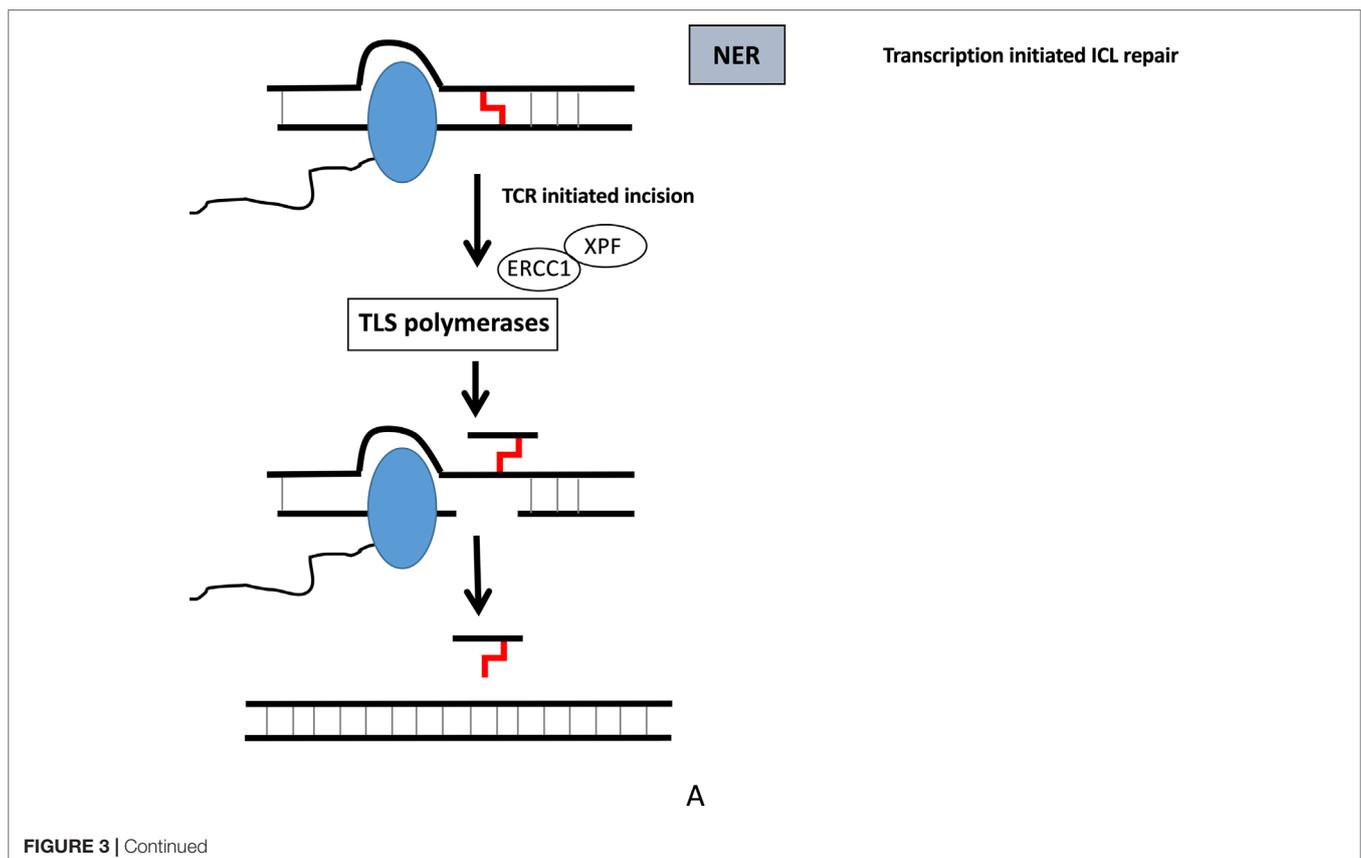
normal plasma cells in association with increased HRR activity (Shammas et al., 2009; Roddam et al., 2010). The proliferation assessment based on propidium iodide incorporation (plasma cell labeling index) (San Miguel et al., 1995) or the assessment of Ki-67-expressing cells of malignant plasma cells (Alexandrakis et al., 2004) has been shown to be a powerful and independent predictor of survival in MM. (Greipp et al., 1988) The role of HRR in drug resistance of the small fraction of proliferating MM cells may be of particular importance in MM.

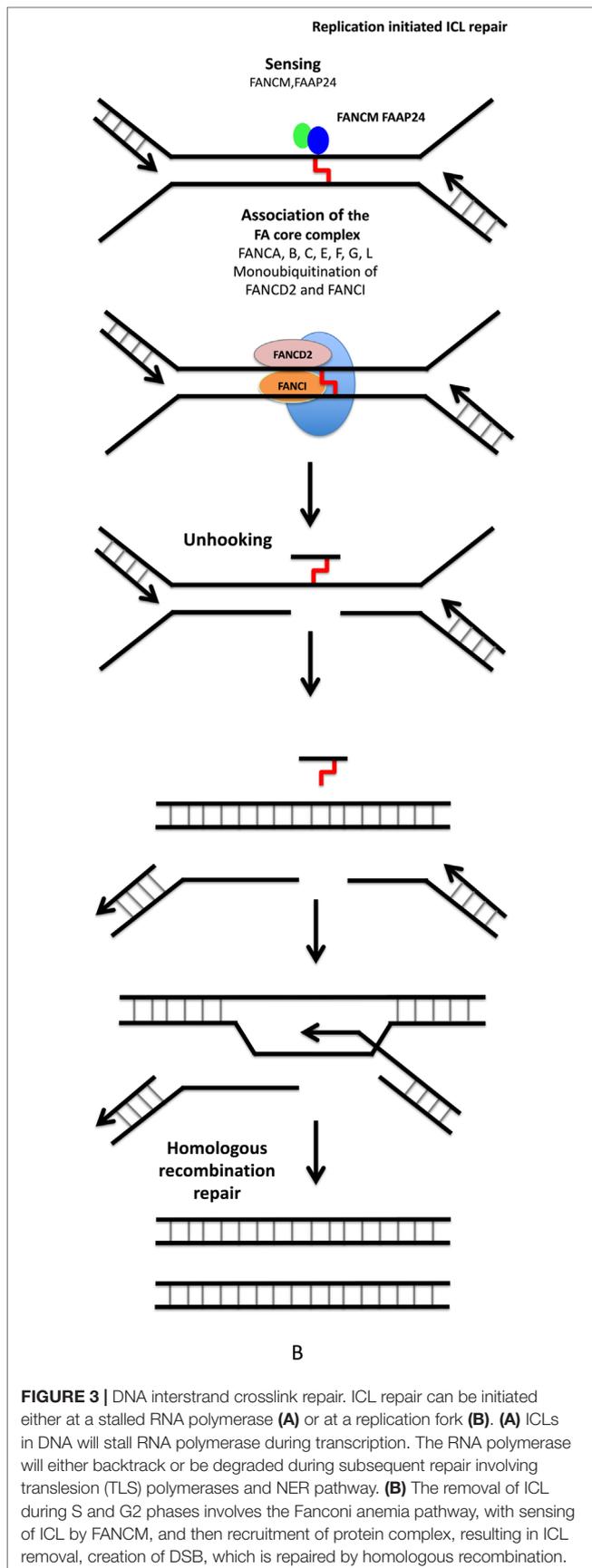
Single-Strand Annealing

SSA is used only when two homologous regions flank the DSB site. In this context, the homologous regions are exposed, and after annealing and cleavage of the DNA overhang, ligation of the ends results in the deletion of the intervening region (Figure 4). According to this process, SSA is inaccurate. Translocations induced by SSA have been reported when DSBs are located in repeats on different chromosomes (Hartlerode and Scully, 2009; Ciccia and Elledge, 2010).

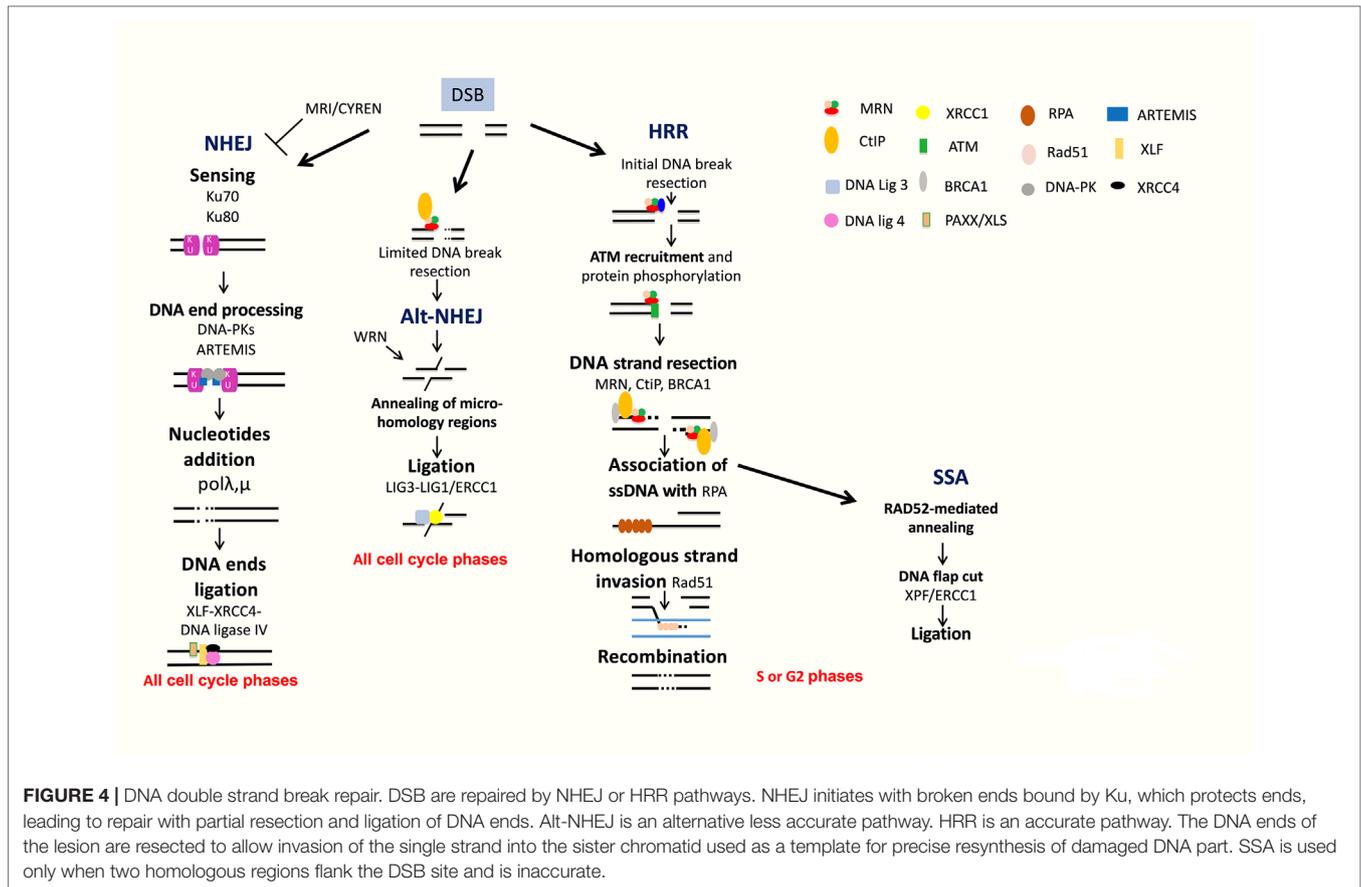
NHEJ DNA Repair

NHEJ does not use significant homology at the broken ends. DSBs are sensed by Ku70/Ku80 that bind and activate DNA-PKcs protein kinase, leading to the recruitment and activation of DNA end-processing enzymes, such as ARTEMIS, template-independent polymerases (polymerases λ and μ), and XLF-XRCC4-DNA ligase 4 complex (Figure 4). More recently,





several new accessory NHEJ factors, including PAXX/XLS and CYREN/MRI, have been described. These factors have been reported to be required for DNA ligation. PAXX/XLS is an accessory related to NHEJ pathway and presenting overlapping functions with XLF (Tadi et al., 2016). CYREN is a cell cycle-dependent inhibitor of NHEJ pathway, promoting repair by HR pathway (Arnoult et al., 2017; Hung et al., 2018). A correlation between polymorphisms or aberrant expressions of *XRCC4*, *XRCC5* (encoding Ku80), *XRCC6* (encoding Ku70), *ARTEMIS*, and *LIG4* (encoding DNA ligase 4) genes and risk of MM development has been reported (Roddam et al., 2002; Hayden et al., 2007; Roddam et al., 2010). Interestingly, high expression of *XRCC5* and *ARTEMIS* genes was associated with a poor prognosis in MM patients (Calimeri et al., 2012). A recent study investigated the functionality of DSB repair in MM cells and identified upregulation of *ARTEMIS*, DNA-PKcs, and *XRCC4* proteins in MM cells (Herrero et al., 2015). NHEJ activity was also significantly elevated in MM cells compared to a normal lymphoblastoid cell line (Herrero et al., 2015). Furthermore, MMSET was recently identified to be involved in DNA damage response as a sensor (Hajdu et al., 2011). MMSET, like RIF1, is an ATM-dependent DDR factor. MMSET accumulates at DSBs, leading to recruitment of 53BP1. Recruitment of 53BP1 to DSBs is dependent on histone H4 methylation by MMSET (Pei et al., 2011). However, two other groups demonstrated that MMSET had no effect on 53BP1 foci formation and H4K20 methylation (Hsiao and Mizzen, 2013; Tuzon et al., 2014). During B-cell development, MMSET is also involved in class-switch recombination accumulating at immunoglobulin gene-switch regions with H3K36me2 and γ H2AX. Furthermore, MMSET depletion led to defects in class-switch recombination (Pei et al., 2013). DSBs generated during CSR modifications of B-lymphocyte Ig genes are mainly repaired by NHEJ (Lieber, 2010). H3K36 methylation, induced by MMSET and other histone methyltransferases, can also influence DNA repair pathway choice (Fnu et al., 2011; Aymard et al., 2014; Carvalho et al., 2014; Pfister et al., 2014). More recently, it was shown that MMSET is necessary for efficient NHEJ and HRR (Shah et al., 2016). MMSET depletion resulted in decreased expression of DNA repair genes and significant reduction in the recruitment of DNA repair proteins at DNA damage sites. MMSET overexpression is associated with increased DNA repair efficiency. Loss of MMSET combined with melphalan treatment leads to decreased tumor mass and increased survival in mice (Shah et al., 2016). MMSET-overexpressing cells demonstrate increased γ H2AX after treatment with DNA-damaging agents, suggesting a role in drug resistance through accelerated DNA repair (Shah et al., 2016). Given the association of t(4;14), involving MMSET, with poor outcome in MM, this potential perturbation of DNA repair may prove highly relevant to disease progression and drug resistance. More recently, Wang and Goldstein (2016) demonstrated that Dicer- and Drosha-dependent diRNAs play a role in guiding molecules to promote the recruitment of MMSET and other proteins to DSB sites. Several groups have demonstrated that H3K36 is the main target of MMSET without effect on H4K20 (Li et al., 2009; Martinez-Garcia et al., 2010; Popovic et al., 2014). Several



studies reported that H3K36 mark favors NHEJ as DNA repair pathway choice (Fnu et al., 2011; Aymard et al., 2014; Carvalho et al., 2014; Pai et al., 2014; Pfister et al., 2014). H3K36me2 increase and H3K27me3 decrease in MMSET-overexpressing cells are associated with a more open chromatin state (Martinez-Garcia et al., 2010; Popovic et al., 2014). This chromatin state is associated with an increased DNA damage when cells are treated with a DNA-damaging agent (Shah et al., 2016). However, DNA repair proteins could access more easily to chromatin and facilitate efficient DNA repair (Ransom et al., 2010; Soria et al., 2012), as observed in cells with high MMSET expressions (Shah et al., 2016).

Alternative NHEJ DNA Repair

Alt-NHEJ is an alternative end-joining pathway that is detected when the classical NHEJ pathway is impaired (Nussenzweig and Nussenzweig, 2007; Mladenov and Iliakis, 2011) (Figure 4). Alt-NHEJ is less accurate, requires more extensive end resection, and frequently uses microhomology. DNA ends are resected up to regions of microhomology, annealed, and ligated by XRCC1/ligases 1 and 3 (Ciccia and Elledge, 2010; Shaheen et al., 2011; Simsek et al., 2011; Boboila et al., 2012). The 53BP1 protein protects DNA ends from nucleolytic degradation and thereby prevents microhomology mediated repair (Bothmer et al., 2010). Moreover, it has been implicated in the chromosomal translocation process involved in lymphoid tumorigenesis

(Nussenzweig and Nussenzweig, 2007; Kotnis et al., 2009). Overexpression of proteins involved in alt-NHEJ, including DNA ligase 3, was also reported in MM cells (Herrero et al., 2015). Activity of this highly mutagenic pathway was identified in MM cells through detection of largest deletions and higher sequence microhomology at DNA lesion sites and could be decreased by alt-NHEJ inhibition (Herrero et al., 2015).

DNA REPAIR AND RESISTANCE TO DNA-DAMAGING AGENTS IN MULTIPLE MYELOMA

Cyclophosphamide is a genotoxic agent inducing ICLs. Melphalan is also a nitrogen mustard that induces ICLs. The vast majority of the lesions induced by melphalan are monoadducts and 5% ICLs (Osborne et al., 1995; Muniandy et al., 2010). The ICL number induced by melphalan in MM cells is correlated with the concentration used (Spanswick et al., 2002). Doxorubicin induces DNA adducts, DSB, free radicals release, and formaldehyde-dependent ICL formation. A study reported that MM cells from patients previously treated by melphalan are able to repair ICLs *in vitro*, whereas MM cells from untreated patients could not (Spanswick et al., 2002). Furthermore, melphalan-induced DNA damage, *in vitro*, in peripheral blood mononuclear cells is a predictor for clinical outcome in patients treated by high-dose

melphalan and ASCT (Dimopoulos et al., 2007). A significant association between polymorphisms of genes involved in DNA repair and melphalan resistance was identified in MM (Dumontet et al., 2010). In a cohort of MM patients treated with high-dose melphalan and ASCT, polymorphisms of *PARP*, *RAD51*, *PCNA*, *OGG1*, *XPC*, *BRCA1*, *ERCC1*, *BARD1*, and *TP53BP1* are associated with the outcome and overall survival of patients (Dumontet et al., 2010). These genes are significantly enriched in genes involved in HRR and NER necessary for ICL repair. NER removes helix-distorting adducts on DNA and contributes to the repair of ICLs. Furthermore, reduced capacity to repair ICL has been associated with NER deficiency (**Figure 3A**) together with higher sensitivity to platinum agents (Kirschner and Melton, 2010) ERCC2 and XRCC3 gene polymorphisms are associated with treatment outcome and drug resistance in patients treated with high-dose melphalan and ASCT (Vangsted et al., 2007). In MM cell lines resistant to melphalan, overexpression or *BRCA1*, *BRCA2*, *FANCA*, *FANCC*, *FANCF*, *FANCL*, and *RAD51C*; upregulation of HRR; and Fanconi pathways were identified. Depletion of *FANCF* could overcome melphalan resistance, and *FANCF* overexpression protects MM cells from melphalan toxicity (Chen et al., 2005). FA pathway appears as a key player in DNA-damaging agent resistance. NF- κ B, constitutively activated in MM (Demchenko and Kuehl, 2010; Chapman et al., 2011), was shown to transcriptionally regulate FA pathway genes (Yarde et al., 2009). According to that, proteasome inhibitors affect DNA repair pathways in MM through several mechanisms. Proteasome inhibitors block NF- κ B-mediated transcriptional activation of FA genes (Yarde et al., 2009). PIs also inhibit the decrease in FANCD2 monoubiquitination and the expression of FANCD2 proteins (Jacquemont and Taniguchi, 2007). PIs also target HRR by blocking NBS1, *BRCA1*, phospho-ATM, and Rad51 recruitment (Murakawa et al., 2007). PI treatment reduces the pool of available nuclear ubiquitin and impede FANCD2 and H2AX ubiquitination (Neri et al., 2011b). Bortezomib treatment may prevent DNA resection through inhibiting proteasomal degradation of proteins involved in chromatin relaxation (Jacquemont and Taniguchi, 2007; Murakawa et al., 2007). This mechanism impedes the recruitment of RPA onto ssDNA (Jacquemont and Taniguchi, 2007; Murakawa et al., 2007). This effect of PIs on DNA repair could explain the synergistic activity of PI treatment with high-dose melphalan before ASCT (Roussel et al., 2010). MMSET was also involved in melphalan drug resistance through significant increases of DNA repair efficiency (Shah et al., 2016). Several studies demonstrated a role of BER in DNA-damaging agent resistance in MM. APE1 depletion was reported to sensitize MM cells to melphalan treatment (McNeill et al., 2009). More recently, gene expression profile analyses revealed a prognostic value of genes involved in FA (*RMII*, *FANCI*, and *FANCA*), NER (*PCNA*, *RPA3*, *LIG3*, *POLD3*, *ERCC4*, *POMD1*, *ERCC1*, and *ERCC5*), NHEJ [*WHSC1* (MMSET), *RIF1*, *XRCC5* (Ku80), *PNKP*, and *POLL*], MMR (*EXO1* and *MSH2*), and HRR (*EXO1*, *BLM*, *RPA3*, *RAD51*, *MRE11A*, and *ATM*) pathways in MM (Kassambara et al., 2014). Based on these prognostic genes, NHEJ, HRR, FA, and NER gene-based risk scores were created, allowing identification of high-risk MM patients in two independent cohorts of patients

treated with high-dose melphalan and ASCT (Kassambara et al., 2014). Furthermore, a DNA repair risk score, incorporating all of the 22 DNA repair prognostic genes presented a strong prognostic value for both event-free survival and overall survival in two independent cohorts of MM patients. This gene-based DNA repair score remained an independent prognostic factor when tested together with known prognostic factors, including previously published gene expression profiling (GEP)-based risk scores, t(4;14), del17p and with standard clinical prognostic factors, ISS, b2m, and albumin. These DNA repair scores represent powerful tools to develop synthetic lethality approaches and exploit the addiction of MM cells to a specific DNA repair pathway (Kassambara et al., 2014).

RECQ helicases playing a role in repair of damaged replication forks, DNA damage response, and homologous recombination are also involved in cancer cell drug resistance (Futami et al., 2008a; Futami et al., 2008b; Arai et al., 2011; Viziteu et al., 2016). In MM, high RECQ1 expression is associated with resistance to genotoxic agents and poor prognostic value in several independent cohorts of MM patients (Viziteu et al., 2017). MM cells overexpressing RECQ1 are able to repair DNA breaks induced by genotoxic agents more efficiently, conferring drug resistance (Viziteu et al., 2017). Interestingly, the abnormal overexpression of RECQ1 in MM is linked to aberrant methylation of miR-203. DNA methylation inhibitor treatment induces upregulation of miR-203 followed by RECQ1 downregulation and hypersensitivity to treatments (Viziteu et al., 2017).

In MM patients characterized by 1q21 amplification, high expression of ILF2 has also been reported to participate in resistance to genotoxic agents through regulation of DNA repair gene splicing transcription (Marchesini et al., 2017).

MUTAGENIC EFFECTS OF CHEMOTHERAPY IN MULTIPLE MYELOMA

Genotoxic agents can induce mutagenic effects. DNA alkylating agents and intercalating agents have been associated with mutations signatures (Szikriszt et al., 2016). According to these effects, chemotherapy could cause harmful DNA damage associated with new driver genomic abnormalities, providing selective advantage, drug resistance, and higher relapse risk (Weinhold et al., 2016). Alkylating agents also modify the redox potential of cancer cells. This mechanism has been reported for melphalan and bendamustine that increase the level of reactive oxygen species (ROS), inhibit the activity of thioredoxin reductase, and activate p53 pathway (Witte et al., 2005; Surget et al., 2014). Interestingly, reduced glutathione (GSH) protects MM cells from melphalan-induced toxicity without affecting the ability of melphalan to induce DNA damage (Gourzones et al., 2019). Deregulation of genes involved in response to oxidative stress is associated with a poor outcome and Melphalan resistance in MM (Gourzones et al., 2019). The DNA damage accumulation resulting from survival advantage mediated by the redox system upregulation could participate in new genomic driver events and poor outcome in MM (Gourzones et al., 2019). In this context,

the selective pressure of chemotherapy may result in new driver mutations associated with MM cell subclonal selection and drug resistance, as illustrated with double hit events involving tumor suppressor genes, including *TP53* and *FAM46C* in MM patients at relapse (Weinhold et al., 2016). Overexpression of DNA repair machinery could also result in new genomic events especially if nonaccurate pathways like alt-NHEJ are involved (Gourzones-Dmitriev et al., 2013; Viziteu et al., 2017).

SYNTHETIC LETHALITY THERAPEUTIC STRATEGIES TO OVERCOME DRUG RESISTANCE

DNA repair pathways and other pathways are deregulated in many MM patients to provide adaptive mechanisms and support drug resistance. Targeting DNA repair pathways may potentiate the efficacy of current drugs and overcome drug resistance and represents an opportunity to develop synthetic lethality approaches. Synthetic lethal strategies are currently developed in the context of the DNA damage response with the finding that PARP inhibitors are specifically toxic to HRR defective cancer cells (Bryant et al., 2005; Farmer et al., 2005). Because PI treatment was reported to induce BRCAness in MM cells, (Neri et al., 2011b) this strategy was used for MM cells. Cotreatment of MMCs with bortezomib and PARP1 inhibitor resulted in DSB accumulation and MMC killing (Neri et al., 2011b). Novel compounds targeting DNA repair pathways are being clinically evaluated in patients with cancer to induce synthetic lethality (Shaheen et al., 2011; O'Connor, 2015) (Table 2). These inhibitors targeting DNA repair pathways or cell cycle checkpoints (PARP1, DNA-PK,

ATM, Ataxia telangiectasia and Rad3 related (ATR), MGMT, APE, CHK1, CHK2) could be useful to target MMCs in combination with DNA-damaging drugs and reverse drug resistance. The choice of which DNA repair inhibitor combine with the different DNA-damaging agents used in MM is related to align the DNA damage induced by the DNA-damaging agent with the DDR repair mechanism. PARP inhibitor was shown to reverse melphalan resistance in human myeloma cell lines, and combination with melphalan has synergistic association to FA and HRR pathways (Xiong et al., 2015). ATR inhibitor also demonstrated significant anti-MM toxicity in the context of MYC-induced replicative stress (Cottini et al., 2015). Interestingly, combination of ATR inhibitor with piperlongumine, a compound increasing ROS levels, exacerbates oncogene-induced DNA damage and shows a synergistic effect (Cottini et al., 2015). CHK1/2 inhibitor AZD7762 was shown to potentiate the effects of melphalan, bendamustine, and doxorubicin in p53-deficient MM cell lines (Landau et al., 2012). Cyclin-dependent kinase (CDK) inhibitors have been identified to impair HRR and sensitize cancer cells to DNA-damaging agents (Jorda et al., 2011; Raghavan et al., 2012). CDK1 and 2 phosphorylate BRCA1 with an important role in the formation of BRCA1 and RAD51 foci and HRR (Ruffner et al., 1999; Johnson et al., 2011). CDK deregulation is a hallmark of MM, and CDK inhibitors are currently under clinical evaluation in MM (Bergsagel et al., 2005; Kumar et al., 2015; Niesvizky et al., 2015). Alagpulinsa et al. (2016) demonstrated that dinaciclib, an inhibitor of CDK, impairs HRR in MM cells and sensitizes them to PARP inhibitor without toxicity to normal B cells. Maes et al. (2014) reported DNA damage induction by DNA methyltransferase inhibitor decitabine in MM cells and stimulation of HRR and NHEJ pathways. Interestingly, combination with HDAC inhibitor or RAD51 inhibitor significantly enhances decitabine-mediated toxicity (Maes et al., 2014). These combinations could be of therapeutic interest in MM with conventional DNA-damaging agents. Several other epigenetic drugs have been shown to enhance the effect of DNA repair inhibitors in other cancers (Orta et al., 2014; Wu et al., 2015). Histone lysine methyltransferase inhibitors could prevent the BRCA1/BARD1 complex at DSBs (Wu et al., 2015).

Achieving success in the clinic with targeted DNA repair inhibitors in combination with DNA-damaging agents will need identification of the MM cell-specific deregulation associated with susceptibility to a DNA repair inhibitor. The GEP-based DNA repair scores recently reported could be of interest in stratifying MM patients and target the addiction of malignant plasma cells to a specific DNA repair pathway. In addition, maximizing the therapeutic window by identifying the correct dose and schedule will be important. In this context, persistence of viable malignant MM cells, within bone marrow, 7 days after high-dose melphalan and ASCT has been reported in two thirds of the patients (Caraux et al., 2012). This therapeutic window could be exploited for residual MM cell eradication before ASCT, taking advantage of new immune therapies presenting favorable toxicity profiles. Furthermore, modulating the redox system would be of therapeutic interest to optimize the use of genotoxic agents in MM (Gourzones et al., 2019).

TABLE 2 | DNA damage response inhibitors in clinical development.

| Target | Compound |
|------------------------------|--|
| BER | |
| PARP | Olaparib Rucaparib Niraparib Talazoparib Veliparib |
| APE1 | Methoxyamine |
| HRR | |
| ATR | VX-970 AZD6738 |
| NHEJ | |
| DNA-PKcs | MSC2490484A CC-115 |
| Checkpoint inhibitors | |
| CHK1 | GDC-0575 MK-8776 |
| CHK1 and CHK2 | LY2606368 PF-00477736 |
| ATM | AZD0156 |
| WEE1 | AZD11775 |

CONCLUSION

Genotoxic agents cause DNA damage that can be recognized by DNA damage response pathways. The accumulation of unrepaired DNA lesions is associated with MM cell apoptosis. MM cells evading DNA damage-induced apoptosis may acquire new genomic alterations that could confer selective advantage and drug resistance. Several mechanisms associated with MMC resistance to genotoxic agents have been described, underlining the MM's endemic heterogeneous landscape. These findings provide several therapeutic strategies to overcome drug resistance and limit mutagenic effects of genotoxic agents in MM. According to this heterogeneity, adopting precision medicine into clinical practice has the potential to improve MM disease management and treatment. Furthermore, another limitation arises from adverse toxicity on normal cells and tissues, underlining the

need to better understand the drug's mode of action to optimize between efficacy and harmful effects.

AUTHOR CONTRIBUTIONS

CG and CB participated in the writing of the paper. JM supervised the writing of the paper.

FUNDING

This work was supported by grants from the Institut National du Cancer, INCA, Paris, France, PLBIO15-256, and PLBIO18; 2018-160, ANR (TIE-Skip; 2017-CE15-0024-01), FFRMG, AF3M, Institut Universitaire de France and SIRIC Montpellier (INCa-DGOS-Inserm 6045).

REFERENCES

- Aguilera, A., and Gomez-Gonzalez, B. (2008). Genome instability: a mechanistic view of its causes and consequences. *Nat. Rev. Genet.* 9 (3), 204–217. doi: 10.1038/nrg2268
- Alagpulinsa, D. A., Ayyadevara, S., Yaccoby, S. and Shmookler Reis, R. J. (2016). A Cyclin-Dependent Kinase Inhibitor, Dinaciclib, Impairs Homologous Recombination and Sensitizes Multiple Myeloma Cells to PARP Inhibition. *Mol. Cancer Ther.* 15 (2), 241–250. doi: 10.1158/1535-7163.MCT-15-0660
- Alexandrakis, M. G., Passam, F. H., Kyriakou, D. S., Dambaki, K., Niniraki, M., and Stathopoulos, E. (2004). Ki-67 proliferation index: correlation with prognostic parameters and outcome in multiple myeloma. *Am. J. Clin. Oncol.* 27 (1), 8–13. doi: 10.1097/01.coc.0000045810.91816.41
- Arai, A., Chano, T., Futami, K., Furuichi, Y., Ikebuchi, K., Inui, T., et al. (2011). RECQL1 and WRN proteins are potential therapeutic targets in head and neck squamous cell carcinoma. *Cancer Res.* 71 (13), 4598–4607. doi: 10.1158/0008-5472.CAN-11-0320
- Arnoult, N., Correia, A., Ma, J., Merlo, A., Garcia-Gomez, S., Maric, M., et al. (2017). Regulation of DNA repair pathway choice in S and G2 phases by the NHEJ inhibitor CYREN. *Nature* 49 (7673), 548–552. doi: 10.1038/nature24023
- Aymard, F., Bugler, B., Schmidt, C. K., Guillou, E., Caron, P., Briois, S., et al. (2014). Transcriptionally active chromatin recruits homologous recombination at DNA double-strand breaks. *Nat. Struct. Mol. Biol.* 21 (4), 366–374. doi: 10.1038/nsmb.2796
- Barlogie, B., Tricot, G., Rasmussen, E., Anaissie, E., Van Rhee, F., Zangari, M., et al. (2006). Total therapy 2 without thalidomide in comparison with total therapy 1: role of intensified induction and posttransplantation consolidation therapies. *Blood.* 107 (7), 2633–2638. doi: 10.1182/blood-2005-10-4084
- Barlogie, B., Tricot, G. J., Van Rhee, F., Angtuaco, E., Walker, R., Epstein, J., et al. (2006). Long-term outcome results of the first tandem autotransplant trial for multiple myeloma. *Br. J. Haematol.* 135 (2), 158–164. doi: 10.1111/j.1365-2141.2006.06271.x
- Bergsagel, P. L., Kuehl, W. M., Zhan, F., Sawyer, J., Barlogie, B., and Shaughnessy, J., Jr. (2005). Cyclin D dysregulation: an early and unifying pathogenic event in multiple myeloma. *Blood.* 106 (1), 296–303. doi: 10.1182/blood-2005-01-0034
- Boboila, C., Oksenysh, V., Gostissa, M., Wang, J. H., Zha, S., Zhang, Y., et al. (2012). Robust chromosomal DNA repair via alternative end-joining in the absence of X-ray repair cross-complementing protein 1 (XRCC1). *Proc. Natl. Acad. Sci. U.S.A.* 109 (7), 2473–2478. doi: 10.1073/pnas.1121470109
- Bothmer, A., Robbiani, D. F., Feldhahn, N., Gazumyan, A., Nussenzweig, A., and Nussenzweig, M. C. (2010). 53BP1 regulates DNA resection and the choice between classical and alternative end joining during class switch recombination. *J. Exp. Med.* 207 (4), 855–865. doi: 10.1084/jem.20100244
- Braggio, E., Kortum, K. M., and Stewart, A. K. (2015). SnapShot: multiple myeloma. *Cancer Cell.* 28 (5), 678 e671. doi: 10.1016/j.ccell.2015.10.014
- Bret, C., Klein, B., and Moreaux, J. (2013). Nucleotide excision DNA repair pathway as a therapeutic target in patients with high-risk diffuse large B cell lymphoma. *Cell Cycle* 12 (12), 1811–1812. doi: 10.4161/cc.25115
- Brown, J. S., O'Carrigan, B., Jackson, S. P., and Yap, T. A. (2017). Targeting DNA repair in cancer: beyond PARP Inhibitors. *Cancer Discov.* 7 (1), 20–37. doi: 10.1158/2159-8290.CD-16-0860
- Bryant, H. E., Schultz, N., Thomas, H. D., Parker, K. M., Flower, D., Lopez, E., et al. (2005). Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature* 434 (7035), 913–917. doi: 10.1038/nature03443
- Caldecott, K. W. (2008). Single-strand break repair and genetic disease. *Nat. Rev. Genet.* 9 (8), 619–631. doi: 10.1038/nrg2380
- Calimeri, T., Fulciniti, M., Lin, J., Samur, M. K., Calkins, A. S., Vahia, A. V., et al. (2012). Aberrant non-homologous end joining in multiple myeloma: a role in genomic instability and as potential prognostic marker. *Blood.* 120 (21), 2932–2932.
- Caraux, A., Vincent, L., Bouhya, S., Quittet, P., Moreaux, J., Requirand, G., et al. (2012). Residual malignant and normal plasma cells shortly after high dose melphalan and stem cell transplantation. Highlight of a putative therapeutic window in Multiple Myeloma? *Oncotarget* 3 (11), 1335–1347. doi: 10.18632/oncotarget.650
- Carvalho, S., Vitor, A. C., Sridhara, S. C., Martins, F. B., Raposo, A. C., Desterro, J. M., et al. (2014). SETD2 is required for DNA double-strand break repair and activation of the p53-mediated checkpoint. *eLife.* 3, e02482. doi: 10.7554/eLife.02482
- Cavo, M., Tacchetti, P., Patriarca, F., Petrucci, M. T., Pantani, L., Galli, M., et al. (2010). Bortezomib with thalidomide plus dexamethasone compared with thalidomide plus dexamethasone as induction therapy before, and consolidation therapy after, double autologous stem-cell transplantation in newly diagnosed multiple myeloma: a randomised phase 3 study. *Lancet* 376 (9758), 2075–2085. doi: 10.1016/S0140-6736(10)61424-9
- Chapman, M. A., Lawrence, M. S., Keats, J. J., Cibulkis, K., Sougnez, C., Schinzel, A. C., et al. (2011). Initial genome sequencing and analysis of multiple myeloma. *Nature* 471 (7339), 467–472. doi: 10.1038/nature09837
- Chen, Q., Van der Sluis, P. C., Boulware, D., Hazlehurst, L. A., and Dalton, W. S. (2005). The FA/BRCA pathway is involved in melphalan-induced DNA interstrand cross-link repair and accounts for melphalan resistance in multiple myeloma cells. *Blood.* 106 (2), 698–705. doi: 10.1182/blood-2004-11-4286
- Ciccia, A., and Elledge, S. J. (2010). The DNA damage response: making it safe to play with knives. *Mol. Cell.* 40 (2), 179–204. doi: 10.1016/j.molcel.2010.09.019
- Cives, M., Ciavarella, S., Rizzo, F. M., De Matteo, M., Dammacco, F., and Silvestris, E. (2013). Bendamustine overcomes resistance to melphalan in myeloma cell lines by inducing cell death through mitotic catastrophe. *Cell Signal.* 25 (5), 1108–1117. doi: 10.1016/j.cellsig.2013.01.020
- Condomines, M., Veyrune, J. L., Larroque, M., Quittet, P., Latry, P., Lugagne, C., et al. (2010). Increased plasma-immune cytokines throughout the high-dose melphalan-induced lymphodepletion in patients with multiple myeloma: a window for adoptive immunotherapy. *J. Immunol.* 184 (2), 1079–1084. doi: 10.4049/jimmunol.0804159
- Cottini, F., Hideshima, T., Suzuki, R., Tai, Y. T., Bianchini, G., Richardson, P. G., et al. (2015). Synthetic Lethal Approaches Exploiting DNA Damage in Aggressive Myeloma. *Cancer Discov.* 5 (9), 972–987. doi: 10.1158/2159-8290.CD-14-0943

- Curtin, N. J. (2012). DNA repair dysregulation from cancer driver to therapeutic target. *Nat. Rev. Cancer* 12 (12), 801–817. doi: 10.1038/nrc3399
- Curtin, N. J. (2013). Inhibiting the DNA damage response as a therapeutic manoeuvre in cancer. *Br. J. Pharmacol.* 169 (8), 1745–1765. doi: 10.1111/bph.12244
- Dadacaridou, M., Papanicolaou, X., Maltesas, D., Megalaki, C., Patos, P., Panteli, K., et al. (2007). Dexamethasone, cyclophosphamide, etoposide and cisplatin (DCEP) for relapsed or refractory multiple myeloma patients. *J. BUON*. 12 (1), 41–44.
- Dalton, W. S. (1997). Detection of multidrug resistance gene expression in multiple myeloma. *Leukemia* 11 (7), 1166–1169. doi: 10.1038/sj.leu.2400724
- Damiano, J. S., Cress, A. E., Hazlehurst, L. A., Shtil, A. A., and Dalton, W. S. (1999). Cell adhesion mediated drug resistance (CAM-DR): role of integrins and resistance to apoptosis in human myeloma cell lines. *Blood*. 93 (5), 1658–1667.
- Deans, A. J., and West, S. C. (2011). DNA interstrand crosslink repair and cancer. *Nat. Rev. Cancer* 11 (7), 467–480. doi: 10.1038/nrc3088
- Demchenko, Y. N., and Kuehl, W. M. (2010). A critical role for the NFκB pathway in multiple myeloma. *Oncotarget* 1 (1), 59–68. doi: 10.18632/oncotarget.109
- Di Marzo, L., Desantis, V., Solimando, A. G., Ruggieri, S., Annesse, T., Nico, B., et al. (2016). Microenvironment drug resistance in multiple myeloma: emerging new players. *Oncotarget*. 7, 60698–60711 doi: 10.18632/oncotarget.10849
- Dimopoulos, M. A., Souliotis, V. L., Anagnostopoulos, A., Bamia, C., Pouli, A., Baltadakis, I., et al. (2007). Melphalan-induced DNA damage *in vitro* as a predictor for clinical outcome in multiple myeloma. *Haematologica* 92 (11), 1505–1512. doi: 10.3324/haematol.11435
- Dobbelstein, M., and Sorensen, C. S. (2015). Exploiting replicative stress to treat cancer. *Nat. Rev. Drug Discov.* 14 (6), 405–423. doi: 10.1038/nrd4553
- Dumontet, C., Landi, S., Reiman, T., Perry, T., Plesa, A., Bellini, I., et al. (2010). Genetic polymorphisms associated with outcome in multiple myeloma patients receiving high-dose melphalan. *Bone Marrow Transplant* 45 (8), 1316–1324. doi: 10.1038/bmt.2009.335
- Farmer, H., McCabe, N., Lord, C. J., Tutt, A. N., Johnson, D. A., Richardson, T. B., et al. (2005). Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature* 434 (7035), 917–921. doi: 10.1038/nature03445
- Fnu, S., Williamson, E. A., De Haro, L. P., Brenneman, M., Wray, J., Shaheen, M., et al. (2011). Methylation of histone H3 lysine 36 enhances DNA repair by nonhomologous end-joining. *Proc. Natl. Acad. Sci. U.S.A.* 108 (2), 540–545. doi: 10.1073/pnas.1013571108
- Fordham, S. E., Matheson, E. C., Scott, K., Irving, J. A., and Allan, J. M. (2011). DNA mismatch repair status affects cellular response to Ara-C and other anti-leukemic nucleoside analogs. *Leukemia* 25 (6), 1046–1049. doi: 10.1038/leu.2011.38
- Friedberg, E. C. (2001). How nucleotide excision repair protects against cancer. *Nat. Rev. Cancer* 1 (1), 22–33. doi: 10.1038/35094000
- Friedenberg, W. R., Rue, M., Blood, E. A., Dalton, W. S., Shustik, C., Larson, R. A., et al. (2006). Phase III study of PSC-833 (valsopodar) in combination with vincristine, doxorubicin, and dexamethasone (valsopodar/VAD) versus VAD alone in patients with recurring or refractory multiple myeloma (E1A95): a trial of the Eastern Cooperative Oncology Group. *Cancer* 106 (4), 830–838. doi: 10.1002/cncr.21666
- Futami, K., Kumagai, E., Makino, H., Goto, H., Takagi, M., Shimamoto, A., et al. (2008a). Induction of mitotic cell death in cancer cells by small interference RNA suppressing the expression of RecQL1 helicase. *Cancer Sci.* 99 (1), 71–80. doi: 10.1111/j.1349-7006.2007.00647.x
- Futami, K., Kumagai, E., Makino, H., Sato, A., Takagi, M., Shimamoto, A., et al. (2008b). Anticancer activity of RecQL1 helicase siRNA in mouse xenograft models. *Cancer Sci.* 99 (6), 1227–1236. doi: 10.1111/j.1349-7006.2008.00794.x
- Gennerly, A. R., Cant, A. J., and Jeggo, P. A. (2000). Immunodeficiency associated with DNA repair defects. *Clin. Exp. Immunol.* 121 (1), 1–7. doi: 10.1046/j.1365-2249.2000.01257.x
- Gourzones, C., Bellanger, C., Lamure, S., Gadacha, O. K., De Paco, E. G., Vincent, L., et al. (2019). antioxidant defenses confer resistance to high dose melphalan in multiple myeloma cells. *Cancers (Basel)* 11 (4): E439. doi: 10.3390/cancers11040439
- Gourzones-Dmitriev, C., Kassambara, A., Sahota, S., Reme, T., Moreaux, J., Bourquard, P., et al. (2013). DNA repair pathways in human multiple myeloma: role in oncogenesis and potential targets for treatment. *Cell Cycle* 12 (17), 2760–2773. doi: 10.4161/cc.25951
- Greipp, P. R., Katzmann, J. A., O'Fallon, W. M., and Kyle, R. A. (1988). Value of beta 2-microglobulin level and plasma cell labeling indices as prognostic factors in patients with newly diagnosed myeloma. *Blood*. 72 (1), 219–223.
- Grogan, T. M., Spier, C. M., Salmon, S. E., Matzner, M., Rybski, J., Weinstein, R. S., et al. (1993). P-glycoprotein expression in human plasma cell myeloma: correlation with prior chemotherapy. *Blood*. 81 (2), 490–495.
- Hajdu, I., Ciccia, A., Lewis, S. M., and Elledge, S. J. (2011). Wolf-Hirschhorn syndrome candidate 1 is involved in the cellular response to DNA damage. *Proc. Natl. Acad. Sci. U.S.A.* 108 (32), 13130–13134. doi: 10.1073/pnas.1110081108
- Hanawalt, P. C., and Spivak, G. (2008). Transcription-coupled DNA repair: two decades of progress and surprises. *Nat. Rev. Mol. Cell. Biol.* 9 (12), 958–970. doi: 10.1038/nrm2549
- Hartlerode, A. J., and Scully, R. (2009). Mechanisms of double-strand break repair in somatic mammalian cells. *Biochem. J.* 423 (2), 157–168. doi: 10.1042/BJ20090942
- Hassen, W., Kassambara, A., Reme, T., Sahota, S., Seckinger, A., Vincent, L., et al. (2014). Drug metabolism and clearance system in tumor cells of patients with multiple myeloma. *Oncotarget*. 6, 6431–6447 doi: 10.18632/oncotarget.3237
- Hayden, P. J., Tewari, P., Morris, D. W., Staines, A., Crowley, D., Nieters, A., et al. (2007). Variation in DNA repair genes XRCC3, XRCC4, XRCC5 and susceptibility to myeloma. *Hum. Mol. Genet.* 16 (24), 3117–3127. doi: 10.1093/hmg/ddm273
- Hazlehurst, L. A., Damiano, J. S., Buyuksal, I., Pledger, W. J., and Dalton, W. S. (2000). Adhesion to fibronectin *via* beta1 integrins regulates p27kip1 levels and contributes to cell adhesion mediated drug resistance (CAM-DR). *Oncogene* 19 (38), 4319–4327. doi: 10.1038/sj.onc.1203782
- Herrero, A. B., San Miguel, J., and Gutierrez, N. C. (2015). Deregulation of DNA double-strand break repair in multiple myeloma: implications for genome stability. *PLoS One* 10 (3), e0121581. doi: 10.1371/journal.pone.0121581
- Hsiao, K. Y., and Mizzen, C. A. (2013). Histone H4 deacetylation facilitates 53BP1 DNA damage signaling and double-strand break repair. *J. Mol. Cell Biol.* 5 (3), 157–165. doi: 10.1093/jmcb/mjs066
- Hung, P. J., Johnson, B., Chen, B. R., Byrum, A. K., Bredemeyer, A. L., Yewdell, W. T., et al. (2018). MRI Is a DNA Damage Response Adaptor during Classical Non-homologous End Joining. *Mol. Cell*. 71 (2), 332–342 e338. doi: 10.1016/j.molcel.2018.06.018
- Jacquemont, C., and Taniguchi, T. (2007). Proteasome function is required for DNA damage response and fanconi anemia pathway activation. *Cancer Res.* 67 (15), 7395–7405. doi: 10.1158/0008-5472.CAN-07-1015
- Jasin, M., and Rothstein, R. (2013). Repair of strand breaks by homologous recombination. *Cold Spring Harb. Perspect. Biol.* 5 (11), a012740. doi: 10.1101/cshperspect.a012740
- Johnson, N., Li, Y. C., Walton, Z. E., Cheng, K. A., Li, D., Rodig, S. J., et al. (2011). Compromised CDK1 activity sensitizes BRCA-proficient cancers to PARP inhibition. *Nat. Med.* 17 (7), 875–882. doi: 10.1038/nm.2377
- Jorda, R., Havlicek, L., Mcnae, I. W., Walkinshaw, M. D., Voller, J., Sturc, A., et al. (2011). Pyrazolo[4,3-d]pyrimidine bioisostere of roscovitine: evaluation of a novel selective inhibitor of cyclin-dependent kinases with antiproliferative activity. *J. Med. Chem.* 54 (8), 2980–2993. doi: 10.1021/jm200064p
- Karran, P., and Bignami, M. (1994). DNA damage tolerance, mismatch repair and genome instability. *Bioessays* 16 (11), 833–839. doi: 10.1002/bies.950161110
- Kassambara, A., Gourzones-Dmitriev, C., Sahota, S., Reme, T., Moreaux, J., Goldschmidt, H., et al. (2014). A DNA repair pathway score predicts survival in human multiple myeloma: the potential for therapeutic strategy. *Oncotarget* 5 (9), 2487–2498. doi: 10.18632/oncotarget.1740
- Kathawala, R. J., Gupta, P., Ashby, C. R., Jr., and Chen, Z. S. (2015). The modulation of ABC transporter-mediated multidrug resistance in cancer: a review of the past decade. *Drug Resist Updat.* 18, 1–17. doi: 10.1016/j.drug.2014.11.002
- Kirschner, K., and Melton, D. W. (2010). Multiple roles of the ERCC1-XPF endonuclease in DNA repair and resistance to anticancer drugs. *Anticancer Res.* 30 (9), 3223–3232.
- Kotnis, A., Du, L., Liu, C., Popov, S. W., and Pan-Hammarstrom, Q. (2009). Non-homologous end joining in class switch recombination: the beginning of the end. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 364 (1517), 653–665. doi: 10.1098/rstb.2008.0196
- Kumar, S. K., Laplant, B., Chng, W. J., Zonder, J., Callander, N., Fonseca, R., et al. (2015). Dinaciclib, a novel CDK inhibitor, demonstrates encouraging single-agent activity in patients with relapsed multiple myeloma. *Blood* 125 (3), 443–448. doi: 10.1182/blood-2014-05-573741

- Ladetto, M., Pagliano, G., Ferrero, S., Cavallo, F., Drandi, D., Santo, L., et al. (2010). Major tumor shrinking and persistent molecular remissions after consolidation with bortezomib, thalidomide, and dexamethasone in patients with autografted myeloma. *J. Clin. Oncol.* 28 (12), 2077–2084. doi: 10.1200/JCO.2009.23.7172
- Landau, H. J., Mcneely, S. C., Nair, J. S., Comenzo, R. L., Asai, T., Friedman, H., et al. (2012). The checkpoint kinase inhibitor AZD7762 potentiates chemotherapy-induced apoptosis of p53-mutated multiple myeloma cells. *Mol. Cancer Ther.* 11 (8), 1781–1788. doi: 10.1158/1535-7163.MCT-11-0949
- Li, Y., Trojer, P., Xu, C. F., Cheung, P., Kuo, A., Drury, W. J., et al. (2009). The target of the NSD family of histone lysine methyltransferases depends on the nature of the substrate. *J. Biol. Chem.* 284 (49), 34283–34295. doi: 10.1074/jbc.M109.034462
- Lieber, M. R. (2010). NHEJ and its backup pathways in chromosomal translocations. *Nat. Struct. Mol. Biol.* 17 (4), 393–395. doi: 10.1038/nsmb0410-393
- Lindahl, T. (1993). Instability and decay of the primary structure of DNA. *Nature* 362 (6422), 709–715. doi: 10.1038/362709a0
- Lohr, J. G., Stojanov, P., Carter, S. L., Cruz-Gordillo, P., Lawrence, M. S., Auclair, D., et al. (2014). Widespread genetic heterogeneity in multiple myeloma: implications for targeted therapy. *Cancer Cell.* 25 (1), 91–101. doi: 10.1016/j.ccr.2013.12.015
- Maes, K., De Smedt, E., Lemaire, M., De Raeve, H., Menu, E., Van Valckenborgh, E., et al. (2014). The role of DNA damage and repair in decitabine-mediated apoptosis in multiple myeloma. *Oncotarget* 5 (10), 3115–3129. doi: 10.18632/oncotarget.1821
- Mahtouk, K., Moreaux, J., Hose, D., Reme, T., Meissner, T., Jourdan, M., et al. (2010). Growth factors in multiple myeloma: a comprehensive analysis of their expression in tumor cells and bone marrow environment using Affymetrix microarrays. *BMC Cancer.* 10, 198. doi: 10.1186/1471-2407-10-198
- Marchesini, M., Ogoti, Y., Fiorini, E., Aktas Samur, A., Nezi, L., D'anca, M., et al. (2017). ILF2 Is a Regulator of RNA Splicing and DNA Damage Response in 1q21-Amplified Multiple Myeloma. *Cancer Cell.* 32 (1), 88–100 e106. doi: 10.1016/j.ccell.2017.05.011
- Martinez-Garcia, E., Popovic, R., Min, D. J., Sweet, S. M., Thomas, P. M., Zamdborg, L., et al. (2010). The MMSET histone methyl transferase switches global histone methylation and alters gene expression in t(4;14) multiple myeloma cells. *Blood* 117, 211–220 doi: 10.1182/blood-2010-07-298349
- Martinez-Lopez, J., Blade, J., Mateos, M. V., Grande, C., Alegre, A., Garcia-Larana, J., et al. (2011). Long-term prognostic significance of response in multiple myeloma after stem cell transplantation. *Blood* 118 (3), 529–534. doi: 10.1182/blood-2011-01-332320
- McNeill, D. R., Lam, W., DeWeese, T. L., Cheng, Y. C., and Wilson, D. M. 3rd. (2009). Impairment of APE1 function enhances cellular sensitivity to clinically relevant alkylators and antimetabolites. *Mol. Cancer Res.* 7 (6), 897–906. doi: 10.1158/1541-7786.MCR-08-0519
- Mladenov, E., and Iliakis, G. (2011). Induction and repair of DNA double strand breaks: the increasing spectrum of non-homologous end joining pathways. *Mutation Res.* 711 (1–2), 61–72. doi: 10.1016/j.mrfmmm.2011.02.005
- Moreau, P., Avet-Loiseau, H., Facon, T., Attal, M., Tiab, M., Hulin, C., et al. (2011). Bortezomib plus dexamethasone versus reduced-dose bortezomib, thalidomide plus dexamethasone as induction treatment before autologous stem cell transplantation in newly diagnosed multiple myeloma. *Blood.* 118 (22), 5752–5758. doi: 10.1182/blood-2011-05-355081
- Muniandy, P. A., Liu, J., Majumdar, A., Liu, S. T., and Seidman, M. M. (2010). DNA interstrand crosslink repair in mammalian cells: step by step. *Crit. Rev. Biochem. Mol. Biol.* 45 (1), 23–49. doi: 10.3109/10409230903501819
- Murakawa, Y., Sonoda, E., Barber, L. J., Zeng, W., Yokomori, K., Kimura, H., et al. (2007). Inhibitors of the proteasome suppress homologous DNA recombination in mammalian cells. *Cancer Res.* 67 (18), 8536–8543. doi: 10.1158/0008-5472.CAN-07-1166
- Neben, K., Lokhorst, H. M., Jauch, A., Bertsch, U., Hielscher, T., Van Der Holt, B., et al. (2012). Administration of bortezomib before and after autologous stem cell transplantation improves outcome in multiple myeloma patients with deletion 17p. *Blood.* 119 (4), 940–948. doi: 10.1182/blood-2011-09-379164
- Neri, P., Ren, L., Azab, A. K., Brentnall, M., Gratton, K., Klimowicz, A. C., et al. (2011a). Integrin beta7-mediated regulation of multiple myeloma cell adhesion, migration, and invasion. *Blood.* 117 (23), 6202–6213. doi: 10.1182/blood-2010-06-292243
- Neri, P., Ren, L., Gratton, K., Stebner, E., Johnson, J., Klimowicz, A., et al. (2011b). Bortezomib-induced “BRCAness” sensitizes multiple myeloma cells to PARP inhibitors. *Blood.* 118 (24), 6368–6379. doi: 10.1182/blood-2011-06-363911
- Niesvizky, R., Badros, A. Z., Costa, L. J., Ely, S. A., Singhal, S. B., Stadtmauer, E. A., et al. (2015). Phase 1/2 study of cyclin-dependent kinase (CDK)4/6 inhibitor palbociclib (PD-0332991) with bortezomib and dexamethasone in relapsed/refractory multiple myeloma. *Leuk. Lymphoma.* 56 (12), 3320–3328. doi: 10.3109/10428194.2015.1030641
- Noborio-Hatano, K., Kikuchi, J., Takatoku, M., Shimizu, R., Wada, T., Ueda, M., et al. (2009). Bortezomib overcomes cell-adhesion-mediated drug resistance through downregulation of VLA-4 expression in multiple myeloma. *Oncogene* 28 (2), 231–242. doi: 10.1038/onc.2008.385
- Nussenzweig, A., and Nussenzweig, M. C. (2007). A backup DNA repair pathway moves to the forefront. *Cell* 131 (2), 223–225. doi: 10.1016/j.cell.2007.10.005
- O'Connor, M. J. (2015). Targeting the DNA Damage Response in Cancer. *Mol. Cell.* 60 (4), 547–560. doi: 10.1016/j.molcel.2015.10.040
- Orta, M. L., Hoglund, A., Calderon-Montano, J. M., Dominguez, I., Burgos-Moron, E., Visnes, T., et al. (2014). The PARP inhibitor Olaparib disrupts base excision repair of 5-aza-2'-deoxycytidine lesions. *Nucleic Acids Res.* 42 (14), 9108–9120. doi: 10.1093/nar/gku638
- Osborne, M. R., Wilman, D. E., and Lawley, P. D. (1995). Alkylation of DNA by the nitrogen mustard bis(2-chloroethyl)methylamine. *Chem. Res. Toxicol.* 8 (2), 316–320. doi: 10.1021/tx00044a018
- Pai, C. C., Deegan, R. S., Subramanian, L., Gal, C., Sarkar, S., Blaikley, E. J., et al. (2014). A histone H3K36 chromatin switch coordinates DNA double-strand break repair pathway choice. *Nat. Commun.* 5, 4091. doi: 10.1038/ncomms5091
- Park, S., Lee, S. J., Jung, C. W., Jang, J. H., Kim, S. J., Kim, W. S., et al. (2014). DCEP for relapsed or refractory multiple myeloma after therapy with novel agents. *Ann. Hematol.* 93 (1), 99–105. doi: 10.1007/s00277-013-1952-5
- Pei, H., Zhang, L., Luo, K., Qin, Y., Chesi, M., Fei, F., et al. (2011). MMSET regulates histone H4K20 methylation and 53BP1 accumulation at DNA damage sites. *Nature* 470 (7332), 124–128. doi: 10.1038/nature09658
- Pei, H., Wu, X., Liu, T., Yu, K., Jelinek, D. F., and Lou, Z. (2013). The histone methyltransferase MMSET regulates class switch recombination. *J. Immunol.* 190 (2), 756–763. doi: 10.4049/jimmunol.1201811
- Pfister, S. X., Ahrabi, S., Zalmas, L. P., Sarkar, S., Aymard, F., Bachrati, C. Z., et al. (2014). SETD2-dependent histone H3K36 trimethylation is required for homologous recombination repair and genome stability. *Cell Rep.* 7 (6), 2006–2018. doi: 10.1016/j.celrep.2014.05.026
- Podar, K., Chauhan, D., and Anderson, K. C. (2009). Bone marrow microenvironment and the identification of new targets for myeloma therapy. *Leukemia* 23 (1), 10–24. doi: 10.1038/leu.2008.259
- Popovic, R., Martinez-Garcia, E., Giannopoulou, E. G., Zhang, Q., Zhang, Q., Ezponda, T., et al. (2014). Histone methyltransferase MMSET/NSD2 alters EZH2 binding and reprograms the myeloma epigenome through global and focal changes in H3K36 and H3K27 methylation. *PLoS Genet.* 10 (9), e1004566. doi: 10.1371/journal.pgen.1004566
- Poruchynsky, M. S., Komlodi-Pasztor, E., Trostel, S., Wilkerson, J., Regairaz, M., Pommier, Y., et al. (2015). Microtubule-targeting agents augment the toxicity of DNA-damaging agents by disrupting intracellular trafficking of DNA repair proteins. *Proc. Natl. Acad. Sci. U.S.A.* 112 (5), 1571–1576. doi: 10.1073/pnas.1416418112
- Raghavan, P., Tumati, V., Yu, L., Chan, N., Tomimatsu, N., Burma, S., et al. (2012). AZD5363, an inhibitor of Cdk1, 2, and 9, enhances the radiosensitivity of non-small cell lung carcinoma cells. *Int. J. Radiat. Oncol. Biol. Phys.* 84 (4), e507–e514. doi: 10.1016/j.ijrobp.2012.05.035
- Ransom, M., Dennehey, B. K., and Tyler, J. K. (2010). Chaperoning histones during DNA replication and repair. *Cell* 140 (2), 183–195. doi: 10.1016/j.cell.2010.01.004
- Ray, A., Ravillah, D., Das, D. S., Song, Y., Nordstrom, E., Gullbo, J., et al. (2016). A novel alkylating agent Melflufen induces irreversible DNA damage and cytotoxicity in multiple myeloma cells. *Br. J. Haematol.* 174 (3), 397–409. doi: 10.1111/bjh.14065
- Roddam, P. L., Rollinson, S., O'Driscoll, M., Jeggo, P. A., Jack, A., and Morgan, G. J. (2002). Genetic variants of NHEJ DNA ligase IV can affect the risk of developing multiple myeloma, a tumour characterised by aberrant class switch recombination. *J. Med. Genet.* 39 (12), 900–905. doi: 10.1136/jmg.39.12.900
- Roddam, P. L., Allan, J. M., Dring, A. M., Worrillow, L. J., Davies, F. E., and Morgan, G. J. (2010). Non-homologous end-joining gene profiling reveals

- distinct expression patterns associated with lymphoma and multiple myeloma. *Br. J. Haematol.* 149 (2), 258–262. doi: 10.1111/j.1365-2141.2010.08088.x
- Rossi, J. F., Fegueux, N., Lu, Z. Y., Legouffe, E., Exbrayat, C., Bozonnat, M. C., et al. (2005). Optimizing the use of anti-interleukin-6 monoclonal antibody with dexamethasone and 140 mg/m² of melphalan in multiple myeloma: results of a pilot study including biological aspects. *Bone Marrow Transplant* 36 (9), 771–779. doi: 10.1038/sj.bmt.1705138
- Roussel, M., Moreau, P., Huynh, A., Mary, J. Y., Danho, C., Caillot, D., et al. (2010). Bortezomib and high-dose melphalan as conditioning regimen before autologous stem cell transplantation in patients with *de novo* multiple myeloma: a phase 2 study of the Intergroupe Francophone du Myelome (IFM). *Blood*. 115 (1), 32–37. doi: 10.1182/blood-2009-06-229658
- Ruffner, H., Jiang, W., Craig, A. G., Hunter, T., and Verma, I. M. (1999). BRCA1 is phosphorylated at serine 1497 *in vivo* at a cyclin-dependent kinase 2 phosphorylation site. *Mol. Cell. Biol.* 19 (7), 4843–4854. doi: 10.1128/MCB.19.7.4843
- San Miguel, J. F., Garcia-Sanz, R., Gonzalez, M., Moro, M. J., Hernandez, J. M., Ortega, F., et al. (1995). A new staging system for multiple myeloma based on the number of S-phase plasma cells. *Blood* 85 (2), 448–455.
- Shah, M. Y., Martinez-Garcia, E., Phillip, J. M., Chambliss, A. B., Popovic, R., Ezponda, T., et al. (2016). MMSET/WHSC1 enhances DNA damage repair leading to an increase in resistance to chemotherapeutic agents. *Oncogene*. 35, 5905–5915 doi: 10.1038/onc.2016.116
- Shaheen, M., Allen, C., Nickloff, J. A., and Hromas, R. (2011). Synthetic lethality: exploiting the addiction of cancer to DNA repair. *Blood* 117 (23), 6074–6082. doi: 10.1182/blood-2011-01-313734
- Shammas, M. A., Shmookler Reis, R. J., Koley, H., Batchu, R. B., Li, C., and Munshi, N. C. (2009). Dysfunctional homologous recombination mediates genomic instability and progression in myeloma. *Blood* 113 (10), 2290–2297. doi: 10.1182/blood-2007-05-089193
- Simsek, D., Brunet, E., Wong, S. Y., Katyal, S., Gao, Y., Mckinnon, P. J., et al. (2011). Crucial role for DNA ligase III in mitochondria but not in Xrcc1-dependent repair. *Nature* 471 (7337), 245–248. doi: 10.1038/nature09794
- Sonneveld, P., Schmidt-Wolf, I. G., Van Der Holt, B., El Jarari, L., Bertsch, U., Salwender, H., et al. (2001). Cyclosporin A combined with vincristine, doxorubicin and dexamethasone (VAD) compared with VAD alone in patients with advanced refractory multiple myeloma: an EORTC-HOVON randomized phase III study (06914). *Br. J. Haematol.* 115 (4), 895–902. doi: 10.1046/j.1365-2141.2001.03171.x
- Sonneveld, P., Ciucu, S., Weijermans, P., Beksac, M., Neuwirtova, R., Solbu, G., et al. (2012). Bortezomib induction and maintenance treatment in patients with newly diagnosed multiple myeloma: results of the randomized phase III HOVON-65/GMMG-HD4 trial. *J. Clin. Oncol.* 30 (24), 2946–2955. doi: 10.1200/JCO.2011.39.6820
- Soria, G., Polo, S. E., and Almouzni, G. (2012). Prime, repair, restore: the active role of chromatin in the DNA damage response. *Mol. Cell.* 46 (6), 722–734. doi: 10.1016/j.molcel.2012.06.002
- Spanswick, V. J., Craddock, C., Sekhar, M., Mahendra, P., Shankaranarayana, P., Hughes, R. G., et al. (2002). Repair of DNA interstrand crosslinks as a mechanism of clinical resistance to melphalan in multiple myeloma. *Blood* 100 (1), 224–229. doi: 10.1182/blood.V100.1.224
- Surget, S., Lemieux-Blanchard, E., Maiga, S., Descamps, G., Le Gouill, S., Moreau, P., et al. (2014). Bendamustine and melphalan kill myelomrough reactive oxygen species production and activation of the p53 pathway and do not overcome resistance to each other. *Leuk. Lymphoma*. 55 (9), 2165–2173. doi: 10.3109/10428194.2013.871277
- Szikriszt, B., Poti, A., Pipek, O., Krzystanek, M., Kanu, N., Molnar, J., et al. (2016). A comprehensive survey of the mutagenic impact of common cancer cytotoxics. *Genome Biol.* 17, 99. doi: 10.1186/s13059-016-0963-7
- Tadi, S. K., Tellier-Lebegue, C., Nemoz, C., Drevet, P., Audebert, S., Roy, S., et al. (2016). PAXX Is an Accessory c-NHEJ Factor that Associates with Ku70 and Has Overlapping Functions with XLF. *Cell Rep.* 17 (2), 541–555. doi: 10.1016/j.celrep.2016.09.026
- Tuzon, C. T., Spektor, T., Kong, X., Congdon, L. M., Wu, S., Schotta, G., et al. (2014). Concerted activities of distinct H4K20 methyltransferases at DNA double-strand breaks regulate 53BP1 nucleation and NHEJ-directed repair. *Cell Rep.* 8 (2), 430–438. doi: 10.1016/j.celrep.2014.06.013
- van de Velde, H. J., Liu, X., Chen, G., Cakana, A., Deraedt, W., and Bayssas, M. (2007). Complete response correlates with long-term survival and progression-free survival in high-dose therapy in multiple myeloma. *Haematologica* 92 (10), 1399–1406. doi: 10.3324/haematol.11534
- van Rhee, F., Giral, S., and Barlogie, B. (2014). The future of autologous stem cell transplantation in myeloma. *Blood* 124 (3), 328–333. doi: 10.1182/blood-2014-03-561985
- Vangsted, A., Gimsing, P., Klausen, T. W., Nexø, B. A., Wallin, H., Andersen, P., et al. (2007). Polymorphisms in the genes ERCC2, XRCC3 and CD3EAP influence treatment outcome in multiple myeloma patients undergoing autologous bone marrow transplantation. *Int. J. Cancer*. 120 (5), 1036–1045. doi: 10.1002/ijc.22411
- Vikova, V., Jourdan, M., Robert, N., Requirand, G., Boireau, S., Bruyer, A., et al. (2019). Comprehensive characterization of the mutational landscape in multiple myeloma cell lines reveals potential drivers and pathways associated with tumor progression and drug resistance. *Theranostics* 9 (2), 540–553. doi: 10.7150/thno.28374
- Viziteu, E., Klein, B., Basbous, J., Lin, Y. L., Hirtz, C., Gourzones, C., et al. (2016). RECQ helicases are deregulated in hematological malignancies in association with a prognostic value. *Biomark Res.* 4, 3. doi: 10.1186/s40364-016-0057-4
- Viziteu, E., Klein, B., Basbous, J., et al. (2017). RECQ1 helicase is involved in replication stress survival and drug resistance in multiple myeloma. *Leukemia* 10, 2104–2113. doi: 10.1038/leu.2017.54
- Voorhees, P. M., Orlowski, R. Z., Mulkey, E., Watson, P., Geyer, S., Sanford, B. L., et al. (2015). Long-term outcomes for newly-diagnosed multiple myeloma patients treated with pegylated liposomal doxorubicin and bortezomib: final results of CALGB (Alliance) 10301, a multicentre phase II study. *Br. J. Haematol.* 171 (3), 373–377. doi: 10.1111/bjh.13592
- Wang, Q., and Goldstein, M. (2016). Small RNAs recruit chromatin-modifying enzymes MMSET and Tip60 to reconfigure damaged DNA upon double-strand break and Facilitate Repair. *Cancer Res.* 76 (7), 1904–1915. doi: 10.1158/0008-5472.CAN-15-2334
- Weinhold, N., Ashby, C., Rasche, L., Chavan, S. S., Stein, C., Stephens, O. W., et al. (2016). Clonal selection and double-hit events involving tumor suppressor genes underlie relapse in myeloma. *Blood* 128 (13), 1735–1744. doi: 10.1182/blood-2016-06-723007
- Williams, R. S., Williams, J. S., and Tainer, J. A. (2007). Mre11-Rad50-Nbs1 is a keystone complex connecting DNA repair machinery, double-strand break signaling, and the chromatin template. *Biochem. Cell Biol.* 85 (4), 509–520. doi: 10.1139/O07-069
- Witte, A. B., Anestel, K., Jerremalm, E., Ehrsson, H., and Arner, E. S. (2005). Inhibition of thioredoxin reductase but not of glutathione reductase by the major classes of alkylating and platinum-containing anticancer compounds. *Free Radic. Biol. Med.* 39 (5), 696–703. doi: 10.1016/j.freeradbiomed.2005.04.025
- Wu, W., Nishikawa, H., Fukuda, T., Vittal, V., Asano, M., Miyoshi, Y., et al. (2015). Interaction of BARD1 and HP1 Is Required for BRCA1 Retention at Sites of DNA Damage. *Cancer Res.* 75 (7), 1311–1321. doi: 10.1158/0008-5472.CAN-14-2796
- Xiong, T., Wei, H., Chen, X., and Xiao, H. (2015). PJ34, a poly(ADP-ribose) polymerase (PARP) inhibitor, reverses melphalan-resistance and inhibits repair of DNA double-strand breaks by targeting the FA/BRCA pathway in multidrug resistant multiple myeloma cell line RPMI8226/R. *Int. J. Oncol.* 46 (1), 223–232. doi: 10.3892/ijo.2014.2726
- Yang, H. H., Ma, M. H., Vescio, R. A., and Berenson, J. R. (2003). Overcoming drug resistance in multiple myeloma: the emergence of therapeutic approaches to induce apoptosis. *J. Clin. Oncol.* 21 (22), 4239–4247. doi: 10.1200/JCO.2003.06.001
- Yarde, D. N., Oliveira, V., Mathews, L., Wang, X., Villagra, A., Boulware, D., et al. (2009). Targeting the Fanconi anemia/BRCA pathway circumvents drug resistance in multiple myeloma. *Cancer Res.* 69 (24), 9367–9375 doi: 10.1158/0008-5472.CAN-09-2616.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Gourzones, Bret and Moreaux. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.