First Case of Visceral Leishmaniasis Caused by Leishmania martiniquensis

Bernard Liautaud, Nicolas Vignier, Charline Miossec, Yves Plumelle, Moumini Kone, Delphine Delta, Christophe Ravel, André Cabie, Nicole Desbois

To cite this version:

Bernard Liautaud, Nicolas Vignier, Charline Miossec, Yves Plumelle, Moumini Kone, et al.. First Case of Visceral Leishmaniasis Caused by Leishmania martiniquensis. American Journal of Tropical Medicine and Hygiene, American Society of Tropical Medicine and Hygiene, 2015, 92 (2), pp.317-319. 10.4269/ajtmh.14-0205. hal-02188699

HAL Id: hal-02188699
https://hal.umontpellier.fr/hal-02188699

Submitted on 25 May 2021
**Abstract.** We report the first case of visceral leishmaniasis (VL) caused by *Leishmania martiniquensis* in the Caribbean, which until now, was known only to cause cutaneous leishmaniasis. The disease presented with fatigue, anemia, and hepatosplenomegaly in a 61-year-old man with human immunodeficiency virus (HIV) infection who was receiving antiretroviral therapy. Diagnosis was made by bone marrow biopsy. VL is life-threatening, and its emergence in the Caribbean is of concern.

**INTRODUCTION**

Intracellular protozoa of the *Leishmania* genus, mainly transmitted by sandflies, are the causative agents of leishmaniasis. More than 12 million people are currently infected worldwide. Among its different clinical presentations, visceral leishmaniasis (VL) is life-threatening. Human immunodeficiency virus (HIV) infection is one of the major risk factors for developing VL and reported in 2–12% of all cases.1,2

The new autochthonous and divergent *L. (L.) martiniquensis* n. sp. was first isolated in 1995; its taxonomical position was established in 2002, and it was named in 2014.3–5 This species is up to now restricted to Martinique and has been only reported in patients with cutaneous lesions. We report the first case of VL caused by this parasite in an immunocompromised HIV-infected patient.

**CASE REPORT**

A 61-year-old heterosexual Caribbean male was diagnosed with acquired immunodeficiency syndrome (AIDS) prurigo in 2006. He worked as a painter and a coconut picker and seller. He was born and had always lived in Martinique, except from 1994 to 2001, when he had lived in Guadeloupe. He had traveled to northern Europe and Haiti. His sole medical history was hypertension. He had never used intravenous drugs. At the time of HIV diagnosis, no opportunistic infection was found. Immunological, virological, and therapeutic data are summarized in Table 1.

Combination antiretroviral therapy (cART) was introduced in May of 2007, with significant reduction of HIV viral load and increased CD4+ cell count 1 month later. However, at 1 year, although there was clinical improvement of prurigo and continued viral suppression, CD4 counts were decreasing. Genotypic testing for HIV-1 drug resistance on an initial sample had shown no transmitted resistance, and drug monitoring revealed normal absorption. Despite cART regimen switching in January of 2010, the CD4 counts continued to decline, and the patient remained virally suppressed (Table 1).

He progressively developed hepatosplenomegaly and a normochromic normocytic regenerative anemia of < 10 g/dL. The patient had no fever but reported permanent fatigue. Sulfamethoxazole/trimethoprim (SMX-TMP) toxicity was hypothesized because of a reduced serum folate level, but folinic acid supplementation failed to correct it. A bone marrow biopsy performed in November 2011 revealed inrathistiocytic parasites consistent with the amastigote forms of *Leishmania* spp. (Figure 1).

The diagnosis was confirmed by *Leishmania* polymerase chain reaction (PCR) realized on whole blood with RV1/RV2 probes targeting a kinetoplastic DNA locus (145 bp).7 The molecular identification based on the ribosomal 18S RNA locus analysis gave a 100% identity with the sequence of the MHOM/MQ/92/MAR1 strain (GenBank accession number AF303938.1), a divergent *Leishmania* strain described for the first time in Martinique in 1995 and recently named *L. martiniquensis*.4,5 A retrospective analysis of several sera from our patient stored since 2007 detected high levels of antileishmanial antibodies by indirect immunofluorescence assay (IFA) and enzyme-linked immunosorbent assay (ELISA) using *L. infantum* antigen (Table 1). Immunoblot revealed two specific bands of 14 and 16 kDa, confirming the specificity of these antibodies.

The patient was treated with liposomal amphotericin B (4 mg/kg equal to 300 mg/day from day 1 to 5 and on days 10, 17, 24, 31, and 38 for a cumulative dose of 3 g) and also required a blood transfusion. Symptoms rapidly improved, with decreasing hepatosplenomegaly and disappearance of fatigue, and the patient remained asymptomatic more than 1 year later. Significant increases of hemoglobin level and CD4+ cell count above 350/mm3 followed as well as a negative blood *Leishmania* PCR 20 months later. The patient did not receive secondary prophylaxis.

**DISCUSSION**

Leishmaniasis is endemic in Central and South America but rarely occurs in the Caribbean. The first cases of presumed autochthonous cutaneous leishmaniasis (CL) in Martinique were diagnosed based on direct analysis...
examination of skin smears and consisted of localized CL in immunocompetent patients, except for one case of diffuse CL in an HIV-infected patient. More recently, seven additional CL cases (six of them were unpublished) were found to be caused by a new *Leishmania* species. None of these cases presented visceral dissemination. This parasite was identified by both molecular and isoenzymatic techniques and found to be a member of the *Leishmania* subgenus at the base of the phylogenetic tree. The new *Leishmania* taxon was recently described and named *L. (L.) martiniquensis* n. sp. Capacity of visceralization and dissemination has been shown in a murine model. This is the first reported case of VL caused by *L. martiniquensis*, which highlights the possibility of dissemination in immunocompromized patients of this new species as has been reported for other dermotropic leishmanias.

VL is uncommon in the Caribbean. In Guadeloupe, three cases of possibly autochthonous VL have been observed. Two cases were presumed to be autochthonous based on epidemiological data but without species identification. In 2008, a third case of VL was diagnosed in Guadeloupe in an immunocompetent patient, but it was caused by *L. infantum* (unpublished data). No other autochthonous VL cases have been reported elsewhere in the Caribbean. Our patient most probably acquired leishmaniasis in Martinique, because *L. martiniquensis* has only been reported on this island, but late reactivation of an infection acquired in Guadeloupe or Haiti cannot be ruled out.

Although patients coinfected with VL and HIV commonly are very symptomatic, our patient was clinically well for a long time, but his immunological pattern mimicked that of failure of cART. Cases of VL initially presenting with isolated CD4+ cell count drops are uncommon.

VL/HIV coinfection is usually characterized by significantly lower cure rates of leishmaniasis and higher drug toxicity, relapse, and mortality rates than infection with VL in HIV-seronegative individuals. Although post-treatment increase in CD4+ cells is considered greatly predictive of relapse-free evolution, relapses have been reported in well-controlled HIV patients on cART, thus mandating prolonged follow-up. The patient has continued to do well with no relapse 1 year post-VL treatment.

Research tracks have been paved to identify the parasitic cycle, sandfly vectors, and host reservoirs of *Leishmania*.

---

**Table 1**

Summary of biological and clinical data and history of treatment of a patient coinfected by HIV and autochthonous *Leishmania* in Martinique

<table>
<thead>
<tr>
<th>Date</th>
<th>HIV-1 viral load (copies/mL)</th>
<th>CD4 count (cells/mL)</th>
<th>Lymphocytes (cells/mL)</th>
<th>Total leukocytes (cells/mL)</th>
<th>Hemoglobin (g/dL)</th>
<th>Platelet count (× 10⁹/L)</th>
<th>Clinical manifestations</th>
<th>Others investigations</th>
<th>BM</th>
<th>PCR</th>
<th>bPCR</th>
<th>Serology</th>
<th>Antibiotic</th>
<th>cART</th>
<th>Amphotericin B</th>
</tr>
</thead>
<tbody>
<tr>
<td>January 2007</td>
<td>1.6 × 10⁶</td>
<td>186</td>
<td>1,280</td>
<td>4,580</td>
<td>13</td>
<td>166</td>
<td>HSM and WL</td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td>Yes</td>
</tr>
<tr>
<td>May 2007</td>
<td>4.85</td>
<td>218</td>
<td>2,210</td>
<td>5,530</td>
<td>11</td>
<td>214</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>January 2008</td>
<td>2.0 × 10⁶</td>
<td>218</td>
<td>2,285</td>
<td>4,200</td>
<td>10</td>
<td>206</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>August 2008</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>February 2009</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>June 2009</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>January 2010</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>August 2010</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>January 2011</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>November 2011</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>February 2012</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>June 2012</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>January 2013</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>July 2013</td>
<td>&lt; 20</td>
<td>4.5</td>
<td>1,830</td>
<td>3,650</td>
<td>9</td>
<td>241</td>
<td></td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>+</td>
<td>SMX-TMP</td>
<td>+</td>
<td></td>
</tr>
</tbody>
</table>

**Figure 1.** Intrahistiocytic amastigote forms of *Leishmania* on a bone marrow specimen from the patient (May Grunwald Giemsa × 1,000).
species in the Caribbean. *Lutzomyia atroclavata* has been identified in Martinique, Guadeloupe, and the Virgin Islands; black rats (*Rattus rattus*), mongooses (*Herpestes auropunctatus*), marsupials (*Didelphis marsupialis*), and canids are all potential animal reservoirs that should be investigated.\(^8\),\(^16\)

The emergence of *L. martiniquensis* infection with the possibility of visceral extension could be of concern in the Caribbean region, where the prevalence of HIV infection is high.

Received April 6, 2014. Accepted for publication October 6, 2014.

Published online November 17, 2014.

Acknowledgments: We are indebted to Brigitte Roche, who facilitated, in 1992, the identification of the first *Leishmania* MHOM/MQ/92/MAR1 strain. We are grateful to Ms. Marlene Ouca for technical assistance in management of stored sera samples and Patrick Hochedez and Vanessa Rouzier for reviewing the manuscript.

Authors’ addresses: Bernard Liautaud and Nicolas Vignier, Department of Infectious and Tropical Diseases, University Hospital of Fort-de-France, Fort-de-France, Martinique, French West Indies, E-mails: Bliautaud1@gmail.com and vigniernicolas@yahoo.fr. Charline Miossec and Nicole Desbois, Department of Parasitology and Mycology, University Hospital of Fort-de-France, Fort-de-France, Martinique, French West Indies, E-mails: charlinemiossec@hotmail.fr and nicole.desbois@chu-fortdefrance.fr. Yves Plumelle, Moumini Kone, and Delphine Delta, Department of Hematology, University Hospital of Fort-de-France, Fort-de-France, Martinique, French West Indies, E-mails: yves.plumelle@chu-fortdefrance.fr, mt.kone@chu-fortdefrance.fr, and delphine.delta@gmail.com. Christophe Ravel, Université Montpellier 1, Montpellier, France, and Regional University Hospital of Montpellier, UMR5290, French National Reference Center for Leishmaniasis, University Hospital of Montpellier, Montpellier, France, E-mail: christophe.ravel@univ-montp1.fr. André Cabie, Department of Infectious and Tropical Diseases and Inserm CIE802, University Hospital of Fort-de-France, Fort-de-France, Martinique, French West Indies, E-mail: andre.cabie@chu-fortdefrance.fr.

**REFERENCES**


