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Bisphenol A in eggs impairs the long-term stress performance of rainbow trout in two generations

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1 **ABSTRACT**

2 Salmonids are ecologically, economically and culturally important fish species in North
3 America, but whether contaminants in the environment may play a role in their population
4 decline is unclear. We tested the hypothesis that BPA deposition in eggs, mimicking a maternal
5 transfer scenario, compromises the stress axis functioning and target tissues stress response in
6 two generations of a model salmonid species, the rainbow trout (*Oncorhynchus mykiss*). Eggs
7 were enriched with 0, 4 or 40 ng BPA, fertilized, and reared in clean water for two generations.
8 The fish were subjected to an acute stressor after a year in both generations to test their stress
9 performances. Trout raised from BPA-enriched eggs showed impaired stressor-mediated plasma
10 cortisol and lactate response in the F1 and F2 generation, respectively. Key genes involved in
11 cortisol biosynthesis in the head kidney, as well as stress- and growth-related transcripts in the
12 liver and muscle were impacted either in the F1 and/or F2 generations. Our results underscore
13 the long-term impact associated with BPA in eggs, mimicking a maternal transfer scenario, on
14 the stress performance of trout in two generations. The results highlight the need for developing
15 novel biomarkers to predict long-term and generational toxicities in salmonids.

16

17

18 **KEYWORDS:** Salmonids, BPA, Cortisol, Stress response, Gene expression, Transcriptomics

19 INTRODUCTION

20 Bisphenol A (BPA), an organic compound used in the production of plastics and epoxy
21 resins, is ubiquitously distributed in the aquatic environment with mounting evidence of its
22 impact on the endocrine system of animals ^{1,2}. Global production of BPA has increased
23 substantially over the years, and over 500 tons of BPA are released into the environment
24 annually ². A large body of work has provided insight into the toxicities of BPA in both aquatic
25 and terrestrial animals ^{1,3}. In addition, maternal transfer of BPA has been reported in humans,
26 rodents and fish ³⁻⁶, but the long-term developmental effects are far from clear. Recent studies
27 have also described that exposure to BPA during critical early developmental periods may lead
28 to stable epigenetic modifications that are passed on to the next generation ^{7,8}.

29 As in mammals, BPA is an estrogen mimic in fishes and impacts reproduction ⁹⁻¹¹.
30 Recently studies also highlight developmental toxicities related to growth and stress response
31 activation in fish ¹²⁻¹⁴. Furthermore, multigenerational impact of BPA on reproduction was
32 shown in a model small-bodied fish with short life spans and generation times ⁹; however, no
33 information currently exists on multigenerational impacts of BPA in ecologically relevant fish
34 species with longer life spans. Indeed, the potential for chemicals to cause adverse effects that
35 persists in multiple generations are of concern, as it highlights the profound and sustained
36 environmental health dysfunction ^{15,16}, especially when observed in ecologically-relevant
37 species.

38 The physiological response to stressors is highly conserved among vertebrates ^{17,18}, as an
39 evolutionary consequence of its crucial role in animals fitness. Any perturbations in the cortisol
40 stress response, as seen with contaminant exposure ¹⁹, may negatively impact growth and
41 development ^{20,21} and survival of the animal ^{22,23}. In anadromous salmonids, stress axis function

42 and its ability to respond to an acute stressor are also considered a good determinant of
43 reproductive outcome and progenies fitness^{23,24}. In addition, studies have shown that cortisol
44 plays a significant role in the upstream migration of salmonids to their spawning grounds^{25,26}.
45 Therefore, ability of fish to display a normal stress axis activity provides a good marker of global
46 health of the species in a given environment¹⁹.

47 Rainbow trout (*Oncorhynchus mykiss*) is considered an excellent model for toxicological
48 studies^{27,28}, and a model salmonid given its genotypic resemblance with other migratory
49 salmonids^{29,30}. Our companion studies in trout recently showed that BPA in eggs, mimicking
50 maternal transfer of this contaminant, affects the ontogeny of growth and stress response in the
51 F1 generation^{13,14}. Also, we showed changes in growth and metabolism during development in
52 the two generation of trout raised from BPA-enriched eggs²¹. However, we have not shown
53 before whether the stress performance, a key aspect of animal fitness, is impacted in multiple
54 generations by BPA exposure in this species. Against this backdrop, we tested the hypothesis
55 that BPA deposition in trout eggs, mimicking a maternal transfer scenario, compromises the
56 long-term stress performance of the progeny in two generations. Plasma cortisol response to an
57 acute stressor, the head kidney capacity to produce cortisol in response to adrenocorticotrophic
58 hormone (ACTH) stimulation *in vitro*, and the transcript abundance of corticosteroidogenic
59 genes in the head kidney were used as markers of stress axis activity, while plasma glucose,
60 lactate levels and tissue glycogen content were measured as indicators of metabolic stress
61 response^{17,31}. In addition, changes in transcript abundances of stress- and growth-related genes,
62 and epigenetic markers in the liver and muscle in response to acute stressor exposure were used
63 as biomarkers of target tissue responses.

64

65 MATERIALS AND METHODS

66 Experimental Animals and Treatments

67 The experimental details, including BPA exposure, fish maintenance and breeding of fish to
68 obtain F1 and F2 generations have been published already ^{14,21}. Briefly, pooled oocytes from four
69 females were fertilized with pooled milt from four male rainbow trout (3+ year class brood
70 stock). Ovarian fluid was also collected from these four females for BPA treatment. Pooled
71 oocytes were immersed in 50 ml of ovarian fluid containing vehicle alone (<0.01% ethanol;
72 control group) or vehicle containing BPA at 3 or 30 $\mu\text{g ml}^{-1}$ for 3 h at 6-8 °C with gentle shaking
73 every 30 min. After the exposure, the oocytes were mixed with 1-2 ml of milt for fertilization,
74 after which the embryos were rinsed several times with clean water. This treatment resulted in
75 an egg BPA content of 4 and 40 ng egg^{-1} in the 3 and 30 $\mu\text{g ml}^{-1}$ exposure groups, respectively ¹⁴.
76 The embryos were maintained in a Heath chamber incubator receiving clean groundwater at a
77 rate of 10 l min^{-1} (6-8 °C). Larvae were maintained in the incubator for a week after hatch, after
78 which they were moved to holding tanks (3 × 200 l tanks per treatment; n=277-299 larvae per
79 replicate) receiving flow-through water at a rate of 10 l min^{-1} , under a 12h L: 12h D photoperiod.
80 At 1 year, fish were sampled before and after an acute stress challenge (see below).

81 To study the BPA effects in the second generation, oocytes from the F1 generation adult
82 female rainbow trout, kept separate based on the F0 egg BPA concentration, were fertilized with
83 pooled milt from a stock of unexposed male rainbow trout. There was no detectable BPA in the
84 eggs of any treatment groups. The experimental condition and fish rearing was similar to the F1
85 generation trout, except only one tank per treatment was maintained for the F2 generation. At 1
86 year, trout in the F2 generation were also sampled before and after an acute stress challenge (see
87 below). Experiments were conducted at the Alma Research Station (ARS) (Alma, ON, Canada),

88 and the experimental procedures were approved by the Animal Care and Use Committees at the
89 University of Guelph and the University of Waterloo, and adhered to the Canadian Council on
90 Animal Care guidelines for humane animal use.

91 **Stress Sampling**

92 Trout (82.2 ± 5.0 g) from F1 and F2 generations were sampled 365 days post fertilization
93 (dpf) to investigate the effects of BPA in eggs on long-term stress performance in trout. We
94 examined primary and secondary stress response in control and trout raised from BPA
95 accumulated eggs after an acute stress challenge. The stressor consisted of a 3 min handling
96 disturbance, which elicited a transient rise in plasma cortisol levels, as described previously¹².
97 Food was withheld 48 h prior to the commencement of the stress experiment. Fish were sampled
98 either prior to the stressor protocol (0 h time-point) or at 1, 4 and 24 h post-stressor exposure.
99 Fish were euthanized with buffered Tricaine methanesulfonate (MS-222) and blood was
100 collected by caudal severance in tubes containing EDTA as the anticoagulant. Blood samples
101 were centrifuged at 5000 x g for 5 min to separate plasma and stored at -80 °C for later analysis
102 of cortisol, glucose and lactate levels. Tissues (head kidney, liver and muscle) were quickly
103 excised, flash frozen in dry ice, and stored at -80 °C until transcript analysis. We measured the
104 physiological markers of stress response (plasma cortisol, glucose and lactate levels), along with
105 the molecular markers of stress response in the liver and muscle (glucocorticoid receptor 1
106 [*gr1*], glucocorticoid receptor 2 [*gr2*], and mineralocorticoid receptor [*mr*]) and head kidney
107 (genes related to corticosteroid biosynthesis: melanocortin 2 receptor (*mc2r*), cytochrome P450
108 side-chain cleavage (*p450scc*) and steroidogenic acute regulatory protein (*star*) in the two
109 generations of trout. Also, molecular markers of growth (insulin-like growth factor-1 [*igf1*],
110 insulin-like growth factor-2 [*igf2*], insulin-like growth factor 1a receptor [*igf1ra*], insulin-like

111 growth factor 1b receptor [*igf1rb*], growth hormone receptor 1 [*gh1r*], growth hormone receptor
112 2 [*gh2r*) and epigenetics (DNA methyltransferase 1 [*dnmt1*], DNA methyltransferase 2 [*dnmt2*)
113 and liver specific methionine adenosyltransferase 1 alpha [*mat1a*]) were measured in the liver
114 and muscle in the two generations of trout before and after an acute handling stress challenge.

115 **In Vitro Cortisol Production**

116 Cortisol production was measured as previously described ³², with minor modifications.
117 Briefly, head kidney tissue, containing interrenal steroidogenic cells, was removed from
118 unstressed trout from control and BPA treated groups in both F1 and F2 generations (n= 5-6) and
119 placed in a petri dish containing Hank's buffer. The tissue was finely minced, washed in Hank's
120 buffer three times to remove any blood clots, and equally distributed into 24 well plates (3 wells
121 per fish). Tissues slices were pre-incubated for 2 h at 13 °C with gentle shaking to equilibrate.
122 The tissue from each fish was then exposed to either fresh buffer only (no stimulus group) or
123 fresh buffer containing 0.5IU ml⁻¹ ACTH for 4 h at 13 °C, with gentle shaking. The
124 concentration of ACTH chosen was based on a previous study ³². At the end of the exposure,
125 samples were collected, quickly centrifuged at 13,000 × g for 1 min, and supernatant stored
126 frozen at -80 °C for later cortisol determination. Lactate dehydrogenase (LDH) leakage was used
127 to confirm tissue viability ³², and there was no effect on tissue viability due to the incubation
128 protocol.

129 **Plasma, Medium and Tissue Analyses**

130 Cortisol analysis in the plasma and medium (*in vitro* assay) were carried out by
131 radioimmunoassay (RIA) as described previously ³³. Plasma glucose and lactate levels were
132 measured enzymatically as described previously ^{34,35}. Liver glycogen content was determined by

133 measuring glucose levels before and after amyloglucosidase hydrolysis as described before ³⁵,
134 while protein was measured using the bicinchoninic acid method using bovine serum albumin as
135 the standards ³⁵.

136 **Tissue Transcript Analysis**

137 Tissue RNA extraction, cDNA synthesis and the quantitative real-time PCR (qPCR)
138 protocol have been described in detail previously ¹². Briefly, the transcript levels were analyzed
139 using the iQTM SYBR[®] green supermix fluorescent dye with the CFX96 TouchTM Real-Time
140 PCR Detection System (Bio-Rad, Hercules, CA). Each sample was assayed in duplicate and the
141 following thermal cycling protocol was followed: 2 min at 94 °C; 40 cycles of: 30 s at 95 °C,
142 followed by 30 s at the melting temperature for each gene (Supporting Information [SI] Table 1);
143 1 min at 95 °C; 1 min at 55 °C, followed by melt curve analysis starting from 55- 95 °C in
144 increments of 0.5 °C every 10 s. Copy number for each gene was determined using plasmid
145 standard curves previously established in our laboratory following the protocol described
146 previously ¹². All samples were assayed for the genes of interest and for the housekeeping gene,
147 elongation factor 1 α (*ef1 α*), which did not change between treatments.

148 **Transcriptome Analysis of Stress Related Genes in the Liver**

149 Liver stress transcriptome of 4 fish prior to the stressor exposure per treatment in both
150 generations were described using expression results from a previous study ²¹, deposited into the
151 Gene Expression Omnibus (GEO) database (Accession #: GSE94281;
152 <http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE94281>). Protocol for identification,
153 annotation, and enrichment of differentially expressed genes between treatments in 365 dpf F1
154 and F2 generations was described previously ²¹. For the present study, only genes differentially

155 expressed in at least one of BPA treatment and enriched for the GO term “response to stress”
156 (GO:0006950) were used.

157 **Statistical Analysis**

158 Statistical analyses were performed by use of SigmaPlot 13.0 software (Systat Software
159 Inc., San Jose, CA, USA). All transcript abundance results were presented as fold change
160 compared to the treatment control (BPA0) at time 0 h. Heatmaps represent mean values per
161 treatment for each time point pre and post-stressor exposures within each generation. All
162 heatmaps were plotted using the function heatmap.2 from the gplots package in R³⁶. A two way
163 analysis of variance (ANOVA) followed by Holm–Sidak post hoc test was used to determine
164 significant effect of BPA exposure and handling stressor on plasma cortisol, glucose, and lactate
165 concentrations, liver glycogen content, and transcript abundance of genes in the liver, muscle and
166 head kidney tissues. When there was a significant interaction between BPA exposure and
167 handling stressor, a one-way ANOVA followed by Holm–Sidak post hoc test was used to
168 separately test the effect of BPA exposure or time on those parameters. Data were log-
169 transformed wherever necessary to meet the assumptions of normality and equal variance. Only
170 non-transformed data are shown in the figures. Figures were plotted either using SigmaPlot 13.0
171 or R 3.3.1 (<http://cran.r-project.org/>). A probability level of $p < 0.05$ was considered significant.
172 All data (except transcript abundance and liver transcriptome) are shown as mean \pm standard
173 error of the mean (S.E.M.). Transcript abundance results with mean \pm S.E.M can be found in the
174 SI Tables 2, 3 and 4.

175 **RESULTS**

176 **Plasma Cortisol Response**

177 There was a significant interaction between BPA exposure and acute handling stressor on
178 plasma cortisol concentrations in the F1 generation of trout. BPA treatment did not alter resting
179 plasma cortisol concentration at 365 dpf in the F1 generation (Fig. 1A). However, BPA
180 significantly impacted the ability of these fish to respond to an acute handling stressor (Fig. 1A).
181 Trout from the control treatment group showed a significantly greater cortisol response 1 h after
182 an acute stressor exposure, which returned to unstressed levels at 4 h post-stressor exposure (Fig.
183 1A). Although F1 generation trout from the 4 ng BPA group showed a plasma cortisol response
184 to stress similar to that of the control group, the cortisol levels at 1h after the acute handling
185 stressor was significantly lower in that group when compared to the controls. In the F1 40 ng
186 BPA group, the cortisol levels 1 h post-stress were not significantly different than those of
187 unstressed fish (Fig. 1A). In the F2 generation, acute handling stressor significantly increased
188 plasma cortisol levels at 1 h post-stressor and this steroid level dropped to basal level at 4 and 24
189 h post-stressor exposure in all treatment groups (Fig. 1B). BPA had no significant impact on
190 either the unstressed or stressed levels of plasma cortisol concentrations in the F2 generation
191 (Fig. 1B).

192 **Interrenal Cortisol Production and Transcript Abundance**

193 To determine if BPA accumulation in eggs impacts cortisol production capacity, this
194 steroid production was monitored in the head kidney of F1 and F2 rainbow trout following *in*
195 *vitro* stimulation with ACTH (Figs. 1C & D). When compared to the un-stimulated head kidney
196 tissues, ACTH stimulation for 4 h significantly increased cortisol levels in all treatment groups,
197 and this response was not modified by BPA accumulation in eggs (Fig. 1C). A similar response
198 was also seen in the F2 generation head kidney tissues and the basal or ACTH-stimulated
199 cortisol production was not altered by BPA in eggs (Fig. 1D).

200 Significantly greater transcript abundance of *mc2r*, *p450scc* and *star* was observed in the
201 head kidney tissues of BPA40 group compared to the control group in the F1 generation trout
202 (Fig. 1E). Acute handling stressor had no effects on *mc2r* and *p450scc* transcript abundance, but
203 it significantly upregulated transcript abundance of *star* in all treatment groups at 24 h post-stress
204 when compared to expression of *star* in the pre-stressed trout (Fig. 1E).

205 In the F2 generation, BPA had no significant effect on *mc2r* and *p450scc* transcript
206 abundance in the head kidney of trout (Fig. 1E). Acute handling stressor significantly
207 upregulated transcript abundance of *p450scc* at 1 and 4 h post-stressor when compared to
208 unstressed fish. However, BPA treatment impacted the stressor-mediated transcript abundance of
209 *star* in the BPA40, but not the BPA4 group (Fig. 1E). In the 40 ng BPA group, *star* mRNA
210 abundance was upregulated by approximately 2-fold at 1 h post-stress when compared to
211 controls at the same time point and to unstressed individuals from the same treatment group (Fig.
212 1E & SI Table 2).

213 **Plasma Secondary Stress Response**

214 There were no significant effects of BPA on plasma glucose and lactate concentrations in
215 the F1 generation fish at 365 dpf (Fig. 2A & B). The acute stressor significantly increased
216 plasma glucose concentration at 4 h in all groups. Similarly, plasma lactate concentrations were
217 increased by 2-3 fold at 1h post-stress in all treatment groups (Fig. 2B). Liver glycogen content
218 was significantly lower in 24 h post-stress trout from all the treatment groups when compared to
219 the unstressed and 1h post-stress trout, but this was not impacted by BPA treatment (Fig. 2C).

220 In the F2 generation, BPA had no effect on plasma glucose levels in trout from unstressed
221 or post-stressor groups (Fig. 2D). Acute stressor significantly decreased the plasma glucose

222 concentrations 24 h after post stress in all treatment groups when compared to the 0, 1 and 4h
223 trout (Fig. 2D). There was a significant main effect of BPA on stressor-mediated plasma lactate
224 concentrations in the F2 generation trout (Fig. 2E). Similar to F1 generation trout, plasma lactate
225 was significantly greater 1h after the acute handling stressor in all treatment groups. Ancestral
226 exposure to 40 ng BPA increased plasma lactate concentrations by approximately 2-fold in F2
227 trout when compared to the controls and the 4 ng BPA group (Fig. 2E). Liver glycogen
228 concentrations were significantly lower 24 h after post-stress in all treatment groups when
229 compared to the unstressed and 1h post-stress trout, but this was not impacted by BPA treatment
230 (Fig. 2F).

231 **Liver Stress Transcriptomics and Targeted Genes Expression**

232 The effect of BPA accumulation on stress related genes was determined in the liver of
233 unstressed F1 and F2 generation trout using a transcriptomics approach²¹. Only differentially
234 expressed genes related to the GO term ‘response to stress’ were selected for this study (Fig. 3A
235 - D & SI Table 5). There were a total of 35 and 66 stress-related genes that were differentially
236 expressed between at least one BPA treatment compared to the control in F1 and F2 generations,
237 respectively (Fig. 3A, C & D). 17 of those differentially expressed genes were identical in both
238 F1 and F2 generations (Fig. 3A). Based on gene ontology terms, the six most represented
239 biological functions in the F1 generation trout were defense response, innate immune response,
240 response to organic substance, cellular nitrogen compound metabolic process, macromolecule
241 metabolic process and regulation of cellular process. In the F2 generation, the six most
242 represented biological functions were defense response, regulation of cellular process,
243 macromolecule metabolic process, innate immune response, signal transduction, and cellular

244 macromolecule metabolic process. The majority of differentially expressed stress-related genes
245 in the F1 and F2 generations trout participated in the defense response (Fig. 3B).

246 The stressor-mediated growth and stress related transcript changes were also assessed
247 using qPCR. In the F1 generation trout, 40 ng BPA egg accumulation upregulated transcript
248 abundance of *igf1* and *igf2* (Fig. 4A). Acute handling stressor significantly increased (~1.5 to 3
249 fold) transcript abundance of *igf1*, *igf2* and *gh2r* in all treatment groups at all time-points post-
250 stressor exposure (Fig. 4A & SI Table 3). Transcript abundance of *gh1r* was significantly
251 increased (~2 to 3 fold) in all groups at 24h post-stress when compared to the unstressed trout
252 (Fig. 4A & SI Table 3). There were no interactive effects of BPA accumulation in eggs and acute
253 handling stressor on transcript abundance of *igf1ra* and *igf1rb* in trout (Fig. 4A). Also, BPA had
254 no significant effect on the expression of stress related genes (*gr1*, *gr2* and *mr*) in the trout liver
255 (Fig. 4A). However, an acute handling stressor significantly increased *gr1* and *gr2* (~1.5 to 4
256 fold), but not *mr*, transcript levels in all the post-stress treatment groups when compared to the
257 unstressed trout (Fig. 4A & SI Table 3).

258 In the F2 40 ng BPA group, transcript abundance of genes involved in growth (*igf2* and
259 *igf1ra*), and stress response (*gr1* and *mr*) were modified (Fig. 4B). Transcript levels of *igf1ra*
260 were significantly increased, while those for *igf2* were significantly decreased in pre and post
261 stress time periods when compared to the control trout (Fig. 4B). A 50 % reduction in transcript
262 abundance of *igf1ra* was observed at 1, 4 and 24 h post stress trout from all treatment groups
263 when compared to unstressed fish. Acute handling stressor, but not maternal ancestral exposure
264 to BPA, significantly increased *igf1* transcript abundance at 24 h post stress in trout from all
265 treatment groups. In the liver of F2 40 ng BPA group, there was a significant upregulation of *mr*
266 and downregulation of *gr1* transcript abundance in both pre and post stress time points when

267 compared to the control trout (Fig. 4B). An acute stressor significantly increased *gr1* transcript
268 abundance in the liver of F2 generation trout from all treatment groups when compared to the
269 unstressed individuals, but *gr2* transcript abundance significantly increased only at 1h post-stress
270 when compared to the unstressed fish (Fig. 4B).

271 **Muscle Transcript Abundance**

272 BPA accumulation in eggs had no effect on transcript abundance of genes related to
273 growth (*igflrb*, *gh1r*, *gh2r* and *igflra*) and stress response (*mr*, *gr2* and *gr1*) in the muscle of F1
274 rainbow trout (Fig. 5A). *Igf1* and *2* are not expressed in muscle tissues and hence we did not
275 measure transcript abundance of those genes. An acute handling stressor significantly
276 upregulated transcript abundance of all above mentioned genes in the muscle at all post-stressor
277 time-points when compared to the unstressed trout (Fig. 5A).

278 In the F2 generation, elevated transcript abundance of muscle *igflrb* was noticed in
279 BPA4 and 40 groups compared to the controls (Fig. 5B). Also, BPA40 group had significantly
280 higher *igflra* and *mr* compared to the control group. An acute handling stressor significantly
281 increased transcript abundance of *igflrb*, *gh1r*, *gh2r*, *igflra*, *mr* and *gr2* in trout muscle at all
282 post-stressor time-points when compared to the unstressed trout (Fig. 5B).

283 **Epigenetic Markers in the Liver and Muscle**

284 BPA accumulation in eggs did not significantly affect the transcript abundance of liver
285 epigenetic markers, including *dnmt1*, *dnmt2* and liver specific *mat1a*, in the F1 generation (Fig.
286 4A). An acute handling stressor challenge significantly increased *dnmt2* transcript levels at 4 and
287 24 h post-stressor in the liver of trout from all treatment groups when compared to the unstressed
288 trout (Fig. 4A). No significant changes were observed in *mat1a* and *dnmt1* transcript abundance

289 (Fig. 4A). In the F2 generation, BPA treatment significantly increased liver *dnmt2*, but not *mat1a*
290 and *dnmt1* transcript levels only in the BPA40 group compared to the controls (Fig. 4B). The
291 acute handling stressor, regardless of BPA treatment, reduced the transcript abundance of *dnmt1*
292 at all post-stressor time-points compared to the unstressed trout (Fig. 4B). The transcript
293 abundance of *dnmt2* was significantly higher only at 4 h post-stressor time-point when compared
294 to the unstressed trout, while *mat1a* was not affected by the stress challenge.

295 BPA accumulation in eggs had no significant effect on the transcript abundance of
296 muscle *dnmt1* and *dnmt2* in the F1 generation (Fig. 5A). An acute handling stressor significantly
297 upregulated the transcript abundance of muscle *dnmt1*, but not *dnmt2*, at all time-points post-
298 stressor compared to the unstressed trout (Fig. 5A). In the F2 generation, trout from the BPA40
299 group demonstrated significantly greater increase in transcript abundance of muscle *dnmt1*
300 compared to the control group, but not such treatment effect was noticed for *dnmt2* (Fig. 5B). An
301 acute handling stressor significantly up regulated transcript abundance of muscle *dnmt1*, but not
302 *dnmt2* in post stress trout from all treatment groups when compared to the unstressed group (Fig.
303 5B).

304 **DISCUSSION**

305 The most significant finding of this study was that BPA accumulation in eggs, mimicking
306 maternal transfer of this contaminant, altered the acute stress performances in two generations of
307 rainbow trout. The longer-term and generations effects in plasma stress parameters and the target
308 tissue molecular effects were more evident in the 40 ng BPA per egg compared to the 4 ng BPA
309 per egg groups. The teleost stress axis functioning is highly conserved³⁷, and we have shown
310 previously that the developmental programming of the cortisol stress axis was disrupted by BPA
311 accumulation in eggs in the F1 generation^{12,14}. To our knowledge this is the first study to

312 demonstrate BPA impact not only on the plasma stress response, but also acute stress-related
313 transcript changes in the liver, muscle and head kidney of trout in successive generations. The
314 concentrations of BPA reported in trout embryos in the present study are environmentally
315 realistic, as similar BPA concentrations were found in wild fish and zooplankton collected from
316 the BPA-impacted sites^{38,39}. In addition, recent studies have provided evidence of maternal
317 transfer of BPA from the exposed adult female fish to eggs^{3,6}, and hence understanding the
318 generational toxicities of maternally deposited BPA is highly relevant from a risk assessment
319 stand-point.

320 In our study, the accumulated BPA in eggs was rapidly cleared during embryogenesis
321 with levels below detection at hatch (42 dpf)¹³. The low level exposure of BPA during early
322 embryogenesis was shown previously to impact the developmental programming of the growth
323 and stress axes¹²⁻¹⁴. This disruption of stress axis development in early life stages may have
324 played a role in the altered stressor-mediated plasma cortisol and/or metabolite levels seen in the
325 1 yr old fish in the F1 and F2 generations. The suppression of stressor-induced plasma cortisol
326 response in the BPA40 group is consistent with an earlier study showing a similar response in the
327 F1 generation¹². The lower steroid response corresponded with an upregulation of genes
328 encoding proteins critical for steroid biosynthesis in the interrenal tissue in BPA40 group in the
329 F1 generations. A similar mismatch in steroidogenic gene expression and cortisol output was
330 also observed in progenies (baseline group only) of sockeye salmon (*Oncorhynchus nerka*)
331 exposed to maternal stress⁴⁰, and 65 dpf trout exposed to BPA during embryogenesis¹⁴,
332 suggesting contaminant impact on the transcript stability or turnover. However, our results reveal
333 that the attenuation of the cortisol response in the F1 generation may not be due to disruption in
334 steroid biosynthesis as the BPA fish were able to evoke an ACTH-stimulated cortisol response

335 similar to that of the control group (Fig. 1C). This suggests that BPA impacts the hypothalamus-
336 pituitary axis development, leading to disruption in either the CRF and/or ACTH production.
337 This notion was supported by a recent study demonstrating that BPA modifies both CRH and
338 ACTH transcripts in rats ⁴¹, and such changes may affect the developmental programming of the
339 cortisol stress axis. The developmental impact of BPA on cortisol stress functioning seen in F1
340 generation was not transferred to the F2 generation of trout in the present study. This was also
341 the case with the secondary stress response indicators, including plasma glucose and liver
342 glycogen content, as BPA accumulation in eggs did not modify the stressor-mediated changes in
343 these parameters in the F1 and F2 generation trout. However, this was not the case with stressor-
344 mediated plasma lactate level, which was higher in the BPA40 group in the F2 generation. As
345 elevated lactate level is an indicator of anaerobic metabolism and altered secondary stress
346 response in fish ^{31,42}, our results suggest adverse effects on muscle energy metabolism in the F2
347 generation in response to ancestral exposure to BPA (see below).

348 In the present study liver transcriptomic analysis revealed that a total of 35 and 66 stress
349 related genes were differentially expressed between at least one BPA treatment and the control (0
350 BPA) in F1 and F2 generations, respectively. In the F2 generation liver, approximately a two-
351 fold increase in stress-related genes, including genes related to host defense, regulation of
352 cellular process and macromolecule metabolic process, suggest that BPA impacts on molecular
353 programming events are more evident in the F2 generation trout ²¹. The majority of differentially
354 expressed stress-related genes in both F1 and F2 generations participate in the host defense
355 response. Examples of differentially expressed stress genes with host defense response functions
356 include, among others, mx proteins, retinoic acid inducible protein-i, interferon inducible mx
357 proteins, toll-like receptors genes. All these genes participate in immune response and are

358 essential for limiting viral and bacterial infection or diseases⁴³. A bidirectional communication
359 between stress axis and immune system has been reported in fish⁴⁴ and modification of this
360 communication by contaminants may lead to organismal level impact. A number of previous
361 studies have demonstrated that exposure to pollutants or stress can modify the host immune
362 response, which increases susceptibility of salmonids to both viral and bacterial infections⁴⁵⁻⁴⁷.
363 Future studies should test the hypothesis that whether developmental exposure to BPA in trout
364 increases the risk of infections in exposed generations and their progenies, and the underlying
365 mechanism needs to be elucidated.

366 While the hormonal regulation of growth has been extensively reviewed⁴⁸, the alterations
367 in the response of somatotrophic axis genes to acute stressor exposure is far from clear. Our
368 results reveal for the first time acute changes in the stress- and growth-related transcripts in the
369 liver and muscle of fish in response to an acute handling stressor. The majority of transcripts
370 were upregulated at 1 h and they stayed elevated over the 24 h period after an acute stressor, the
371 only exception was liver *igflra* in the F2 generation that was significantly downregulated post-
372 stressor exposure. Acute handling stress challenge in Coho salmon (*Oncorhynchus kisutch*)
373 increased hepatic *igfl* expression without an increase in *ghr* at 1.5 h post-stressor, but transcript
374 levels of the two genes were dropped to the control levels at 16 h post-stressor challenge⁴⁹. As in
375 Coho salmon, we observed similar trend in *igfl* and *ghr1* gene expression in the liver of F1
376 generation trout at 1 h, but we did not observe a drop in transcript abundance of those genes at 24
377 h, suggesting that the temporal profile of *ghr1* and *igfl* genes respond differentially to an acute
378 stressor challenge. We saw greater transcript abundance of growth-related genes in trout muscle
379 after an acute handling stress challenge. Greater muscle GHR protein expression was reported in
380 fish after heat shock⁵⁰. On the other hand, no change in *ghr1* and down regulation of *ghr2* were

381 observed in muscle of *Pampus argenteus* underwent a handling stress challenge⁵¹. Collectively,
382 the results suggest that the transcript abundance of growth-related genes in the liver and muscle
383 are modified by acute stressor in fish, and these changes may be related to acute stress hormone
384 stimulation of muscle metabolism^{20,31}.

385 Our results reveal that BPA in eggs disrupt the acute stressor-mediated changes in
386 molecular growth targets in the liver and muscle of the trout progeny in two generations. Given
387 these tissues are the major metabolic targets for stress hormones action during acute stress
388 recovery and adaptation in fish^{31,52}, the results suggest a compromised stress performance. The
389 lower transcript abundance of *igf2* and a trend for reduced transcript abundance of *igf1* in the
390 liver, as well as a significant increase in transcript abundance of *igf1ra* and *igf1rb* in the muscle
391 of F2 generation trout in the BPA40 group supports disruption in molecular programming of the
392 growth axis by BPA, as studies have shown that stressors impair growth axis development and
393 function^{20,21}. Similarly, both *gr1* and *mr* were differentially expressed in the liver of F2
394 generation trout from the BPA40 group underscoring possible changes in target tissue stress
395 steroid responsiveness that are evident in the F2 generation. The overall increase in transcript
396 abundance in the muscle and liver in the BPA group reflects a higher tissue metabolic demand,
397 as transcription and/or translation are energy demanding^{20,52}, and contributes to the increased
398 energy demand during stress in fish^{31,52}. Our results suggest that BPA accumulation in eggs
399 disrupts the metabolic adjustments that are essential during acute stress adaptation⁵², leading to
400 the proposal that the overall stress performance will be compromised by BPA even in the F2
401 generation of trout.

402 Recent studies have suggested BPA-induced epigenetic modifications for generational
403 toxicities in mammals^{8,53}, and we propose a similar mode of action for the observed

404 modification of stress response in F2 generation trout. To this end, we investigated the impact of
405 BPA in eggs on transcript abundance of genes involved in epigenetic modification in both liver
406 and muscle of the F1 and F2 generations trout before and after an acute handling stress
407 challenge. We observed a significant upregulation of *dnmt2* in the liver and *dnmt1* in the muscle
408 of F2 generations trout from the BPA40 group. Role of *dnmt1* in DNA methylation is well
409 established whereas *dnmt2* has a weak DNA methylation activity but is involved in the transfer
410 RNA (tRNA) methylation^{8,53-55}. Methylation of tRNA by *dnmt2* has been demonstrated to
411 promote tRNA stability as it protects tRNA against ribonuclease cleavage during thermal or
412 chemical stress⁵⁴. tRNA-derived small RNAs has been suggested to trigger gene silencing⁵⁵,
413 suggesting that altered *dnmt2*-induced tRNA methylation during stress may indirectly affect gene
414 expression. In addition, a role for *dnmt2* in transgenerational epigenetic modification was
415 recently demonstrated in mice⁵⁶. Hyper DNA methylation of the promotor region of a gene
416 causes transcriptional repression, and this may be involved in the down-regulation of *igf2* and
417 *gr1* in the liver of F2 generation trout from the BPA40 group, but this needs to be further tested
418 and validated. However, we also saw a significant number of growth and stress response genes
419 upregulated in both liver and muscle of F2 generation trout from the BPA40 group, leading to
420 the proposal that epigenetics mechanisms other than DNA methylation, including histone
421 modification⁵⁷, may also be involved in the BPA-induced generational toxicities in trout. Taken
422 together, epigenetic modification, including DNA methylation may potentially be involved in the
423 BPA-induced generational toxicities in fish.

424 Overall, the study provided evidence that BPA (~ 40 ng) accumulation in eggs,
425 mimicking a maternal transfer scenario, leads to impairment of the primary and secondary stress
426 response over two generations of rainbow trout. Given the importance of trout as a model species

427 to investigate stressor effects in salmonids ^{27,58}, observed BPA-induced generational impairment
428 in the stress response may be reflective of the potential impact this environmental contaminant
429 may exert on salmonid fitness, including reducing their ability to respond to additional stressors
430 such as climate change, pollution, disease or predation. To this end, studies have shown that
431 exposure to endocrine disruptors affect fish performances, including development, stress
432 reactivity, behaviour, disease susceptibility, reproduction and fitness ^{12,16,40,47,59}. However,
433 toxicities associated with parental and ancestral exposures are not currently included in the
434 ecological risk assessment framework. Our finding that BPA accumulation in eggs can have
435 long-term and multigenerational adverse effects, in spite of complete lack of tissue contaminant
436 burden ^{12,13}, suggests that the current risk assessment framework may not protect aquatic animals
437 against chemicals such as BPA in contaminated sites. Our study underscores the need for
438 developing biomarkers to predict generational toxicities in aquatic animals, and include that
439 information in ecological risk assessment for management of such chemicals.

440

441 **ASSOCIATED CONTENT**

442 **Supporting Information**

443 Information on primer sequences (Table 1), mean values of stress-related genes in head kidney
444 (Table 2), and mean values of growth, stress and epigenetic-related genes in liver (Table 3) and
445 muscle (Table 4) of the F1 and F2 generations trout, and the gene ontology (GO) term
446 description of differentially expressed stress-related genes in the liver of two generations of trout
447 (Table 5).

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465 **Reference**

- 466 (1) Canesi, L.; Fabbri, E. Environmental Effects of BPA Focus on Aquatic Species. *Dose-Response*
 467 **2015**, *13* (3), 1559325815598304.
- 468 (2) USEPA. Bisphenol A Action Plan (CASRN 80-05-7). 2010.
- 469 (3) Oehlmann, J.; Schulte-Oehlmann, U.; Kloas, W.; Jagnytsch, O.; Lutz, I.; Kusk, K. O.;
 470 Wollenberger, L.; Santos, E. M.; Paull, G. C.; Look, K. J. W. V.; et al. A Critical Analysis of the
 471 Biological Impacts of Plasticizers on Wildlife. *Philos. Trans. R. Soc. B Biol. Sci.* **2009**, *364* (1526),
 472 2047–2062.
- 473 (4) Balakrishnan, B.; Henare, K.; Thorstensen, E. B.; Ponnampalam, A. P.; Mitchell, M. D. Transfer
 474 of Bisphenol A across the Human Placenta. *Am. J. Obstet. Gynecol.* **2010**, *202* (4), 393.e1-393.e7.
- 475 (5) Moors, S.; Diel, P.; Degen, G. H. Toxicokinetics of Bisphenol A in Pregnant DA/Han Rats after
 476 Single I.v. Application. *Arch. Toxicol.* **2006**, *80* (10), 647–655.
- 477 (6) Takao, Y.; Oishi, M.; Nagae, M.; Kohra, S.; Arizono, K. Bisphenol A Incorporated into Eggs from
 478 Parent Fish Persists for Several Days. *J. Health Sci.* **2008**, *54* (2), 235–239.
- 479 (7) Kundakovic, M.; Champagne, F. A. Epigenetic Perspective on the Developmental Effects of
 480 Bisphenol A. *Brain. Behav. Immun.* **2011**, *25* (6), 1084–1093.
- 481 (8) Xin, F.; Susiarjo, M.; Bartolomei, M. S. Multigenerational and Transgenerational Effects of
 482 Endocrine Disrupting Chemicals: A Role for Altered Epigenetic Regulation? *Semin. Cell Dev.*
 483 *Biol.* **2015**, *43*, 66–75.
- 484 (9) Bhandari, R. K.; vom Saal, F. S.; Tillitt, D. E. Transgenerational Effects from Early
 485 Developmental Exposures to Bisphenol A or 17 α -Ethinylestradiol in Medaka, *Oryzias Latipes*. *Sci.*
 486 *Rep.* **2015**, *5*, 9303.
- 487 (10) Kang, I. J.; Yokota, H.; Oshima, Y.; Tsuruda, Y.; Oe, T.; Imada, N.; Tadokoro, H.; Honjo, T.
 488 Effects of Bisphenol a on the Reproduction of Japanese Medaka (*Oryzias Latipes*). *Environ.*
 489 *Toxicol. Chem.* **2002**, *21* (11), 2394–2400.
- 490 (11) Tyl, R. W.; Myers, C. B.; Marr, M. C.; Thomas, B. F.; Keimowitz, A. R.; Brine, D. R.; Veselica,
 491 M. M.; Fail, P. A.; Chang, T. Y.; Seely, J. C.; et al. Three-Generation Reproductive Toxicity Study
 492 of Dietary Bisphenol A in CD Sprague-Dawley Rats. *Toxicol. Sci.* **2002**, *68* (1), 121–146.
- 493 (12) Aluru, N.; Leatherland, J. F.; Vijayan, M. M. Bisphenol A in Oocytes Leads to Growth
 494 Suppression and Altered Stress Performance in Juvenile Rainbow Trout. *PLoS ONE* **2010**, *5* (5),
 495 e10741.
- 496 (13) Birceanu, O.; Servos, M. R.; Vijayan, M. M. Bisphenol A Accumulation in Eggs Disrupts the
 497 Endocrine Regulation of Growth in Rainbow Trout Larvae. *Aquat. Toxicol.* **2015**, *161*, 51–60.
- 498 (14) Birceanu, O.; Mai, T.; Vijayan, M. M. Maternal Transfer of Bisphenol A Impacts the Ontogeny of
 499 Cortisol Stress Response in Rainbow Trout. *Aquat. Toxicol.* **2015**, *168*, 11–18.
- 500 (15) Baker, T. R.; King-Heiden, T. C.; Peterson, R. E.; Heideman, W. Dioxin Induction of
 501 Transgenerational Inheritance of Disease in Zebrafish. *Mol. Cell. Endocrinol.* **2014**, *398* (0), 36–
 502 41.
- 503 (16) King-Heiden, T. C.; Mehta, V.; Xiong, K. M.; Lanham, K. A.; Antkiewicz, D. S.; Ganser, A.;
 504 Heideman, W.; Peterson, R. E. Reproductive and Developmental Toxicity of Dioxin in Fish. *Mol.*
 505 *Cell. Endocrinol.* **2012**, *354* (1–2), 121–138.
- 506 (17) Vijayan, M. M.; Aluru, N.; Leatherland, J. F. Stress Response and the Role of Cortisol. In *Fish*
 507 *Diseases and Disorders Non-infectious Disorders*; Leatherland, J. F., Woo, P. T. K., Eds.; CABI
 508 Press: New York, 2010; pp 182–201.
- 509 (18) Charmandari, E.; Tsigos, C.; Chrousos, G. Endocrinology of the Stress Response. *Annu. Rev.*
 510 *Physiol.* **2005**, *67* (1), 259–284.
- 511 (19) Hontela, A.; Vijayan, M. M. Adrenocortical Toxicology in Fishes. In *Adrenal Toxicology*; Harvey,
 512 P. W., Everett, D., Springall, C. J., Eds.; Target Organ Toxicology Series; CRC Press: Boca Raton,
 513 FL, 2008; pp 233–256.

- 514 (20) Sadoul, B.; Vijayan, M. M. Stress and Growth. In *Biology of Stress in Fish*; Schreck, C. B., Tort,
515 L., Farrell, A. P., Brauner, C. J., Eds.; Fish Physiology; Academic Press: London, UK., 2016; Vol.
516 35, pp 167–206.
- 517 (21) Sadoul, B.; Birceanu, O.; Aluru, N.; Thomas, J. K.; Vijayan, M. M. Bisphenol A in Eggs Causes
518 Development-Specific Liver Molecular Reprogramming in Two Generations of Rainbow Trout.
519 *Sci. Rep.* **2017**, *7* (1), 14131.
- 520 (22) McConnachie, S. H.; Cook, K. V.; Patterson, D. A.; Gilmour, K. M.; Hinch, S. G.; Farrell, A. P.;
521 Cooke, S. J. Consequences of Acute Stress and Cortisol Manipulation on the Physiology,
522 Behavior, and Reproductive Outcome of Female Pacific Salmon on Spawning Grounds. *Horm.*
523 *Behav.* **2012**, *62* (1), 67–76.
- 524 (23) Cook, K. V.; McConnachie, S. H.; Gilmour, K. M.; Hinch, S. G.; Cooke, S. J. Fitness and
525 Behavioral Correlates of Pre-Stress and Stress-Induced Plasma Cortisol Titters in Pink Salmon
526 (*Oncorhynchus Gorbusha*) upon Arrival at Spawning Grounds. *Horm. Behav.* **2011**, *60* (5), 489–
527 497.
- 528 (24) Bonier, F.; Martin, P. R.; Moore, I. T.; Wingfield, J. C. Do Baseline Glucocorticoids Predict
529 Fitness? *Trends Ecol. Evol.* **2009**, *24* (11), 634–642.
- 530 (25) Carruth, L. L.; Dores, R. M.; Maldonado, T. A.; Norris, D. O.; Ruth, T.; Jones, R. E. Elevation of
531 Plasma Cortisol during the Spawning Migration of Landlocked Kokanee Salmon (*Oncorhynchus*
532 *Nerka Kennerlyi*). *Comp. Biochem. Physiol. Toxicol. Pharmacol. CBP* **2000**, *127* (2), 123–131.
- 533 (26) Carruth, L. L.; Jones, R. E.; Norris, D. O. Cortisol and Pacific Salmon: A New Look at the Role of
534 Stress Hormones in Olfaction and Home-Stream Migration. *Integr. Comp. Biol.* **2002**, *42* (3), 574–
535 581.
- 536 (27) Köllner, B.; Wasserrab, B.; Kotterba, G.; Fischer, U. Evaluation of Immune Functions of Rainbow
537 Trout (*Oncorhynchus Mykiss*)—how Can Environmental Influences Be Detected? *Toxicol. Lett.*
538 **2002**, *131* (1), 83–95.
- 539 (28) Thorgaard, G. H.; Bailey, G. S.; Williams, D.; Buhler, D. R.; Kaattari, S. L.; Ristow, S. S.;
540 Hansen, J. D.; Winton, J. R.; Bartholomew, J. L.; Nagler, J. J.; et al. Status and Opportunities for
541 Genomics Research with Rainbow Trout. *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* **2002**,
542 *133* (4), 609–646.
- 543 (29) Danzmann, R. G.; Cairney, M.; Davidson, W. S.; Ferguson, M. M.; Gharbi, K.; Guyomard, R.;
544 Holm, L.-E.; Leder, E.; Okamoto, N.; Ozaki, A.; et al. A Comparative Analysis of the Rainbow
545 Trout Genome with 2 Other Species of Fish (Arctic Charr and Atlantic Salmon) within the
546 Tetraploid Derivative Salmonidae Family (Subfamily: Salmoninae). *Genome* **2005**, *48* (6), 1037–
547 1051.
- 548 (30) Timusk, E. R.; Ferguson, M. M.; Moghadam, H. K.; Norman, J. D.; Wilson, C. C.; Danzmann, R.
549 G. Genome Evolution in the Fish Family Salmonidae: Generation of a Brook Charr Genetic Map
550 and Comparisons among Charrs (Arctic Charr and Brook Charr) with Rainbow Trout. *BMC Genet.*
551 **2011**, *12*, 68.
- 552 (31) Mommsen, T. P.; Vijayan, M. M.; Moon, T. W. Cortisol in Teleosts: Dynamics, Mechanisms of
553 Action, and Metabolic Regulation. *Rev. Fish Biol. Fish.* **1999**, *9* (3), 211–268.
- 554 (32) Sandhu, N.; Vijayan, M. M. Cadmium-Mediated Disruption of Cortisol Biosynthesis Involves
555 Suppression of Corticosteroidogenic Genes in Rainbow Trout. *Aquat. Toxicol.* **2011**, *103* (1–2),
556 92–100.
- 557 (33) Ings, J. S.; Servos, M. R.; Vijayan, M. M. Exposure to Municipal Wastewater Effluent Impacts
558 Stress Performance in Rainbow Trout. *Aquat. Toxicol.* **2011**, *103* (1–2), 85–91.
- 559 (34) Birceanu, O.; Sorensen, L. A.; Henry, M.; McClelland, G. B.; Wang, Y. S.; Wilkie, M. P. The
560 Effects of the Lampricide 3-Trifluoromethyl-4-Nitrophenol (TFM) on Fuel Stores and Ion Balance
561 in a Non-Target Fish, the Rainbow Trout (*Oncorhynchus Mykiss*). *Comp. Biochem. Physiol. Part*
562 *C Toxicol. Pharmacol.* **2014**, *160*, 30–41.
- 563 (35) Vijayan, M. M.; Aluru, N.; Maule, A. G.; Jørgensen, E. H. Fasting Augments PCB Impact on
564 Liver Metabolism in Anadromous Arctic Char. *Toxicol. Sci.* **2006**, *91* (2), 431–439.

- 565 (36) Warnes, G. R.; Bolker, B.; Bonebakker, L.; Gentleman, R.; Liaw, W. H. A.; Lumley, T.; Maechler,
566 M.; Magnusson, A.; Moeller, S.; Schwartz, M.; et al. *Gplots: Various R Programming Tools for*
567 *Plotting Data*; 2016.
- 568 (37) Wendelaar Bonga, S. E. The Stress Response in Fish. *Physiol. Rev.* **1997**, *77* (3), 591–625.
- 569 (38) Corrales, J.; Kristofco, L. A.; Steele, W. B.; Yates, B. S.; Breed, C. S.; Williams, E. S.; Brooks, B.
570 W. Global Assessment of Bisphenol A in the Environment. *Dose-Response* **2015**, *13* (3).
- 571 (39) Staniszevska, M.; Falkowska, L.; Grabowski, P.; Kwaśniak, J.; Mudrak-Cegiołka, S.; Reindl, A.
572 R.; Sokołowski, A.; Szumiło, E.; Zgrundo, A. Bisphenol A, 4-Tert-Octylphenol, and 4-
573 Nonylphenol in the Gulf of Gdańsk (Southern Baltic). *Arch. Environ. Contam. Toxicol.* **2014**, *67*
574 (3), 335–347.
- 575 (40) Sopinka, N. M.; Jeffrey, J. D.; Burnett, N. J.; Patterson, D. A.; Gilmour, K. M.; Hinch, S. G.
576 Maternal Programming of Offspring Hypothalamic–pituitary–interrenal Axis in Wild Sockeye
577 Salmon (*Oncorhynchus Nerka*). *Gen. Comp. Endocrinol.* **2016**, *242*, 30–37.
- 578 (41) Chen, F.; Zhou, L.; Bai, Y.; Zhou, R.; Chen, L. Hypothalamic-Pituitary-Adrenal Axis
579 Hyperactivity Accounts for Anxiety- and Depression-like Behaviors in Rats Perinatally Exposed to
580 Bisphenol A. *J. Biomed. Res.* **2015**, *29* (3), 250–258.
- 581 (42) Thomas, J. K.; Wiseman, S.; Giesy, J. P.; Janz, D. M. Effects of Chronic Dietary
582 Selenomethionine Exposure on Repeat Swimming Performance, Aerobic Metabolism and
583 Methionine Catabolism in Adult Zebrafish (*Danio Rerio*). *Aquat. Toxicol.* **2013**, *130–131*, 112–
584 122.
- 585 (43) Ellis, A. E. Innate Host Defense Mechanisms of Fish against Viruses and Bacteria. *Dev. Comp.*
586 *Immunol.* **2001**, *25* (8), 827–839.
- 587 (44) Weyts, F. A. A.; Cohen, N.; Flik, G.; Verburg-van Kemenade, B. M. L. Interactions between the
588 Immune System and the Hypothalamo-Pituitary-Interrenal Axis in Fish. *Fish Shellfish Immunol.*
589 **1999**, *9* (1), 1–20.
- 590 (45) Collet, B. Innate Immune Responses of Salmonid Fish to Viral Infections. *Dev. Comp. Immunol.*
591 **2014**, *43* (2), 160–173.
- 592 (46) Tort, L. Stress and Immune Modulation in Fish. *Dev. Comp. Immunol.* **2011**, *35* (12), 1366–1375.
- 593 (47) Arkoosh, M. R.; Casillas, E.; Clemons, E.; Kagley, A. N.; Olson, R.; Reno, P.; Stein, J. E. Effect
594 of Pollution on Fish Diseases: Potential Impacts on Salmonid Populations. *J. Aquat. Anim. Health*
595 **1998**, *10* (2), 182–190.
- 596 (48) Wood, A. W.; Duan, C.; Bern, H. A. Insulin-like Growth Factor Signaling in Fish. *Int. Rev. Cytol.*
597 **2005**, *243*, 215–285.
- 598 (49) Nakano, T.; Afonso, L. O. B.; Beckman, B. R.; Iwama, G. K.; Devlin, R. H. Acute Physiological
599 Stress down-Regulates mRNA Expressions of Growth-Related Genes in Coho Salmon. *PLOS*
600 *ONE* **2013**, *8* (8), e71421.
- 601 (50) Kameda, M.; Nakano, T.; Yamaguchi, T.; Sato, M.; Afonso, L. O. B.; Iwama, G. K.; Devlin, R. H.
602 Effects of Heat Shock on Growth Hormone Receptor Expression in Coho Salmon. In *Proceedings*
603 *of the 5th World Fisheries Congress, Yokohama*; Yokohama, Japan, 2008.
- 604 (51) Sun, P.; Yin, F.; Tang, B. Effects of Acute Handling Stress on Expression of Growth-Related
605 Genes in *Pampus Argenteus*. *J. World Aquac. Soc.* **2017**, *48* (1), 166–179.
- 606 (52) Faught, E.; Aluru, N.; Vijayan, M. M. The Molecular Stress Response. In *Biology of Stress in*
607 *Fish*; Schreck, C. B., Tort, L., Farrell, A. P., Brauner, C. J., Eds.; Fish Physiology; Academic
608 Press: London, UK., 2016; Vol. 35, pp 113–166.
- 609 (53) Mileva, G.; Baker, S. L.; Konkle, A. T. M.; Bielajew, C. Bisphenol-A: Epigenetic Reprogramming
610 and Effects on Reproduction and Behavior. *Int. J. Environ. Res. Public Health* **2014**, *11* (7),
611 7537–7561.
- 612 (54) Schaefer, M.; Pollex, T.; Hanna, K.; Tuorto, F.; Meusburger, M.; Helm, M.; Lyko, F. RNA
613 Methylation by Dnmt2 Protects Transfer RNAs against Stress-Induced Cleavage. *Genes Dev.*
614 **2010**, *24* (15), 1590–1595.

- 615 (55) Haussecker, D.; Huang, Y.; Lau, A.; Parameswaran, P.; Fire, A. Z.; Kay, M. A. Human tRNA-
616 Derived Small RNAs in the Global Regulation of RNA Silencing. *RNA* **2010**, *16* (4), 673–695.
- 617 (56) Kiani, J.; Grandjean, V.; Liebers, R.; Tuorto, F.; Ghanbarian, H.; Lyko, F.; Cuzin, F.;
618 Rassoulzadegan, M. RNA-mediated Epigenetic Heredity Requires the Cytosine Methyltransferase
619 Dnmt2. *PLOS Genet.* **2013**, *9* (5), e1003498.
- 620 (57) Bannister, A. J.; Kouzarides, T. Regulation of Chromatin by Histone Modifications. *Cell Res.*
621 **2011**, *21* (3), 381–395.
- 622 (58) Coghlan, S. M.; Ringler, N. H. Temperature-Dependent Effects of Rainbow Trout on Growth of
623 Atlantic Salmon Parr. *J. Gt. Lakes Res.* **2005**, *31* (4), 386–396.
- 624 (59) Heintz, R. A.; Rice, S. D.; Wertheimer, A. C.; Bradshaw, R. F.; Thrower, F. P.; Joyce, J. E.; Short,
625 J. W. Delayed Effects on Growth and Marine Survival of Pink Salmon *Oncorhynchus Gorbuscha*
626 after Exposure to Crude Oil during Embryonic Development. *Mar. Ecol. Prog. Ser.* **2000**, *208*,
627 205–216.

628 **Figure Legends**

629 **Fig. 1. Primary stress response.** Plasma cortisol levels (A and B), head kidney cortisol
630 production (C and D), and transcript abundance of key cortisol biosynthesis genes in head kidney
631 (E) were determined in F1 and F2 trout raised from either the control (0) or BPA-treated (4 and
632 40 ng) eggs in the F0 generation. For A, B and E, time 0 represents changes in variables in the
633 unstressed fish, whereas rest of the time-points (1, 4 and 24 h) represent post-stressor responses.
634 The heatmap represents mean fold changes of key cortisol biosynthesis genes in each treatment
635 groups at each time periods (0, 1, 4, and 24 h) when compared to the unstressed (0 h) control
636 trout. For figure A, different lower case letters denote significant difference between treatment
637 groups across the time periods, while an asterisk represents a significant difference between the
638 control and BPA groups within that time period; for figure B, different lower case letters denote
639 significant difference between the time periods; for figure C and D, different lower case letters
640 denote significant differences between ACTH treated and non-treated groups within each
641 treatment group; and for figure E, different lower case letters denote significant difference
642 between the time periods at the given BPA concentration, an asterisk represents a significant
643 treatment effect, while different uppercase letters denote significant differences within control
644 and BPA treatment groups across the time periods, and a hashtag denotes significant difference
645 between control and given BPA exposed trout at that time-point. All data are shown as mean \pm
646 standard error of the mean (S.E.M.; n = 4-6 samples in each treatment and time points).

647 **Fig. 2. Secondary stress response.** Plasma glucose (A&D), lactate (B&E) and liver glycogen
648 (C&F) levels were determined in F1 and F2 generation trout raised from eggs containing 0, 4 or
649 40 ng BPA. Time '0' represents changes in plasma or liver variables in the unstressed fish from
650 all the treatment groups, whereas the other time points (1, 4 and 24 h) represent post-stressor

651 responses. A two way analysis of variance (ANOVA) followed by Holm–Sidak post hoc test was
652 used to determine significant effect of BPA exposure and time on secondary stress response
653 biomarkers in plasma and liver of two generations of trout (n= 5-6). Different lower case letters
654 denote significant differences with the time periods and an asterisk represents a significant
655 difference between control and given BPA exposed trout.

656 **Fig. 3. Transcriptome analysis of stress related genes in the liver.** Venn diagram of
657 differentially expressed stress-related genes in the liver of F1 and F2 generations trout (A). Only
658 genes differentially expressed in at least one of BPA treatment and enriched for the GO term
659 “response to stress” were selected in our study. Bar graph illustrates the six most represented
660 biological functions of differentially expressed stress response genes in the liver of both
661 generations of trout (B). Differentially expressed stress related genes in F1 (C) and F2 (D)
662 generations trout were shown in the Heatmaps. Each box represents average expression of stress
663 related genes (n=4).

664 **Fig. 4. Liver transcripts of growth- and stress-related genes.** Heatmaps illustrate
665 multigenerational effects of egg BPA accumulation on key growth-, stress- and epigenetics-
666 related genes in the liver of two generations of rainbow trout. Transcript abundance of growth
667 (insulin-like growth factor-1 [*igf1*], insulin-like growth factor-2 [*igf2*], insulin-like growth factor
668 1a receptor [*igf1ra*], insulin-like growth factor 1b receptor [*igf1rb*], growth hormone receptor 1
669 [*gh1r*], growth hormone receptor 2 [*gh2r*]), stress (glucocorticoid receptor 1 [*gr1*],
670 glucocorticoid receptor 2 [*gr2*], and mineralocorticoid receptor [*mr*]) and epigenetics (DNA
671 methyltransferase 1 [*dnmt1*], DNA methyltransferase 2 [*dnmt2*] and liver specific methionine
672 adenosyltransferase 1 alpha [*mat1a*]) related transcripts were measured in trout livers raised from
673 eggs containing 0, 4 and 40 ng BPA before and after an acute handling stress challenge in the F1

674 (A) and F2 (B) generations. Each small box in the heatmaps represents mean fold changes (n=4-
675 6) of key growth, stress and epigenetic related genes in each treatment groups at each time
676 periods (0, 1, 4, and 24h) when compared to the unstressed (0 h) control trout. In the analysis of
677 transcript abundance of growth, stress and epigenetics related genes, different lower case letters
678 denote significant differences with the time periods (two-way ANOVA with Holm–Sidak post
679 hoc test, $p < 0.05$). An asterisk represents a significant difference between control and given
680 BPA exposed trout (two-way ANOVA with Holm–Sidak post hoc test, $p < 0.05$).

681 **Fig. 5. Muscle transcripts of growth- and stress-related genes.** Heatmaps illustrate
682 multigenerational effects of egg BPA accumulation on key growth-, stress- and epigenetics-
683 related genes in the muscle of two generations of rainbow trout. Transcript abundance of growth
684 (insulin-like growth factor 1a receptor [*igflra*], insulin-like growth factor 1b receptor [*igflrb*],
685 growth hormone receptor 1 [*gh1r*], growth hormone receptor 2 [*gh2r*]), stress (glucocorticoid
686 receptor 1 [*gr1*], glucocorticoid receptor 2 [*gr2*], and mineralocorticoid receptor [*mr*]) and
687 epigenetics (DNA methyltransferase 1 [*dnmt1*], DNA methyltransferase 2 [*dnmt2*] related
688 transcripts were measured in trout muscles raised from eggs containing 0, 4 and 40 ng BPA
689 before and after an acute handling stress challenge in the F1 (A) and F2 (B) generations. Each
690 small box in the heatmaps represents mean fold changes (n=4-6) of key growth, stress and
691 epigenetics related genes in each treatment groups at each time periods (0, 1, 4, and 24h) when
692 compared to the unstressed (0 h) control trout. In the analysis of transcript abundance of growth
693 and stress related genes, different lower case letters denote significant differences with the time
694 periods (two-way ANOVA with Holm–Sidak post hoc test, $p < 0.05$). An asterisk represents a
695 significant difference between control and given BPA exposed trout (two-way ANOVA with
696 Holm–Sidak post hoc test, $p < 0.05$).

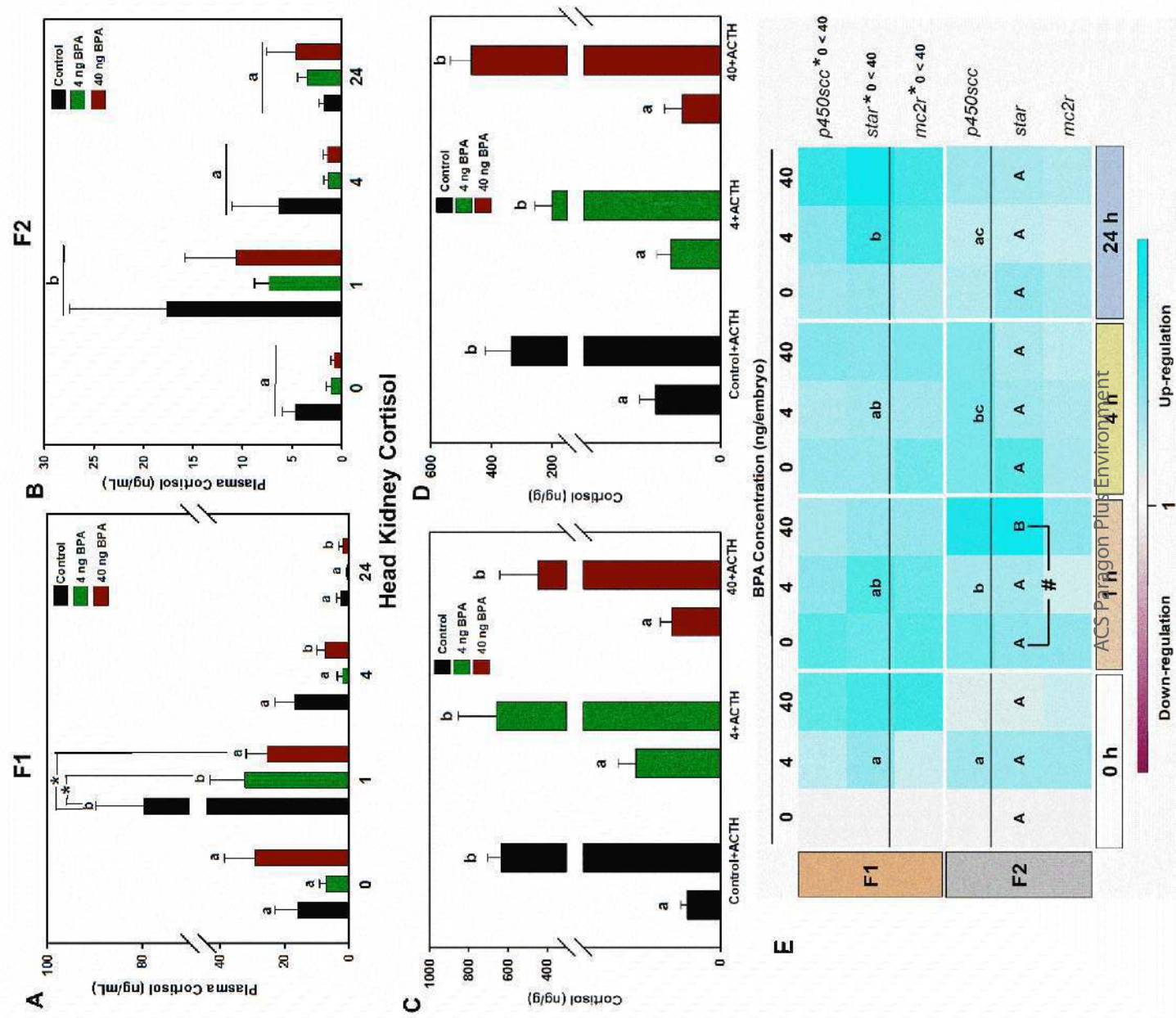
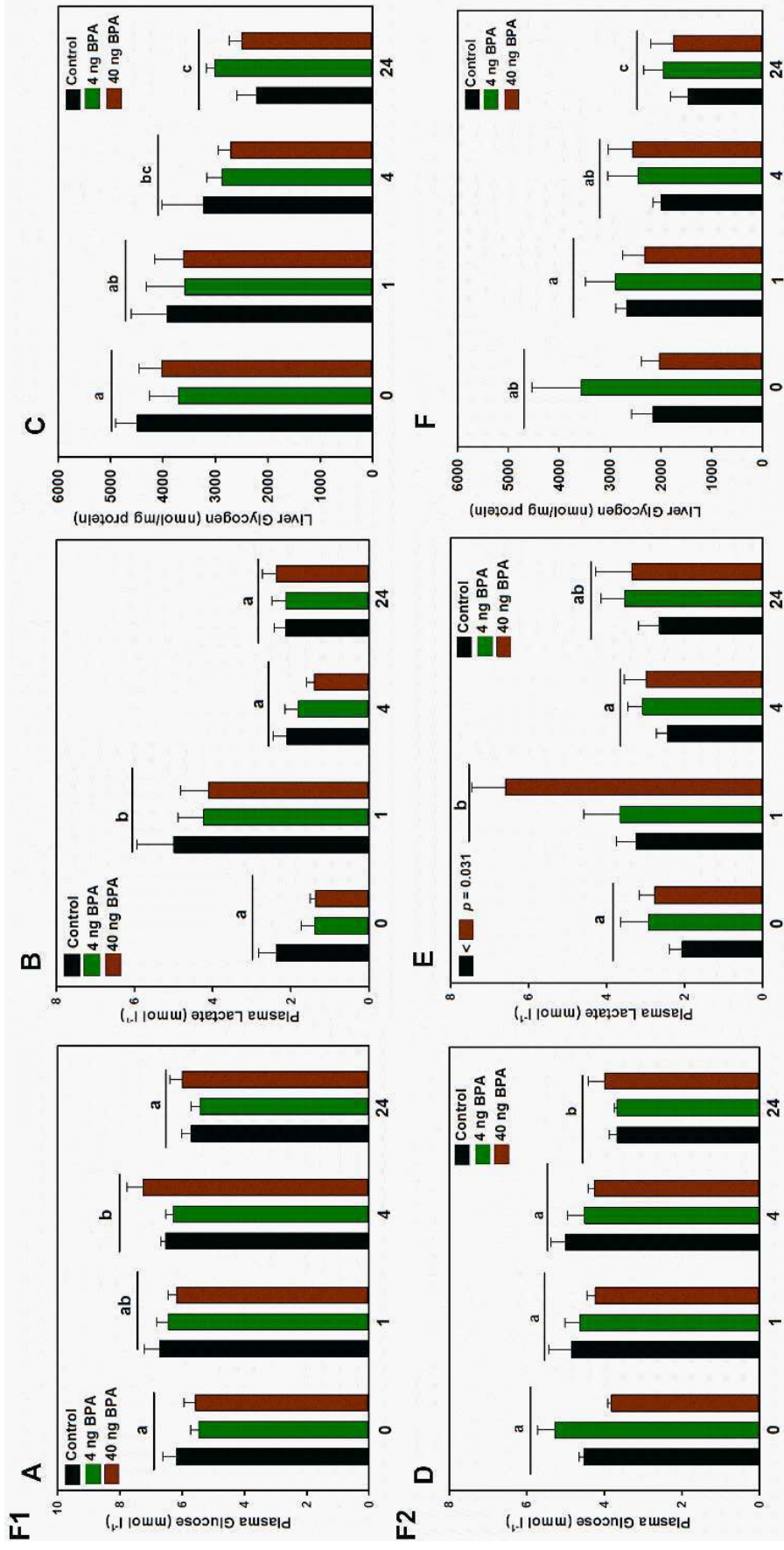
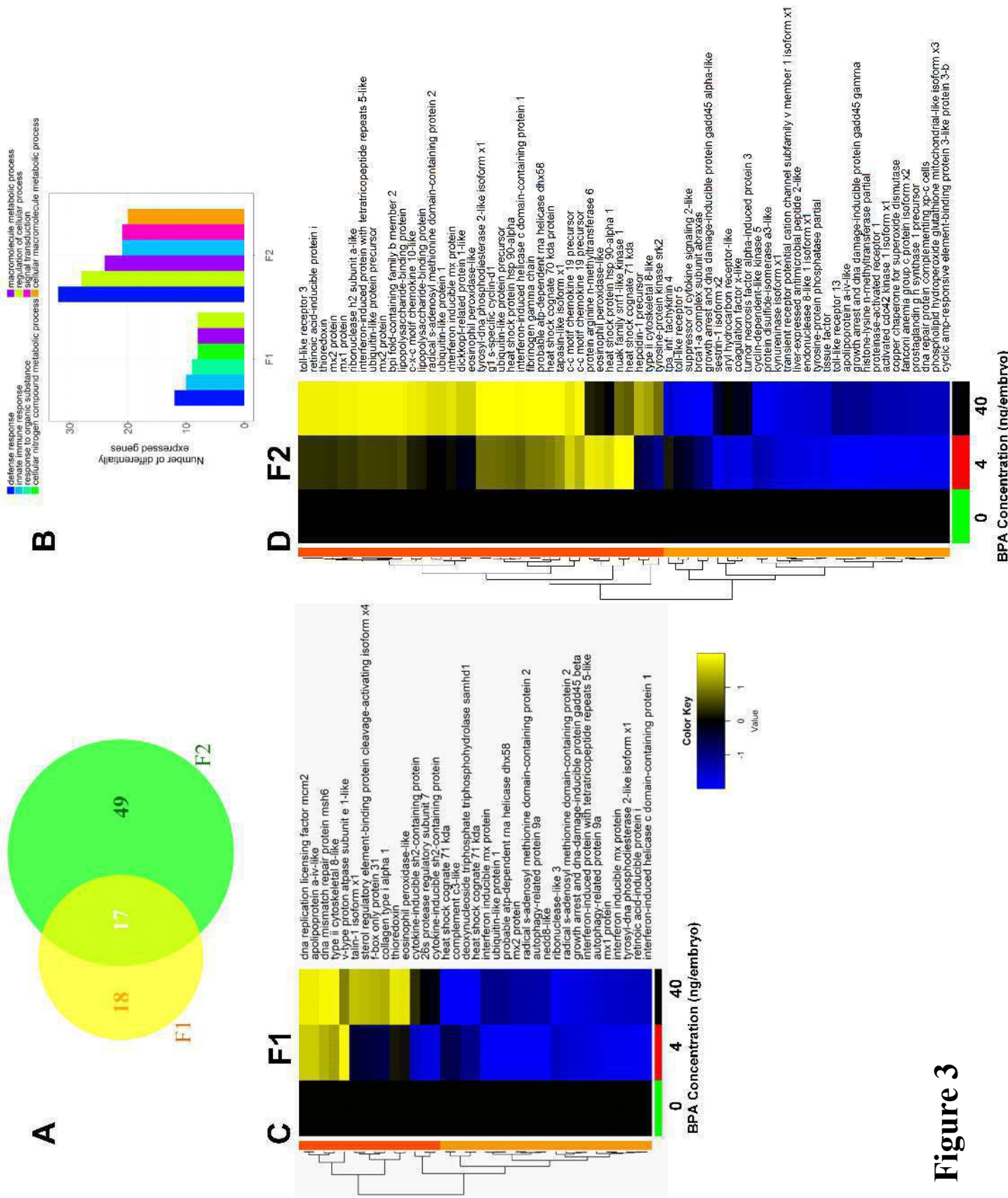


Figure 1





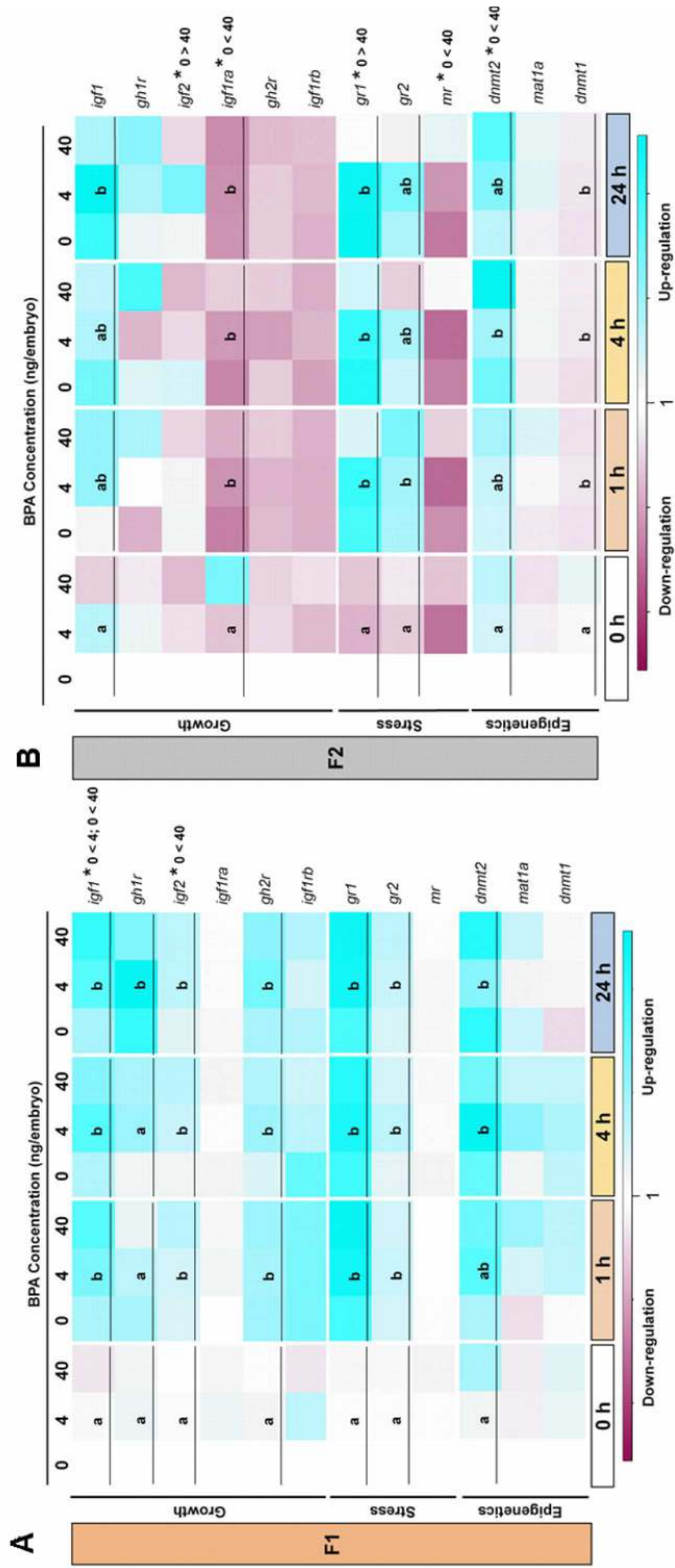


Figure 4

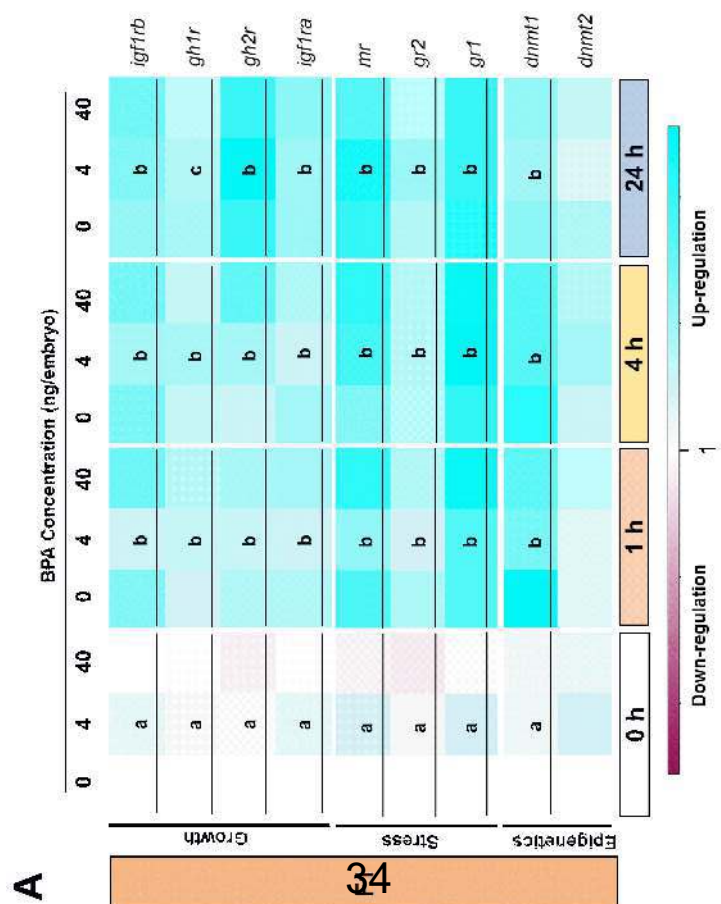
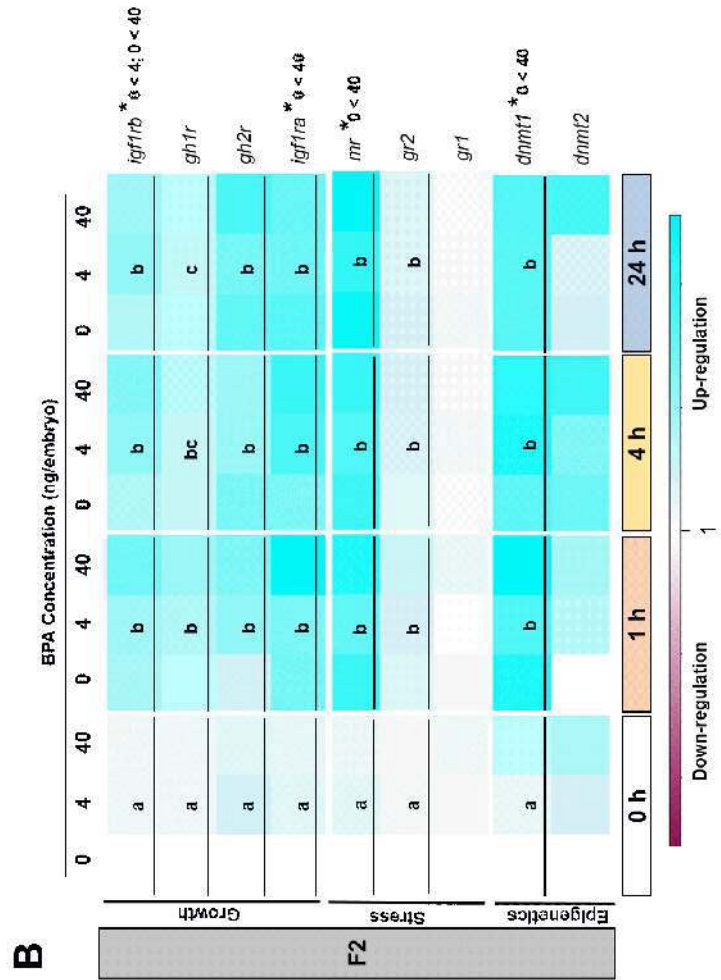


Figure 5

