



HAL
open science

Impact of irradiation on the reproductive traits of field and laboratory *An. arabiensis* mosquitoes

Serge B Poda, Edwige Guissou, Hamidou Maïga, Sévérin N Bimbile-Somda, Jérémie Gilles, Jean-Baptiste Rayaisse, Thierry Lefèvre, Olivier Roux, Roch Dabiré

► **To cite this version:**

Serge B Poda, Edwige Guissou, Hamidou Maïga, Sévérin N Bimbile-Somda, Jérémie Gilles, et al.. Impact of irradiation on the reproductive traits of field and laboratory *An. arabiensis* mosquitoes. Parasites & Vectors, 2018, 11, pp.641. 10.1186/s13071-018-3228-3 . hal-01971501

HAL Id: hal-01971501

<https://hal.umontpellier.fr/hal-01971501>

Submitted on 7 Jan 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH

Open Access



Impact of irradiation on the reproductive traits of field and laboratory *An. arabiensis* mosquitoes

Serge B. Poda^{1,2}, Edwige Guissou^{1,2}, Hamidou Maïga¹, Sévérin N. Bimbile-Somda^{1,3}, Jérémie Gilles³, Jean-Baptiste Rayaisse⁴, Thierry Lefèvre^{1,2}, Olivier Roux^{1,2*} and Roch K. Dabiré^{1†}

Abstract

Background: The sterile insect technique (SIT) aims at suppressing or decreasing insect pest populations by introducing sterile insects into wild populations. SIT requires the mass-production of insects and their sterilization through, for example, radiation. However, both mass-rearing and radiation can affect the life history traits of insects making them less competitive than their wild counterparts. In the malaria mosquito *Anopheles arabiensis*, some progress has been made to improve the mating competitiveness of mass-reared irradiated males. However, to date, no study has explored the relative effects of colonization and irradiation on important reproductive traits in this species. Such data may help to focus research efforts more precisely to improve current techniques.

Methods: Two strains of *An. arabiensis* originating from the same locality were used: one reared in the laboratory for five generations and the second collected as late larval instars in the field prior to experimentation. Pupae were irradiated with 95 Gy and some adult reproductive traits, including insemination rate, fecundity, oviposition behavior, fertility and male survivorship, were assessed in different mating combinations.

Results: Our study revealed the different effects of mosquito strain and irradiation on reproductive processes. The insemination rate was higher in field (67.3%) than in laboratory (54.9%) females and was negatively affected by both female and male irradiation (un-irradiated vs irradiated: 70.2 vs 51.3% in females; 67.7 vs 53.7% in males). Irradiated females did not produce eggs and egg prevalence was lower in the field strain (75.4%) than in the laboratory strain (83.9%). The hatching rate was higher in the field strain (88.7%) than in the laboratory strain (70.6%) as well as in un-irradiated mosquitoes (96.5%) than in irradiated ones (49%). Larval viability was higher in the field strain (96.2%) than in the laboratory strain (78.5%) and in un-irradiated mosquitoes (97.6%) than irradiated ones (52%). Finally, field males lived longer than laboratory males (25.1 vs 20.5 days, respectively).

Conclusions: Our results revealed that both irradiation and colonization alter reproductive traits. However, different developmental stages are not equally affected. It is necessary to consider as many fitness traits as possible to evaluate the efficacy of the sterile insect technique.

Keywords: Egg retention, Fecundity, Fertility, Insemination, Longevity, Malaria vector, Oviposition, Sterile insect technique

* Correspondence: olivier.roux@ird.fr

†Olivier Roux and Roch K. Dabiré contributed equally to this work.

¹Institut de Recherche en Sciences de la Santé (IRSS), Bobo-Dioulasso 01 01 BP 545, Burkina Faso

²MIVEGEC, IRD, CNRS, University of Montpellier, Montpellier, France

Full list of author information is available at the end of the article



Background

Faced with the increase in insecticide resistance, it is necessary to design new tools or improve existing methods to better control vector-borne diseases. During the past decade, there has been renewed interest in the use of the sterile insect technique (SIT) which aims to suppress or decrease insect pest populations by introducing sterile insects into wild populations. The principle is based on the insemination of wild females by sterile, laboratory-reared males resulting in non-viable progeny and hence in a reduction in pest populations [1]. To reach this goal, the SIT requires the mass production of insects, sterilization, gender separation, and the transportation and release of males only into the target areas [2, 3]. Among the techniques available or under investigation to induce sterility (see [4] for an overview), the use of radiation for SIT is recognized as the safest for human health, is environmentally friendly, species-specific, and does not require regulation for use in the field [2, 5, 6]. Moreover, it has proved its efficacy in the field on various insect pest species [1].

To be efficient, the SIT requires releasing a large number of laboratory-reared males which have to be as competitive as wild males to mate with wild females. However, irradiation and insect mass production can reduce the competitiveness of sterile males in several ways [7, 8]. First, radiation induces an oxidative stress which can affect germ cells through lethal DNA mutations. The higher the radiation dose, the higher the rate of mutation. These mutations do not lead to cell death and do not affect gamete or zygote formation but can cause the failure of embryo development [9]. Secondly, oxidative stress can directly affect somatic cells. DNA, protein and lipid oxidation alter cells, tissues and organism functioning which can lead to changes in life history traits [10]. Reduced longevity is the most evident effect of a high irradiation level but sperm production, emergence success, courtship behavior, pheromone production, and the structure of flight muscles can also be affected [7, 8]. Finally, insect colonization and mass rearing over many generations can cause the loss of natural genetic traits. Indeed, as only a fraction of field individuals survive during colonization, laboratory populations undergo genetic bottlenecks resulting in the selection of traits more adapted to insectary conditions [8, 11]. Many reproductive traits such as fecundity, courtship, oviposition, the production of and response to pheromones [11–14], but also other traits involved in mating or oviposition behavior such as flight ability, eye morphology, visual sensitivity and resistance to stress [15–17] have been described as being affected and possibly reducing overall insect vigor in the wild.

Despite recent improvements in both irradiation and laboratory mass-rearing techniques, SIT for *Anopheles* mosquito species has not reached an operational level as it has for some other pest insects [18]. *Anopheles arabiensis* is

one of the main malaria vectors in Africa. Recent studies have provided valuable information on its radiation biology including the effects of variable radiation doses [19, 20] and mosquito developmental stages that should be targeted for irradiation [20–22]. While the impact of radiation on sperm production [23, 24], male competitiveness [19, 21, 25], longevity [22], fertility and fecundity [26] was previously assessed in laboratory conditions, some field and semi-field trials investigated the impact of transportation on mortality [27], the participation of sterile males in swarms and their dispersion capacities [28, 29]. To date, however, there is little information on the relative effects of radiation and laboratory rearing on the life history traits of *An. arabiensis*. Here we compare the effect of irradiation on the reproductive traits of laboratory vs wild *An. arabiensis* from the same locality.

Methods

Mosquitoes

Both field and laboratory *An. arabiensis* mosquitoes were used and originated from Dioulassoba, a central urban area of Bobo-Dioulasso, Burkina Faso. The strain considered as “laboratory” mosquitoes was established from wild gravid *An. arabiensis* females collected indoors in 2015. Females were placed individually in oviposition cups containing tap water. After oviposition, the females were all identified as *An. arabiensis* using PCR as described by Fanello et al. [30]. The larvae were reared in tap water exposed to controlled conditions in the insectaries (27 ± 2 °C, $70 \pm 10\%$ RH, 12L:12D) and fed with Tetramin® Baby Fish Food (Tetrawerke, Melle, Germany) *ad libitum*. Females were blood-fed on rabbit for egg production and reared in insectary conditions. Experiments were conducted in 2016 with the fifth generation of the laboratory strain and with F0 individuals that emerged from wild larvae collected in 2016 (“field” strain).

Mosquito irradiation

Prior to irradiation, *ca.* 20-h-old pupae were randomly assigned to two different plastic cups ($\varnothing = 45$ mm, $h = 85$ mm) at similar densities: an irradiated group and a control group. To ensure uniform radiation dose distribution, pupal densities did not exceed 200 pupae per cup. Only 1cm of water was left to limit radiation absorbance by the water. Pupae were irradiated at a theoretical dose of 92 Gy in a Gamma Cell ^{137}Cs self-contained gamma source at a rate of about 4 Gy/min for 23 min. This dose has been shown to allow a good sterility-competitiveness trade-off in *An. arabiensis* males [22]. The cups were placed at the centre of the chamber to maximize dose uniformity within the batch. A dosimetry system was used to measure the accurate dose received by each batch using Gafchromic® HD-V2 film (Ashland, Bridgewater, NJ, USA) placed on the walls of the cups. After irradiation, the

optical density of the irradiated films was read at both 458 and 590 nm with a dose reader (Dosereader4, Radgen, Budapest, Hungary) and compared to a control. The dose effectively measured was 95.4 ± 0.9 Gy (mean \pm standard error, SE). The control group was handled similarly but was not irradiated. Following irradiation, irradiated and control pupae were placed in $30 \times 30 \times 30$ cm mesh-covered cages separately and kept under standard insectary conditions (27 ± 2 °C; $70 \pm 10\%$ RH; 12L:12D photoperiod). The next day, emerged mosquitoes were separated by gender early in the morning and males and females were kept in separate $30 \times 30 \times 30$ cm cages to prevent mating. They were maintained under insectary conditions and provided with a 5% glucose solution. The number of dead pupae was recorded. Three to four batches of mosquitoes were irradiated on consecutive days for each replicate in order to obtain enough mosquitoes for experiments.

Mosquito traits

A series of mosquito traits playing key roles in determining sexual competitiveness was measured, namely insemination rate, fecundity, fertility and longevity. The insemination rate was defined as the proportion of females with spermatozoa-positive spermathecae. Fecundity was defined as the capacity of females to produce eggs and fertility as their capacity to produce viable offspring. Longevity was considered as the lifespan from the first day in mating cages to death. To gauge the effect of irradiation on these traits, 2-day-old males and females (un-irradiated and irradiated) were assorted to obtain four mating combinations for each mosquito strain (laboratory *vs* field) as follows: both un-irradiated males and females (un-irradiated pairs, hereafter), irradiated males and un-irradiated females, un-irradiated males and irradiated females and both irradiated males and females (irradiated pairs, hereafter) (Additional file 1: Table S1). Males and females were kept together in $20 \times 20 \times 20$ cm cages and provided with a 5% glucose solution for two nights. For each mating combination, two to three cages were used. A mating ratio of two males for one female was used with 40 to 80 males per cage.

On the second night, all of the females were removed from the cages and put into cardboard cups ($\varnothing = 75$ mm, h = 100 mm) for fecundity and fertility assays. The males were kept in their cages and provided with a 2.5% glucose solution every other day and water *ad libitum* every day. Male longevity was assessed by counting and then removing dead individuals every 24 h. To gauge the effect of irradiation on fecundity and fertility, females were provided with two blood meals on a rabbit at two-day intervals. Females that had not fed were discarded after each blood meal. Blood-fed females were kept in the cardboard cups and provided with a 5%

glucose solution. Three days after the second blood meal, the females were placed individually in plastic oviposition cups ($\varnothing = 45$ mm, h = 85mm) containing tap water. Following egg laying, the spermatheca was dissected and the insemination status assessed under a microscope at 400 \times magnification to observe the presence/absence of spermatozoa. Females that did not lay eggs after 10 days were killed and their spermatheca dissected. The ovaries of all of the females were also dissected and the presence/absence of eggs as well as the number of unlayed eggs, if any, were recorded. Laid eggs were kept in plastic cups containing water for one week to allow them to hatch and the presence/absence of viable larvae was recorded. Then, hatched and unhatched eggs were counted to assess fertility. For both field and laboratory mosquitoes, two replicates were performed.

Statistical analysis

All analyses were conducted in R v.2.12.1. The effect of irradiation and mosquito strain on a series of mosquito traits was analyzed using different statistical models summarized in Additional file 1: Table S2. These traits were the emergence rate of adult mosquitoes, female insemination rate (the contents of the spermatheca, i.e. presence/absence of spermatozoa), egg prevalence (the proportion of females that developed eggs in their ovaries), egg load (the number of eggs in gravid females), oviposition rate (the proportion of females that laid at least one egg), laid egg proportion (the number of eggs laid out of the total number of eggs produced), hatching rate (the proportion of egg batches in which at least one egg hatched), hatching proportion (the proportion of eggs that hatched within each egg batch), viable larvae prevalence (the proportion of egg batches in which at least one larva survived more than 3 days) and male survivorship (the length of time between the first day in mating cages and death). The general building of the statistical models was as follows. Mosquito strain (laboratory and field), treatment (irradiated and un-irradiated), insemination status (inseminated and non-inseminated) and their interactions were considered fixed effects. Plastic cups and mosquito cages were considered random effects.

For model selection, we used the stepwise removal of terms, followed by likelihood ratio tests. Term removals that significantly reduced explanatory power ($P < 0.05$) were retained in the minimal adequate model [31].

Results

Emergence rate

A total of 2420 and 3000 pupae were obtained from the field and the laboratory populations, respectively. Half of each group was irradiated (i.e. 1210 and 1500 pupae, respectively); the other halves were used as un-irradiated controls. The mean emergence rate (\pm 95% CI) was $89.5 \pm 1.2\%$

and was not affected by irradiation (un-irradiated: $89.9 \pm 1.7\%$ vs irradiated: $89 \pm 1.8\%$), mosquito strain (field: $90.8 \pm 1.3\%$ vs laboratory: $87.6 \pm 2.2\%$) or their interaction (Table 1).

Insemination rate

A significant negative effect of male and female irradiation on insemination was found (un-irradiated and irradiated: $67.7 \pm 5.2\%$ vs $53.7 \pm 6.1\%$ in males and $70.2 \pm 6\%$ vs $51.3 \pm 5.2\%$ in females). The insemination rate was significantly higher in field females than in laboratory females ($67.3 \pm 5.4\%$ vs $54.9 \pm 5.9\%$). There was no interaction between irradiation and mosquito strain (Fig. 1 and Table 1).

Egg prevalence

Egg prevalence was defined as females that had at least one egg in their ovaries or that had laid at least one egg after two blood meals. We found a significant negative effect of irradiation on females with irradiated females never producing eggs (Fig. 2). The analysis conducted on un-irradiated females only showed that there was a significant effect of female insemination status on egg prevalence with non-inseminated females being less likely to produce eggs than inseminated females ($61.3 \pm 10.1\%$ vs $87 \pm 4.5\%$, respectively; Fig. 2, Table 1). There was a significant effect of mosquito strain on egg prevalence with un-irradiated field females being less likely to produce eggs than un-irradiated laboratory females ($75.4 \pm 6.6\%$ vs $83.9 \pm 6.1\%$, respectively; Fig. 2, Table 1). There was no interaction between irradiation and mosquito strain (Fig. 2, Table 1) or between strain and insemination status (Table 1).

Egg load

In un-irradiated gravid females, the mean egg load (\pm SE) (i.e. the number of laid eggs and/or eggs retained in the ovaries) was not affected by male irradiation (un-irradiated: 98.7 ± 3.9 vs irradiated: 91.3 ± 4.3 eggs) or by mosquito strain (field: 98.5 ± 3.7 vs laboratory: 92.7 ± 4.5 eggs; Fig. 3, Table 1). Egg load was, however, affected by insemination status with inseminated females having significantly more eggs than non-inseminated ones (99.6 ± 3.2 vs 82.5 ± 6.5 eggs, respectively; Table 1, Additional file 1: Figure S1). There was no interaction between irradiation status and mosquito strain (Fig. 3, Table 1), irradiation status and insemination status or between mosquito strain and insemination status (Table 1).

Oviposition rate

The oviposition rate (i.e. females that had laid at least one egg) was affected by insemination status, with non-inseminated females never laying eggs (Additional file 1: Figure S2). As non-inseminated and irradiated females never laid eggs, we analyzed the oviposition rate on un-irradiated gravid and inseminated females only.

Insemination by irradiated males had no effect on the oviposition rate (un-irradiated: $67.7 \pm 8.1\%$ vs irradiated: $62.9 \pm 10.5\%$; Fig. 4a, Table 1). However, laboratory females were more likely to lay eggs than were their field counterparts ($82.4 \pm 7.8\%$ vs $52.9 \pm 9\%$, respectively; Fig. 4a, Table 1). There was no interaction between male irradiation status and mosquito strain (Fig. 4a, Table 1).

Laid egg proportion

Gravid females did not lay their eggs all at once. The proportion of eggs laid per female was significantly affected by mosquito strain with field females retaining eggs more frequently than laboratory females (proportion of eggs laid: $60.1 \pm 7.1\%$ vs $83.7 \pm 5.3\%$, respectively; Fig. 4b, Table 1). Insemination by irradiated males had no effect on egg retention (un-irradiated: $74.8 \pm 6.3\%$ vs irradiated: $65.6 \pm 6.9\%$; Fig. 4b, Table 1). There was no interaction between male irradiation and mosquito strain (Fig 4b, Table 1).

Hatching rate

Hatching rate (i.e. presence of at least one hatched egg in the batches) was negatively affected by insemination by an irradiated male (un-irradiated: $96.5 \pm 3.8\%$ vs irradiated: $49 \pm 13.7\%$; Fig. 5a, Table 1) and by mosquito strain with field mosquitoes having better hatching success than their laboratory counterparts (field: $88.7 \pm 7.8\%$ vs laboratory: $70.6 \pm 10.3\%$; Fig. 5a, Table 1). There was no interaction between male irradiation and mosquito strain (Fig. 5a, Table 1).

Hatching proportion

To gauge more precisely the reproduction potential of partially sterilized males, we also considered the proportion of hatching within egg batches (i.e. the number of hatched eggs out of the total number of eggs laid per batch). Hatching proportion was negatively affected by male irradiation (un-irradiated: $76.1 \pm 8\%$ vs irradiated: $14.3 \pm 6.6\%$) and eggs from laboratory mosquitoes hatched more frequently than those from field mosquitoes (field: $54 \pm 9.3\%$ vs laboratory: $70 \pm 8.6\%$; Fig. 5b, Table 1). There was no interaction between irradiation and mosquito strain (Fig. 5b, Table 1).

Larval prevalence

Male irradiation also reduced the prevalence of viable larvae (fertility) (un-irradiated: $97.6 \pm 3.2\%$ vs irradiated: $52 \pm 19.5\%$). Larval viability was also higher in laboratory than in field mosquitoes (field: $78.5 \pm 10.7\%$ vs laboratory: $96.2 \pm 5.1\%$; Fig. 6, Table 1). There was no interaction between male irradiation and mosquito strain (Fig. 6, Table 1).

Table 1 Effects of mosquito strain, irradiation and insemination status on reproductive traits and survivorship

Factors	χ^2	df	P-value
Emergence rate			
Strain	1.90	1	0.167
Treatment	0.32	1	0.568
Strain: Treatment	0.07	1	0.784
Insemination rate			
Strain	7.83	1	0.005
Male treatment	9.69	1	0.001
Female treatment	18.16	1	<0.001
Strain: Male treatment	1.45	1	1.227
Strain: Female treatment	0.93	1	0.334
Egg prevalence			
Strain	5.45	1	0.019
Treatment ^a	0.25	1	0.611
Insemination status	24.54	1	<0.001
Strain: Treatment	0.00	1	0.994
Strain: Insemination status	1.88	1	0.170
Egg load			
Strain	0.48	1	0.480
Treatment	0.96	1	0.327
Insemination status	5.42	1	0.019
Strain: Treatment	1.36	1	0.243
Strain: Insemination status	0.04	1	0.841
Treatment: Insemination status	2.58	1	0.108
Oviposition rate			
Strain	9.70	1	0.001
Treatment	0.56	1	0.452
Strain: Treatment	0.02	1	0.879
Laid egg proportion			
Strain	4.68	1	0.030
Treatment	1.41	1	0.233
Strain: Treatment	0.16	1	0.688
Hatching rate			
Strain	5.10	1	0.023
Treatment	17.11	1	<0.001
Strain: Treatment	0.98	1	0.321
Hatching proportion			
Strain	5.44	1	0.019
Treatment	64.36	1	<0.001
Strain: Treatment	1.79	1	0.180
Larval prevalence			
Strain	4.83	1	0.027
Treatment	12.35	1	<0.001
Strain: Treatment	1.66	1	0.196

Table 1 Effects of mosquito strain, irradiation and insemination status on reproductive traits and survivorship (Continued)

Factors	χ^2	df	P-value
Survivorship			
Strain	8.21	1	0.004
Treatment	0.54	1	0.458
Strain: Treatment	0.08	1	0.772

Strain (field or laboratory), treatment (un-irradiated or irradiated) and insemination status (inseminated or not)

^aAs irradiated females did not produce eggs, only the treatment of males was considered (starting with egg prevalence and below), bold values are significant

Longevity

Male survivorship was not influenced by irradiation (un-irradiated: 22.8 ± 0.3 vs irradiated: 22.3 ± 0.3 days). However, field males lived longer than those reared in the laboratory (field: 25.1 ± 0.3 vs laboratory: 20.5 ± 0.3 days; Fig. 7, Table 1). There was no interaction between irradiation and mosquito strain (Fig. 7, Table 1).

Figure 8 shows an attempt to estimate the theoretical “realized fertility” rates in females based on our results. It shows that mating pairs including irradiated females had a null fertility rate due to the absence of egg production. In the un-irradiated pairs, field mosquitoes were 1.7 times less likely to produce viable offspring compared to laboratory mosquitoes with a greater probability of failing at both developing and laying eggs. Overall, according to Abbott’s formula [32], the percentage of radiation-induced sterility was about 80%.

Discussion

Insemination rate

The insemination rate was reduced by both radiation and rearing. The dose of radiation used in this study has been determined to be acceptable for a good sterility-competitiveness trade-off in *An. arabiensis* [22, 33]. The insemination rates were similar to those obtained by Helinski et al. [27] in large cages with a lower irradiation dose (70 Gy). Our irradiated field mosquitoes had a higher insemination rate than did their laboratory counterparts, suggesting that laboratory mass rearing might be an important factor affecting the way mosquitoes cope with irradiation effects when it comes to mating [34]. As for *An. albimanus*, irradiated pairs had the lowest insemination rate compared to other mating pairs [35] suggesting that both sexes are affected by irradiation and that both sexes are active in the mating process. Mating behavior, which leads to female insemination, occurs in swarms that consist mainly of males. Virgin females are known to join these swarms to form mating copulae in mid-flight [36]. Swarming behavior is very challenging for both males and females as the competition for a mate is strong. Males are numerous and they wait for females while

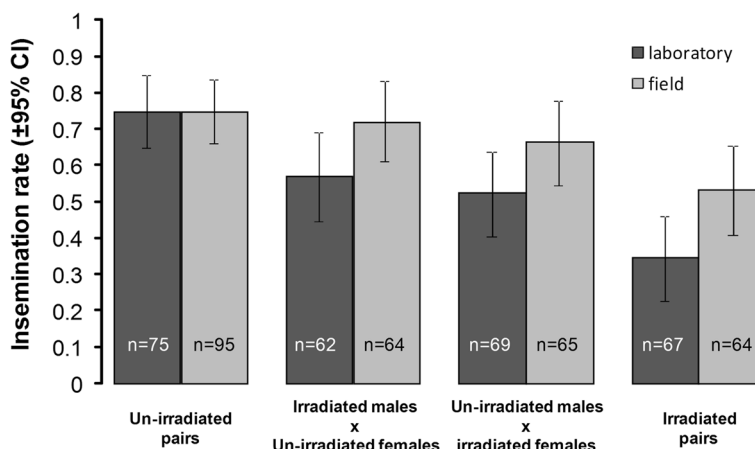


Fig. 1 Effects of mosquito strain and irradiation on female insemination rate. The female insemination rate was assessed after two mating nights for the four mating pair combinations. The numbers in the bars indicate sample size

swarming and sometimes chase each other inside the swarm. Once a female enters into the swarm, recognition between partners and probably the courtship behavior are based on the alteration of male and female wing beat frequencies until they converge into the same harmonic frequency [37]. To perform competitively, both males and females need high energy reserves as well as good flight and hearing abilities. Mosquitoes reared in the laboratory are not under high selective pressures compared to wild mosquitoes. Consequently, some of the laboratory mosquitoes, those that would have never survived or mated in the wild, may not possess enough reserves to compete for females in this very demanding behavior [38, 39]. Moreover, flight ability is also controlled by the thoracic muscles which are formed during metamorphosis. Pupae irradiation, as well as laboratory rearing, have been shown to alter flight muscles in numerous insects ([8] and references therein).

In irradiated mosquitoes, the radiation may have altered their flight muscles so that they are less able to reach the wing beat that coincides with the same harmonic frequency as their partner and, as such, may be ignored during courtship thus preventing insemination. In addition, laboratory rearing does not select for flight capacity and this behavior may not be an advantage in small cages. Pheromone production (if any in *Anopheles* species [40]), which may be responsible for attractiveness and acceptance between the two sexes, might also be counter-selected or altered in both quality and quantity [8]. Similarly, hearing capabilities can be altered by irradiation. The Johnston’s organ located at the base of the antennae, which amplifies the air-borne vibrations produced by wing beats, may be affected by irradiation. No study has yet demonstrated this but some studies have shown that irradiation and mass rearing alter the quality of vision in fruit flies [16] and in New World

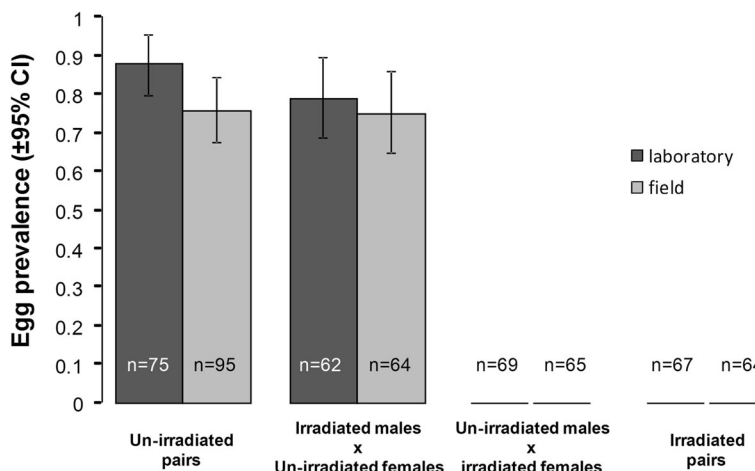


Fig. 2 Effects of mosquito strain and irradiation on egg prevalence. Egg prevalence is assessed through females that had at least one egg in their ovaries or laid at least one egg. The numbers in the bars indicate sample size

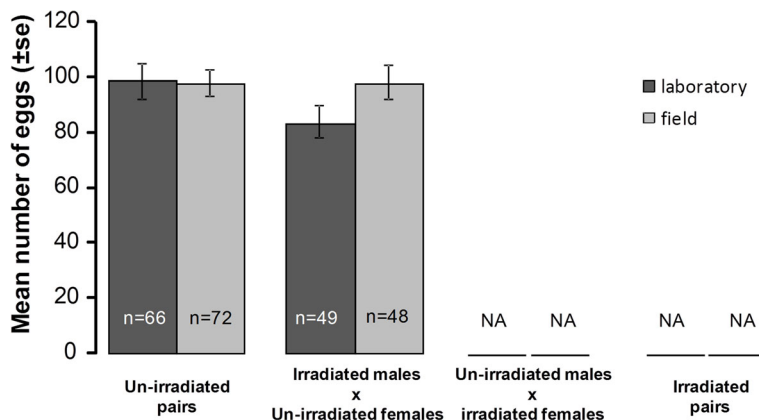


Fig. 3 Effects of mosquito strain and irradiation on mean number of eggs in un-irradiated, gravid females. The number of eggs is the sum of eggs laid or retained in the ovaries per female. The numbers in the bars indicate sample size. NA: no data as irradiated females did not produce eggs

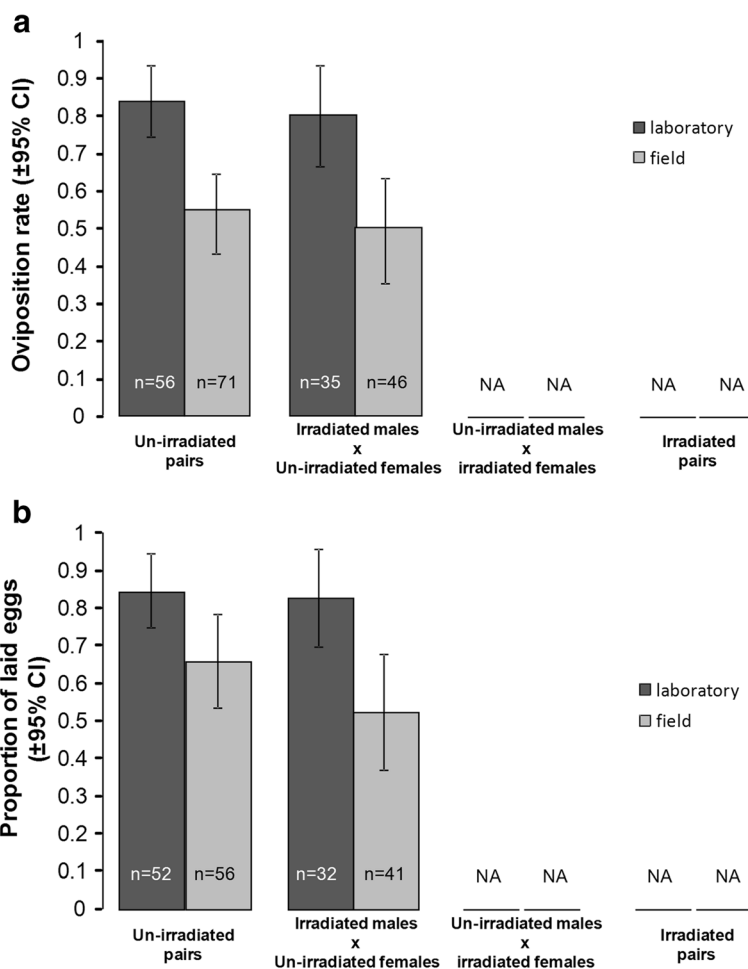


Fig. 4 Effects of mosquito strain and male irradiation on un-irradiated female oviposition behavior: **a** oviposition rate (i.e. females that laid at least one egg) and **b** proportion of eggs laid per female [number of eggs laid out of the total number of eggs produced (laid eggs + retained eggs) per female]. The numbers in the bars indicate sample size. NA: no data as irradiated females did not produce eggs

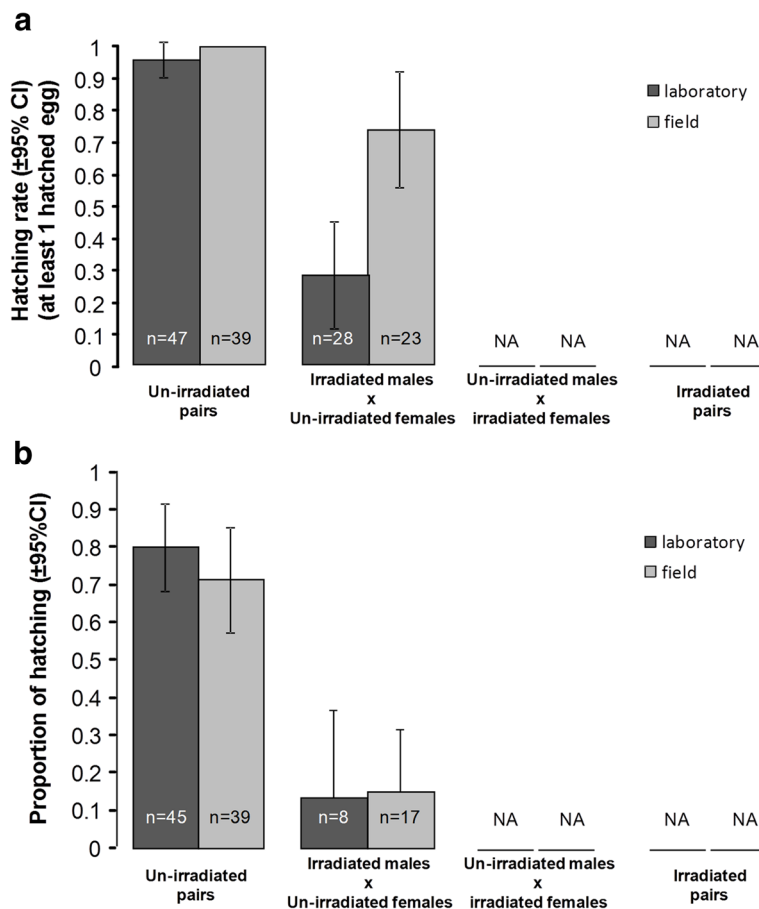


Fig. 5 Effects of mosquito strain and male irradiation on egg hatching. **a** Hatching rate per egg batch (i.e. presence of at least one hatched egg in the batch). **b** Hatching proportion within egg batches (i.e. number of hatched eggs over the total number of eggs laid per batch). The numbers in the bars indicate sample size. NA: no data as irradiated females did not produce eggs

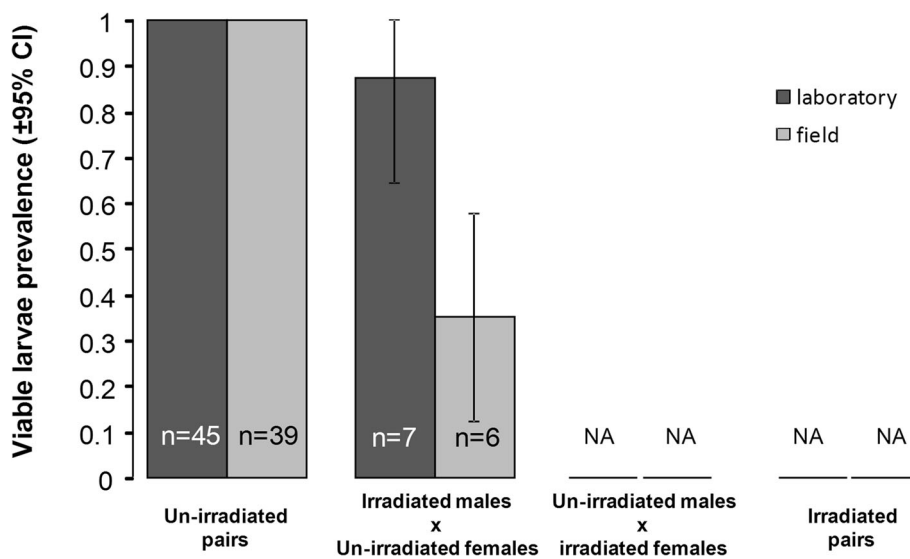
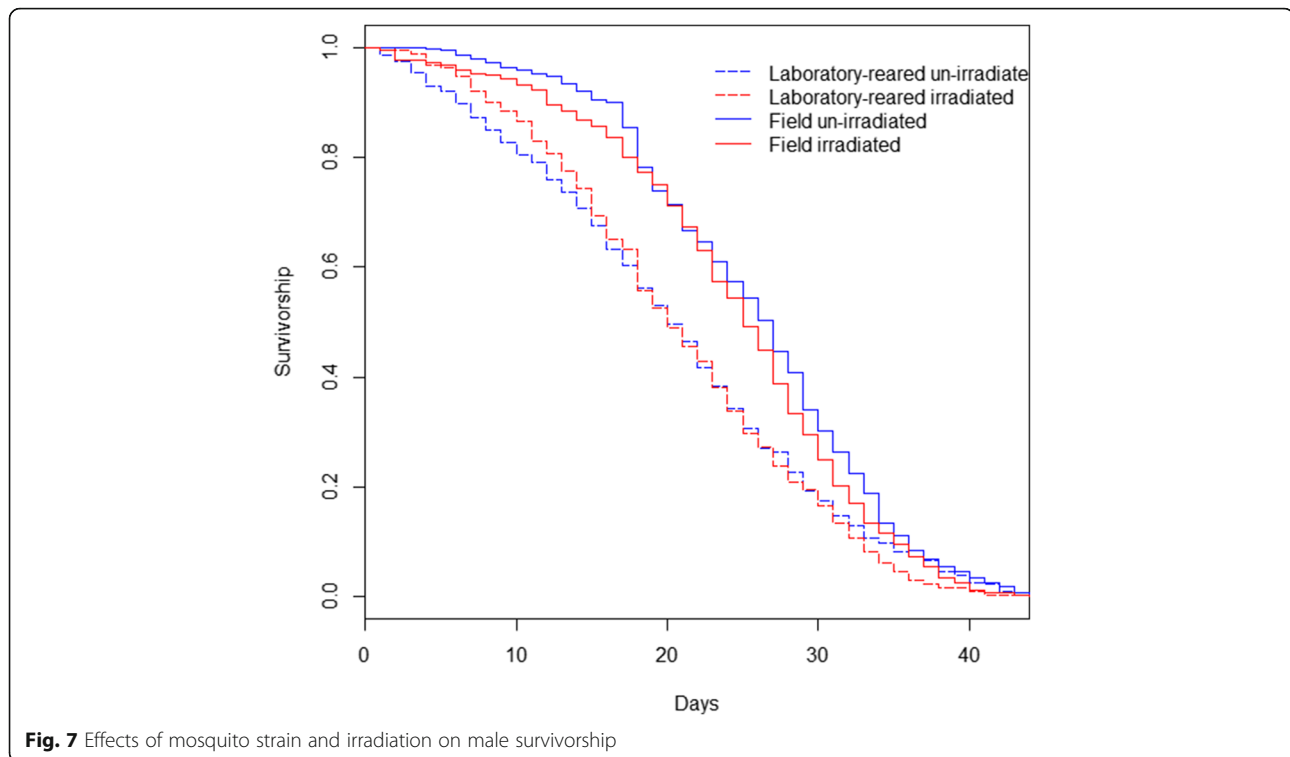


Fig. 6 Effects of mosquito strain and male irradiation on first-instar larva viability. The numbers in the bars indicate sample size. NA: no data as irradiated females did not produce eggs



screwworms [17]. Consequently, it can be expected that other finely tuned senses may also be affected. Sperm production is also negatively affected by irradiation and may be a reason for a lower insemination rate [23, 24]. Altogether, the alteration of energy reserves, flight muscles, hearing capability, and both pheromone and sperm production, may be responsible for the drop in mating efficiency and for lower insemination rates.

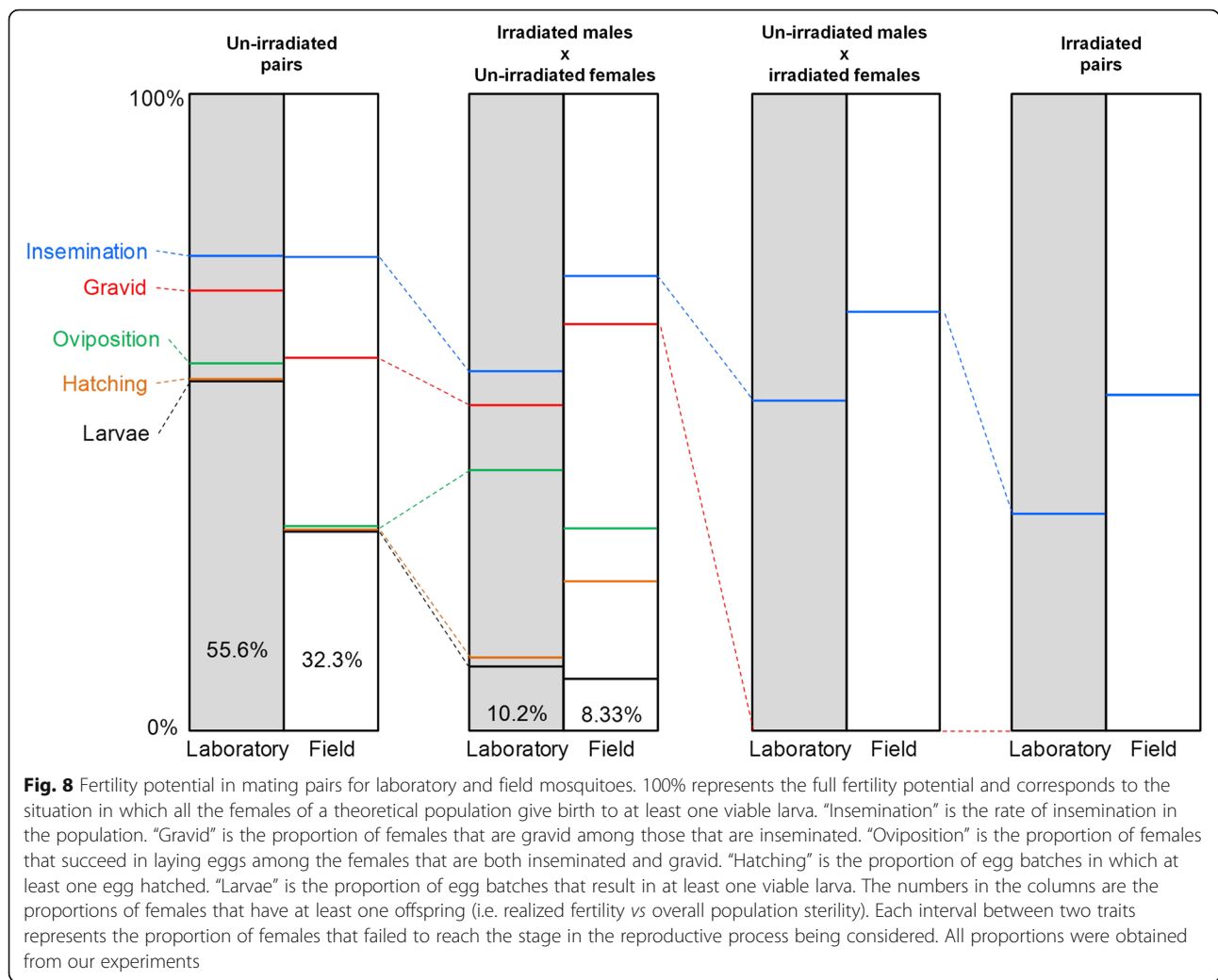
Egg prevalence and egg load

Fecundity was affected in several ways by both irradiation and laboratory rearing. First, irradiated females never produced eggs which is a common observation in irradiated female arthropods and was already observed in *An. arabiensis* [41]. Females are generally more radiosensitive than are males because of the later development of oocytes. Consequently, during irradiation, oocytes that have not yet produced eggs develop lethal mutations which inhibit egg development [42]. An exposition of 20-h-old pupae to a dose of 95Gy resulted in complete female sterilization. Second, field mosquitoes were slightly less likely to develop eggs. Here, differences between optimal rearing conditions and field larval site conditions may be responsible for this difference in female fecundity. Indeed, it has been shown that larval stresses have carry-over effects on adult fertility [43–45]. For example, stresses that affect food intake can impact the teneral reserves acquired during larval development. Teneral reserves are important for the production of the first egg batch in adults. Females that were well fed as

larvae will need only one blood meal to produce their first egg batch while those with low teneral reserves will need two to three blood meals [46] which impacts fecundity during the first gonotrophic cycle. Finally, male irradiation also had an indirect negative effect on female fecundity. Indeed, irradiated males were less likely to inseminate females and non-inseminated females are known to be less fecund than inseminated ones ([47–49]; present study). This higher fecundity in inseminated females is triggered by the transfer of sex peptides and hormones from male accessory glands to the female atrium [47]. Nevertheless, as there was no difference in egg load and egg prevalence between un-irradiated inseminated females mated with irradiated or un-irradiated males, we can speculate that the production and transfer of these sex compounds are not directly affected by irradiation.

Oviposition

Insemination by irradiated males did not affect oviposition. However, the mosquito strain had an impact on their decision to lay eggs. Indeed, while most of our laboratory females tended to lay their eggs all at once, field mosquitoes retained their eggs more frequently or did not lay at all. For organisms without parental care and with limited dispersal ability, oviposition site selection is a critical choice for reproductive success [50]. For mosquitoes, factors including larval competition, food quantity, predation or desiccation risk may be responsible for delayed oviposition or egg retention until more suitable conditions



can be found [51–53]. Consequently, wild females are selected to be choosy thereby providing the best growing place for their progeny while laboratory females have been selected to oviposit on artificial substrates or in small containers. Thus, in our laboratory conditions, field females may be reluctant to oviposit in tap water, which is poor in nutrients, and retained their eggs more frequently than their laboratory counterparts.

Hatching and larva viability

Only about 50% of the control females were fully sterilized when mated with irradiated males (no hatching). Nevertheless, within fertile female batches, only 15% of the eggs hatched. The radiation-induced sterility was about 80% which is very similar to results found in a previous study using the same radiation dose [22]. In addition, among females that produced fertile egg batches, only 50% of the batches produced at least one viable first-instar larva. This shows that induced mutations may be lethal at any developmental stages and, thus, that such studies should

consider the entire larval development cycle until adult emergence to provide a more realistic picture of induced sterility [42]. To date, for mosquitoes, most interpretations are based on egg hatchability and thus probably underestimate the real efficiency of partially sterilizing radiation doses. A follow-up of larval mortality may help to optimize the competitiveness-sterility trade-off in irradiated males. Finally, no interaction between mosquito origin and radiation treatment was found for hatching rate or for larvae viability which means that neither of the two mosquito origins was better at coping with radiation than the other.

Longevity

The effect of some stresses can be amplified when associated with a diet-based stress but even in our stressful design in which glucose was available only every other day, irradiation did not affect male longevity [3, 43]. However, field males survived longer than laboratory males. In the wild, the longer a male survives, the more it can mate. Longer longevity is therefore selected for. Conversely, laboratory-

reared mosquitoes experienced different selective pressures which may keep weak males in the population. Therefore our results may be a consequence of laboratory rearing in which laboratory population longevity is impeded by the artificial selection of some weak males.

Our study includes two limitations. First, our laboratory mosquitoes were recently colonized (five generations) and we can speculate that larger differences may be expected with older colonies. Secondly, our experiments were conducted in small cages (insemination) and small vials (artificial substrate for oviposition) which are more favorable to laboratory-reared mosquitoes compared to field mosquitoes and make it difficult to extrapolate as to how laboratory mosquitoes will respond once released into the wild.

Conclusions

Overall, our study reveals the different impacts of mosquito strain and irradiation on reproductive processes. The different steps in progeny production, from insemination to larval survivorship, were not affected to the same magnitude or in the same way for laboratory and wild mosquitoes. Despite these differences, the final potential reproductive success of both irradiated, laboratory-reared and irradiated, field mosquitoes were similar (10.2 vs 8.33%, respectively; Fig. 8). This highlights the need to consider as many fitness traits as possible when evaluating the efficiency of the sterile insect technique. As parental effects may also play a role in the reproductive success of the progeny, such studies should be extended to examine progeny fecundity and fertility. Finally, irradiated females never produced eggs. If irradiated females are released they will not contribute to maintaining population density. However, they may still be potential vectors of malaria. Because current methods of sexing are not fully efficient [54], further studies are needed to determine if irradiated females are competent vectors.

Additional files

Additional file 1: Figure S1. Effect of insemination on egg number in un-irradiated females. The number of eggs is the sum of eggs laid or retained in the ovaries per female. The numbers in the bars indicate sample size. * $P < 0.05$. **Figure S2.** Effect of insemination on oviposition rate in un-irradiated females (i.e. females that laid at least one egg). The numbers in the bars indicate sample size. *** $P < 0.001$. **Table S1.** Mating combinations. **Table S2.** Statistical models used in data analyses. (DOCX 54 kb)

Abbreviations

L:D: Light:dark; PCR: Polymerase chain reaction; RH: Relative humidity; SIT: Sterile insect technique

Acknowledgements

We are grateful to Fulgence Da for mosquito rearing, to Hervé Janvier Tapsoba for his help with the experiments and to the team of the Centre International de Recherche-Développement sur l'élevage en zone sub-

humide Bobo-Dioulasso (CIRDES) for assistance and providing access to an irradiator. We would also like to thank Andrea Yockey-Dejean for proofreading the paper.

Funding

Financial support for this study was provided by a grant from the IAEA to RKD (RFQ23855). The funder had no role in study design, data collection or analysis, preparation of the manuscript or the decision to publish it.

Availability of data and materials

The datasets supporting the conclusions of this article are included in the article and its additional files. The raw datasets are available from the corresponding author upon reasonable request.

Authors' contributions

OR and TL conceived and designed the study. SP and OR drafted the manuscript. OR analyzed the data. SP and EG conducted the experiments. JBR provided access to irradiation facilities. OR and RKD supervised the study. All authors read, revised and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Institut de Recherche en Sciences de la Santé (IRSS), Bobo-Dioulasso 01 01 BP 545, Burkina Faso. ²MIVEGEC, IRD, CNRS, University of Montpellier, Montpellier, France. ³Joint FAO/IAEA Division of Nuclear Techniques in Food and Agriculture, Insect Pest Control Laboratory, Vienna, Austria. ⁴Centre International de Recherche-Développement sur l'élevage en zone Subhumide (CIRDES), Bobo-Dioulasso 01 01 BP 454, Burkina Faso.

Received: 8 June 2018 Accepted: 23 November 2018

Published online: 17 December 2018

References

- Dyck VA, Hendrichs J, Robinson AS. Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management. Dordrecht, Netherlands: Springer; 2005.
- Knipling EF. Sterile-male method of population control. *Science*. 1959;130: 902–4.
- Helinski ME, Parker AG, Knols BGJ. Radiation biology of mosquitoes. *Malaria J*. 2009;8:S6.
- Bourtzis K, Lees RS, Hendrichs J, Vreysen MJB. More than one rabbit out of the hat: radiation, transgenic and symbiont-based approaches for sustainable management of mosquito and tsetse fly populations. *Acta Trop*. 2016;157:115–30.
- Lees RS, Gilles JRL, Hendrichs J, Vreysen MJB, Bourtzis K. Back to the future: the sterile insect technique against mosquito disease vectors. *Curr Opin Insect Sci*. 2015;10:156–62.
- Oliva CF, Vreysen MJB, Dupé S, Lees RS, Gilles JRL, Gouagna L-C, et al. Current status and future challenges for controlling malaria with the sterile insect technique: technical and social perspectives. *Acta Trop*. 2014; 132(Suppl.):S130–S159.
- Perez-Staples D, Shelly TE, Yuval B. Female mating failure and the failure of 'mating' in sterile insect programs. *Entomol Exp App*. 2013;146:66–78.
- Calkins CO, Parker AG. Sterile insect quality. In: Dyck VA, Hendrichs J, Robinson AS, editors. Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management. Dordrecht, Netherlands: Springer; 2005. p. 269–296.
- LaChance LE. The induction of dominant lethal mutations in insects by ionizing radiation and chemicals - as related to the sterile male technique

- of insect control. In: Wright JW, Pal R, editors. Genetics of Insect Vectors of Disease. Amsterdam: Elsevier; 1967. p. 617–50.
10. Monaghan P, Metcalfe NB, Torres R. Oxidative stress as a mediator of life history trade-offs: mechanisms, measurements and interpretation. *Ecol Lett*. 2009;12:75–92.
 11. Baeshen R, Ekechukwu NE, Toure M, Paton D, Coulibaly M, Traoré SF, et al. Differential effects of inbreeding and selection on male reproductive phenotype associated with the colonization and laboratory maintenance of *Anopheles gambiae*. *Malar J*. 2014;13:19.
 12. Cayol JP. Changes in sexual behavior and life history traits of tephritid species caused by mass-rearing processes. In: Aluja M, Norrbom AL, editors. Fruit Flies (Tephritidae): Phylogeny and Evolution of Behavior. Boca Raton, FL: CRC Press; 2000. p. 843–60.
 13. Miyatake T, Shimizu T. Genetic correlations between life-history and behavioural traits can cause reproductive isolation. *Evolution*. 1999;53:201–8.
 14. Minks AK. Decreased sex pheromone production in an in-bred stock of the summer fruit tortrix moth *Adoxophyes orana*. *Entomol Exp App*. 1971;14:361–4.
 15. Nelson RL, Milby MM. Dispersal and survival of field and laboratory strains of *Culex tarsalis* (Diptera: Culicidae). *J Med Entomol*. 1980;17:146–50.
 16. Agee HR, Park ML. Use of the electroretinogram to measure the quality of vision of the fruit fly. *Envir Lett*. 1975;10:171–6.
 17. Bush GL, Neck R, Kitto GB. Screwworm eradication: inadvertent selection for noncompetitive ecotypes during mass rearing. *Science*. 1976;193:491–3.
 18. Benelli G. Research in mosquito control: current challenges for a brighter future. *Parasitol Res*. 2015;114:2801–5.
 19. Helinski MEH, Knols BGJ. Mating competitiveness of male *Anopheles arabiensis* mosquitoes irradiated with a partially or fully sterilizing dose in small and large laboratory cages. *J Med Entomol*. 2008;45:698–705.
 20. Ndo C, Yamada H, Damiens DD, N'do S, Seballos G, JRL G. X-ray sterilization of the *An. arabiensis* genetic sexing strain 'ANO IPCL1' at pupal and adult stages. *Acta Trop*. 2014;131:124–8.
 21. Helinski MEH, Knols BGJ. The influence of late-stage pupal irradiation and increased irradiated: un-irradiated male ratio on mating competitiveness of the malaria mosquito *Anopheles arabiensis* Patton. *Bull Entomol Res*. 2009;99:317–22.
 22. Helinski MEH, Parker AG, Knols BGJ. Radiation-induced sterility for pupal and adult stages of the malaria mosquito *Anopheles arabiensis*. *Malar J*. 2006;5:41.
 23. Damiens D, Vreysen MJB, Gilles JRL. *Anopheles arabiensis* sperm production after genetic manipulation, dieldrin treatment, and irradiation. *J Med Entomol*. 2013;50:314–6.
 24. Helinski MEH, Knols BGJ. Sperm quantity and size variation in un-irradiated and irradiated males of the malaria mosquito *Anopheles arabiensis* Patton. *Acta Trop*. 2009;109:64–9.
 25. Yamada H, Vreysen MJB, Gilles JRL, Munhenga G, Damiens DD. The effects of genetic manipulation, dieldrin treatment and irradiation on the mating competitiveness of male *Anopheles arabiensis* in field cages. *Malar J*. 2014;13:318.
 26. Krafus ES, Davidson G. Production of semisterile mutants in the *Anopheles gambiae* s.l. species complex. *Iowa State J Res*. 1987;62:85–97.
 27. Helinski MEH, Hassan MM, El-Motasim WM, Malcolm CA, Knols BGJ, El-Sayed B. Towards a sterile insect technique field release of *Anopheles arabiensis* mosquitoes in Sudan: irradiation, transportation, and field cage experimentation. *Malar J*. 2008;7:65.
 28. Ageep TB, Damiens D, Alsharif B, Ahmed A, Salih EHO, Ahmed FTA, et al. Participation of irradiated *Anopheles arabiensis* males in swarms following field release in Sudan. *Malar J*. 2014;13:484.
 29. Hassan MM, Zain HM, Basheer MA, Elhaj H-EF, El-Sayed BB. Swarming and mating behavior of male *Anopheles arabiensis* Patton (Diptera: Culicidae) in an area of the sterile insect technique project in Dongola, northern Sudan. *Acta Trop*. 2014;132:S64–S9.
 30. Fanello C, Santolamazza F, della Torre A. Simultaneous identification of species and molecular forms of the *Anopheles gambiae* complex by PCR-RFLP. *Med Vet Entomol*. 2002;16:461–4.
 31. Crawley MJ. *The R book*. Chichester, West Sussex: John Wiley & Sons, Ltd; 2007.
 32. Abbott WS. A method of computing the effectiveness of an insecticide. *J Econ Entomol*. 1925;128:265–7.
 33. Helinski MEH, Hood RC, Knols BGJ. A stable isotope dual-labelling approach to detect multiple insemination in un-irradiated and irradiated *Anopheles arabiensis* mosquitoes. *Parasit Vectors*. 2008;1:9.
 34. Darrow DI. The effect of gamma irradiation on reproduction and life span of the mosquito *Culex tarsalis* Coquillett. *Mosq News*. 1968;28:21–4.
 35. Ali SR, Rozeboom LE. Observations on sterilization of *Anopheles* (C.) *albimanus* Wiedemann by x-irradiation. *Mosq News*. 1972;32:574–9.
 36. Clements AN. *The Biology of Mosquitoes. Sensory Reception and Behaviour*, Vol. 2. Wallingford: CAB; 1999.
 37. Pennetier C, Warren B, Dabire KR, Russell IJ, Gibson G. "Singing on the wing" as a mechanism for species recognition in the malarial mosquito *Anopheles gambiae*. *Curr Biol*. 2010;20:131–6.
 38. Maiga H, Dabire RK, Lehmann T, Tripet F, Diabate A. Variation in energy reserves and role of body size in the mating system of *Anopheles gambiae*. *J Vector Ecol*. 2012;37:289–97.
 39. Maiga H, Niang A, Sawadogo S, Dabiré RK, Lees RS, Gilles J, et al. Role of nutritional reserves and body size in *Anopheles gambiae* males mating success. *Acta Trop*. 2014;132:S102–S7.
 40. Pitts RJ, Mozūraitis R, Gauvin-Bialecki A, Lempérière G. The roles of kairomones, synomones and pheromones in the chemically-mediated behaviour of male mosquitoes. *Acta Trop*. 2014;132(Suppl.):S26–S34.
 41. Dandolo LC, Kemp A, Koekemoer LL, Munhenga G. Effect of ionising (gamma) radiation on female *Anopheles arabiensis*. *Trans R Soc Trop Med H*. 2017;111:38–40.
 42. Bakri A, Mehta K, Lance DR. Sterilizing insects with ionizing radiation. In: Dyck VA, Hendrichs J, Robinson AS, editors. *Sterile Insect Technique: Principles and Practice in Area-Wide Integrated Pest Management*. Dordrecht, Netherlands: Springer; 2005. p. 233–68.
 43. Roux O, Vantaux A, Roche B, Yameogo KB, Dabiré KR, Diabaté A, et al. Evidence for carry-over effects of predator exposure on pathogen transmission potential. *Proc Biol Sci*. 2015;282:20152430.
 44. Vantaux A, Ouattara I, Lefèvre T, Dabiré KR. Effects of larvicidal and nutritional stresses on *Anopheles gambiae* development, survival and competence for *Plasmodium falciparum*. *Parasit Vectors*. 2016;9:226.
 45. Vantaux A, Lefèvre T, Cohuet A, Dabiré KR, Roche B, Roux O. Larval nutritional stress affects vector life history traits and human malaria transmission. *Sci Rep*. 2016;6:36778.
 46. Briegel H. Fecundity, metabolism, and body size in *Anopheles* (Diptera: Culicidae). vectors of malaria. *J Med Entomol*. 1990;27:839–50.
 47. Klowden MJ, Russell RC. Mating affects egg maturation in *Anopheles gambiae* Giles (Diptera: Culicidae). *J Vector Ecol*. 2004;29:135–9.
 48. Klowden MJ, Chambers GM. Male accessory gland substances activate egg development in nutritionally stressed *Aedes aegypti* mosquitoes. *J Insect Physiol*. 1991;37:721–6.
 49. Gabrieli P, Kakani EG, Mitchell SN, Mameli E, Want EJ, Mariezcurrena Anton A, et al. Sexual transfer of the steroid hormone 20E induces the postmating switch in *Anopheles gambiae*. *Proc Natl Acad Sci USA*. 2014;111:16353–8.
 50. Bentley MD, Day JF. Chemical ecology and behavioral aspects of mosquito oviposition. *Ann Rev Entomol*. 1989;34:401–21.
 51. Silberbush A, Blaustein L. Mosquito females quantify risk of predation to their progeny when selecting an oviposition site. *Funct Ecol*. 2011;25:1091–5.
 52. Wasserberg G, White L, Bullard A, King J, Maxwell R. Oviposition site selection in *Aedes albopictus* (Diptera: Culicidae): Are the effects of predation risk and food level independent? *J Med Entomol*. 2013;50:1159–64.
 53. Zahir N, Rau ME. Oviposition attraction and repellency of *Aedes aegypti* (Diptera: Culicidae) to waters from conspecific larvae subjected to crowding, confinement, starvation, or infection. *J Med Entomol*. 1998;35:782–7.
 54. Gilles JRL, Schetelig MF, Scolari F, Marec F, Capurro ML, Franz G, et al. Towards mosquito sterile insect technique programmes: exploring genetic, molecular, mechanical and behavioural methods of sex separation in mosquitoes. *Acta Trop*. 2014;132:S178–S87.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

