EcoMnOx, a Biosourced Catalyst for Selective Aerobic Oxidative Cleavage of Activated 1,2-Diols

Vincent Escande, * Chun Ho Lam, Claude Grison, and Paul T. Anastas*

Supporting Information

Total pages: 12; total figures: 14; total tables: 2

General methods	S2
XPS analysis	S3
Figure S1. XPS survey spectrum of EcoMnOx.	S3
Figure S2. XPS spectrum of Mn3s.	S4
ICP-AES analyses	S5
Table S1. ICP-AES analysis of EcoMnOx catalysts prepared from different lots of <i>Grevillea exul</i> subsp. <i>rubiginosa</i> .	S5
Characterization of EcoMnOx after reaction	S6
Table S2. ICP-AES analysis of EcoMnOx (EcoMnOx batch #3) before and after oxidative cleavage of <i>me</i> hydrobenzoin.	<i>so-</i> S6
Figure S3. SEM images of EcoMnOx (EcoMnOx batch #3): a), b) before oxidative cleavage of <i>meso</i> -hydrobenzoin and c), d) after oxidative cleavage of <i>meso</i> -hydrobenzoin	S6
Figure S4. EDX spectra of EcoMnOx (EcoMnOx batch #3): a) before oxidative cleavage of <i>meso</i> -hydrobenzoin and b) after oxidative cleavage of <i>meso</i> -hydrobenzoin	S7
GC chromatograms	S8
Figure S5. Oxidative cleavage of meso-hydrobenzoin 1a analyzed after 1 h reaction.	S8
Figure S6. Oxidative cleavage of (<i>R</i> , <i>R</i>)-hydrobenzoin 1b analyzed after 3 h reaction.	S8
Figure S7. Oxidative cleavage of 4,4'-dimethoxyhydrobenzoin 1c analyzed after 1 h reaction.	S8
Figure S8. Oxidative cleavage of 2,2'-dichlorohydrobenzoin 1d analyzed after 1 h reaction	S9
Figure S9. Oxidative cleavage of 4,4'-difluorohydrobenzoin 1e analyzed after 1 h reaction	S9
Figure S10. Oxidative cleavage of 2,3-diphenyl-2,3-butanediol 1f analyzed after 24 h reaction	S9
Figure S11. Oxidative cleavage of benzopinacol 1g analyzed after 1 h reaction.	. S10
Figure S12. Oxidative cleavage of 1-phenyl-1,2-propanediol 1h analyzed after 24 h reaction	. S10
Figure S13. Oxidative cleavage of 1,2-di-2-furanyl-1,2-ethanediol 1j analyzed after 24 h reaction	. S10
Figure S14. Oxidative cleavage of 1,6-diphenyl-1,5-hexadiene-3,4-diol 1k analyzed after 3 h	. S11
E-factor calculations	. S12
Procedure for EcoMnOx-catalyzed meso-hydrobenzoin oxidative cleavage	. S12
Calculations	. S12
References	. S12

General methods

All common reagents and solvents were obtained from Sigma-Aldrich and Fisher Scientific, and were used without any further purification. Non-commercially available 1,2-diols were prepared by reduction of corresponding commercial diketones according to a known procedure.¹ Products were characterized by ¹H/¹³C NMR spectroscopy and by GC, by comparison of their retention times with those of pure standards, and analytical data were in accordance with those previously published. NMR characterization was performed with an Agilent DD2 400 MHz NMR spectrometer. GC were recorded on a Shimadzu GC-2010 Plus, equipped with an Agilent J&W GC column, DB-5, 60 m x 0.25 mm x 0.25 µm, using He as carrier gas, with a flame ionization detector (FID). All analyses were performed including dodecane as internal standard. FTIR spectra were recorded on a FTIR/Raman Thermo Nicolet 6700 spectrometer, in ATR mode. ICP-AES measurements were performed using the Perkin Elmer Optima 3000. Samples of 10 mg were digested and dissolved in 2% HNO₃ acid, and were quantified against external standards. SEM analyses were performed using the Hitachi SU-70 field emission microscope. Samples were pre-dried in vacuum overnight, and were examined with a 10kV bean power under ultra-high vacuum without surface treatment. Energy-dispersive X-ray spectroscopy analysis (EDX) was performed using a Zeiss EVO HD15 microscope coupled with an Oxford X-MaxN EDX detector. XRPD measurements were performed on a Rigaku SmartLab X-ray Diffractometer equipped with a Cu line-focus sealed tube and a NaI SC70 scintillation detector. Measurements were made with a 40 kV, 44 mA beam, with an incident slit of 2/3 deg and a length limiting slit of 5.00 mm, in the range 20 from 10° to 90° locked couple scan type, a step size of 0.05° and a scan speed of 10.33 deg/min. XPS analysis was performed using a ThermoScientific ESCALAB 250 instrument. Spectra were collected using a monochromatic Al X-ray source. A low energy electron flood and top-side contact were used for charge neutralization. Survey spectra were collected using a pass energy of 150 eV. Multiplex composition scans were acquired with 20 eV pass energy. All dynamic light scattering (DLS) samples were suspended in methanol (1 mg/3 mL) and were sonicated for 5 minutes prior analysis. DLS was performed at an angle of 90 degrees (ALV-5000, ALV-GmbH) using an incident laser of wavelength 532 nm (coherent). 30 runs were recorded per sample, with a duration of 30 seconds per run. All DLS measurements were carried out at room temperature.

XPS analysis



Figure S1. XPS survey spectrum of EcoMnOx.

XPS analysis shows the oxide nature of the polymetallic material.



Figure S2. XPS spectrum of Mn3s.

In spite of obscuration by the interfering Mg peak, Mn3s multiplet splitting (4.6 eV) is characteristic of Mn^{IV} .²

ICP-AES analyses

Type of catalyst	G. exul subsp. rubiginosa	Elements wt% (±sd)							
#batch number	Location of harvest (latitude and longitude)	Fe	Mn	Ni	Ca	Mg	Al	Na	К
EcoMn #1	« Col de Mouirange » (-22.225415, 166.650282)	0.9 (±0.008)	6.7 (±0.04)	0.04 (±0.0004)	10.4 (±0.2)	5.3 (±0.09)	0.2 (±0.02)	2.0 (±0.1)	4.3 (±0.008)
EcoMnOx #1	« Col de Mouirange » (-22.225415, 166.650282)	0.3 (±0.006)	14.1 (±0.4)	n.d. ^a	28.2 (±0.2)	28.7 (±0.3)	n.d. ^a	0.5 (±0.09)	n.d. ^a
EcoMn #2	« Monts Kouanenoa » (-22.203329, 166.699232)	0.7 (±0.003)	6.5 (±0.04)	0.2 (±0.0009)	7.9 (±0.05)	5.4 (±0.07)	0.3 (±0.03)	0.8 (±0.01)	3.8 (±0.05)
EcoMnOx #2	« Monts Kouanenoa » (-22.203329, 166.699232)	1.3 (±0.01)	10.0 (±0.06)	0.07 (±0.002)	18.7 (±0.1)	29.5 (±0.01)	0.6 (±0.01)	n.d. ^a	n.d. ^a
EcoMn #3	« Rivière des Pirogues » (-22.187158, 166.721983)	0.3 (±0.005)	2.7 (±0.05)	0.02 (±0.0004)	12.1 (±0.4)	2.5 (±0.02)	0.1 (±0.01)	2.0 (±0.03)	4.6 (±0.07)
EcoMnOx #3	« Rivière des Pirogues » (-22.187158, 166.721983)	1.0 (±0.005)	8.9 (±0.05)	0.04 (±0.0001)	22.5 (±0.2)	8.3 (±0.05)	0.5 (±0.01)	n.d. ^a	n.d. ^a

 Table S1. ICP-AES analysis of EcoMnOx catalysts prepared from different lots of Grevillea exul subsp. rubiginosa.

^an.d.: not detected.

Comparison of several batches of EcoMn and their corresponding EcoMnOx, prepared from *Grevillea exul* subsp. *rubiginosa* harvested in different locations, shows some variation in elemental content, due to the natural origin of the biomass feedstock.

However, because the catalytic efficiency of EcoMnOx relies only on Mn content (other metal elements are spectator elements, as shown in Table 3), the different batches of EcoMnOx display the same performances when used in 10 mol% Mn for aerobic oxidative cleavage of *meso*-hydrobenzoin **1a**.

Characterization of EcoMnOx after reaction

Table S2. ICP-AES analysis of EcoMnOx (EcoMnOx batch #3) before and after oxidative cleavage o	f
neso-hydrobenzoin.	

Type of catalyst	<i>G. exul</i> subsp.	Elements wt% (±sd)							
#batch number	Location of harvest (latitude and longitude)	Fe	Mn	Ni	Ca	Mg	Al	Na	K
EcoMnOx #3 (before reaction)	« Rivière des Pirogues » (-22.187158, 166.721983)	1.0 (±0.005)	8.9 (±0.05)	0.04 (±0.0001)	22.5 (±0.2)	8.3 (±0.05)	0.5 (±0.01)	n.d. ^a	n.d. ^a
EcoMnOx #3 (after reaction ^b)	« Rivière des Pirogues » (-22.187158, 166.721983)	1.2 (±0.01)	8.8 (±0.09)	0.05 (±0.005)	24.2 (±0.06)	8.2 (±0.04)	0.5 (±0.01)	n.d. ^a	n.d. ^a

^an.d.: not detected. ^bCatalyst recovered after centrifugation, rinsed with 2 x 4 mL of ethanol, and then dried in oven under air at 105 °C for 2 h.



Figure S3. SEM images of EcoMnOx (EcoMnOx batch #3): a), b) before oxidative cleavage of *meso*-hydrobenzoin and c), d) after oxidative cleavage of *meso*-hydrobenzoin.



Figure S4. EDX spectra of EcoMnOx (EcoMnOx batch #3): a) before oxidative cleavage of *meso*-hydrobenzoin and b) after oxidative cleavage of *meso*-hydrobenzoin.

GC chromatograms



Figure S5. Oxidative cleavage of meso-hydrobenzoin 1a analyzed after 1 h reaction.



Figure S6. Oxidative cleavage of (*R*,*R*)-hydrobenzoin 1b analyzed after 3 h reaction.



Figure S7. Oxidative cleavage of 4,4'-dimethoxyhydrobenzoin 1c analyzed after 1 h reaction.



Figure S8. Oxidative cleavage of 2,2'-dichlorohydrobenzoin 1d analyzed after 1 h reaction.



Figure S9. Oxidative cleavage of 4,4'-difluorohydrobenzoin 1e analyzed after 1 h reaction.



Figure S10. Oxidative cleavage of 2,3-diphenyl-2,3-butanediol 1f analyzed after 24 h reaction.



Figure S11. Oxidative cleavage of benzopinacol 1g analyzed after 1 h reaction.



Figure S12. Oxidative cleavage of 1-phenyl-1,2-propanediol 1h analyzed after 24 h reaction.



Figure S13. Oxidative cleavage of 1,2-di-2-furanyl-1,2-ethanediol 1j analyzed after 24 h reaction.



Figure S14. Oxidative cleavage of 1,6-diphenyl-1,5-hexadiene-3,4-diol 1k analyzed after 3 h.

E-factor calculations

Procedure for EcoMnOx-catalyzed *meso*-hydrobenzoin oxidative cleavage

A mixture of *meso*-hydrobenzoin (428.5 mg, 2.0 mmol), EcoMnOx (10 mol% Mn) and 1-butanol (1 mL) was added in a 7 mL flask, equipped with a O_2 balloon. The reaction was heated to 100 °C under magnetic stirring (400 rpm) for 1 h. After reaction, the catalyst was separated by filtration and washed with two 1 mL portions of 1-butanol. Evaporation of the combined 1-butanol fractions afforded pure benzaldehyde in 90% yield (382 mg, 3.6 mmol).

Calculations

$$E = \frac{\sum_{i} m(raw \ material_{i}) - m_{p}}{m_{p}}$$

 $(m_p: \text{mass of product. Solvents are included assuming 90% recovery, catalysts are ignored if recovered and reused³)$

$$E = \frac{0.429 + 0.243 - 0.382}{0.382} = 0.8$$

References

1. Anju, K. S.; Ramakrishnan, S.; Thomas, A. P.; Suresh, E.; Srinivasan, A., 9,10,19,20-Tetraarylporphycenes. *Org. Lett.* **2008**, *10* (24), 5545-5548.

2. Junta, J. L.; Hochella, M. F., Manganese (II) oxidation at mineral surfaces: A microscopic and spectroscopic study. *Geochim. Cosmochim. Acta* **1994**, *58* (22), 4985-4999.

3. Sheldon, R. A., The E Factor: fifteen years on. *Green Chem.* **2007**, *9* (12), 1273-1283.