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1 **Characterization of diblock copolymers by capillary electrophoresis: From**
2 **electrophoretic mobility distribution to distribution of composition**

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10 **Abstract**

11 Free solution capillary-electrophoresis (CE) is a powerful separation technique for the
12 characterization of diblock copolymers. In this work, four series of double-hydrophilic anionic and
13 cationic block copolymers, namely, poly(acrylamide)-*block*-poly(acrylic acid) (PAM-*b*-PAA),
14 poly(acrylamide)-*block*- poly((3-acrylamidopropyl)trimethylammonium chloride) (PAM-*b*-PAPTAC),
15 poly(ethylene oxide)-*block*-poly(acrylic acid) (PEO-*b*-PAA) and poly(poly(ethylene glycol) methyl
16 ether acrylate)-*block*-poly(acrylic acid) (P(PEGA)-*b*-PAA), were synthesized by reversible addition-
17 fragmentation chain transfer (RAFT) polymerization and characterized by CE. The electrophoretic
18 mobility distributions of the copolymers were transformed into distributions of composition ratio by
19 introducing a retardation parameter, X_{exp} , that represents the hydrodynamic drag retardation due to the
20 neutral block of the copolymer. A linear correlation between X_{exp} and the ratio of the degrees of
21 polymerization of each blocks was experimentally established and was consistent with the model of
22 electrophoretic mobility of composite macromolecules with hydrodynamic coupling. Finally, the
23 comparison of the distributions between the different copolymer families was significantly improved
24 by considering the distributions in composition ratio compared to the electrophoretic mobility
25 distributions, since it takes into account the differences in solvation, expansion and drag force
26 according to the chemical nature of the blocks.

27 1 Introduction

28 The characterization of diblock copolymers by separation techniques is challenging but of
29 primary importance to check their purity and to characterize their distributions in size and in chemical
30 composition.¹ Block copolymers prepared by reversible deactivation radical polymerization frequently
31 contain homopolymer impurities.^{2, 3} These include dead chains from termination reactions during
32 polymerization of the first block, as well as the products of side-reactions such as chain transfer to
33 solvent or monomer during polymerization of the second block. The growth of the second block in
34 copolymer synthesis is often evidenced by a decrease of the elution time in size-exclusion
35 chromatography (SEC) as a result of the increased hydrodynamic radius^{4, 5}. However, this only holds if
36 the second block contributes significantly to the hydrodynamic radius of the diblock copolymer.
37 Getting more quantitative information can be challenging especially in aqueous SEC. Different
38 solvation properties between the blocks of a block copolymer can lead to coelution of polymers of
39 different mass in SEC, resulting in inaccuracy in the obtained molar masses.⁶ Additionally,
40 interactions with the stationary phase³ may lead to HPLC-type elution which is dependent on the
41 chemical composition of the polymer as well as its size. For diblock copolymer SEC, particular elution
42 conditions are generally required and size distributions should be expressed in terms of hydrodynamic
43 radius (and not molar mass) due to the difference in chemical composition / solvation of the two
44 blocks⁶. The proportion of each monomer in a copolymer can be obtained by liquid chromatography
45 under critical conditions LCCC, also known as LC-PEAT, for the point of exclusion-adsorption
46 transition for neutral blocks^{7, 8}. The critical conditions for LCCC (or LC-PEAT) are usually difficult to
47 find and are very sensitive to small changes in mobile phase composition and/or temperature.

48 An alternative separation technique for charged copolymers is free solution capillary
49 electrophoresis (CE)^{2, 3, 7-9}. The electrophoretic separation of charged homopolymers from diblock
50 copolymers is generally easily obtained in free solution CE. Moreover, for self-assembling diblock
51 copolymers, CE can also separate micelles from unimers^{2, 7, 8} and allows studying the impact of added
52 surfactant on the copolymer micelles^{2, 7}. In the presence of cationic blocks, experimental difficulties

53 arise from polymer adsorption onto the wall of silica based capillaries. The characterization of cationic
54 diblock copolymers requires the use of a neutrally coated³, or positively charged capillary¹⁰.
55 Another challenging issue in the characterization of diblock copolymers by CE is to extract the
56 distribution in composition of the copolymers from the electropherogram. Raw electropherograms can
57 be transformed into distributions of effective mobility, or of any other related parameter, provided that
58 the relationship between effective mobility and the considered parameter is known¹¹. The
59 determination of polymer dispersity via the variance of the chemical composition distribution¹¹ or via
60 the calculation of the ratio of moments of the distribution has also been studied¹. A key point to
61 achieve such electropherogram transformation is to have a reliable relation between the effective
62 mobility of the diblock copolymer and the degrees of polymerization of each block and thus, to the
63 chemical composition of the copolymer³. The electrophoretic mobility of a diblock copolymer is
64 generally expressed as a weighted average of the mobilities of different subunits constituting the
65 copolymer¹². The choice of the subunits and the corresponding weights have been described in the
66 literature as depending on the conformations of each block and on the hydrodynamic coupling regime
67 between the two blocks¹²⁻¹⁵. More recently, Chubynsky and Slater studied in more detail the “end-
68 effect” (i.e. the fact that the ends of the copolymer chain are more hydrodynamically exposed to the
69 solvent)¹⁶ and the effect of polymer stiffness on the electrophoretic modeling¹⁷. The electrophoretic
70 models of composite objects^{12, 13} which are relevant for diblock copolymers, were also applied to end-
71 labeled free solution electrophoresis (ELFSE)^{14, 15}, which consists in attaching a monodisperse neutral
72 block (drag-tag) to a polydisperse biopolyelectrolyte (for instance, for DNA sequencing in free
73 solution^{15, 18}), or conversely, in attaching a monodisperse polyelectrolyte to a polydisperse neutral
74 polymer (for instance, for size-based neutral polymer characterization¹⁴). In this way, the dependence
75 of electrophoretic mobility with the molar mass of the end-labelled composite object is obtained in
76 free solution due to the variation of the charge-to-friction ratio.

77 Double-hydrophilic block copolymers (DHBC) are block copolymers containing two
78 hydrophilic segments. DHBCs on their own are completely soluble in water and do not self-assemble
79 in dilute conditions. DHBCs can still retain an amphiphilic character and this can lead to self-
80 organization at the meso-scale in concentrated conditions¹⁹. They can undergo morphological

81 transitions induced by external stimuli²⁰ in dilute solution. The great development of reversible
82 deactivation radical polymerization²¹⁻²⁴ in the last two decades allows tailoring the stimuli-
83 responsiveness (e.g. to changes in pH, temperature, ionic strength, or light) of these polymers by
84 controlling both the nature of monomers and the degree of polymerization of the blocks. When one
85 block is a polyelectrolyte, DHBCs can undergo micellization by electrostatic complexation in the
86 presence of an oppositely charged polyelectrolyte. These properties lead to a wide range of
87 applications such as control of crystallization of inorganic compounds²⁵, drug delivery²⁶ or template
88 for ordered mesoporous materials²⁷. For this last application of DHBC, the asymmetry ratio, defined as
89 the ratio of degrees of polymerization of both blocks, is of crucial interest since it determines the
90 structure of the DHBC-templated mesoporous materials.

91 It is the aim of the present work to characterize the composition of DHBC by CE, with particular
92 attention to the asymmetry ratio. The approach has been applied to series of anionic and cationic
93 DHBCs, namely, poly(acrylamide)-*block*-poly(acrylic acid) (PAM-*b*-PAA), poly(acrylamide)-*block*-
94 poly((3-acrylamidopropyl)trimethylammonium chloride) (PAM-*b*-PAPTAC), poly(ethylene oxide)-
95 *block*-poly(acrylic acid) (PEO-*b*-PAA) and poly(poly(ethylene glycol) methyl ether acrylate)-*block*-
96 poly(acrylic acid) (P(PEGA)-*b*-PAA). These DHBC have been synthesized in aqueous medium by
97 reversible addition-fragmentation transfer (RAFT) polymerization.

98 In the next section, different models for the electrophoretic mobility of composite objects such as
99 block copolymers are briefly reviewed. In the third section, the synthesis of the copolymers and the
100 experimental conditions of their characterization by CE are reported. The results of this work are
101 presented in section 4, where we describe a method to transform the distribution of electrophoretic
102 mobility into a distribution of the ratio of degree of polymerization of both blocks.

103 **2 Mobility of diblock copolymers constituted of a polyelectrolyte and a neutral block**

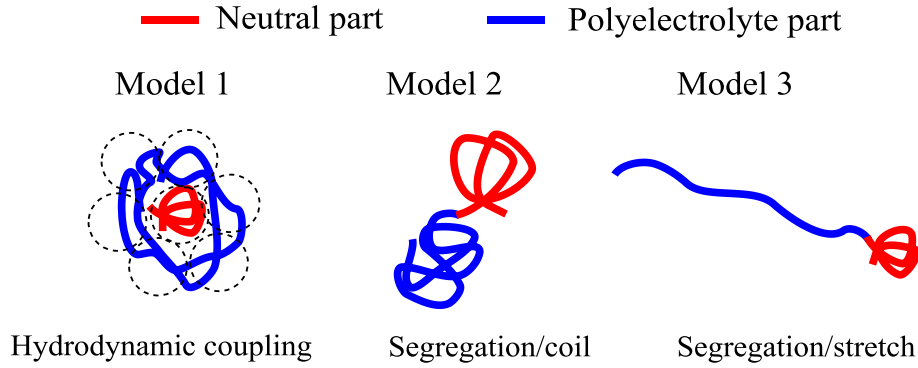


Figure 1. Representation of possible conformations of a double hydrophilic block copolymer composed of a neutral Gaussian coil (in red) linked to a polyelectrolyte block (in blue). In model 1, the polyelectrolyte chain is a coil in hydrodynamic interaction with the neutral Gaussian coil. In model 2, the polyelectrolyte and the neutral polymer separate into two coils. In model 3, the polyelectrolyte is fully stretched and has no hydrodynamic coupling with the neutral coil. Adapted from²⁸

104 In this section, we present different possible electrophoretic mobility models that are relevant
 105 for diblock copolymers composed of a polyelectrolyte part attached to a neutral polymer coil. These
 106 theoretical models were developed by Desruisseaux et al²⁸, building on previous work by Long et al.¹².
 107 ¹³ Figure 1 shows the different possible conformations that can be encountered for DHBC,
 108 corresponding to the different models presented below.

109 **2.1 Model 1: hydrodynamic coupling**

110 In Model 1, hydrodynamic coupling between the polyelectrolyte part and the neutral coil is
 111 taken into account. The polyelectrolyte block of the DHBC is composed of N_{blob} equivalent blobs of a
 112 size equivalent to the hydrodynamic radius of the neutral coil $R_h^{neutral}$. If $R_h^{neutral}$ is larger or equal to
 113 the Debye length, Long et al.²⁹ demonstrated that the electrophoretic mobility of the DHBC composite
 114 object composed of $N_{blob} + 1$ subunits of equal size, is given by the number-average of the
 115 electrophoretic mobilities calculated on all the equivalent blobs constituting the object. The
 116 electrophoretic mobility of the DHBC, $\mu_{ep,1}^{diblock}$, is thus given by²⁸:

$$117 \quad \mu_{ep,1}^{diblock} = \frac{\sum_{i=0}^{N_{blob}} \mu_{ep}^i}{N_{blob} + 1} = \frac{N_{blob} \mu_{ep}^0 + \mu_{ep}^{neutral}}{N_{blob} + 1} = \frac{\mu_{ep}^0}{1 + \frac{\alpha}{DP_0}} \quad (1)$$

118 where μ_{ep}^i is the effective mobility of the i^{th} entity (or blob) constituting the DHBC, DP_0 is the degree
 119 of polymerization of the polyelectrolyte block, α is the number of charged monomers per blob, μ_{ep}^0 is
 120 the effective electrophoretic mobility of the polyelectrolyte part (alone) and $\mu_{ep}^{neutral}$ is the
 121 electrophoretic mobility of the neutral part ($\mu_{ep}^{neutral} = 0$). The number of equivalent blobs in the
 122 polyelectrolyte chain is given by $N_{blob} = \frac{DP_0}{\alpha}$. Note that N_{blob} (and $\mu_{ep,1}^{diblock}$) depends on the
 123 polyelectrolyte persistence length, and thus, on the ionic strength. Equation (1) neglects the so-called
 124 end-effect¹⁶. Please note that subscript and superscript 0 refer to the polyelectrolyte block, for
 125 consistency with ref 28.

126 2.2 Models without hydrodynamic coupling

127 2.2.1 Model 2: polyelectrolyte chain in coil conformation

128 Model 2 in Figure 1 corresponds to the segregation of the neutral polymer coil from the
 129 polyelectrolyte coil. In the absence of hydrodynamic coupling between the two parts, and if the
 130 polyelectrolyte chain does not stretch during electrophoresis (i.e. at sufficiently low electric field), the
 131 electrophoretic mobility of the DHBC, $\mu_{ep,2}^{diblock}$, is given by the average electrophoretic mobility of the
 132 two parts weighted by their hydrodynamic friction coefficient¹². Using Stokes equation for spherical
 133 objects, $\mu_{ep,2}^{diblock}$ is expressed as^{13, 28}:

$$134 \quad \mu_{ep,2}^{diblock} = \frac{\sum_{i=0}^1 \gamma_i \mu_{ep}^i}{\sum_{i=0}^1 \gamma_i} = \frac{\mu_{ep}^0}{1 + \frac{R_h^{neutral}}{R_h^0}} \quad (2)$$

135 where γ_i is the friction coefficient of the i^{th} part constituting the DHBC, R_h^{neutral} is the hydrodynamic
 136 radius of the neutral coil, R_h^0 is the hydrodynamic radius of the polyelectrolyte block.

137 2.2.2 Model 3: polyelectrolyte chain in fully stretched conformation

138 When the polyelectrolyte coil is stretched under the concomitant influence of the electric field
 139 and the opposed drag force due to the presence of the neutral coil, it can reach a fully extended
 140 conformation as depicted in Figure 1 (Model 3). Stokes law is no longer appropriate for the frictional
 141 coefficient of the polyelectrolyte chain, and the electrophoretic mobility of the DHBC, $\mu_{ep,3}^{\text{diblock}}$, is
 142 given by²⁷ :

$$143 \mu_{ep,3}^{\text{diblock}} = \frac{\sum_{i=0}^1 \gamma_i \mu_{ep}^i}{\sum_{i=0}^1 \gamma_i} = \frac{\mu_{ep}^0}{1 + \frac{2R_h^{\text{neutral}}}{b_0 DP_0} \ln DP_0} \quad (3)$$

144 where b_0 is the size of a charged monomer in the polyelectrolyte chain. Equation (3) assumes that the
 145 friction coefficient γ_{rod} of the stretched polyelectrolyte cylinder is averaged on all orientations relative
 146 to the flow direction, and is given by:

$$147 \gamma_{rod} = \frac{3\pi\eta b_0 DP_0}{\ln(DP_0)} \quad (4)$$

148 where η is the viscosity of the solvent.

149 Even if the electrical field is not strong enough to stretch the polyelectrolyte block (hydrodynamic
 150 segregation), the polyelectrolyte contour length may still be shorter than the persistence length of the
 151 polyelectrolyte. In this situation, the segregation between the neutral and the polyelectrolyte parts is
 152 sterically obtained, but the electrophoretic mobility is still described by equation (3)²⁷.

153 3 Experimental

154 3.1 Chemicals

155 Ammonium persulfate (APS, 98%) and sodium formaldehyde sulfoxylate dehydrate (NaFS, 98%)
156 were purchased from Acros organics. Poly(ethylene glycol) methyl ether acrylate ($M_n = 480$ g/mol, 8.5
157 EO units on average) and (3-acrylamidopropyl)trimethylammonium chloride (APTAC) aqueous
158 solution (75 wt.%) were purchased from Sigma Aldrich (Saint-Quentin-Fallavier, France) and used as
159 received. Acrylic acid from the same suppliers was distilled under vacuum at room temperature. PEO
160 $M_n=5000$ g.mol⁻¹, $D = 1.04$ was purchased from. 4,4'-azobiscyanopentanoic acid (ACPA, Aldrich,
161 98%) and 2,2-Azobis(isobutyramidine) dihydrochloride (AIBA, Sigma-Aldrich, 97%) were used as
162 received.

163
164 For EC experiments, background electrolytes were prepared in ultra-pure water purified on a Millipore
165 system (Molsheim, France) from tris(hydroxymethyl)aminomethane (TRIS, 99,9%, Merck), 4-
166 Morpholinoethanesulphonic acid (MES, >99%, Acros Organics), and 2-[Bis(2-hydroxyethyl)amino]-
167 2-(hydroxymethyl)propane-1,3-diol (BIS-TRIS, >99%, Acros Organics). Anisic acid (99,5%),
168 ammediol (99,5%) used as markers for detection were purchased from Sigma-Aldrich.

169

170 3.2 Double-hydrophilic block copolymer synthesis

171 This section describes the synthesis of the copolymers. The synthetic pathway, the size exclusion
172 chromatograms and the ¹H NMR spectra are given in SI as well as the temporal electropherograms.

173 3.2.1 Synthesis of poly(acrylamide)-*b*-poly(acrylic acid)

174 Aqueous RAFT/MADIX polymerization of AA

175 Polyacrylamide macro RAFT/MADIX agent (PAM-Xa, $M_n = 5000$ g.mol⁻¹) was synthesized according
176 the procedure described by Layrac et al.³⁰ Synthesis of PAM₇₀-*b*-PAA₂₀ was performed as follows³¹:
177 PAM-based chain transfer agent (PAM-Xa,) (15.76 g, 3.029 mmol), acrylic acid (4.24 g; 58.77 mmol),
178 AIBA (0.0821 g, 0,3 mmol) and water (42 g) (solids = 30.6%) were introduced in a round bottom

179 flask. The mixture was degassed with argon at room temperature for 30 min and then placed in a
180 thermostated oil bath at 65°C under argon for 2 hours. Conversion was quantitative, acrylic acid traces
181 were eliminated by dialysis (MWCO 1000 Da) and pH-metric monitoring. The polymer solution was
182 then freeze dried and a white powder was obtained. Four DHBC were synthesized according to this
183 procedure: PAM₇₀-*b*-PAA₂₀, PAM₁₄₀-*b*-PAA₄₀, PAM₁₄₀-*b*-PAA₆₀ and PAM₁₄₀-*b*-PAA₈₀ (see Table 1).

184 **3.2.2 Synthesis of poly(acrylamide)-*b*-poly((3-acrylamidopropyl)trimethylammonium** 185 **chloride)**

186 **Aqueous redox RAFT/MADIX polymerization of APTAC**

187 Synthesis of PAM₇₀-*b*-PAPTAC₃₀ was performed as follows: two aqueous solutions of NaFS (5%w)
188 (1.23 g of solution, 0.4 mmol) and NaPS (5%w) (2.37 mg of solution, 0.5 mmol) were prepared.
189 PAM-Xa macroxanthate (9.97 g, 1.99 mmol), APTAC monomer (13.3 g of solution, 0.057 mmol), and
190 water (50 g) (solids = 25.4%) were introduced in a round bottom flask. The pH of the mixture was first
191 adjusted at 2 with hydrochloric solution 1M. Then the mixture was degassed with argon at room
192 temperature for 30 min and placed in a thermostated oil bath at 25°C under argon. Both solution of
193 NaFS and NaPS were introduced in the round bottom flask and the reaction mixture was stirred for 3
194 hours. Monomer traces were eliminated with dialysis (MWCO 1000 Da) and conductivity monitoring.
195 The polymer solution was then lyophilized and a white powder was obtained. This redox process at
196 25°C was developed after the paper of Sutton et al.⁹ to minimize the formation of dead chains. Four
197 DHBC were synthesized according to this procedure: PAM₇₀-*b*-PAPTAC₃₀, PAM₇₀-*b*-PAPTAC₆₀,
198 PAM₁₄₀-*b*-PAPTAC₆₀ and PAM₁₄₀-*b*-PAPTAC₁₂₀ (see Table 1).

199 **3.2.3 Synthesis of poly(ethylene oxide)-*b*-poly(acrylic acid)**

200 **Aqueous RAFT polymerization of AA**

201 Synthesis of PEO₁₀₅-*b*-PAA₂₀ was performed as follows: PEO₁₀₅-CTA (poly(ethylene oxide)-chain
202 transfer agent) macro RAFT agent was obtained following the procedure published by Bathfield et
203 al.³². PEO₁₀₅-CTA, (10.53 g, 2.19 mmol), ACPA (0.123 mg, 0,439 mmol), acrylic acid (5.5 g, 76
204 mmol) and deionized water (29.5 mL) (solids=35.4%) were introduced in a Schlenk tube equipped
205 with a magnetic stirrer. The mixture was degassed by five freeze-evacuate-thaw cycles and then heated

206 for 42 hours at 75°C under nitrogen in a thermostated oil bath. Final conversion = 72%. Monomer
207 conversion was determined by ¹H NMR spectroscopy, using a Bruker 400MHZ spectrometer. Samples
208 for analysis by NMR were prepared by adding 0.6 mL of D₂O to 0.1 mL of polymerization medium.
209 Once the reaction was complete, the solvent was evaporated, and then the DHBC was dissolved in a
210 minimum amount of dichloromethane before being precipitated twice in a large volume of cold diethyl
211 ether. It was then recovered by filtration, and finally dried under vacuum overnight before analysis by
212 SEC and ¹H-NMR. SEC was performed in DMF-LiBr after methylation³³ with
213 trimethylsilyldiazomethane. Four DHBC were synthesized according to this procedure: PEO₁₀₅-*b*-
214 PAA₂₀, PEO₁₀₅-*b*-PAA₃₀, PEO₂₁₀-*b*-PAA₄₀ and PEO₂₁₀-*b*-PAA₅₀ (see Table 1). The degrees of
215 polymerization of the commercial starting PEO have been determined by ¹H NMR³² and are presented
216 in Table 1.

217 **3.2.4 Synthesis of poly(acrylic acid)-*b*-poly(poly(ethylene glycol) methyl ether acrylate)**

218 **Aqueous RAFT polymerization of AA and PEGA**

219 5,7-dithia-6-thio-4-methyl-4-cyanodecanoic acid (CTPPA) was obtained by reaction of ACPA
220 with bis(propylsulfanylthiocarbonyl) disulfide according to literature.³⁴ Synthesis of PAA₂₁-*b*-
221 P(PEGA)₁₂ was performed as follows: PAA-CTPPA chain transfer agent was synthesized according to
222 the process described in literature³⁵: in a round bottom flask, CTPPA (0.8 g, 2.57 mmol, purity=89%),
223 acrylic acid (3.9g, 54.1 mmol), ACPA (0,072 g, 0.26 mmol) and half of the amount of water (7.5 g)
224 are stirred until dissolution of CTPPA. The remaining water (7.5 g) was introduced and the mixture
225 was degassed with argon for 40 min. The mixture was then heated in an oil bath at 70°C for 5.5 h. For
226 the synthesis of PAA-*b*-P(PEGA), ACPA (0.0715 g; 0.26 mmol) and poly(ethylene glycol) methyl
227 ether acrylate (12.8 g, 26.4 mmol) were added to the PAA-CTPPA reaction medium and the mixture
228 was degassed with argon for 40 min. The mixture was then heated in an oil bath at 70°C for 5 h.
229 Conversion was followed by ¹H NMR. At the end of the polymerization, water was evaporated under
230 reduced pressure and the polymer washed with diethyl ether. NMR sample preparation: 0.6 mL of D₂O
231 was added to 0.1 mL of polymerization medium and quenched in liquid nitrogen. Two DHBC were
232 synthesized according to this procedure: P(PEGA)₁₂-*b*-PAA₂₁ and P(PEGA)₂₂-*b*-PAA₄₅ (see Table 1).

233 **Table 1:** Presentation of the chemical structure and the different DHBC samples synthesized and studied in this
 234 work. The subscripts in the names correspond to the degree of polymerization of each block. $MW_{neutral}$ and MW_0
 235 are the molar masses of the neutral and of the polyelectrolyte block respectively.

Type of DHBC	$MW_{neutral}$ - MW_0	Chemical structure
	as determined by NMR	
PAM ₇₀ - <i>b</i> -PAA ₂₀	5k-1.4k	
PAM ₁₄₀ - <i>b</i> -PAA ₄₀	10k-2.8k	
PAM ₁₄₀ - <i>b</i> -PAA ₆₀	10k-4.4k	
PAM ₁₄₀ - <i>b</i> -PAA ₈₀	10k-5.6k	
PEO ₁₀₅ - <i>b</i> -PAA ₂₀	5k-1.4k	
PEO ₁₀₅ - <i>b</i> -PAA ₃₀	5k-2.2k	
PEO ₂₁₀ - <i>b</i> -PAA ₄₀	10k-2.8k	
PEO ₂₁₀ - <i>b</i> -PAA ₅₀	10k-3.6k	
PAM ₇₀ - <i>b</i> -PAPTAC ₃₀	5k-5k	
PAM ₇₀ - <i>b</i> -PAPTAC ₆₀	5k-10k	
PAM ₁₄₀ - <i>b</i> -PAPTAC ₆₀	10k-10k	
PAM ₁₄₀ - <i>b</i> -PAPTAC ₁₂₀	10k-20k	
P(PEGA) ₁₂ - <i>b</i> -PAA ₂₁	5.7k-1.5k	
P(PEGA) ₂₂ - <i>b</i> -PAA ₄₅	10.6k-3.2k	

236 3.3 Capillary electrophoresis

237 *Instrumentation and method*

238 Capillary electrophoresis experiments were performed on an Agilent 7100 capillary electrophoresis
 239 instrument with a diode array UV detector. Fused silica capillaries of 50/375 μm I.D./O.D. with
 240 polyimide outer coating (cat. no. TSP050375) were from Polymicro Technologies (Phoenix, AZ,

241 USA). Capillary dimensions were 38.5 cm long (30 cm to detection window). New capillaries were
242 conditioned by performing the following washes at 1 bar: 1M NaOH for 30 min and water for 15 min.
243 The temperature of the capillary cartridge was set at 25 °C.

244 In the case of PAM-*b*-PAA, an electrolyte consisting of 20 mM MES and 14 mM ammediol pH 6.5
245 was used. 0.1 g/L anisic acid was added in the sample as a mobility marker. The same background
246 electrolyte was used for PAM-*b*-PAPTAC, but with a different mobility marker (imidazole 0.1 g/L). In
247 the case of PEO-*b*-PAA and P(PEGA)-*b*-PAA, an electrolyte constituted of 6 mM anisic acid and 12
248 mM BIS-TRIS, pH 6.5 was used as buffer, with MES at 0.5 g/L as mobility marker in the case of
249 P(PEGA)-*b*-PAA.

250 All copolymers were dissolved in water at a concentration of 5 g/L. Samples were injected
251 hydrodynamically on the inlet side of the capillary by applying 30 mbar for 5 s. Separations were
252 carried out by applying a +20 kV voltage. For PAM-*b*-PAA, PEO-*b*-PAA and P(PEGA)-*b*-PAA and
253 PAM-*b*-PAPTAC, detection was realized at 192 +/- 2 nm (reference off).

254 For PAM-*b*-PAA, PEO-*b*-PAA and P(PEGA)-*b*-PAA, the capillary was rinsed between each run by
255 flushing the capillary for 2 min with the background electrolyte, 2 min with 0.1 M NaOH, 2 min with
256 ultra-pure water and 2 min with background electrolyte. For the analysis of the cationic polymer
257 PAM-*b*-PAPTAC, and in order to reduce the adsorption on the capillary wall, surface of the capillary
258 was modified using UltraTrol™ LN (Target Discovery, Inc., Palo Alto, CA), which is a commercial
259 neutral semi-permanent coating based on polyacrylamide derivatives. The coating procedure was
260 performed using the following successive flushes: methanol for 2 min at 1 bar, water for 2 min at
261 3 bar, 1 M NaOH for 2 min at 3 bar, 0.1 M NaOH for 2 min at 1 bar, 1 M HCl for 5 min at 1 bar, water
262 for 5 min at 1 bar, UltraTrol™ LN solution for 5 min at 1 bar, wait for 5 min, water for 2 min at 1 bar.
263 Prior to each analysis of PAM-*b*-PAPTAC, the capillary was rinsed with the background electrolyte
264 for 2 min at 1 bar.

265

266 **Electropherogram data treatment**

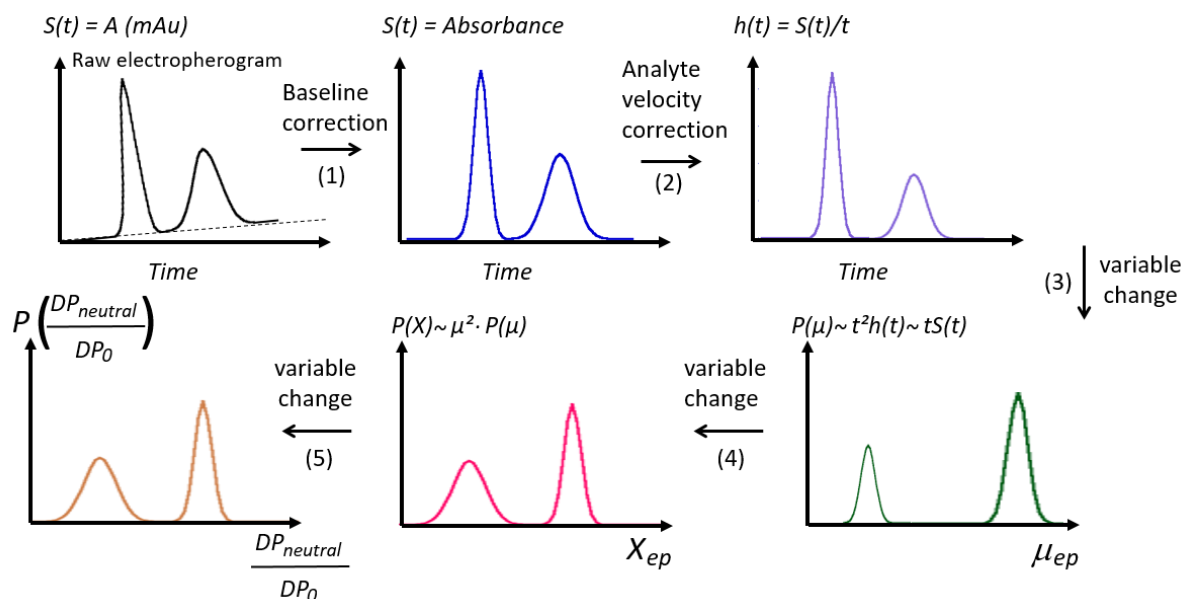


Figure 2. General scheme for changing a time-scale electropherogram into a mobility-scale and X_{exp} -scale distributions. The raw electropherogram is first corrected from baseline shift (1). The time-scale electropherogram is then corrected from the differences in analyte velocities (2). The time-corrected electropherogram is converted into the effective mobility-scale distribution (3). The mobility-scale electropherogram is changed into a X_{exp} -scale distribution (4) and finally to a compositional $\frac{DP_{\text{neutral}}}{DP_0}$ ratio (5).

$S(t)$ is the UV absorbance signal (in mAU). $h(t)$ is the time-corrected UV absorbance. $P(\mu_{\text{ep}})$ is the effective mobility distribution. $P(X_{\text{exp}})$ is the distribution in X_{exp} (see section 4.2) and $P(\frac{DP_{\text{neutral}}}{DP_0})$ is the distribution in $\frac{DP_{\text{neutral}}}{DP_0}$.

$\frac{DP_{\text{neutral}}}{DP_0}$. Adapted from¹¹ for the characterization of diblock copolymers.

267 **Scale transformations.** In this section, the transformations of the electropherograms into distributions
 268 of the parameter of interest are described following a previously published protocol¹¹. Briefly,
 269 experimental raw time-scale electropherograms, were first corrected from any baseline shift using
 270 Origin (Origin 2016, OriginLab, USA) as depicted in step 1, Figure 2. For quantitative purpose, the
 271 absorbance signal $S(t)$ was next divided by the migration time (t) to correct the differences in analyte

272 velocity (Figure 2, step 2)¹¹. Next, the time-corrected electropherogram $h(t)$ was changed into an
 273 effective mobility distribution $P(\mu_{ep}) = t \times S(t)$ (Figure 2, step 3)¹¹, which requires the
 274 transformations of both the x and y axis¹¹. Note that μ_{ep} is obtained by equation (5):

$$275 \quad \mu_{ep} = \frac{lL}{V} \left(\frac{1}{t} - \frac{1}{t_{eo}} \right) \quad (5)$$

276 where l is the effective capillary length, L is the total capillary length, t is the migration time, t_{eo} is the
 277 EOF marker migration time and V is the separation voltage.

278 **Moments of the electrophoretic mobility distribution** The average effective mobility of the diblock
 279 copolymer $\overline{\mu_{ep}^{diblock}}$ was obtained by integration of the peak of the copolymer in the effective mobility
 280 scale according to:

$$281 \quad \overline{\mu_{ep}^{diblock}} = \frac{\int P(\mu_{ep}) \mu_{ep} d\mu_{ep}}{\int P(\mu_{ep}) d\mu_{ep}} \approx \frac{\sum_i P(\mu_{ep,i}) \mu_{ep,i} (\mu_{ep,i+1} - \mu_{ep,i})}{\sum_i P(\mu_{ep,i}) (\mu_{ep,i+1} - \mu_{ep,i})} \quad (6)$$

282 where integration is carried out over the peak. In practice the integration is done numerically and the i
 283 index represents the digitized experimental data points. The summation is carried out over values of
 284 $P(\mu_{ep,i})$ greater than the median of the base line added to its standard deviation. Calculation of
 285 $\overline{\mu_{ep}^{diblock}}$ was performed using Excel 2016 (Microsoft, USA), following the discrete form of equation
 286 (6). Variance of the diblock electrophoretic mobility was obtained by the following equation:

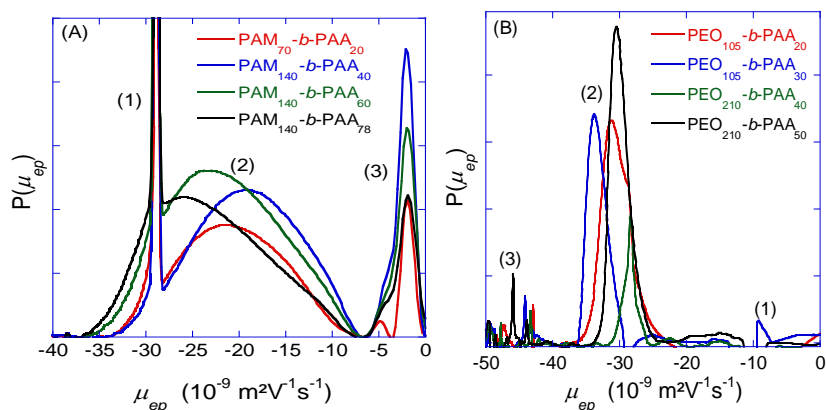
$$287 \quad \sigma_{\mu}^2 = \frac{\int P(\mu_{ep}) \left(\mu_{ep} - \overline{\mu_{ep}^{diblock}} \right)^2 d\mu_{ep}}{\int P(\mu_{ep}) d\mu_{ep}} = \frac{\sum_i P(\mu_{ep,i}) (\mu_{ep,i} - \overline{\mu_{ep}^{diblock}})^2 (\mu_{ep,i+1} - \mu_{ep,i})}{\sum_i P(\mu_{ep,i}) (\mu_{ep,i+1} - \mu_{ep,i})} \quad (7)$$

288

289 4. Results and discussion

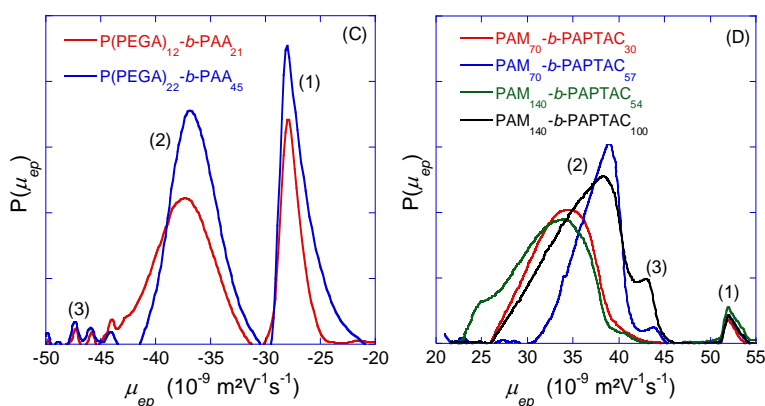
290 4.1. Electrophoretic separation of the DHBC

291 The main goal of the CE characterization is to provide information about the chemical composition
292 distribution of the DHBC and about the purity of the DHBC in terms of possible presence of
293 homopolymers. The separation of the DHBC by CE requires an appropriate background electrolyte,
294 depending on the nature of the copolymer. For copolymers absorbing in UV (i.e. those with a PAM
295 neutral block), direct UV detection was possible and a background electrolyte based on 20 mM MES
296 and 14 mM ammonium dihydrogen phosphate at pH 6.5 was used, with a UV detection at 192 nm. For DHBC copolymers that
297 do not absorb UV enough to ensure sensitivity (i.e. PEO-*b*-PAA or P(PEGA)-*b*-PAA), an indirect
298 detection mode based on a 6 mM anisic acid and 12 mM BIS TRIS at pH 6.5 was used. At this pH
299 about 60% of the carboxylic acid groups of the PAA are ionized, and this ensures appropriate
300 selectivity of separation between PAA homopolyelectrolyte and the DHBC. Uncoated fused silica
301 capillary was used for the characterization of all anionic DHBC. Semi-permanent UltraTrollLN neutral
302 coating was used for the characterization of the cationic PAM-*b*-PAPTAC DHBC, to avoid any
303 copolymer adsorption on the capillary surface. To correct the apparent mobility from the
304 electroosmotic mobility, a mobility marker (anisic acid for PAM-*b*-PAA, MES for P(PEGA)-*b*-PAA,
305 and imidazolium for PAM-*b*-PAPTAC) of known effective mobility ($\mu_{ep, MES} = -28$ TU (where TU,
306 Tiselius Unit, stands for $10^{-9} \text{ m}^2\text{V}^{-1}\text{s}^{-1}$) and $\mu_{ep, imidazolium} = 52$ TU) was co-injected. For PEO-*b*-PAA, the
307 electroosmotic mobility was estimated from the electroosmotic flow (EOF) peak. The distributions of
308 effective mobility (DEM) of PAM-*b*-PAA, PEO-*b*-PAA, P(PEGA)-*b*-PAA and PAM-*b*-PAPTAC are
309 displayed in Figure 3.



310

311



312

313 **Figure 3.:** Distributions of effective electrophoretic mobility obtained for PAM-*b*-PAA (A), PEO-*b*-
 314 PAA (B), P(PEGA)-*b*-PAA (C), and PAM-*b*-PAPTAC (D).-Electrophoretic conditions: fused silica
 315 capillary (A, B, C) or coated with UltraTrolLNTM (D), 50 μm I.D. \times 38.5 cm (effective length, 30 cm).
 316 Electrolytes: 20 mM MES, 14 mM ammediol, pH 6.5 (A, D); 6 mM anisic acid, 12 mM BIS TRIS, pH
 317 6.5 (B, C). Applied voltage: +20 kV. Hydrodynamic injection: 30 mbar, 5 s. Direct (A, D) or indirect
 318 (B, C) UV detection at 192 \pm 2 nm. Temperature: 25 $^{\circ}\text{C}$. Samples: 5 g/L DHBC. Assignment of the
 319 peaks: PAM-*b*-PAA (A): anisic acid (1), DHBC (2), PAM homopolymer (3); PEO-*b*-PAA (B): system
 320 peak (1), DHBC (2), PAA oligomers (3); P(PEGA)-*b*-PAA (C): MES (1), DHBC (2), PAA oligomers
 321 (3); PAM-*b*-PAPTAC (D): Imidazolium (1), DHBC (2), PAPTAC (3). The degree of polymerization
 322 of each block is specified on the graph.

323 The DEM in the series PAM-*b*-PAA (Figure 3A) show three peaks, two sharp at -29 TU (peak 1) and
 324 -2 TU (peak 3) and one broad (peak 2) between -7 and -37.5 TU, the latter being assigned to the

325 copolymer of interest. The peak at -29 TU is assigned to anisic acid (electrophoretic mobility marker)
326 and the one at -2 TU corresponds to homopolymer of PAM. The non-zero electrophoretic mobility of
327 the PAM homopolymer is explained by the incorporation of the negatively charged initiator 4,4'-
328 azobis(4-cyanopentanoic) acid. The mass proportion of this PAM population of dead chains has been
329 quantified by external calibration based on time-corrected peak areas using direct injections of PAM
330 solutions of known concentration in the same condition as the DHBC. The proportion of PAM
331 homopolymer amounts to 21wt% in the solid form polymer sample for PAM₇₀-*b*-PAA₂₀, 15% for
332 PAM₁₄₀-*b*-PAA₄₀ and PAM₁₄₀-*b*-PAA₆₀ and 8% for PAM₁₄₀-*b*-PAA₇₈. The copolymer peak is broad
333 and, as expected, the DEM shifts further from zero as the proportion of charged monomer increases in
334 the composition of the copolymers. This can be verified by comparing the average electrophoretic
335 mobility value $\overline{\mu_{ep}^{diblock}}$ (given in Table 2) which varies between -19.5 TU and -24.1 TU from PAM₁₄₀-
336 *b*-PAA₄₀ to PAM₁₄₀-*b*-PAA₇₈. The greater the average molar mass of the polymer, the more dispersed
337 its electrophoretic mobility, as demonstrated by the standard deviation σ_{μ} which varies from 3.9 TU
338 for PAM-*b*-PAA 70-20, to 6.2 TU for PAM-*b*-PAA 140-60.

339 The DEM of PEO based copolymers are presented in Figure 3B for linear PEO and Figure 3C for PEO
340 grafted polyacrylates (P(PEGA)). Three populations are observed in both series: several small peaks
341 associated with large electrophoretic mobility at -45TU are assigned to short oligomers of PAA, the
342 weight percent of which is estimated to be lower than 10%. The sensitivity of the UV detection is too
343 low to conclude about the presence / absence of PEO or P(PEGA) in the DHBC. The least mobile
344 species at -28TU in Figure 3C corresponds to the MES used as mobility marker. The peak at
345 intermediate values of mobility corresponds to the DHBC. The electrophoretic mobility of PEO-*b*-
346 PAA ($\overline{\mu_{ep}^{diblock}}$ ranging from -28 to -33 TU, Table 2) is significantly closer to zero than that of
347 P(PEGA)-*b*-PAA copolymers ($\overline{\mu_{ep}^{diblock}}$ ranging from -36 to -38 TU, Table 2), although the molar
348 masses are close. This is because P(PEGA), a comb-like polymer, is more compact than linear PEO of
349 the same molar mass. As a consequence, the drag force due to the neutral block is lower for P(PEGA)
350 than for PEO.

351 As for PAM-*b*-PAPTAC copolymer, PAPTAC homopolymer was detected at about +43TU, as a
352 shoulder merged in the copolymer distribution, only for the $DP_{neutral}/DP_0$ equal to 140/100 and 70/57
353 samples. Figure 3D displays DEM ranged between +22 and +41 TU, with higher effective mobilities
354 for the DHBC of highest charge. Comparison of PAM-*b*-PAA series with PAM-*b*-PAPTAC series
355 illustrates the importance of the nature of the blocks on the drag effect of the neutral block. This effect
356 is discussed in more detail in section 4.2.

357 As a general trend, electrophoretic mobilities of DHBC are always closer to zero than those of the
358 homopolyelectrolyte which are: $\mu_{PAA} = -42$ TU; $\mu_{PAPTAC} = +44$ TU, and the electrophoretic mobility of
359 the DHBC increases as the proportion of charged monomers in the DHBC increases (see Figure SI
360 20). In terms of EM dispersion, the least dispersed series is the P(PEGA)-*b*-PAA, with relative
361 standard deviation of EM $\sigma_{\mu} / \overline{\mu_{ep}^{diblock}}$ between 6.5% to 9%, followed by the PEO-*b*-PAA (
362 $\sigma_{\mu} / \overline{\mu_{ep}^{diblock}} \sim 5\%$ to 16%) and the most disperse series is the PAM-*b*-PAA series with 20% to 30%
363 relative standard deviation of the electrophoretic mobility. This dispersion in mobility results from
364 both the level of control of the polymerization, and from the spatial extension of the polymer in the
365 solvent. It can be explained by the chemistry of the RAFT polymerization (chain transfer agent R-
366 SC(S)Z where Z is the activating group and R is the leaving group), for which the polymerization of
367 acrylates is better controlled by dithiobenzoates (Z: -SC(S)Ph) ($\mathfrak{D} < 1.2$) than by ethyl xanthate (Z: -
368 SC(S)OEt) ($\mathfrak{D} > 1.3$). Besides, the 2-phenylacetate ester of PEO is a better homolytic leaving group
369 than PAM.³⁶ Furthermore, the synthesis of PAM-*b*-PAA cumulates two successive RAFT/MADIX
370 polymerizations of AM and AA, whereas the synthesis of PEO-*b*-PAA starts from a narrow PEO-CTA
371 ($\mathfrak{D} = 1.04$) (PEO obtained by anionic polymerization) to perform a single RAFT polymerization of
372 AA. This is consistent with the higher dispersity of PAM-*b*-PAA compared to PEO-*b*-PAA³¹. In
373 addition, in reversible-deactivation radical polymerization³⁷, at full conversion, the dispersity as
374 defined by the ratio of the weight average molar mass over the number average molar mass, decreases
375 when the DP increases³⁸:

$$D = 1 + \frac{1}{DP} + \frac{1}{C_{ex}} \quad (8)$$

where DP is the targeted polymerization degree and C_{ex} is the degenerative chain transfer constant between dormant and active chains^{39, 40}, which is consistent with a higher dispersity for a shorter poly(acrylic acid) block in PEO-*b*-PAA.

To get a better description of the copolymer distribution, it would be interesting to get a distribution of a new parameter which is directly related to the chemical composition of the copolymer instead of the electrophoretic mobility, which is not linearly dependent on the copolymer composition. This is the purpose of the two following sections.

384

	$DP_{neutral}$ DP_0	$\mu_{ep}^{diblock}$ peak max (TU)	$\overline{\mu_{ep}^{diblock}}$ integration (TU)	σ_{μ} (TU)	X_{exp} peak max	$\overline{X_{exp}}$ by integration	$\sigma_{X_{exp}}$
PAM-<i>b</i>-PAA	70-20	-22	-19.9	3.89	0.61	1.12	0.45
	140-40	-19.1	-19.5	5.44	0.74	1.26	0.73
	140-60	-23.3	-21.4	6.19	0.58	1.05	1.0
	140-78	-25.3	-24.1	5.68	0.41	0.99	0.56
PEO-<i>b</i>-PAA	105-20	-31	-30.2	4.83	0.28	0.33	0.12
	105-30	-33.8	-33.2	2.45	0.18	0.21	0.06
	210-40	-28	-28.4	1.36	0.40	0.41	0.06
	210-50	-30.5	-30	1.97	0.30	0.34	0.64
P(PEGA)-<i>b</i>-PAA	21-11.5	-37.6	-38.6	3.61	0.059	0.043	0.09
	44.9-22.2	-37	-36.6	2.41	0.079	0.098	0.08

70-30	34.6	33.9	3.18	0.19	0.25	0.12
70-60	39.1	37.1	3.2	0.08	0.11	0.05
140-60	34.2	32	4.1	0.21	0.32	0.39
140-120	38.5	36.3	4.1	0.09	0.164	0.13

385 **Table 2.** Electrophoretic mobility $\mu_{ep}^{diblock}$ at peak maximum and average value $\overline{\mu_{ep}^{diblock}}$ obtained by peak
386 integration, standard deviation of the electrophoretic mobility distribution σ_{μ} , X_{exp} value at peak maximum and
387 average value $\overline{X_{exp}}$ obtained by peak integration, standard deviation of the X_{exp} distribution $\sigma_{X_{exp}}$ of all DHBC
388 studied in this work. Note that the peak of anisic acid was first subtracted before integration for the calculation of
389 $\overline{\mu_{ep}^{diblock}}$ and σ_{μ} for PAM-*b*-PAA.

390

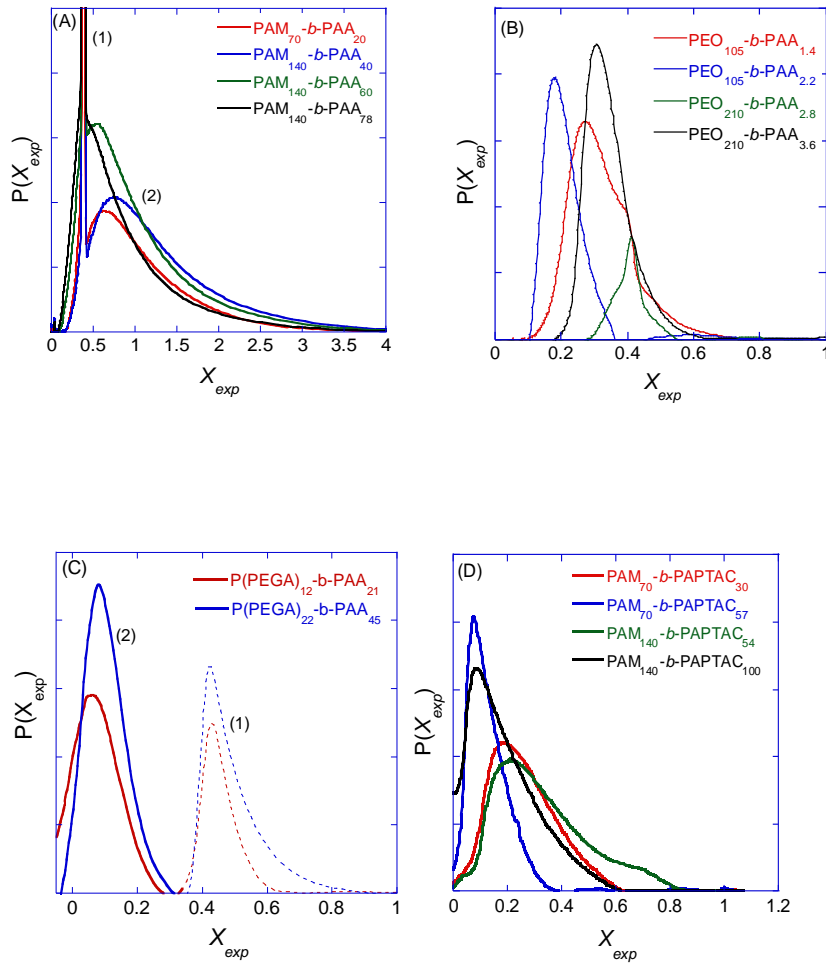
391 4.2. Change of variable from μ_{ep} to the retardation parameter X_{exp}

392 The drag effect of the neutral block can be expressed by the retardation parameter X_{exp} defined as :

$$393 \quad X_{exp} = \frac{\mu_{ep}^0 - \mu_{ep}^{diblock}}{\mu_{ep}^{diblock}} = \frac{\mu_{ep}^0}{\mu_{ep}^{diblock}} - 1 \quad (9)$$

394 where μ_{ep}^0 is the electrophoretic mobility of the homopolyelectrolyte; $\mu_{ep}^{diblock}$ is the electrophoretic
395 mobility of the copolymer. X_{exp} expresses the relative decrease of mobility due to the presence of the
396 neutral block. It is positive and increases as the drag effect increases.

397



398

399

400

401 **Figure 4.** Distribution of retardation parameter X_{exp} for PAM-*b*-PAA (A) PEO-*b*-PAA (B), P(PEGA)-
 402 *b*-PAA (C), and PAM-*b*-PAPTAC (D). Experimental conditions as in Figure 3 X_{exp} was determined
 403 using eq. (8), eq. (12) and $\mu_{ep,PAA}^0 = -42TU$; $\mu_{ep,PAPTAC}^0 = 44 TU$. Assignment of the peaks: PAM-*b*-
 404 PAA (A): anisic acid (1), DHBC (2); P(PEGA)-*b*-PAA (C): MES (1), DHBC (2). The degree of
 405 polymerization of each block is specified on the graph.

406

407 The new experimental variable, X_{exp} , not only points out the friction due to the neutral block but is also
 408 more directly related to the composition of the DHBC. Introducing equation (9) in the various
 409 expressions of the electrophoretic mobility (equations 1 to 3), X_{exp} , can be expressed as a function of

410 the ratio between the degrees of polymerization of the neutral $DP_{neutral}$ and the charged blocks DP_0 .

411 Taking into account the hydrodynamic coupling (model 1), X_{exp} reads:

$$412 \quad X_{model,1} = \frac{1}{N_{blob}} = \frac{\alpha}{DP_0} \quad (10a)$$

413 and can be further developed as a function of the Kuhn lengths of the neutral block, b_{K_1} , and of the

414 polyelectrolyte, b_{K_0} :^{15, 41}

$$415 \quad X_{model,1} = \alpha_1 \frac{DP_{neutral}}{DP_0} \quad (10b)$$

$$416 \quad \text{with } \alpha_1 = \frac{b_1 b_{K_1}}{b_0 b_{K_0}} \quad (10c)$$

417 where b_1 is the neutral monomer size. The Kuhn statistical segment length (which is twice the
418 persistence length) is a measure of the polymer stiffness. Parameter α_1 in Equation (10b) is a relative
419 friction coefficient and it is non-dimensional. Since the polyelectrolyte is generally stiffer than the
420 neutral block, α_1 is often much smaller than unity¹⁵.

421 As for model 2, it is clear from equation (2) that the X parameter is directly expressed as the ratio of
422 the hydrodynamic radius of each block:

$$423 \quad X_{model,2} = \frac{R_h^{neutral}}{R_h^0} \quad (11a)$$

424 which can be rewritten as a function of the degrees of polymerization of each block by:

$$425 \quad X_{model,2} = \frac{C_1 DP_{neutral}^{a_1}}{C_0 DP_0^{a_0}} \quad (11b)$$

426 where C_1 (resp. C_2) and a_1 (resp. a_2) are, respectively, the prefactors and exponents for the neutral
427 (resp. charged) block in the relationship between R_h and DP . Note that a_0 and a_1 are supposed to be
428 close to 0.5-0.6 for coil conformations, and slightly higher for more extended conformations.

429 As for model 3 (see equation (3)), the X parameter is directly expressed as:

$$430 \quad X_{\text{model},3} = \frac{2R_h^{\text{neutral}} \ln DP_0}{b_0 DP_0} \quad (12a)$$

431 Injecting R_h^{neutral} in equation (9a) leads to:

$$432 \quad X_{\text{model},3} = \frac{2C_1 DP_{\text{neutral}}^{a_1} \ln DP_0}{b_0 DP_0} \quad (12b)$$

433 Finally, equations (10b) (11b) and (12b) demonstrates that, whatever the considered model, the X
 434 parameter is related to a compositional ratio between the neutral and the charged blocks with,
 435 however, different scaling dependences with the DP of each block, and with an additional logarithmic
 436 term in model 3. It is worth noting that in the case of hydrodynamic coupling (model 1), the newly
 437 introduced variable varies linearly with the ratio of degree of polymerization of the two blocks.

438 The distributions of the X_{exp} parameter are simply deduced from the distribution of electrophoretic
 439 mobility using the following equation (Figure 2, step 4)¹¹:

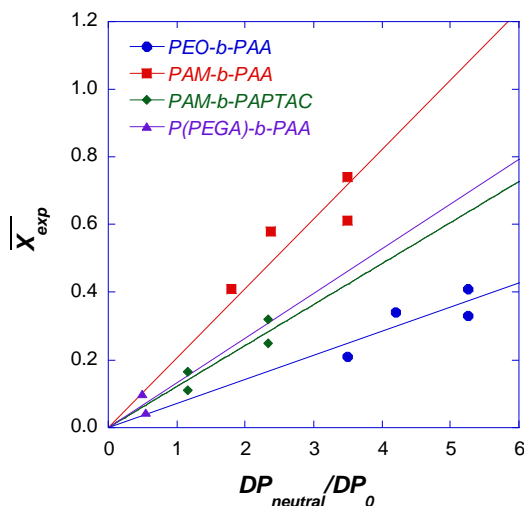
$$440 \quad P(X_{\text{exp}}) = \left| \frac{1}{\frac{\partial X_{\text{ep}}}{\partial \mu_{\text{ep}}}} \right| P(\mu_{\text{ep}}) = \mu_{\text{ep}}^2 P(\mu_{\text{ep}}) \quad (13)$$

441 All the X_{exp} distributions are presented in Figure 4. Since μ_{ep} and X_{exp} are not linearly related, the
 442 change of the variable from μ_{ep} to X_{exp} modify the form of the distribution. The different moments
 443 characterizing the distribution of X_{exp} are calculated using similar relations as equations (6) and (7) and
 444 are reported in Table 2.

445 X_{exp} range varies between 0.1-3 for PAM-*b*-PAA, 0.1-0.7 for PEO-*b*-PAA, 0-0.3 for P(PEGA)-*b*-PAA
 446 and 0-0.8 for PAM-*b*-PAPTAC. The dispersion of the retardation parameter X_{exp} expressed as $\sigma_{X_{\text{exp}}}$
 447 values (Table 2) follows the following order: P(PEGA)-*b*-PAA < PEO-*b*-PAA ~ PAM-*b*-PAPTAC <
 448 PAM-*b*-PAA. The dispersion of the retardation parameter cannot be interpreted as a dispersity in

449 molar mass or in composition since the retardation parameter will change with these chemical features
 450 in a way that depends on the conformation of the blocks. So, a further step is needed to get the
 451 composition dispersion.

452 **4.3. Change of variable from X_{exp} to chemical composition ratio**



453
 454 **Figure 5.** Plot of \overline{X}_{exp} against $\frac{DP_{neutral}}{DP_0}$. \overline{X}_{exp} was determined by using eq.(6) after replacing μ_{ep} by
 455 $X_{exp} \cdot \frac{DP_{neutral}}{DP_0}$ was obtained by NMR (see Table 1). Least-square linear regressions provide the following
 456 experimental slopes: 0.071+/-0.005 for PEO-b-PAA; 0.12+/-0.01 for PAM-b-PAPTAC; 0.13 +/- 0.01
 457 for P(PEGA)-b-PAA; 0.21+/-0.02 for PAM-b-PAA.

458
 459 To go further in the interpretation and in the process of the experimental data, it is crucial to identify
 460 the model which is best adapted to describe the electrophoretic behavior of the DHBC investigated in
 461 this work. To assess the validity of model 1 (hydrodynamic coupling between blocks, see section 2.3),
 462 \overline{X}_{exp} was plotted against $\frac{DP_{neutral}}{DP_0}$ in Figure 5 for the four DHBC families.

463 **Table 3.** Characteristic parameters of neutral and charged blocks constituting the DHBC studied in this work.
 464 M_w are expressed in g/mol.

	b_l (nm)	b_{k_1} (nm)	$R_h^{neutral}$ (nm) ^a	465
PAM	0.25	0.6 ⁴²	$0.01447 \times M_w^{0.57}$ ⁴³	466
PEO	0.3 ⁴⁴	0.74 ⁴⁴	$0.02398 \times M_w^{0.53}$ ⁴⁵	467
	b_0 (nm)	b_{k_0} (nm)	R_h^0 (nm)	468
PAA	0.25	2.5 ⁴⁶	$0.007906 \times M_w^{0.585}$ ²	469
PAPTAC	approximated as PAA			470

471

472 ^a From Mark-Houwink parameters using $R_h = \left(\frac{3[\eta]M}{10\pi N_A} \right)^{1/3}$, where $[\eta]$ is the intrinsic viscosity and N_A is
473 the Avogadro number. from ref⁴⁶ (see Table 1 and Figure 13(a) herein).

474

475 The results are consistent with model 1 which predicts a linear dependence of the retardation
476 parameter $\overline{X_{exp}}$ on the ratio of degree of polymerization $\frac{DP_{neutral}}{DP_0}$. The slopes determined from the
477 graph in Figure 5 correspond to the parameter α_1 in equation (10b) which can be calculated from
478 equation (10c). Experimentally, the numerical values of the slopes $\alpha_{1,exp}$ are in the range of ~0.1-0.2.
479 Taking the characteristic parameters (Kuhn lengths, monomer dimensions) given in Table 3 leads to
480 $\alpha_1=0.24$ (vs $\alpha_{1,exp}=0.21\pm 0.02$ experimentally obtained) for PAM-*b*-PAA, $\alpha_1=0.36$ (vs $\alpha_{1,exp}=0.071\pm$
481 0.005) for PEO-*b*-PAA, and $\alpha_1=0.24$ (vs $\alpha_{1,exp}=0.12\pm 0.01$) for PAM-*b*-PAPTAC. Theoretical values
482 of α_1 are in a reasonably good agreement with the experimental ones, knowing the uncertainty on the
483 persistence length (notably for the polyelectrolyte blocks) and monomer sizes. As for P(PEGA)-*b*-
484 PAA, we only get an estimation of $\alpha_{1,exp}=0.13$, since the P(PEGA) Kuhn length is not available in the
485 literature. From Figure 5, we can conclude that the linear correlation between $\overline{X_{exp}}$ and $\frac{DP_{neutral}}{DP_0}$ is
486 confirmed and that model 1 (with hydrodynamic coupling between the two blocks) can be used to

487 transform the X_{exp} distributions into compositional $\frac{DP_{\text{neutral}}}{DP_0}$ distributions. As for models 2 and 3, they

488 lead to poor correlations between theoretical $X_{\text{model},i}$ versus experimental $\overline{X_{\text{exp}}}$ values (see Figure 6).

489 The knowledge of α_l provides the last relation necessary to carry on the general scheme presented in
490 Figure 2 to its end and which leads to the distribution of ratio of chemical composition. In practice, we
491 used $\alpha_{l,\text{exp}}$ obtained in Figure 5 together with equation (10b) to transform the data of Figures 4A to 4D
492 into the distributions presented in Figures 7A to 7D, using the following equation:

$$493 \quad P\left(\frac{DP_{\text{neutral}}}{DP_0}\right) = \frac{P(X)}{\frac{\partial\left(\frac{DP_{\text{neutral}}}{DP_0}\right)}{\partial X}} = \alpha_l P(X) \quad (14)$$

494 Since X_{exp} and $\frac{DP_{\text{neutral}}}{DP_0}$ are linearly correlated, the shapes of both distributions are similar. However,

495 reading $\frac{DP_{\text{neutral}}}{DP_0}$ axis, which corresponds to a compositional ratio, is more convenient for the

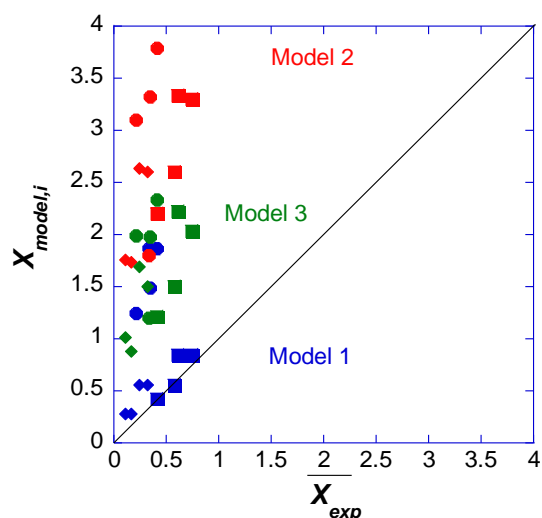
496 practitioners than keeping the X_{exp} scale. Moreover, and as previously anticipated, since the α_l

497 coefficients are different from one DHBC to another, the distribution in $\frac{DP_{\text{neutral}}}{DP_0}$ allows a better

498 comparison between them. On the whole, the dispersion of the composition ratio are in the order of:

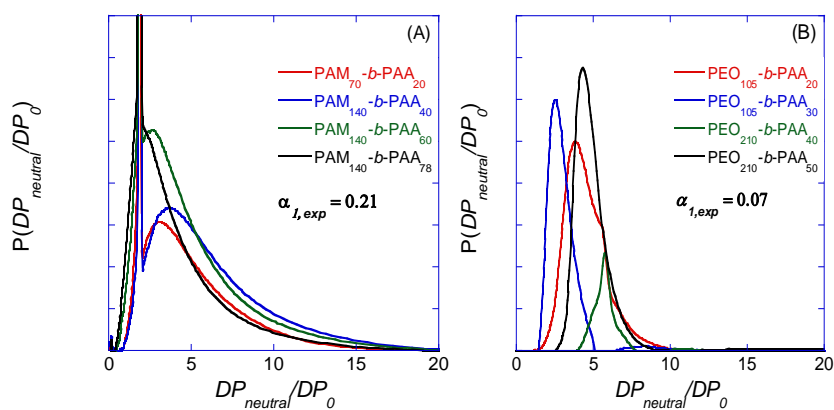
499 P(PEGA)-*b*-PAA < PAM-*b*-PAPTAC ~ PEO-*b*-PAA < PAM-*b*-PAA.

500

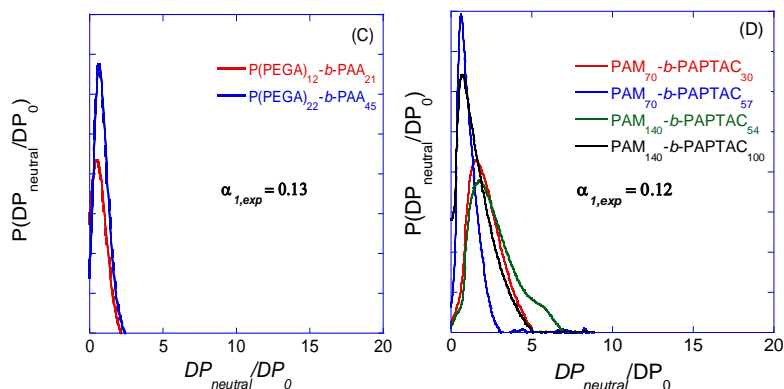


501
 502 **Figure 6.** Comparison between $X_{model,i}$ and \overline{X}_{exp} for the three different models using theoretical
 503 parameters given in Table 3. PAM-*b*-PAPTAC (◆); PEO-*b*-PAA (●); PAM-*b*-PAA (■). \overline{X}_{exp} was
 504 determined by integration of the DHBC peak (in *X* scale). For all $X_{model,i}$ calculations, theoretical
 505 $DP_{neutral}$ and DP_0 were used. $X_{model,1}$ was determined according to equations (10b) and (10c), $X_{model,2}$
 506 according to equation (11a) and $X_{model,3}$ according to equations (12a), with the characteristic
 507 numerical parameters given in Table 3.

508



509



510

511 **Figure 7.** Distribution of composition in terms of the ratio of the degrees of polymerization of both
 512 blocks for PAM-*b*-PAA (A), PEO-*b*-PAA (B), P(PEGA)-*b*-PAA (C), and PAM-*b*-PAPTAC (D).

513 Experimental conditions as in Figure 3. $\frac{DP_{neutral}}{DP_0}$ was determined using eq. (10b) and $P\left(\frac{DP_{neutral}}{DP_0}\right)$ was

514 obtained using eq. (14). In (C), the MES peak has been removed. In (A), the sharp peak is a mobility
 515 marker (anisic acid) and was deleted before peak integration.

516

517 Conclusion

518 In an effort to make information provided by capillary electrophoresis more directly useful for
 519 polymer chemists, a protocol was proposed to convert electrophoretic mobility distributions of double
 520 hydrophilic block copolymers into distributions of chemical composition ratios. This ratio of
 521 composition is expressed as the ratio of the degrees of polymerization of each block $\frac{DP_{neutral}}{DP_0}$. To get

522 this composition ratio, we have introduced the retardation parameter X which takes into account the
 523 drag force exerted by the neutral block on the polyelectrolyte. The distribution of X that characterizes a
 524 DHBC is readily obtained from the experimental electropherogram and the relation between X and the
 525 ratio of DP . The latter is available from different models for electrophoretic mobility of composite
 526 objects. A linear relation has been found experimentally between the retardation parameter X and the
 527 ratio of DP s, within each of the four families of DHBC studied in this work. This result is consistent
 528 with the model of electrophoretic mobility of Long *et al.* that takes into account hydrodynamic

529 coupling, although the prefactors are slightly overestimated. Our experimental findings not only
530 support the theoretical prediction, but also facilitate the last step of data transformation, from
531 distribution of X into distributions of ratios of DP .

532 The dispersions in composition are in the order of: P(PEGA)-*b*-PAA < PEO-*b*-PAA ~ PAM-*b*-
533 PAPTAC < PAM-*b*-PAA. Therefore, we can conclude that in the DHBC families, the PAM block
534 leads to broader composition ratio distributions compared to a PEO block, when associated to a PAA
535 block. Similarly, the P(PEGA) block lead to less disperse composition ratio distributions compared to
536 a PEO block, when associated to a PAA block. Finally, PAA associated with PAM leads to broader
537 composition ratio distributions compared to PAPTAC associated with PAM. The relatively low
538 composition dispersity of the PEO-PAA block copolymer is most likely due to the low dispersity of
539 the PEO block, prepared by anionic polymerization, and the use of a dithiobenzoate chain transfer
540 agent, which has a higher chain transfer constant and thus gives narrower molar mass distributions
541 than the xanthate chain transfer agent used to prepare the PAM-PAA and PAM-PAPTAC block
542 copolymers. The use of a trithiocarbonate chain transfer agent and a relatively short P(PEGA) block
543 leads to a fairly narrow composition distribution for P(PEGA)-PAA block copolymers.

544 Finally, the transformation of electrophoretic mobility distributions into composition ratio
545 distributions significantly improved the comparison of the distributions between the different
546 copolymer families, since it takes into account the differences in expansion and drag force according
547 to the chemical nature of the blocks.

548 **Supporting Information.**

549 Synthetic pathway, SEC and NMR characterizations, raw electropherograms are provided for all
550 dibloc copolymers studied in this work.

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677

678 **For TOC only:**

Composition distributions by Capillary Electrophoresis

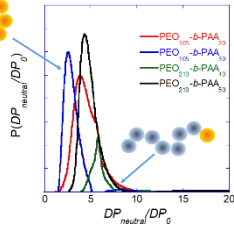
neutral and charged diblock copolymers

- Retardation parameter:

$$X_{sp} = \frac{\mu_{sp}^0}{\mu_{sp}^{block}} - 1$$

- Composition ratio:

$$\frac{DP_{neutral}^0}{DP_0}$$



679

680