Malaria parasite clearance from patients following artemisinin-based combination therapy in Côte d’Ivoire
Offianan Toure, Tiacoh Landry, Serge Brice Assi, Antoinette Kone, Eric Gbessi, Berenger Ako, Baba Coulibaly, Bouakary Kone, Oumar Ouattara, Sylvain Beourou, et al.

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Malaria parasite clearance from patients following artemisinin-based combination therapy in Côte d’Ivoire

Introduction: Parasite clearance is useful to detect artemisinin resistance. The aim of this study was to investigate parasite clearance in patients treated with artesunate + amodiaquine (AS + AQ) and artemether + lumefantrine (AL): the two artemisinin-based combination therapies (ACTs) recommended in the first-line treatment of uncomplicated malaria in Côte d’Ivoire.

Methods: This study was conducted in Bouaké, Côte d’Ivoire, from April to June 2016. Patients aged at least 6 months with uncomplicated malaria and treated with AS + AQ or AL were hospitalized for 3 days, and follow-up assessments were performed on days 3, 7, 14, 21, 28, 35, and 42. Blood smears were collected at the time of screening, pre-dose, and 6-hour intervals following the first dose of administration until two consecutive negative smears were recorded, thereafter at day 3 and follow-up visits. Parasite clearance was determined using the Worldwide Antimalarial Resistance Network’s parasite clearance estimator. The primary end points were parasite clearance rate and time.

Results: A total of 120 patients (57 in the AS + AQ group and 63 in the AL group) were randomized among 298 patients screened. The median parasite clearance time was 30 hours (IQR, 24–36 hours), for each ACT. The median parasite clearance rate had a slope half-life of 2.36 hours (IQR, 1.85–2.88 hours) and 2.23 hours (IQR, 1.74–2.63 hours) for AS + AQ and AL, respectively. The polymerase chain reaction-corrected adequate clinical and parasitological response was 100% and 98.07% at day 42 for AS + AQ and AL, respectively.

Conclusion: Patients treated with AS + AQ and AL had cleared parasites rapidly. ACTs are still efficacious in Bouaké, Côte d’Ivoire, but continued efficacy monitoring of ACTs is needed.

Keywords: Plasmodium falciparum, ACTs, parasite clearance, Côte d’Ivoire

Introduction
Malaria remains a serious public health problem in sub-Saharan Africa despite the reduction in morbidity and mortality observed in recent years.1 The advances in the control of malaria are partly related to the use of artemisinin-based combination therapies (ACTs) in almost all endemic countries in combination with other strategies.

However, the emergence and spread of artemisinin-resistant malaria parasite populations in Southeast Asian countries and the possibility of extension to other endemic areas are serious threats.2–4 The possibility that artemisinin resistance might spread or emerge independently elsewhere necessitates careful surveillance.5

The WHO recommends that the efficacy of the first- and second-line antimalarial drugs should be regularly assessed for early detection and prevention of the spread of resistant parasite populations.6
Several reliable methods exist currently to search for possible emergence of resistance to artemisinin derivatives. Surveillance of artemisinin resistance to date has relied on in vivo studies to measure early clearance of peripheral parasitaemia by microscopy and k13 propeller gene mutations. 

The phenotype of artemisinin resistance is characterized by a significant delay in parasite clearance following the initiation of therapy. Thus, many studies on the k13 propeller marker gene for resistance to artemisinin conducted in sub-Saharan Africa have shown no evidence of resistance mutation which is associated with artemisinin resistance in Southeast Asia. Therefore, parasite clearance rates are important measures of antimalarial drug efficacy and particularly important in the assessment of artemisinin resistance. The parasite clearance estimator (PCE) developed by Worldwide Antimalarial Resistance Network (WARN) seemed to be an accurate and reliable method for the early detection of artemisinin resistance. This method is uses by sampling multiple times (6, 8, or 12 hours) a day at measured time points to estimate the rate of parasite clearance.

A number of clinical trials have already been conducted in different parts of Côte d’Ivoire on artesunate + amodiaquine (AS + AQ) and artemether + lumefantrine (AL) efficacy and safety from 2005 to 2016 in Malaria Control Program sentinel sites. Results from these studies have provided evidence that the two ACTs remain efficacious in the treatment of Plasmodium falciparum uncomplicated malaria. Polymerase chain reaction (PCR)-corrected 28-day adequate clinical and parasitological response (ACPR) for each ACT was more than 95% with no early clinical failures. However, in Bouake, which is one of the newest sentinel sites of the National Malaria Control Program (NMCP) of Côte d’Ivoire having high malaria transmission, no efficacy data of AS + AQ and AL are available since the use of these ACTs in this region. Therefore, this study was designed to assess parasite clearance and antimalarial drug efficacy in patients treated with AS + AQ and AL in Bouake, Center of Côte d’Ivoire.

Study population
All patients aged at least 6 months with uncomplicated falciparum malaria were screened. The WHO’s standard efficacy testing protocol was used. Briefly, patients with monospecific P. falciparum infestation confirmed by microscopy with parasite density between 2,000 and 200,000 asexual parasites/μL of blood, axillary temperature of ≥37.5°C, or history of fever over the past 24 hours, body weight ≥5 kg, and who were able to take oral medications and follow study procedures, were included after informed consent was obtained from participate, parents, or legal guardian. Patients with severe malaria symptoms, symptoms of severe malnutrition, or chronic diseases or with mixed infection were excluded.

Sample size
The sample size was calculated using the WHO guidelines on the assessment of antimalarial drugs. Population size was determined according to the following criteria: the proportion of probable clinical failures with the antimalarial combinations studied should not be higher than 10%, for a level of confidence of 95% and a precision of 10%, taking into account patients who were excluded or lost to follow up. Using these criteria, a minimum of 50 patients was required in each treatment arm.

Clinical procedures
All included patients were hospitalized for 3 days (day 0, 1, 2), and follow-up assessments were performed on day 3, 7, 14, 21, 28, 35, and 42. Physical examination, vital signs, body temperature, and clinical assessment were performed at screening and at all follow-up visits. Body temperature was recorded at 6-hour intervals following the first dose of study medication until temperature normalized and remained normal for 24 hours and at every visit thereafter.

Drug administration
Patients were randomly assigned to receive either AS + AQ or AL. Both treatments were 3-day oral regimens dosed by
weight according to the manufacturer’s instructions: AS + AQ 5 to <9 kg: one tablet/day of AS 25 mg/AQ 67.5 mg; 9 to <18 kg: one tablet/day of AS 50 mg/AQ 135 mg; 18 to <36 kg: one tablet/day of AS 100 mg/AQ 270 mg; ≥36 kg: two tablets/day of AS 100 mg/AQ 270 mg. AS + AQ were administered once daily in children and twice daily in adult participants.

AL tablet strength was 20 mg artemether/120 mg lumefantrine: 5 to <15 kg: one tablet/dose; 15 to <25 kg: two tablets/dose; 25 to <35 kg: three tablets/dose; ≥35 kg four tablets/dose. AL was administrated twice a day. If vomiting occurred within 30 minutes following the first drug dose, a redosing was performed. If the patient vomited within 30 minutes of repeated dosing, he or she was withdrawn from the trial and given rescue medication according to the NMCP’s malaria treatment guidelines.

**Laboratory procedures**

**Parasitological assessments**

Thick/thin blood smears were collected at the time of screening, pre-dose, and at day 0- and 6-hour intervals following the first dose of administration until two consecutive negative smears were recorded, thereafter at day 3 and follow-up visits. Blood smears were prepared using Giemsa staining (10%). Thick film was examined with a binocular microscope with an oil immersion objective lens to quantify the parasitaemia. Parasitaemia was measured by counting the number of asexual parasites and leucocytes in 200 high power fields based on a putative count of 8,000 leucocytes/mL of blood.

Two qualified independent microscopists read all Giemsa-stained slides. Discordant readings were reexamined by a third qualified independent microscopist.

Discordance was defined as differences between the first and second microscopist regarding parasite density >25%, species diagnosis or any difference that affected recruitment or study outcome. Final parasite density was the average of the two most concordant counts.

**PCE**

The PCE established by WWARN was used to determine parasite clearance rate, parasite clearance time (PCT), slope half-life, and the time needed for parasitaemia to fall by 50%, 90%, 95%, and 99% (PC50, PC90, PC95, and PC99, respectively).

WWARN’s PCE is an uniform, reliable, and accurate method to estimate malaria parasite clearance based on the linear portion of the slope of the log-parasitaemia vs time relationship.15,21

This standardized approach provides information that enables the detection of early changes in *P. falciparum* sensitivity to artemisinins and support timely responses in treatment guidelines when needed.

**Parasite genotyping**

To distinguish between recrudescence and reinfection, blood spots on Whatman®3 MM filter paper (Whatman International Ltd, Maidstone, UK) were prepared before treatment and after day 7 in the event of reappearance of parasites, confirmed by microscopy. PCR genotyping was performed following the standard protocol as previously described on the basis of *P. falciparum* merozoite surface protein 1 (*msp1*), merozoite surface protein 2 (*msp2*) and glutamate-rich protein (*glurp*).6,22,23 Possible outcomes were new infection or recrudescence. A “new infection” is a subsequent occurring parasitaemia in which all the alleles of the parasites from the posttreatment sample are different from those in the admission sample, for one or more loci tested. In a “recrudescence,” at least one allele at each locus should be in common for both paired samples.

**Study end points**

The primary end points were PCT and parasite clearance rate. The PCT was defined as the time in hours from the first treatment dose to the first of two consecutive thick blood films that were negative for asexual *P. falciparum* parasites after checking 200 oil immersion fields. The secondary outcomes were the PCR-corrected ACPR at day 42, fever clearance times, time in hours from the start of treatment at which the temperature first decreased to <37.5°C and remained below 37.5°C for 24 hours.

**Statistical analyses**

SPSS Version 17 (SPSS Inc., Chicago, IL, USA) was used for data management and analysis. Frequencies were compared by either chi-squared or Fisher’s exact tests, as appropriate, and continuous variables by Student’s t-tests.

Distribution of parasite clearance rate constants and slope half-lives were generated by the WWARN’s PCE (http://www.w.Warn.org/research/parasite-clearance-estimator).

**Ethical issues**

The study was conducted according to the local laws and regulations, International Conference on Harmonization-Good Clinical Practice (ICH-GCP). The protocol was reviewed and approved by the Comité National d’Éthique de la Recherche de Côte d’Ivoire [National Research Ethic Committee of Côte d’Ivoire]. Written informed consent was...
obtained from each participant or legal guardians before any sample collection. In the case of an illiterate patient, his/her thumb impression and signature of an independent witness were obtained. Patients providing assent were <18 year old who were able to understand and complied with study procedures in addition to parents’ or legal guardians’ written informed consent.

**Results**

**Trial profile**

The trial profile is shown in Figure 1. From April to June 2016, 298 patients with suspected malaria were screened for eligibility in the clinical trial. A total of 120 patients were eligible and randomized in the AS + AQ (57) and AL (63) groups. The reasons for ineligibility were low-density parasitaemia (<2,000 parasites/μL of blood), very high-density parasitaemia (>200,000 parasites/μL of blood), low probability of completing follow-up, inability to take oral drugs, and unwillingness to consent to the study. Six patients (two in the AS + AQ group and four in the AL group) defaulted as a result of withdrawal and/or lost to follow up. Primary end points were reached by 54 and 55 patients in the AS + AQ and AL groups, respectively.

**Baseline characteristics of study population**

The study population consisted of 44.4% males and 55.6% females in the AL group. In the AS + AQ group, 40.3% of patients were males. The median age was 7±0.67 years in the AL group and 8.7±1.0 years in the AS + AQ group. Mean weight was 22.1±1.65 kg in the AL group and 23.3±1.5 kg in the AS + AQ group. Both treatment groups were comparable in terms of baseline demographic, clinical, and laboratory characteristics except for hemoglobin and age (Table 1).

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**Figure 1** Trial profile.
Primary end points

The parasite clearance rate was assessed in 54 and 55 patients in the AS+AQ group and AL group, respectively. The median PCT in this study was 30 hours (IQR, 24–36 hours), for each ACT, and the median parasite clearance rate had a slope half-life of 2.36 hours (IQR, 1.85–2.88 hours) and 2.23 hours (IQR, 1.74–2.63 hours) for AS+AQ and AL, respectively (Table 2).

Secondary end points

No early treatment failure (ETF) was observed in either of the study groups. Day 42 crude ACPRs were 96.4% and 83.3% in the AS+AQ and AL groups, respectively. Twelve patients who completed 42-day follow-up had reappearance of parasitaemia (ten in the AL group, two in the AS+AQ group) between days 14 and 42 after the initial clearance of parasitaemia. PCR genotyping was successfully performed on all blood samples. PCR-corrected ACPR at day 42 after PCR correction in per-protocol population was 100% and 98.07% for AS + AQ and AL, respectively. This rate was 100% for each ACTs at day 28. Among those febrile on inclusion, the median fever clearance time was 6 hours (IQR, 6–6 hours) for each drug (Table 2).

Discussion

ACT as recommended by the WHO for the treatment of uncomplicated *P. falciparum* malaria has been adopted as the first-line therapy in most malaria-endemic countries.

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**Table 1** Baseline demographic characteristics

<table>
<thead>
<tr>
<th>Parameters</th>
<th>AL (n=63)</th>
<th>AS + AQ (n=57)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex, n (%) male</td>
<td>28 (44.4)</td>
<td>23 (40.3)</td>
<td>0.78</td>
</tr>
<tr>
<td>Age (years), mean ± SD</td>
<td>7.0±0.67</td>
<td>8.7±1.0</td>
<td>0.16</td>
</tr>
<tr>
<td>Age group</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1–5 years, n (%)</td>
<td>30 (47.6)</td>
<td>17 (29.8)</td>
<td>0.07</td>
</tr>
<tr>
<td>6–15 years, n (%)</td>
<td>27 (42.8)</td>
<td>38 (66.6)</td>
<td>0.01</td>
</tr>
<tr>
<td>&gt;15 years, n (%)</td>
<td>06 (09.5)</td>
<td>02 (03.5)</td>
<td>0.34</td>
</tr>
<tr>
<td>Weight (kg), mean ± SD</td>
<td>22.1±1.65</td>
<td>23.3±1.5</td>
<td>0.59</td>
</tr>
<tr>
<td>Pf. (parasitemia/ml), geometric mean ± SD</td>
<td>71.320±2.8</td>
<td>59.437±2.8</td>
<td>0.16</td>
</tr>
<tr>
<td>Anemia (hemoglobin level &lt;10), n (%)</td>
<td>20 (31.7)</td>
<td>10 (17.5)</td>
<td>0.11</td>
</tr>
<tr>
<td>Fever (axillary temperature ≥37.5°C), n (%)</td>
<td>63 (100)</td>
<td>57 (100)</td>
<td>1b</td>
</tr>
<tr>
<td>Body temperature among febrile patients (°C), mean±SD</td>
<td>39.2±0.08</td>
<td>39.1±0.07</td>
<td>0.30</td>
</tr>
</tbody>
</table>

Notes: Asexual parasites per microliter. *Chi-squared test. Student’s t-test.

**Table 2** Primary and secondary end points

<table>
<thead>
<tr>
<th>End point</th>
<th>AL (n=63)</th>
<th>AS + AQ (n=57)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Primary end point</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Parasite clearance rate (n)</td>
<td>55</td>
<td>54</td>
<td></td>
</tr>
<tr>
<td>Lag phase (hours), (n); median (range)</td>
<td>(29); 6.0 (6–12)</td>
<td>(31); 6.0 (6–12)</td>
<td>0.77</td>
</tr>
<tr>
<td>Parasite clearance rate constant (per hour), median (IQR)</td>
<td>0.29 (0.24–0.37)</td>
<td>0.31 (0.26–0.40)</td>
<td>0.17</td>
</tr>
<tr>
<td>Slope half-life (hours), median (IQR)</td>
<td>2.36 (1.85–2.88)</td>
<td>2.23 (1.74–2.63)</td>
<td>0.20</td>
</tr>
<tr>
<td>PCT (hours), n</td>
<td>55</td>
<td>54</td>
<td></td>
</tr>
<tr>
<td>PCT, median (IQR)</td>
<td>30.0 (24.0–36.0)</td>
<td>30.0 (24.0–36.0)</td>
<td>0.22</td>
</tr>
<tr>
<td>PC50, median IQR</td>
<td>8.93 (4.37–11.84)</td>
<td>7.95 (4.65–10.46)</td>
<td>0.77</td>
</tr>
<tr>
<td>PC90, median IQR</td>
<td>14.34 (10.58–17.78)</td>
<td>14.02 (10.65–17.10)</td>
<td>0.51</td>
</tr>
<tr>
<td>PC95, median IQR</td>
<td>16.65 (13.11–20.69)</td>
<td>16.0 (12.78–19.69)</td>
<td>0.44</td>
</tr>
<tr>
<td>PC99, median IQR</td>
<td>22.19 (18.75–25.84)</td>
<td>21.32 (18.46–25.07)</td>
<td>0.34</td>
</tr>
<tr>
<td>Secondary end points</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treatment outcomes, n</td>
<td>63</td>
<td>57</td>
<td></td>
</tr>
<tr>
<td>LPF (PCR corrected), n</td>
<td>01</td>
<td>00</td>
<td>1</td>
</tr>
<tr>
<td>PCR-corrected cure rate (K–M)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>At day 28, n (%) (95% CI)</td>
<td>63 (100)</td>
<td>57 (100)</td>
<td>1</td>
</tr>
<tr>
<td>At day 42, n (%) (95% CI)</td>
<td>62 (98.07) (88.07–99.67)</td>
<td>57 (100)</td>
<td>1</td>
</tr>
<tr>
<td>FCT (hours), (n); median (IQR)</td>
<td>(63); 6 (6–6)</td>
<td>(57); 6 (6–6)</td>
<td>0.49</td>
</tr>
</tbody>
</table>

Notes: *PC50, 50% parasite clearance rate; PC90, 90% parasite clearance rate; PC95, 95% parasite clearance rate; PC99, 99% parasite clearance rate.* As estimated by the PCE (WWARN).

Abbreviations: AL, artemether + lumefantrine; AS + AQ, artesunate + amodiaquine; FCT, fever clearance time; K–M, Kaplan–Meier; LPF, late parasitological failure; PCE, parasite clearance estimator; PCR, polymerase chain reaction; PCT, parasite clearance time; WWARN, Worldwide Antimalarial Resistance Network.
Monitoring the efficacy of malaria treatment and studies related to K13 propeller mutations have become essential to detect resistance to ACTs as early as possible.

A significant decrease in the rate of parasite clearance and delayed PCT following treatment with artemisinin and increased failure rates following ACT treatments provided definitive evidence of resistance in Southeast Asia.8,9,24 The median PCT in this study was 30 hours (IQR, 24–36 hours), for each ACT, and the median parasite clearance rate had a slope half-life of 2.36 hours (IQR, 1.85–2.88 hours) and 2.23 hours (IQR, 1.74–2.63 hours) for AS + AQ and AL, respectively.

A Phase III trial related to PCT (sampling every 6 hours) from patients treated with AL conducted in the south of Côte d’Ivoire in 2012 and 2014 demonstrated a rapid PCT of 24 hours.20,25

It seemed that the current PCT found in the center of the country is higher than in the south. This could be due to patients’ immunity related to transmission which is more intense in the south, drug pressure, the type of AL used, human genetics factors, or both.

Most of clinical trials conducted in Côte d’Ivoire since the introduction of AS + AQ and AL as the first-line treatment from 2005 to 2015 measured parasitaemia either daily or only on days 0, 2, and 3 as per WHO guidelines. Results from these studies demonstrated an estimated PCT of 48 hours for both AS + AQ and AL.18,19,26 However, these estimations of PCT have some limitations. Determination of PCT based on daily sampling lacks precision since the exact time of parasite count is not recorded. In addition, this time could vary by several hours depending on the timing of the visit to the clinic at inclusion and during the follow-up days. Accurate estimations of parasite clearance rates with WWARN’s PCE are suitable but need more frequent samplings which are difficult to implement during routine monitoring of the efficacy of ACTs.

The proportion of patients with persistent parasitaemia on day 3 after ACTs provides a useful indicator to be considered as a simple and readily measure in the setting of drug efficacy surveillance studies.27

The delayed parasite clearance at 72 hours is an in vivo predictor of subsequent treatment failure with ACTs26 and an indicator of choice for the routine monitoring of suspected artemisinin resistance in P. falciparum.

In the current study, none of the patients presented parasites on day 3. In a study conducted in 2009 in Côte d’Ivoire, delay in parasite clearance occurred in one patient treated with AL, suggesting probably a decreased sensitivity.26

Data related to PCT or proportion of patients with parasitaemia at day 3 from this current study and the previous one showed that malaria patients in Côte d’Ivoire cleared parasites rapidly after ACTs. These results are consistent with those reported from several sub-Saharan African countries.28–30 An individual patient data analysis of parasite clearance conducted on a large sample (n=15,000) of uncomplicated malaria treatments showed that rapid P. falciparum clearance continues to be achieved in sub-Saharan African patients treated with ACT.31

The artemisinin component is mostly responsible for the rapid parasite clearance. Studies conducted in Western Cambodia on artemisinin alone have reported 84 hours for PCT which was related to artemisinin resistance.12,32,33 The PCT with artemisinin observed in African patients is therefore much lower than that found in Western Cambodia.34

In a study conducted in Mali where parasitaemia was recorded every 8 hours in patients treated for uncomplicated P. falciparum with AS for 7 days and where there was no evidence of delayed parasitemia, the observed median PCT was 32 hours.35

Several factors could explain this difference. Indeed, artemisinin has been widely used in Africa since 2000, compared to more than 30 years in Cambodia, and is mainly available as co-formulated ACT.8,36 The other factors are related to host immunity and transmission reservoir of asymptomatic individual. These factors serve as obstacles to the selection of artemisinin resistance.37 Factors such as drug concentration, pretreatment parasite density, host malaria-specific immunity, independently of intrinsic drug-related effects, may alter or increase effects of drug on parasites.30–32

Malaria-specific immunity factor plays an important role in parasite clearance. A study conducted in the west of Côte d’Ivoire showed that glutamate rich protein and circumsporozoite protein IgG antibody responses in Man region in the west contribute to AS + AQ and AL treatment success and suggest that antimalarial immunity plays an important role in early parasitological responses.38–40

Efficacy data from the current study showed a high cure rate for both regimens after a standard day 42 follow-up. PCR-corrected ACPR at day 42 in the per-protocol population was 100% and 98.07% for AS + AQ and AL, respectively. Both treatments were well above the WHO recommended 90% threshold for treatments in use.4 Crude clinical treatment failure occurred in ten and two participants in the AL and AS + AQ groups, respectively. Our findings are in line with results from previous studies that showed that ACTs had a high cure rate and short PCT in malaria-endemic areas in Côte d’Ivoire.18–20
The study has some limitations. Pharmacology of drug action, parasite biology, and human immunity were not performed.

**Conclusion**

Patients treated with AS + AQ and AL cleared malaria parasites quickly in Bouake region. Data from this study support evidence that ACTs are still efficacious in this part of Côte d’Ivoire, but continued efficacy monitoring of ACTs is still needed.

**Acknowledgments**

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**Author contributions**

All the authors participated in design, implementation, analysis, or interpretation of the study. OAT was involved in all phases of the study and had full access to all the data in the study. SBA, AAK, FR, CR, and SB participated in the design and supervised the study. TNL, AAK, BK, EAG, and OO were responsible for conducting field studies and coordinating study procedures. BAA and BC performed PCR genotyping, and EAG performed the statistical analysis. The manuscript was drafted by OAT; substantial input came from all investigators. All authors contributed toward data analysis, drafting and critically revising the paper, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

**Disclosure**

The authors report no conflicts of interest in this work.

**References**


