



HAL
open science

Genetic consequences of long-term isolation for the last French population of *Eryngium viviparum* 1

Pauline Rascle, Elodie Flaven, Frédéric Bioret, Sylvie Magnanon, Erwan Glemarec, Eric Imbert, Sébastien Gallet

► **To cite this version:**

Pauline Rascle, Elodie Flaven, Frédéric Bioret, Sylvie Magnanon, Erwan Glemarec, et al.. Genetic consequences of long-term isolation for the last French population of *Eryngium viviparum* 1. 2019. hal-02172011

HAL Id: hal-02172011

<https://hal.umontpellier.fr/hal-02172011>

Preprint submitted on 10 Jul 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Genetic consequences of long-term isolation for the last French population of *Eryngium viviparum*
2 J.Gay (Apiaceae).

3

4 Pauline Rascle^{a*}, Elodie Flaven^b, Frédéric Bioret^a, Sylvie Magnanon^c, Erwan Glemarec^c, Sébastien
5 Gallet^a, Eric Imbert^b

6

7 ^a EA7462: Géoarchitecture, Territoires, Urbanisation, Biodiversité, Environnement

8 Université de Bretagne Occidentale-Brest, France

9 ^b ISEM

10 Université de Montpellier, CNRS, IRD, EPHE-Montpellier, France.

11 ^c Conservatoire Botanique National de Brest, France

12 *corresponding author

13

14 ABSTRACT

15

16 *Eryngium viviparum* (Apiaceae) is an endangered endemic plant of the Atlantic region of Europe,
17 growing in seasonally flooded sites. The species is characterized by a highly disjunct distribution.

18 Indeed, it occurs only in a few sites in the North-West part of the Iberian Peninsula and in a single

19 locality in France. In order to improve the conservation status of *Eryngium viviparum* in France, a
20 conservation program has been implemented, which plans reintroduction actions. Before considering

21 such an operation, genetic studies are conducted in order to determine the genetic status of the last

22 French population and to identify the genetic source that should be considered for the best

23 reintroduction strategy. Using microsatellite markers, we documented the genetic structure of the last

24 French population, and compared its genetic diversity with ten Iberian populations, which cover the

25 three geographic regions where the species occurs. As expected, the French population of *Eryngium*
26 *viviparum* present a very low genetic diversity due to bottleneck and to geographical isolation. The
27 evolutionary potential appears very low, with no private allele in this population. Furthermore, this
28 population is highly differentiated from the Iberian populations, both for genetic variation and
29 ecological niche. These results imply new questions about the conservation of *Eryngium viviparum* in
30 France, especially for management and reintroduction, which are aimed to favor genetic diversity and
31 to avoid extinction.

32

33 *KEYWORDS*

34

35 Conservation genetics - Endangered species - *Eryngium viviparum* - Isolated population -
36 Microsatellites

37 INTRODUCTION

38

39 Ongoing habitat destruction, mainly induced by human activities, is described as “the most serious
40 threat to biological extinction crisis” by causing a reduction and fragmentation of species geographic
41 range (Wilcox & Murphy, 1985; Saunders, Hobbs & Margules, 1991; Sala *et al.*, 2000). Consequently,
42 natural populations became smaller and more isolated in anthropogenic landscapes (McGarigal &
43 Cushman, 2002; Fahrig, 2003), and more sensitive to demographic and genetic stochasticity (Young,
44 Boyle & Brown, 1996; Lowe *et al.*, 2005; Ouborg, Vergeer & Mix, 2006; Honnay & Jacquemyn, 2007).

45 As gene flow appears to be restricted in fragmented species, an erosion of genetic diversity within
46 isolated populations and an increase of the genetic divergence with other populations are expected
47 (Young & Clarke, 2000; Newman & Tallmon, 2001). In the initial stage after the isolation, both the
48 number of polymorphic loci and the number of alleles per locus decrease, due to bottleneck and
49 genetic drift (Ellstrand & Elam, 1993; Young *et al.*, 1996). If isolation persists, the population can
50 experience a significant decrease of vigor and fecundity due to the increased inbreeding and the
51 accumulation of deleterious alleles (Lynch, Conery & Burger, 1995; Higgins & Lynch, 2001; Keller &
52 Waller, 2002; Reed & Frankham, 2003). In the short-term, the fragmented population becomes more
53 vulnerable face to environmental stochasticity, and population extinction risk can drastically increase
54 (Huenneke, 1991; Young *et al.*, 1996). On the long-term, genetic depletion reduces the population
55 ability to adapt to any environmental change (Barrett & Kohn, 1991). Genetic drift, consequent to
56 isolation, also contributes to increase the genetic differentiation between the isolated population and
57 populations in the core distribution of the species (Pironon *et al.*, 2016). Small populations are often
58 more particularly affected (Leimu *et al.*, 2006; Richards, 2000), which can sometimes lead to the
59 extinction of the population (Lande & Barrowclough, 1987; Spielman, Brook & Frankham, 2004). The
60 species vulnerability face to these negative consequences also varies according to any life-history trait
61 that reduces the effective population size such as a short-life cycle (Young *et al.*, 1996), and selfing

62 (Hamrick & Godt, 1989). In contrast, the ability to reproduce clonally may buffer the genetic events
63 (Gitzendanner & Soltis, 2000; Honnay & Bossuyt, 2005).

64 Integrating genetic variation has become a key element of practical conservation and adapted
65 management (Holsinger & Gottlieb, 1991; Hamrick & Godt, 1996; Rieseberg & Swensen, 1996;
66 Escudero, Iriondo & Torres, 2003) and is particularly relevant when reintroduction or reinforcement
67 are planned (Mistretta, 1994; Havens, 1998; Falk *et al.*, 2006; Neale, 2012). For example, genetic
68 studies are used to identify the most appropriate source of plant material for reintroduction operations
69 (Haig, 1998; Petit, Mousadik & Pons, 1998; Lawrence & Kaye, 2011), giving the advantage to suitable
70 level of genetic diversity (Breed *et al.*, 2013). Using the closest geographic population as source
71 material for reinforcement is often recommended (McKay *et al.*, 2005), while using multiple source
72 populations appears relevant when populations exhibit a low genetic variability (Vergeer *et al.*, 2005;
73 Maschinski *et al.*, 2013). However, it should be noted that, despite guidelines and recommendations,
74 a few number of reintroduction success, or reinforcement, have been recorded yet in plant species
75 (Godefroid & Vanderborcht, 2011).

76 *Eryngium viviparum* J. Gay, is a *priority species* of the European Habitats Directive and the Bern
77 Convention. It occurs in vernal pools, highly specific of these open habitats maintained by the
78 temporary flooded system and traditional farming practices (Jovet, 1939). In the 1980's, *Eryngium*
79 *viviparum* experienced a rapid population decline throughout its natural range, especially in France,
80 leading to a highly fragmented Ibero-Armorican distribution (Fig. 1). The reduction of suitable habitats,
81 due to change in farming practices and urbanization, is a significant threat for the survival of the species
82 (Magnanon, Hardegen & Guillevic, 2013). In France, it is considered as *critically endangered* (Olivier,
83 1995) as only one population remains. For these reasons, several protection measures has been
84 applied since 1987, including annual monitoring, demographic studies, and conservation management
85 of the population. More recently, a National Action Plan (NAP) was implemented, which supports the
86 long-term conservation of *Eryngium viviparum* in France (Magnanon *et al.*, 2013). This program
87 includes notably the restoration of extinct populations. This latter aspect implies to define the best

88 genetic sources to be reintroduced. Despite the strong isolation of the French population and its
89 reduced spatial distribution (0.1 ha), annual census showed an increasing population between 1994
90 and 2016 (from 1 500 to 10 000 individuals), including an average of 30% of flowering plants over the
91 summer (Guillevic, unpublished data). However, considering the clonal ability of this species, the
92 effective population size should be significantly lower than the demographic one.

93 The aim of this study is to investigate the genetic status of the remaining *Eryngium viviparum* French
94 population. Using microsatellites markers, specifically designed for the study, we compared the within
95 population genetic diversity between the French population and the Iberian populations (Spain and
96 Portugal), to infer consequences of isolation. According to the theoretical models in population
97 genetics, it is expected that this population has a low genetic diversity. We also identified which Iberian
98 population is the most genetically related to the French one, in order to provide practical
99 recommendations for the reintroduction plan. Under these objectives, ecological differentiation
100 between *Eryngium viviparum* populations is also evaluated, based on climatic and vegetation data.

101 MATERIAL AND METHODS

102 *STUDY SPECIES*

103
104
105
106 *Eryngium viviparum* J. Gay (Apiaceae) is a diploid species, occurring in temporary ponds. These
107 monocarpic species combines sexual and clonal reproduction. The flowers are hermaphroditic and
108 pollinated by insects. Numerous clonal rosettes emerge at the root plate level of the mother plants
109 and on the flower stems. Seed germination and clonal plants development occur mostly in Autumn.
110 The dispersion appears limited as clonal individuals and seedlings are mainly observed at the foot of
111 the mother plants, causing a patchy distribution.

112 The species is distributed in the north-west of France (Brittany) and north-west part of the Iberian
113 peninsula in three distinct geographical groups (North, Central and South, Fig. 1). Only one population
114 is known in France, occurring in the protected area of "les Quatre-chemins" (Belz, department of

115 Morbihan). This population is nowadays strongly isolated, whereas in the 80's, about 40 populations
116 were known (Magnanon *et al.*, 2013). The North Iberian populations also experienced a decline in the
117 last decades, due to habitat modification (Romero, Ramil & Rubinos, 2004).

118

119 *SAMPLING DESIGN AND MICROSATELLITE ANALYSES*

120

121 Leaves were sampled in 2014 on 247 individuals from 11 populations (Fig. 1). Sampled individuals were
122 sufficiently spaced from each other, to reduce the probability of sampling identical genotypes. Samples
123 were dried and stored in silica gel. The number of samples per population ranged from 15 to 27
124 individuals for the Iberian populations, depending on population size, and 37 individuals were sampled
125 from the French population (Table 1).

126

127 Genomic DNA was extracted from dried leaves using the CTAB protocol from Doyle & Doyle (1990).
128 The genotypes of each individual were characterized using 7 operational microsatellite markers (Table
129 2) specifically developed for *Eryngium viviparum* by the biotechnology company "Genoscreen" (Lille,
130 France). Microsatellite loci were isolated by Titanium pyrosequencing (Malausa *et al.*, 2011) and
131 designed using QDD pipeline (Megléczy *et al.*, 2009). The PCR were processed performing two reactions,
132 multiplexing markers based on size compatibility, and using fluorescent labeling of the forward primers
133 (Applied Biosystems). The PCR were carried out in a final volume of 10 μ L, including 1 μ L of the
134 extracted DNA, 0.2 μ L of the forward and reverse primers, and 2X QIAGEN Multiplex Master Mix (5 μ L,
135 QIAGEN, France). All the microsatellites amplifications were performed using a thermocycler
136 (Eppendorf Pro) under the following temperature conditions: initial denaturation at 95°C for 15 min,
137 followed by 30 cycles of 94°C for 30 seconds (denaturation), 90 seconds at 60°C (annealing) and 60
138 seconds at 72°C (elongation), finished by the final extension step at 60°C during 30 min. The sizes of

139 PCR products were analysed by electrophoresis using a 24 capillary Genetic analyser (ABI3500XL,
140 Applied Biosystems). The raw data were visualized with GeneMapper 5.0 (Applied Biosystems).

141

142 *POPULATION SIZE AND ECOLOGY*

143

144 Individual censuses were performed each year in the French population, but no comparable estimation
145 exists for Iberian populations. Therefore population size was estimated using the population surface
146 and the abundance of *Eryngium viviparum* individuals evaluated by Glemarec *et al.* (2017), and was
147 summarized in 5 classes: 1: < 500 individuals; 2: 500 to 2 000 individuals; 3: 2 000 to 10 000 individuals;
148 4: 10 000 to 50 000 individuals; 5: > 50 000 individuals. Geographic distance to the nearest existent
149 population was estimated from GPS coordinates and used as connectivity indice (Table 1).

150

151 Ecological distances among populations were considered using both climatic and vegetation data.
152 Three climatic data were extracted from WorldClim 2 database (Hijmans *et al.*, 2005; Trabucco &
153 Zomer, 2009) with 30 arc second spatial resolution (about 1km): maximum and minimum annual
154 temperature (T_{MAX} and T_{MIN} , respectively) and annual precipitation (PP, Table 1). Mean values between
155 1970 and 2000 were used for each population.

156 For eight of the sampled populations (S_1 , S_2 , C_1 , C_3 , C_4 , N_1 , N_2 and FRENCH), vegetation communities
157 have been characterized by Glemarec *et al.* (2017) according to the phytosociological approach. From
158 this dataset, presence/absence of co-occurring species have been extracted (Supplementary Material,
159 Table S1), as they appeared relevant to characterized global ecological factors (Gillet, 2000). These
160 qualitative data have been preferred to abundance/dominance values since species abundances vary
161 according to the management regime which is very heterogeneous among *Eryngium viviparum*
162 populations.

163

164 DATA ANALYSES

165
166 *MICROSATELLITE POLYMORPHISM AND DIVERSITY PARAMETERS*

167
168 The presence of null alleles was checked using Micro-Checker 2.2.3 (Van Oosterhout *et al.*, 2004).
169 Linkage disequilibrium for each pair of loci within each population and the conformity to Hardy-
170 Weinberg equilibrium were tested with a significance level of 5% using GENEPOP 4.0 (Rousset, 2008).
171 When multiple tests were involved the sequential Bonferroni correction was applied to adjust
172 significance values (Rice, 1989).

173 Several genetic diversity parameters, including average number of alleles per locus, the observed
174 heterozygosity, the unbiased expected heterozygosity, and the fixation index were computed using
175 GENETIX 4.05 (Belkhir *et al.*, 1996). Correlations between population size and diversity parameters,
176 and between connectivity and diversity, were tested using Spearman ranks correlations (significance
177 level of 0.05).

178
179 *PAST DEMOGRAPHY DYNAMICS*

180
181 Demographic changes in effective population sizes were inferred using the Migraine software (Version
182 0.5, Leblois *et al.*, 2014). A detailed procedure can be found in Zenboudji *et al.* (2016). Briefly, we
183 estimated two parameters: the ancestral $\theta_{anc}=4N_{anc}\mu$, and the actual $\theta_{act}=4N_{act}\mu$, where N_{act} is the
184 current effective population size, N_{anc} is the ancestral population size, and μ is the mutation rate per
185 locus per generation. The parameter $Nratio=N_{act}/N_{anc}$ allows to detect, either a reduction (ratio <1) or
186 an expansion (ratio >1) in population size. For each $Nratio$ estimated, its 95 % confidence intervals was
187 used to test for significant difference with 1. All Migraine runs were done using the pGSM model for

188 mutation model (Leblois *et al.*, 2014), with 2000 trees per iteration and 512 points per tree and 9
189 iterations.

190

191 *DIFFERENTIATION AMONG POPULATIONS*

192

193 Pairwise F_{ST} values were calculated with GENEPOP 4.0 (Rousset, 2008). Isolation by distance was tested
194 using Mantel test between $F_{ST}/(1-F_{ST})$ and log (linear distance), with a significance level of 5%.
195 Furthermore, we used Nei's D_A distances (Nei, Tajima & Tatenno, 1983), to study the relationship among
196 populations and groups of populations. Overall genetic differentiation was evaluated using a principal
197 components analysis (PCA) using the "adegenet" R package (Jombart, 2008). Bayesian analysis of
198 genetic structure was also applied using STRUCTURE v.2.3.4 (Pritchard, Stephens & Donnelly, 2000).
199 The admixture models were performed for eleven independent runs ($K=1$ to 11), with 10 replicates at
200 each value of K . Each run consisted of 5 000 Markov Chain Monte Carlo (MCMC) repetitions following
201 a burn-in period of 5 000 iterations. The optimum value of K was determined according to the $\Delta(K)$
202 method developed by Evanno, Regnaut & Goudet (2005) with the STRUCTURE HARVESTER tool (Earl
203 & vanHoldt, 2012).

204

205 Ecological data were used to compute a dissimilarity matrix among the 8 concerned populations using
206 the R "Vegan" package (Oksanen *et al.*, 2016; R Development Core Team 2016). We used the Gower
207 coefficient (Gower, 1971), since it allows to combine continuous variables (climatic data) and binary
208 data (vegetation data).

209

210 Correlations among genetic distances ($F_{ST}/(1-F_{ST})$ and Nei's D_A), geographical distances (log-
211 transformed, and obtained from GPS coordinates) and ecological distances were tested, at an alpha
212 level of 0.05, using Mantel tests implemented in the R "Vegan" package (R Development Core Team,

213 2016; Oksanen *et al.*, 2016) with 1000 permutations. Following Guillot & Rousset (2013), we did not
214 perform partial Mantel tests.

215

216 RESULTS

217

218 *MICROSATELLITE POLYMORPHISM*

219

220 The number of alleles per locus varied from 5 (pmEV01) to 19 (pmEV04). Only the locus pmEV05
221 showed sign of null alleles with frequencies ranging from 0.08 to 0.41. This locus is the only one
222 showing a significant heterozygote deficiency ($P < 0.01$, Table 2).

223 Among the 231 tests used to detect for linkage disequilibrium between loci, only three were significant
224 after the Bonferroni correction: pmEV04/pmEV17, pmEV04/pmEV09, and pmEV05/pmEV09 ($P <$
225 0.001). Hardy-Weinberg equilibrium was tested at each locus for each population and only three tests
226 (out of 77) showed a significant deviation from Hardy-Weinberg expectations after Bonferroni
227 correction for loci pmEV04 and pmEV05 in the N_3 population and locus pmEV05 in N_2 population ($P <$
228 0.001). This is congruent with presence of null alleles for pmEV05.

229

230 *GENETIC DIVERSITY WITHIN POPULATIONS*

231

232 As expected, the French population showed the lowest values for the three parameters estimating the
233 within-population diversity (Table 3). The French population was the only one with a significant
234 heterozygote deficit ($F_{IS}=0.17$, $P < 0.05$, Table 3). Genetic diversity was quite similar among the Iberian
235 populations, with the exception of the smallest population (S_2), which had the lowest genetic diversity
236 (Table 3), but which was nevertheless twice that in the French population. These results are congruent

237 with the ratio between the number of multilocus genotypes and the sample size (Table 3). The French
238 population showed the lowest value and all the Iberian populations showed high values. However, no
239 similar multilocus genotype was detected across populations.

240 None of the genetic diversity parameters were correlated with population size ($P > 0.05$), with or
241 without the French population. In contrast, distance to the closest population, used as a connectivity
242 index, was correlated to N_A , H_{OBS} and F_{IS} ($R=-0.61$, -0.66 , and 0.69 respectively, $P < 0.05$) when
243 considering the French population, but not anymore when excluding it ($P > 0.05$).

244

245 *PAST DEMOGRAPHIC DYNAMICS*

246

247 According to the N ratio estimates produced by Migraine, no population showed a significant
248 demographic expansion ($P > 0.05$). A bottleneck (N ratio > 1) was detected in most of the sampled
249 populations, and all of them have recovered allelic diversity (Table 3). Consistent with the low diversity,
250 a bottleneck was also detected for the French population (Table 3).

251

252 *DIFFERENTIATION AMONG POPULATIONS*

253

254 *Genetic differentiation*

255

256 Global F_{ST} value (0.29) indicated high differentiation among populations. For the Iberian populations,
257 the genetic differentiation was also high ($F_{ST}=0.24$). Pairwise F_{ST} values ranged from 0.04 to 0.6, and 26
258 values out 55 were > 0.25 (Table 4). Isolation by distance tested using the correlation between $F_{ST}/(1-$
259 $F_{ST})$ and $\log(\text{geographical distance})$ was significant considering the 11 populations ($R=0.65$, $P < 0.01$,
260 Fig. 2), and was also significant excluding the French population ($R=0.41$, $P=0.01$). Correlation between

261 Nei D_A and log (geographical distance) was also significant whether the French population was
262 included, or not, in the analysis ($R > 0.58$, $P < 0.001$).

263

264 The PCA also suggested differentiation between the FRENCH and Iberian populations (Fig. 3A). This
265 observation is also consistent with the structure analysis. According to the $\Delta(K)$ method, the most
266 appropriate value of K given for our data was $K=4$ (Fig. 3B). These four clusters of individuals match the
267 four geographical regions where *Eryngium viviparum* occurs (FRENCH, North, Central and South of
268 Iberian peninsula, Fig. 3C).

269

270 *Ecological differentiation*

271

272 Gower distances ranged from 0.19 (N_2 & N_1) to 0.67 (French & S_1). Genetic and Gower distances were
273 positively correlated using Nei D_A values ($R=0.59$, $P < 0.001$), but not when using linearized F_{ST} ($R=0.43$,
274 $P > 0.05$). These correlations, using both Nei D_A and linearized F_{ST} , were not significant when
275 considering only the Iberian populations ($P > 0.05$).

276 Among populations, the plant communities share common species, characterized by pioneer species
277 growing in seasonally flooded open habitats, such as *Exaculum pusillum* (Lam.) Caruel (Gentianaceae),
278 *Chamaemelum nobile* (L.) All. (Asteraceae), *Baldellia ranunculoides* (L.) Parl. (Alismataceae) and
279 *Eleocharis multicaulis* (Sm.) Desv (Cyperaceae). However some ecological vicariations were observed
280 among *Eryngium viviparum* co-occurring species. For example, *Agrostis canina* L. (Poaceae) was
281 observed only in the French population, while *Agrostis hesperica* Romero Garcia, Blanca López &
282 Morales Torres was identified in North and Central Iberian population, and *Agrostis pourrettii* Willd.
283 only in South Iberian populations.

284

285 Considering only the presence/absence of species, the French population seems closer to North Iberian
286 populations, sharing several Atlantic species, such as *Aristavena setacea* (Huds.) F.Albers & Butzin
287 (Poaceae) and *Galium debile* Desv. (Rubiaceae), which are not reported in the other two clusters. S₁ is
288 the furthest population from the French one, characterized by the occurrence of Mediterranean species
289 such as *Agrostis pourrettii* (Poaceae), *Pulicaria paludosa* Link. (Asteraceae) and *Mentha cervina* L.
290 (Lamiaceae).

291

292

293 DISCUSSION

294

295 *ASSESSMENT OF GENETIC DIVERSITY FOR THE ONLY FRENCH POPULATION*

296

297 Among the 11 studied populations, a reduction of effective size was detected in eight populations.
298 These bottlenecks are consistent with the historical data of the species, known from the botanical
299 literature and herbarium specimens (Magnanon *et al.*, 2013). For instance, in France, since the first
300 description of the species in 1839, 36 populations had been recorded in a limited area in Brittany, and
301 all of them except one, have disappeared. However, the genetic consequences of these bottlenecks
302 appear very different according to the geographic area. Indeed, all the Iberian populations, even the
303 smallest one (<500 individuals) appear to have recovered genetic diversity. In contrast, the French
304 population shows an extremely low level of genetic diversity and an evidence of inbreeding.

305

306 Moreover, only one rare allele was found for the French population for the pmEV09 locus (frequency
307 <5%), and no private allele was observed, suggesting an absence of specificity for this population
308 considering these seven microsatellites. Conversely, some Iberian populations display private alleles
309 and a global high multi-locus allelic diversity, except for S₂, the smallest one, which appeared twice

310 diversified than the French population. The level of genetic diversity detected in the French population
311 appeared especially lower than those noticed in other endangered Apiaceae, such as its congeneric
312 *Eryngium alpinum* L. (Gaudeul & Till-Bottraud, 2008), or other species under similar isolated context
313 (Wiberg *et al.*, 2016; Tamaki, Setsuko & Tomaru, 2016; Aavik *et al.*, 2017).

314 In this study, population size is not correlated with genetic diversity, while genetic diversity
315 and heterozygosity are commonly expected to be positively correlated with population size (Ellstrand
316 & Elam, 1993; Frankham, 1996; Leimu *et al.*, 2006). Among the eleven populations studied, the French
317 population showed the lowest genetic variability, despite a population size estimated to 10 000
318 individuals in 2016, which was considered as intermediate when compared to Iberian populations.
319 Therefore, according to our results, isolation appeared to have more negative influence than
320 population size on the genetic diversity for *Eryngium viviparum*, and connectivity seems to reduce the
321 effect of bottleneck. This isolation effect has been reported in some genetic studies on other plant
322 species (*Eucalyptus albens* Benth., Prober & Brown, 1994; *Anthyllis vulneraria* L., Honnay *et al.*, 2006),
323 which suggest to avoid population fragmentation when possible or to reestablish connectivity among
324 populations. The demographic size of the French population reveals the efficiency of the management
325 strategy to sustain the population. However, the absence of genetic diversity due to the combined
326 effect of bottleneck and spatial isolation could cast doubt on the long-term persistence of the
327 population.

328 The species is also characterized by clonal propagation, which often contributes to decrease genetic
329 diversity (Hamrick & Godt, 1989; Watkinson & Powell, 1993; Young *et al.*, 2002; Vallejo-Marín, Dorken
330 & Barrett, 2010). It could ultimately lead to a monoclonal genotypic pattern (Balloux, Lehmann & De
331 Meeûs, 2003; Honnay & Bossuyt, 2005). However, in some cases, clonal propagation may benefit
332 species under isolated context, by maintaining allelic diversity, polymorphism and heterozygosity, but
333 only on the short-term (Ellstrand & Roose, 1987; Auge *et al.*, 2001; Meloni *et al.*, 2013). The monocarpy
334 (short-lived species) and the poor dispersal ability observed for *Eryngium viviparum* are also factors
335 that are well-known to induce a loss of genetic diversity (Young *et al.*, 1996).

336

337

338

339 *STRONG STRUCTURE BETWEEN GEOGRAPHIC GROUPS*

340

341 Consistent with the low dispersal ability of the species, but despite the putative contribution of genetic
342 drift, isolation by distance revealed that geographical distance appears to be a major component of
343 the genetic differentiation among populations. The French population appears less distant from the
344 North Iberian populations, as suggested by Rodriguez-Gacio *et al.* (2009) with RAPD analyses. This
345 relative genetic proximity is probably the result of a common biogeographical history. Nevertheless,
346 the F_{ST} showed a high differentiation between the French population and the Iberian ones, which is
347 consistent with the spatial isolation. Such high F_{ST} levels are regularly found for rare plants (Maguire *et*
348 *al.*, 2000).

349 Consistently with the geographical distances, bioclimatic data also revealed that the French population
350 is distant from all other populations. This population occurs on temperate hyperoceanic domain, while
351 northern and central Iberian populations are located respectively under temperate oceanic and
352 temperate submediterranean bioclimate (Rivas-Martínez, Rivas-Sáenz & Penas-Merino, 2011).
353 However, Glemarec *et al.* (2017) considered that the French population and the northern Iberian
354 populations occur under the same bioclimatic region, as temperate oceanic, and highlight similar plant
355 associations. In these two regions, *E. viviparum* communities are characterized by Atlantic wetland
356 species which belong to the floristic associations of *Eleocharitetum multicaulis* Allorge 1922 ex Tüxen
357 1937 and *Deschampsio setaceae-Agrostietum caninae* Lemée 1937. The populations S_1 and S_2 located
358 in the southern limit of the *E. viviparum* distribution range, are characterized by a Mediterranean
359 pluvioseasonal oceanic bioclimate (Rivas-Martínez *et al.*, 2011) and the occurrence of Atlantic and
360 Mediterranean species. According to Glemarec *et al.* (2017), these floristic composition may be linked

361 to the associations of *Pulicario uliginosae-Agrostietum salmanticae* Rivas Goday 1956 and *Periballio*
362 *laevis-Illecebretum verticillati* Rivas Goday 1954. These two populations also appeared more
363 genetically differentiated from the other Iberian populations. Therefore, both ecological and genetical
364 data confirm the differentiation between the French population and the Iberian populations, and also
365 between the three Iberian clusters.

366

367 *IMPLICATIONS FOR CONSERVATION*

368

369 With its strong isolation (<500 km), excluding any genetic exchange with other populations, the French
370 population of *Eryngium viviparum* is a suitable model to discuss implications of genetic studies for
371 conservation purposes. The very low genetic diversity, on the edge of monoclonality, can reduce the
372 environmental adaptability of the species (Frankham, 2005). This low allelic diversity can also lead to
373 inbreeding depression and affect the viability of individuals by fixation of deleterious alleles (Lynch *et*
374 *al.*, 1995; Higgins & Lynch, 2001; Keller & Waller, 2002; Reed & Frankham, 2003). Correlation between
375 population size and reduction in fitness is a common pattern for plants (Reed, 2005). Indeed, a
376 decrease of the viability of both seeds and seedlings seems already to occur in the French population
377 (Guillevic, pers. com.). Moreover, *ex situ* germination experiments show lower germination rates for
378 seeds sampled on French individuals (30%) than seeds collected on Iberian individuals (80%, Gautier,
379 pers. com.). These observations combined to our results imply that restoring the genetic diversity in
380 the French population should be considered.

381 However, introduction of new genotypes in an endangered population always remains problematic for
382 stakeholders concerned with the species conservation, arguing that it will alter the genetic identity of
383 the local population (Maurice *et al.*, 2013). An alternative solution is to reestablish connectivity by
384 restoring populations, as planned within the framework of the NAP (Magnanon *et al.*, 2013). The long-
385 term and ambitious objective is to re-create a sustainable metapopulation of *E. viviparum* in Brittany.

386 Nevertheless, guidelines for reintroduction suggest avoiding low genetic diversity (Montalvo *et al.*,
387 1997; Weeks *et al.*, 2011), and studies have shown that reintroducing material from multiple
388 populations can increase translocation success (Vergeer *et al.*, 2005; Maschinski *et al.*, 2013) and
389 restore significantly the genetic diversity (Zavodna *et al.*, 2015). This would suggest that in absence of
390 other French sources, reintroduction could be implemented using Iberian genetic material. It is also
391 generally recommended choosing source material from the nearest populations (Brown & Marshall,
392 1995; McKay *et al.*, 2005; Bottin *et al.*, 2007), in order to avoid environmental maladaptation. As the
393 French population clearly appears strongly ecologically distant from the Iberian populations, it should
394 be necessary to test for a possible outbreeding depression between French and Iberian populations
395 before any reintroductions (Yardeni *et al.*, 2016). Populations from the Northern part of the Iberian
396 distribution range should be favored, as they present the lesser distance for both ecological and
397 genetic data. Experimental crossing should also be considered beforehand, to test the viability of first
398 generations, and their adaptability to survive in the natural habitat of *E.viviparum*.

399

400 CONCLUSION

401

402 This study brings new elements of the genetic status of *Eryngium viviparum*, an important requirement
403 for the French National Action Plan. Seven microsatellites markers were developed and validated
404 specifically for the genetic characterization of *E. viviparum*. Our results clearly contribute to guide
405 management and restoration operations for the species, and suggest that the restoration of extinct
406 populations is a priority. Experimental reintroductions, using only the French genotypes, had already
407 occurred in order to optimize technical modalities and to identify factors that influence the success of
408 *E. viviparum* reintroductions (Rasclé *et al.*, 2018). However, the origin of the material must be clarified
409 by some complementary studies in order to test the viability and the environmental adaptation of
410 offspring from controlled crosses.

411

412 AKNOWLEDGEMENTS

413

414 First, we are grateful the two anonymous reviewers for providing helpful comments. Financial support
415 was provided by the Regional Department for the Environment of Brittany (DREAL Bretagne) and the
416 Water Agency Loire-Bretagne, as part of the French National Action Plan in favor of *Eryngium*
417 *viviparum*, and by the University of Brest. This study could not occur without the CNPN (Conseil
418 national de la protection de la nature), the DDTM Morbihan (Direction départementale des territoires
419 et de la mer), the IBADER (Instituto de Biodiversidade Agraria e Desenvolvimento Rural, Lugo) and the
420 ICNF (Instituto de Conservação da Natureza e das Florestas, Portugal), which authorized the sample
421 collection. This work would also not be realized without the NGO Bretagne Vivante reserve disposal,
422 nor without the management applied since 1991, which has allowed the conservation of *Eryngium*
423 *viviparum* in France. We thank Yvon Guillevic, the reserve manager, for its assistance and its important
424 suggestions. We are also truly grateful to: Pr Pablo Ramil-Rego, Javier Ferreiro Da Costa, Marco Rubinos
425 Román, and Boris Alejandro Hinojo Sánchez (IBADER); Serafín González Prieto and Alejandra Couto-
426 Vásquez (Sociedad Gallega de Historia Natural, SGHN); and to Carla Marisa Quaresma (ICNF); which
427 accompanied us among the studied Iberian populations. Microsatellites genotyping was carried out at
428 the Platform Genseq of the LabEx “Centre Méditerranéen Environnement Biodiversité” (ISEM,
429 Montpellier, France) with the help of Frédérique Cerqueira and Erick Desmarais. We also thank
430 Raphael Leblois for his help to produce Migraine results.

431

432 REFERENCES

433

434 **Aavik T, Talve T, Thetloff M, Uuemaa E, Oja T. 2017.** Genetic consequences of landscape change for

435 rare endemic plants - A case study of *Rhinanthus osiliensis*. *Biological Conservation* **210**: 125-135.

436 **Auge H, Neuffer B, Erlinghagen F, Grupe R, Brandl R. 2001.** Demographic and random amplified
437 polymorphic DNA analyses reveal high levels of genetic diversity in a clonal violet. *Molecular Ecology*
438 **10(7)**: 1811-1819.

439 **Balloux F, Lehmann L, De Meeûs T. 2003.** The population genetics of clonal and partially clonal
440 diploids. *Genetics* **164(4)**, 1635-1644.

441 **Barrett SH, Kohn J. 1991.** Genetic and evolutionary consequences of small population size in plants:
442 implications for conservation. In Falk DA, Holsinger KE, eds. *Genetics and conservation of rare plants*.
443 New York: Oxford University Press, 3-30.

444 **Belkhir K, Borsa P, Chikhi L, Raufaste N, Bonhomme F. 1996.** GENETIX 4.05, logiciel sous Windows™
445 pour la génétique des populations, v4. 05. *CNRS UMR 5171, Laboratoire génome, populations,*
446 *interactions, France.*

447 **Bottin L, Le Cadre S, Quilichini A, Bardin P. Moret J, Machon N. 2007.** Re-establishment trials in
448 endangered plants: A review and the example of *Arenaria grandiflora*, a species on the brink of
449 extinction in the Parisian region (France). *Ecoscience* **14(4)**: 410-419.

450 **Breed MF, Stead MG, Ottewell KM, Gardner MG, Lowe AJ. 2013.** Which provenance and where? Seed
451 sourcing strategies for revegetation in a changing environment. *Conservation Genetics* **14(1)**: 1-10.

452 **Brown AHD, Marshall DR. 1995.** A basic sampling strategy: theory and practice. In *Collecting plant*
453 *genetic diversity: technical guidelines*. CAB International, Wallingford, 75-91.

454 **Doyle JJ, Doyle JL. 1990.** Isolation of Plant DNA from fresh tissue. *Focus* **12**: 13-15.

455 **Earl DA, VanHoldt BM. 2012.** STRUCTURE HARVESTER: a website and program for visualizing
456 STRUCTURE output and implementing the Evanno method. *Conservation Genetics Resources* **4(2)**: 359-
457 361.

458 **Ellstrand NC, Roose ML. 1987.** Patterns of genotypic diversity in clonal plant species. *American Journal*
459 *of Botany* **74(1)**: 123-131.

460 **Ellstrand NC, Elam DR. 1993.** Population genetic consequences of small population size: Implications

461 for plant conservation. *Annual Review of Ecology Evolution and Systematics* **24**: 217-242.

462 **Escudero A, Iriondo JM, Torres ME. 2003.** Spatial analysis of genetic diversity as a tool for plant
463 conservation. *Biological Conservation* **113(3)**: 351-365.

464 **Evanno G, Regnaut S, Goudet J. 2005.** Detecting the number of clusters of individuals using the software
465 STRUCTURE: a simulation study. *Molecular ecology* **14(8)**: 2611-2620.

466 **Fahrig L. 2003.** Effects of Habitat Fragmentation on Biodiversity. *Annual Review of Ecology, Evolution,*
467 *and Systematics* **34(1)**: 487-515.

468 **Falk DA, Richards CM, Montalvo AM, Knapp EE. 2006.** Population and ecological genetics in
469 restoration ecology. In Palmer MA, Zedler JB, Falk DA, eds. *Foundations of Restoration Ecology*.
470 Washington, D.C: Island press, 14-41.

471 **Frankham R. 1996.** Relationship of genetic variation to population size in wildlife. *Conservation Biology*
472 **10(6)**: 1500-1508.

473 **Frankham R. 2005.** Stress and adaptation in conservation genetics. *Journal of Evolutionary Biology*
474 **18(4)**: 750-755.

475 **Gaudeul M, Till-Bottraud I. 2008.** Genetic structure of the endangered perennial plant *Eryngium*
476 *alpinum* (Apiaceae) in an alpine valley. *Biological Journal of the Linnean Society* **93(4)**: 667-677.

477 **Gillet F. 2000.** La phytosociologie synusiale intégrée: guide méthodologique. *Technical report,*
478 *Laboratoire d'écologie végétale et de phytosociologie.* Neuchâtel University, France.

479 **Gitzendanner M, Soltis P. 2000.** Patterns of genetic variation in rare and widespread plant congeners.
480 *American Journal of Botany* **87(6)**: 783-792.

481 **Glemarec E, Magnanon S, Rasclé P, Ramil-Rego P, Rodríguez Guitián MA, Ferreiro Da Costa J, Bioret**
482 **F, Gallet S. 2017.** La phytosociologie, un outil d'aide à la conservation d'*Eryngium viviparum*, J.Gay.
483 *Documents phytosociologiques série 3, in press.*

484 **Godefroid S, Vanderborght T. 2011.** Plant reintroductions: the need for a global database. *Biodiversity*
485 *and Conservation* **20(14)**: 3683-3688.

486 **Gower JC. 1971.** A general coefficient of similarity and some of its properties. *Biometrics* **27(4)**: 857-

487 871.

488 **Guillot G, Rousset F. 2013.** Dismantling the Mantel tests. *Methods in Ecology and Evolution* **4(4)**: 336-
489 344.

490 **Haig SM. 1998.** Molecular contributions to conservation. *Ecology* **79(2)**: 413-425.

491 **Hamrick JL, Godt MJW. 1989.** Allozyme diversity in plant species. In Brown AHD, Clegg MT, Kahler AL,
492 Weir BS, eds. *Plant population genetics, breeding and genetic resources*. Sunderland, MA: Sinauer, 43-
493 64.

494 **Hamrick JL, Godt MJW. 1996.** Conservation genetics of endemic plant species. In Avise JC, Hamrick JL,
495 eds. *Conservation Genetics: Case Histories from Nature*. New York: Chapman & Hall, 281-304.

496 **Havens K. 1998.** The genetics of plant restoration. *Restoration and Management Notes* **16(1)**: 68-72.

497 **Higgins K, Lynch M. 2001.** Metapopulation extinction caused by mutation accumulation. *Proceedings*
498 *of the National Academy of Sciences* **98(5)**: 2928-2933.

499 **Hijmans RJ, Cameron SE, Parra JL, Jones PG, Jarvis A. 2005.** The WorldClim interpolated global
500 terrestrial climate surfaces. Ver. 1.3. <http://www.worldclim.org>.

501 **Holsinger KE, Gottlieb LD. 1991.** Conservation of rare and endangered plants: principles and prospects.
502 In Falk DA, Holsinger KE, eds. *Genetics and conservation of rare plants*. New York: Oxford University
503 Press, 195-208.

504 **Honnay O, Bossuyt B. 2005.** Prolonged clonal growth: Escape route or route to extinction? *Oikos*
505 **108(2)**: 427-432.

506 **Honnay O, Jacquemyn H. 2007.** Susceptibility of common and rare plant species to the genetic
507 consequences of habitat fragmentation. *Conservation biology* **21(3)**: 823-31.

508 **Honnay O, Coart E, Butaye J, Adriaens D, Van Glabeke S, Roldán-Ruiz I. 2006.** Low impact of present
509 and historical landscape configuration on the genetics of fragmented *Anthyllis vulneraria* populations.
510 *Biological Conservation* **127(4)**: 411-419.

511 **Hueneke LF. 1991.** Ecological implications of genetic variation in plant populations. In Falk DA,
512 Holsinger KE, eds. *Genetics and conservation of rare plants*. New York: Oxford University, 31-44.

513 **Jombart T. 2008.** Adegnet: a R package for the multivariate analysis of genetic markers.
514 *Bioinformatics* 24(11): 1403-1405.

515 **Jovet P. 1939.** Notes sur quelques plantes de Carnac (Morbihan). *Bulletin de la société française*
516 *d'échanges de plantes*: 1-6.

517 **Keller LF, Waller DM. 2002.** Inbreeding effects in wild populations. *Trends in Ecology and Evolution*
518 **17(5)**: 230-241.

519 **Lande R, Barrowclough G. 1987.** Effective population size, genetic variation, and their use in
520 population management. In Soulé ME, eds. *Viable populations for conservation*. Cambridge University
521 press, 87-124.

522 **Lawrence BA, Kaye TN. 2011.** Reintroduction of *Castilleja levisecta*: effects of ecological similarity,
523 source population genetics, and habitat quality. *Restoration Ecology* **19(2)**: 166-176.

524 **Leblois R, Pudlo P, Néron J, Bertaux F, Reddy Beeravolu C, Vitalis R, Rousset F. 2014.** Maximum-
525 likelihood inference of population size contractions from microsatellite data. *Molecular Biology and*
526 *Evolution* **31(10)**: 2805-2823.

527 **Leimu R, Mutikainen P, Koricheva J, Fischer M. 2006.** How general are positive relationships between
528 plant population size, fitness and genetic variation? *Journal of Ecology* **94(5)**: 942-952.

529 **Lowe AJ, Boshier D, Ward M, Bacles CFE, Navarro C. 2005.** Genetic resource impacts of habitat loss
530 and degradation; reconciling empirical evidence and predicted theory for neotropical trees. *Heredity*
531 **95(4)**: 255-273.

532 **Lynch M, Conery J, Burger R. 1995.** Mutation accumulation and the extinction of small populations.
533 *The American Naturalist* **146(4)**: 489-518.

534 **Magnanon S, Hardegen M, Guillevic Y. 2013.** *Plan national d'actions en faveur du panicaut vivipare,*
535 *Eryngium viviparum J. Gay. 2012-2017.* Paris.

536 **Maguire TL, Saenger P, Baverstock P, Henry R. 2000.** Microsatellite analysis of genetic structure in the
537 mangrove species *Avicennia marina* (Forsk.) Vierh. (Avicenniaceae). *Molecular Ecology* **9(11)**: 1853-
538 1862.

539 **Malausa T, Gilles A, Megléc E, Blanquart H, Duthoy S, Costedoat C, Dubut V, Pech N, Castagnone-**
540 **Sereno P, Délye C, Feau N, Frey P, Gauthier P, Guillemaud T, Hazard L, Le Corre V, Lung-Escarmant B,**
541 **Malé PJG, Ferreira S, Martin J.F. 2011.** High-throughput microsatellite isolation through 454 GS-FLX
542 Titanium pyrosequencing of enriched DNA libraries. *Molecular Ecology Resources* **11(4)**: 638-644.

543 **Maschinski J, Wright SJ, Koptur S, Pinto-Torres E.C. 2013.** When is local the best paradigm? Breeding
544 history influences conservation reintroduction survival and population trajectories in times of extreme
545 climate events. *Biological Conservation* **159**: 277-284.

546 **Maurice AC, Abdelkrim J, Cisel M, Zavodna M, Bardin P, Matamoro A, Dumez R, Machon N. 2013.**
547 Mixing plants from different origins to restore a declining population: ecological outcomes and local
548 perceptions 10 years later. *PLoS ONE* **8(1)**: e50934.

549 **McGarigal K, Cushman S.A. 2002.** Comparative evaluation of experimental approaches to the study of
550 habitat fragmentation effects. *Ecological Applications* **12(2)**: 335-345.

551 **McKay JK, Christian CE, Harrison S, Rice KJ. 2005.** 'How local is local?' - A review of practical and
552 conceptual issues in the genetics of restoration. *Restoration Ecology* **13(3)**: 432-440.

553 **Megléc E, Costedoat C, Dubut V, Gilles A, Malausa T, Pech N, Martin JF. 2009.** QDD: A user-friendly
554 program to select microsatellite markers and design primers from large sequencing projects.
555 *Bioinformatics* **26(3)**: 403-404.

556 **Meloni M, Reid A, Caujapé-Castells J, Marrero Á, Fernández-Palacios JM, Mesa-Coelo RA, Conti E.**
557 **2013.** Effects of clonality on the genetic variability of rare, insular species: The case of *Ruta microcarpa*
558 from the Canary Islands. *Ecology and Evolution* **3(6)**: 1569-1579.

559 **Mistretta O. 1994.** Genetics of species re-introductions: applications of genetic analysis. *Biodiversity*
560 *and Conservation* **3(2)**: 184-190.

561 **Montalvo AM, Williams SL, Rice KJ, Buchmann SL, Cory C, Handel SN, Nabhan GP, Primack R,**
562 **Robichaux R.H. 1997.** Restoration biology: a population biology perspective. *Restoration Ecology* **5(4)**:
563 277-290.

564 **Neale JR. 2012.** Genetic considerations in rare plant reintroduction: practical applications (or how are

565 we doing?). In Maschinski J, Haskins KE, eds. *Plant reintroduction in a changing climate*. Washington
566 DC: Island Press, 71-88.

567 **Nei M, Tajima F, Tateno Y. 1983.** Accuracy of estimated phylogenetic trees from molecular data.
568 *Journal of molecular evolution* **19(2)**: 153-170.

569 **Newman D, Tallmon DA. 2001.** Experimental evidence for beneficial fitness effects of gene flow in
570 recently isolated populations. *Conservation Biology* **15(4)**: 1054-1063.

571 **Oksanen J, Blanchet FG., Kindt R, Legendre P, Minchin PR, O'Hara RB, Simpson GL, Solymos P, Henry**
572 **M, Stevens H, Wagner H. 2016.** Vegan: community ecology package. *R package version 2.4-1*.
573 <https://CRAN.R-project.org/package=vegan>.

574 **Olivier L. 1995.** *Livre rouge de la flore menacée de France. Tome 1 : espèces prioritaires*. Paris: Muséum
575 National d'Histoire Naturelle.

576 **Ouborg NJ, Vergeer P, Mix C. 2006.** The rough edges of the conservation genetics paradigm for plants.
577 *Journal of Ecology* **94(6)**: 1233-1248.

578 **Petit RJ, Mousadik AEL, Pons O. 1998.** Identifying populations for conservation on the basis of genetic
579 markers. *Conservation Biology* **12(4)**: 844-855.

580 **Pironon S, Papuga G, Villellas J, Angert AL, García MB, Thompson JD. 2016.** Geographic variation in
581 genetic and demographic performance: new insights from an old biogeographical paradigm. *Biological*
582 *Reviews* **92(4)**: 1877-1909.

583 **Pritchard J, Stephens M, Donnelly P. 2000.** Inference of population structure using multilocus
584 genotype data. *Genetics* **155(2)**: 945-959.

585 **Prober S, Brown AHD. 1994.** Conservation of the grassy white box woodlands: population genetics and
586 fragmentation of *Eucalyptus albens*. *Conservation Biology* **8(4)**: 1003-1013.

587 **R Core Team. 2016.** *R: A language and environment for statistical computing*. Vienna Austria: R
588 Foundation for Statistical Computing. Retrieved from <https://www.R-project.org/>.

589 **Rasclé P, Bioret F, Magnanon S, Glemarec E, Gautier C, Guillevic Y, Gallet S. 2018.** Identification of
590 success factors for the reintroduction of the critically endangered species *Eryngium viviparum* J. Gay

591 (Apiaceae). *Ecological Engineering* **122**: 112-119.

592 **Reed DH. 2005.** Relationship between population size and fitness. *Conservation biology* **19(2)**: 563-
593 568.

594 **Reed DH, Frankham R. 2003.** Correlation between fitness and genetic diversity. *Conservation Biology*
595 **17(1)**: 230-237.

596 **Rice WR. 1989.** Analyzing tables of statistical tests. *Evolution* **43(1)**: 223.

597 **Richards CM. 2000.** Inbreeding depression and genetic rescue in a plant metapopulation. *The American*
598 *Naturalist* **155(3)**: 383-394.

599 **Rieseberg LH, Swensen SM. 1996.** Conservation genetics of endangered island plants. In Avise AC,
600 Hamrick JL, eds. *Conservation genetics: case histories from nature*. New York: Chapman & Hall, 305-
601 345.

602 **Rivas-Martínez S, Rivas-Sáenz S, Penas-Merino A. 2011.** Worldwide bioclimatic classification system.
603 *Global Geobotany* **1(1)**: 1-638.

604 **Rodríguez-Gacio MC, De Jesús J, Romero MI, Herrera MT. 2009.** Genetic diversity among genotypes
605 of *Eryngium viviparum* (Apiaceae): A plant threatened throughout its natural range. *Botanical Journal*
606 *of the Linnean Society* **159(2)**: 237-244.

607 **Romero MI, Ramil P, Rubinos M. 2004.** Conservation status of *Eryngium viviparum* Gay. *Acta Botanica*
608 *Gallica* **151(1)**: 55-64.

609 **Rousset F. 2008.** GENEPOP'007: A complete re-implementation of the GENEPOP software for Windows
610 and Linux. *Molecular Ecology Resources* **8(1)**: 103-106.

611 **Sala OE, Chapin FS, Armesto JJ, Berlow E, Bloomfield J, Dirzo R, Huber-Sandwald E, Huenneke LF,**
612 **Jackson RB, Kinzig A, Leemans R, Lodge DM, Mooney HA, Oesterheld M, Leroy Poff N, Sykes MT,**
613 **Walker BH, Walker M, Wall DH. 2000.** Global biodiversity scenarios for the year 2100. *Science*
614 **287(5459)**: 1770-1774.

615 **Saunders DA, Hobbs RJ, Margules CR. 1991.** Biological consequences of ecosystem fragmentation: a
616 review. *Conservation Biology* **5(1)**: 18-32.

617 **Spielman D, Brook B, Frankham R. 2004.** Most species are not driven to extinction before genetic
618 factors impact them. *Proceedings of the National Academy of Sciences* **101(42)**: 15261-15264.

619 **Tamaki I, Setsuko S, Tomaru N. 2016.** Genetic diversity and structure of remnant *Magnolia stellata*
620 populations affected by anthropogenic pressures and a conservation strategy for maintaining their
621 current genetic diversity. *Conservation genetics* **17(3)**: 715-725.

622 **Trabucco A, Zomer RJ. 2009.** Global aridity index (global-aridity) and global potential evapo-
623 transpiration (global-PET) geospatial database. *CGIAR Consortium for Spatial Information, CGIAR-CSI*
624 *GeoPortal*.

625 **Vallejo-Marín M, Dorken ME, Barrett SCH. 2010.** The ecological and evolutionary consequences of
626 clonality for plant mating. *Annual Review of Ecology, Evolution, and Systematics* **41(1)**: 193-213.

627 **Van Oosterhout C, Hutchinson WF, Wills DPM, Shipley P. 2004.** MICRO-CHECKER: software for
628 identifying and correcting genotyping errors in microsatellite data. *Molecular Ecology Resources* **4(3)**:
629 535-538.

630 **Vergeer P, Van Den Berg LJJ, Roelofs JGM, Ouborg NJ. 2005.** Single-family versus multi-family
631 introductions. *Plant Biology* **7(5)**: 509-515.

632 **Watkinson AR, Powell JC. 1993.** Seedling recruitment and the maintenance of clonal diversity in plant
633 populations - A computer simulation of *Ranunculus Repens*. *Journal of Ecology* **81(4)**: 707-717.

634 **Weeks AR, Sgro CM, Young AG, Frankham R, Mitchell NJ, Miller KA, Byrne M, Coates DJ, Eldridge**
635 **MDB, Sunnucks P, Breed MF, James EA, Hoffmann AA. 2011.** Assessing the benefits and risks of
636 translocations in changing environments: a genetic perspective. *Evolutionary Applications* **4(6)**: 709-
637 725.

638 **Wiberg RAW, Scobie AR, A'Hara SW, Ennos RA, Cottrell JE. 2016.** The genetic consequences of long
639 term habitat fragmentation on a self-incompatible clonal plant, *Linnaea borealis* L. *Biological*
640 *Conservation* **201**: 405-413.

641 **Wilcox BA, Murphy DD. 1985.** Conservation strategy: The effects of fragmentation on extinction. *The*
642 *American Naturalist* **125(6)**: 879-887.

643 **Yardeni G, Tessler N, Imbert E, Sapir Y. 2016.** Reproductive isolation between populations of *Iris*
644 *atropurpurea* is associated with ecological differentiation. *Annals of Botany* **118(5)**: 971-982.

645 **Young A, Boyle T, Brown T. 1996.** The population genetic consequences of habitat fragmentation for
646 plants. *Trends in Ecology & Evolution* **11(10)**: 413-418.

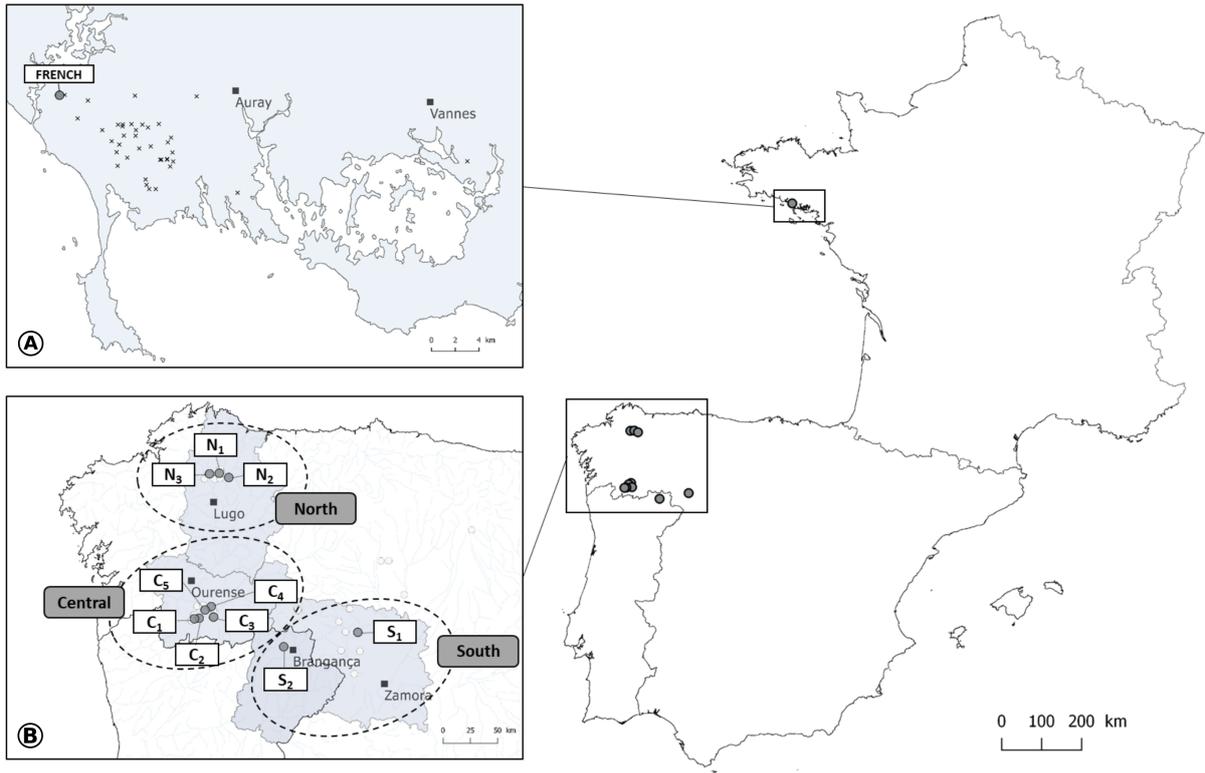
647 **Young AG, Clarke GM. 2000.** Genetics, demography and viability of fragmented populations. Vol. 4.
648 Cambridge: Cambridge University Press.

649 **Young AG, Hill JH, Murray BG, Peakall R. 2002.** Breeding system, genetic diversity and clonal structure
650 in the sub-alpine forb *Rutidosia leiolepis* F. Muell. (Asteraceae). *Biological Conservation* **106(1)**: 71-78.

651 **Zavodna M, Abdelkrim J, Pellissier V, Machon N. 2015.** A long-term genetic study reveals complex
652 population dynamics of multiple-source plant reintroductions. *Biological Conservation* **192**: 1-9.

653 **Zenboudji S, Cheylan M, Arnal V, Bertolero A, Leblois R, Astruc G, Bertorelle G, Pretus JL, Lo Valvo**
654 **M, Sotgiu G, Montgelard C. 2016.** Conservation of the endangered Mediterranean tortoise *Testudo*
655 *hermanni hermanni*: The contribution of population genetics and historical demography. *Biological*
656 *Conservation* **195**: 279-291.

657

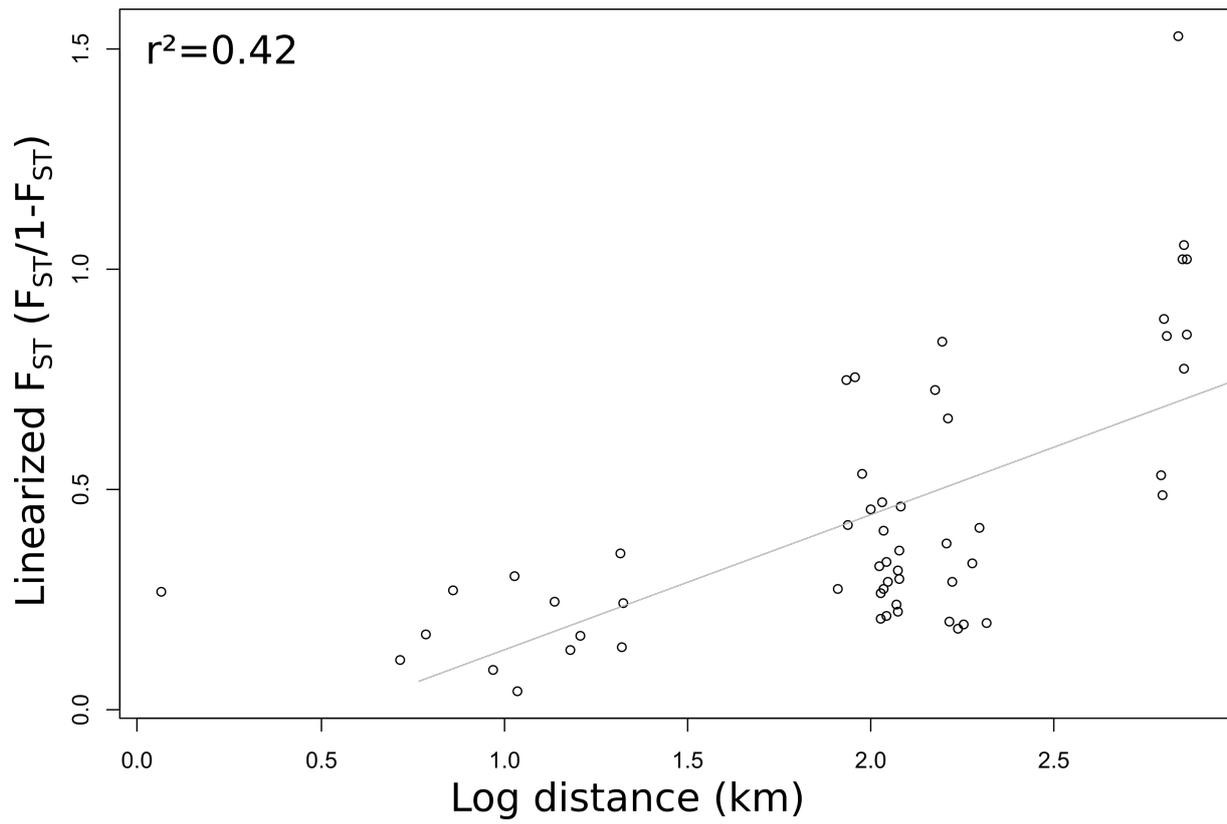


658

659 **Fig. 1.** Global distribution and location of the 11 sampled populations of *E. viviparum*. A. Actual and
 660 historic distribution in France. B. Actual distribution in the Iberian Peninsula. Dotted line ellipses
 661 delimit the three population geographical clusters.

662

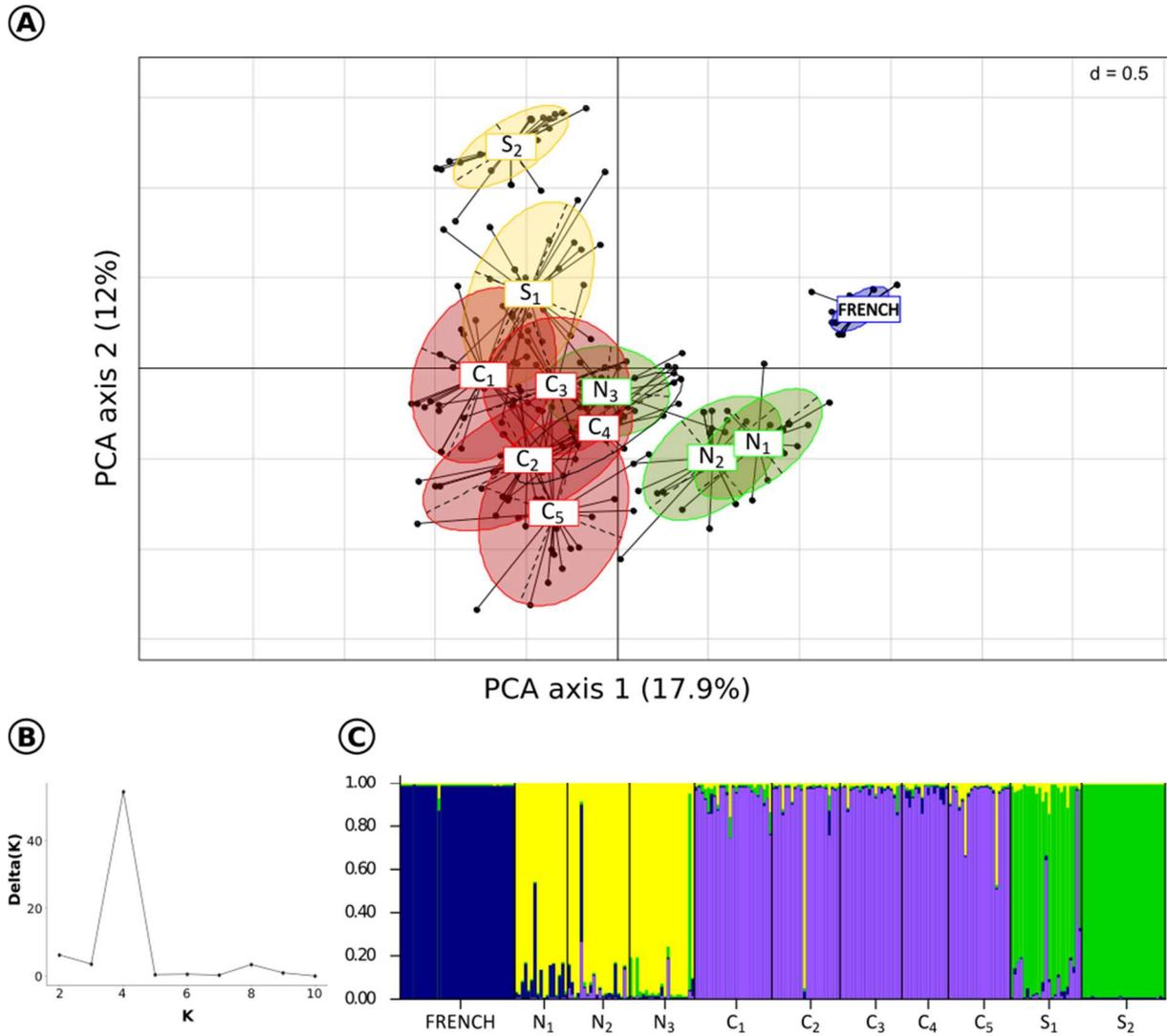
663



664

665 **Fig. 2.** Correlation between F_{ST} ($F_{ST}/1-F_{ST}$) and $\log(\text{geographical distance, in km})$ for pairwise
666 comparisons of the ten *E. viviparum* Iberian populations. r^2 indicates the coefficient of correlation from
667 the Mantel test.

668



669

670 **Fig. 3.** Population structure analyses. a. First plan of the principal components analysis (PCA) of
 671 *Eryngium viviparum*'s genetic structure among eleven populations and based on seven microsatellites
 672 multilocus genotypes. Each point represent the individual genotypes and are connected by lines to the
 673 centroid of the 95% confidence interval ellipse of each population. b. Delta(K) values from K=1 to 11.
 674 c. Average assignment probability of *Eryngium viviparum* individuals for K=4.

Table 1. Population location, sample size and estimated size (see text for details), distance to the nearest population and ecological characteristics. The four climatic data (mean annual temperature= T_M , maximum annual temperature= T_{MAX} , minimum annual temperature= T_{MIN} and annual precipitation=PP) were extracted from Worldclim 2 (Hijmans *et al.*, 2005) based on 30-years average values (from 1970 to 2000). The number of co-occurring species was extracted from Glemarec *et al.* (2017). The populations N₃, C₂ and C₅ were not included in this study. Exact GPS coordinates are not indicated considering the rarity of *Eryngium viviparum*. NA notifies data are not available.

Location	Herbarium voucher	ID population	Sampled size	Population size category	Distance to the nearest existant population (km)	Elevation (m)	Climatic values from Worldclim v.2				N° of co-occurring species	
							T _M (°C)	T _{MAX} (°C)	T _{MIN} (°C)	PP (mm)		
France												
Morbihan	<i>No voucher</i>											
Belz		FRENCH	37	3	516	11	12.1	21.1	3.5	893	20	
NORTH (Spain, Galicia, Lugo) :												
Lagoa de Cospeito	<i>J. Amigo, P. Ramil, M. Rodriguez & J. Izco</i> 39497 [SANT 38490] [LUGO 773]	N ₁	17	2	2.9	457	12.2	22.5	1.6	1357	15	
Bexan	[LUGO 773]	N ₂	20	3	8	401	12.4	22.4	1.5	1329	13	
Fontefria	[LUGO 772]	N ₃	21	2	6	413	12.4	22.4	1.9	1382	NA	
CENTRAL (Spain, Galicia, Ourense) :												
O Toxal	<i>I. Pulgar</i> [SANT 45414]	C ₁	25	4	2.4	613	12.2	23.1	0.9	1602	12	
O Foxos		C ₂	22	5	2.7	616	12.5	24.3	0.4	1638	NA	
Vilaseca	<i>I. Pulgar</i> [SANT 45969]	C ₃	20	5	0.6	617	12.5	24.5	0.4	1718	19	
Veiga de Gomareite		C ₄	15	5	0.7	618	12.4	24.4	0.4	1719	17	
Cardeita	<i>I. Pulgar</i> [SANT 45413]	C ₅	20	4	4.6	618	12.5	24.4	0.2	1698	NA	
SOUTH :												
Ferreira de Abajos (Spain, Castilla y León, Zamora)	<i>P. Bariego</i> 2480 [SANT 60815]	S ₁	23	3	1.3	804	11.6	24.8	-2.8	750	19	
Tras-o-Montes (Portugal, Bragança)	<i>No voucher</i>	S ₂	27	1	33.3	963	11	23.2	-1.9	943	7	

Table 2. Characteristics of the seven microsatellites primers used for genotyping *Eryngium viviparum* populations. T_a: annealing temperature. N_A: average number of alleles. H_{OBS}: observed heterozygosity. H_{EXP}: expected heterozygosity. F_{IS}: intrapopulation fixation index. F_{IS} values statistically different from zero, at the 0.05 level, appeared in bold.

Locus	GenBank accession number	Repeat arrays	Primer sequences (5'-3')	T _a (F/R)	Size range (bp)	Global diversity indices among all populations			
						N _A	H _{OBS}	H _{EXP}	F _{IS}
pmEv01	MK319936	ACA ₂₂	F : AGTATTACTTCTGCCTTTAATATTTTCG R : CATGATTAATTAGATGCTTGAAGATG	60.7/60	219 -295	5	0.19	0.24	-0.0517
pmEv02	MK319937	GT ₂₁	F : TTAGTGTCGCAATGAGCAGC R : GCACCGTTTCCTGTTGGTAT	58.4	75-125	9	0.4	0.52	0.0707
pmEv04	MK319938	GA ₂₁	F : TTGGTGAGGGTTTCGATTTG R : TCACCTCGATTCTTGTGCAT	56.4	124-170	19	0.72	0.91	0.0206
pmEv05	MK319939	GT ₂₀	F : CGCAAGAAATTGCTCCATA R : TGTTGCCAATATGACAGTAACG	56.4/58.4	108-160	11	0.5	0.85	0.1195*
pmEv09	MK319940	ATGT ₁₇	F : CCCACGATTGATCTGCATAG R : TCAGAGGATGTCTCCACAA	58.4	250-340	15	0.43	0.77	0.0455
pmEv10	MK319941	CA ₁₇	F : GTTATGTCACACTTCATGCTGC R : TGCTTCTGCTGTTATCCTCA	60.3	149-178	11	0.52	0.79	-0.0132
pmEv17	MK319942	AC ₁₆	F : ATAAGAGGGGGAAAAGGTGG R : TTAATTGTGTATTCAATGAACCTTCC	58.4	213-227	7	0.58	0.72	-0.0526

Table 3. Genetic diversity (number of alleles per locus= N_A , observed heterozygosity= H_{OBS} , unbiased expected heterozygosity= H_{EXP} , fixation index= F_{IS}) within each population of *Eryngium viviparum* and Migraine outputs of past demographic analyses expressed by N ratio (current effective population size = N_{act} /ancestral population size = N_{anc} , the 95% confidence intervals are given into brackets). Populations with an N ratio significantly different from 1 are in bold. F_{IS} values statistically different from zero, at the 0.05 level, are in bold.

Population	N_A	H_{OBS}	H_{EXP}	F_{IS}	No. of multilocus genotypes/ Sample size	No. of private alleles	N ratio	Demographic event
FRENCH	1.57	0.15	0.18	0.17*	16/37	0	2.2e-11 [1.64e-11 -- 5.46e-06]	bottleneck
N ₁	4.14	0.56	0.53	-0.03	15/17	0	3.69e-05 [2.66e-05 -- 72.18]	stable
N ₂	4.86	0.56	0.59	0.07	18/20	0	8.43 e-04 [9.88e-05 - 0.182]	bottleneck
N ₃	5.28	0.53	0.56	0.08	15/21	0	0.0771 [1.02e-05 -- 3.454]	stable
C ₁	5.71	0.57	0.56	-0.006	22/25	2	2.6e-11 [10e-12 – 2.9e-9]	bottleneck
C ₂	6.00	0.69	0.63	-0.07	21/22	2	1.092 [6.22e-05 – 1.955]	stable
C ₃	6.14	0.69	0.67	-0.005	19/20	4	0.00477 [0.000132 -- 0.816]	bottleneck
C ₄	4.00	0.48	0.50	0.005	15/15	1	0.01 [0.003 -- 0.053]	bottleneck
C ₅	4.00	0.55	0.55	0.02	20/20	0	5.88e-03 [8.07e-05 -- 0.142]	bottleneck
S ₁	5.00	0.59	0.57	-0.02	21/23	2	1.54e-19 [1.65e-20 -- 9.06e-11]	bottleneck
S ₂	2.86	0.30	0.32	0.08	25/27	3	7.14e-05 [3.16e-05 – 0.132]	bottleneck

Table 4. Pairwise F_{ST} values for seven microsatellites markers between the 11 *Eryngium viviparum* populations studied. All values are significantly different from zero at $P < 0.05$.

	FRENCH	N1	N2	N3	C1	C2	C3	C4	C5	S1
N1	0.3273									
N2	0.3476	0.0393								
N3	0.4695	0.2317	0.1946							
C1	0.5055	0.3154	0.2647	0.2393						
C2	0.4597	0.2279	0.1825	0.1925	0.1018					
C3	0.4365	0.2148	0.1696	0.2097	0.1238	0.1184				
C4	0.5053	0.2883	0.2451	0.3204	0.2622	0.1438	0.2105			
C5	0.5127	0.2254	0.1745	0.2504	0.1966	0.0822	0.1466	0.213		
S1	0.4582	0.2917	0.249	0.165	0.1627	0.1553	0.1666	0.2744	0.2241	
S2	0.6045	0.4546	0.4204	0.3973	0.3123	0.3481	0.2944	0.4274	0.4298	0.2154

<i>Polypogon viridis</i> (Gouan) Breistr.	Poaceae	1
<i>Pulicaria paludosa</i> Link	Asteraceae	1
