



**HAL**  
open science

## To boldly gulp: standard metabolic rate and boldness have context-dependent influences on risk-taking to breathe air in a catfish

David Mckenzie, Thiago Belao, Shaun Killen, F. Tadeu Rantin

### ► To cite this version:

David Mckenzie, Thiago Belao, Shaun Killen, F. Tadeu Rantin. To boldly gulp: standard metabolic rate and boldness have context-dependent influences on risk-taking to breathe air in a catfish. *Journal of Experimental Biology*, 2015, 218 (23), pp.3762-3770. 10.1242/jeb.122903 . hal-01920943

**HAL Id: hal-01920943**

**<https://hal.umontpellier.fr/hal-01920943>**

Submitted on 4 Jun 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## CORRECTION

# To boldly gulp: standard metabolic rate and boldness have context-dependent influences on risk-taking to breathe air in a catfish

**David J. McKenzie, Thiago C. Belaõ, Shaun S. Killen and F. Tadeu Rantin**

There is a Correction to *J. Exp. Biol.* **218**, 3762-3770.

The article was originally published under a Company of Biologists Publication Agreement. The authors subsequently selected Open Access publication and the article is now published under a CC-BY license. The article has also been deposited in PubMed Central for immediate release.

The online article and PDF have been amended to reflect the change in licensing.

## RESEARCH ARTICLE

# To boldly gulp: standard metabolic rate and boldness have context-dependent influences on risk-taking to breathe air in a catfish

David J. McKenzie<sup>1,2,\*</sup>, Thiago C. Belão<sup>3</sup>, Shaun S. Killen<sup>4</sup> and F. Tadeu Rantin<sup>1</sup>

## ABSTRACT

The African sharptooth catfish *Clarias gariepinus* has bimodal respiration, it has a suprabranchial air-breathing organ alongside substantial gills. We used automated bimodal respirometry to reveal that undisturbed juvenile catfish ( $N=29$ ) breathed air continuously in normoxia, with a marked diurnal cycle. Air breathing and routine metabolic rate (RMR) increased in darkness when, in the wild, this nocturnal predator forages. Aquatic hypoxia (20% air saturation) greatly increased overall reliance on air breathing. We investigated whether two measures of risk taking to breathe air, namely absolute rates of aerial  $O_2$  uptake ( $\dot{M}_{O_2,air}$ ) and the percentage of RMR obtained from air ( $\% \dot{M}_{O_2,air}$ ), were influenced by individual standard metabolic rate (SMR) and boldness. In particular, whether any influence varied with resource availability (normoxia versus hypoxia) or relative fear of predation (day versus night). Individual SMR, derived from respirometry, had an overall positive influence on  $\dot{M}_{O_2,air}$  across all contexts but a positive influence on  $\% \dot{M}_{O_2,air}$  only in hypoxia. Thus, a pervasive effect of SMR on air breathing became most acute in hypoxia, when individuals with higher  $O_2$  demand took proportionally more risks. Boldness was estimated as time required to resume air breathing after a fearful stimulus in daylight normoxia ( $T_{res}$ ). Although  $T_{res}$  had no overall influence on  $\dot{M}_{O_2,air}$  or  $\% \dot{M}_{O_2,air}$ , there was a negative relationship between  $T_{res}$  and  $\% \dot{M}_{O_2,air}$  in daylight, in normoxia and hypoxia. There were two  $T_{res}$  response groups, 'bold' phenotypes with  $T_{res}$  below 75 min ( $N=13$ ) which, in daylight, breathed proportionally more air than 'shy' phenotypes with  $T_{res}$  above 115 min ( $N=16$ ). Therefore, individual boldness influenced air breathing when fear of predation was high. Thus, individual energy demand and personality did not have parallel influences on the emergent tendency to take risks to obtain a resource; their influences varied in strength with context.

**KEY WORDS:** Bimodal respiration, Energy metabolism, Hypoxia, Personality, Respiratory partitioning, Risk-taking

<sup>1</sup>Department of Physiological Sciences, Federal University of São Carlos, 13565-905, São Carlos SP, Brazil. <sup>2</sup>UMR9190 Centre for Marine Biodiversity Exploitation and Conservation (Marbec), Place Eugène Bataillon, Université Montpellier, Montpellier cedex 5 34095, France. <sup>3</sup>Joint Graduate Program in Physiological Sciences, Federal University of São Carlos – UFSCar/São Paulo State University, UNESP Campus Araraquara, 14801-903, Araraquara SP, Brazil. <sup>4</sup>Institute of Biodiversity, Animal Health and Comparative Medicine, College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, G12 8QQ, UK.

\*Author for correspondence (david.mckenzie@cncs.fr)

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.

Received 27 March 2015; Accepted 28 September 2015

## INTRODUCTION

In many animal species, individuals show wide individual variation in traits of both energy metabolism and personality (Burton et al., 2011; Careau et al., 2008; Sih et al., 2012). It has been argued that this variation persists because it shapes life-history trade-offs between production (growth, maturation, reproduction) and mortality (Biro and Stamps, 2008; Burton et al., 2011; Killen et al., 2011; Quinn et al., 2012; Sih et al., 2012; Stamps, 2007). Foraging has received attention because, although it brings advantages for energy acquisition, it increases risk of predation (Anholt and Werner, 1998; Biro et al., 2004; Mangel and Stamps, 2001). Variation in the intensity of such risky behaviour could reflect physiology, with individuals with higher metabolic rate being able, or driven, to forage more (Killen et al., 2011, 2013). It could also reflect personality, with bolder individuals willing to accept greater risks to secure energetic benefits (Conrad et al., 2011; Sih et al., 2012; Wolf and Weissing, 2012). As variation in both sets of traits would contribute to a similar life-history trade-off, it has been predicted that correlational selection should co-adapt physiology and temperament: animals with higher basal oxygen demands should be bolder (Biro and Stamps, 2010; Careau and Garland, 2012; Jenjan et al., 2013; Réale et al., 2010; Stamps, 2007). There is evidence, however, that reality is more complex and that the relative strength of these drivers of risk-taking can vary with context, such as resource availability or perceived risks of predation (Frost et al., 2013; Killen et al., 2011, 2012, 2013; Thomson et al., 2012).

Air-breathing fishes are interesting models to investigate how individual energy metabolism and personality influence risk-taking to obtain a key resource, oxygen. Oxygen has low solubility in water and aquatic habitats are prone to episodes of hypoxia (Diaz and Breitburg, 2009; Rahn, 1966; Randall, 1982). Many fish species have evolved a capacity to sustain aerobic metabolism by gulping oxygen-rich atmospheric air, to store in air-breathing organs (ABOs) from which oxygen diffuses into the blood (Graham, 1997; Johansen, 1968, 1970; Randall et al., 1981). The species are all 'bimodal' breathers that possess both gills and an ABO, and they differ in their relative reliance upon obtaining oxygen from water versus air (Graham, 1997). Although a capacity to gulp air might seem advantageous, the adaptations are relatively rare among fishes and, it has been argued, one major reason is that rising to the water surface brings significant risk of predation (Chapman and McKenzie, 2009; Kramer et al., 1983; Smith and Kramer, 1986). It follows that, in bimodal fishes, risk-taking to obtain atmospheric air may be driven by an individual's oxygen demand but, at the same time, the tendency to surface may also reflect boldness. Context has profound effects on air breathing in bimodal fishes. A reduced availability of resources – aquatic hypoxia – is a profound stimulant of air breathing (Chapman and McKenzie, 2009; Graham, 1997; Johansen, 1968; Lefevre et al., 2014a,b) but surfacing is

**List of symbols and abbreviations**

ABO	air-breathing organ
CV	coefficient of variation
LMEM	linear mixed-effect model
$M_b$	body mass
$\dot{M}_{O_2,air}$	absolute rate of $O_2$ uptake from air
$\% \dot{M}_{O_2,air}$	percentage of SMR obtained from air
$\dot{M}_{O_2,water}$	absolute rate of $O_2$ uptake from water
RMR	routine metabolic rate
SMR	standard metabolic rate
$T_{res}$	time to resume air breathing after a simulated predator attack

inhibited if fishes perceive a risk of predation (Chapman and McKenzie, 2009; Shingles et al., 2005; Smith and Kramer, 1986).

The African sharpnose catfish *Clarias gariepinus* uses a suprabranchial chamber as an ABO. It also possesses substantial gills and is a ‘facultative’ air breather, meaning that, if denied access to the surface in normoxic water, it can maintain routine metabolic rate (RMR) by gill ventilation alone. Aquatic hypoxia stimulates profound increases in air breathing (Belão et al., 2011). It is a nocturnally active hunter; juveniles seek cover in daylight, apparently to avoid detection by predators (Britz and Pienaar, 1992; Bruton, 1979; Hecht and Pienaar, 1993). Although the catfish can meet routine oxygen requirements by gill ventilation in well-aerated water, it will spontaneously gulp air at irregular intervals (Belão et al., 2011). It has been suggested that such aperiodic air gulping might be a reflex driven by peripheral oxygen chemoreceptors (Milsom, 2012; Shelton et al., 1984) and therefore could reflect individual oxygen demand. Given, however, that the catfish is a facultative air breather (Belão et al., 2011), this risky surfacing in normoxia might also be driven by individual boldness, perhaps to increase the flux of oxygen for aerobic activities such as foraging, digestion and growth.

In a population of individually tagged juvenile catfish ( $N=29$ ), automated bimodal respirometry (Lefevre et al., 2016) revealed continuous air breathing in normoxia, with a marked diurnal cycle. Air breathing and RMR increased during the night when, in the wild, the species forages in darkness. Aquatic hypoxia (20% air saturation) markedly increased proportional reliance on air breathing, contributing more to RMR throughout the diurnal cycle. We then investigated the general hypothesis that individual variation in taking risks to gulp air would show dependence on both standard metabolic rate (SMR) and intrinsic boldness. We investigated two slightly contrasting specific hypotheses: (1) that SMR and boldness are correlated and will exert parallel influences on risk taking to breathe air, or (2) that they act independently and their influences differ in strength with prevailing context. We investigated the influence of resource availability by comparing normoxia with aquatic hypoxia, and the influence of perceived risk of predation by comparing periods when fish were more visible to predators (day versus night).

**MATERIALS AND METHODS****Animals**

Feral wild-caught juvenile *Clarias gariepinus* (Burchill 1822), with a mass of approximately 250 g, were obtained from a farm in São Paulo state and transported to the Department of Physiological Sciences, Federal University of São Carlos (São Carlos, SP, Brazil). There, they were maintained together in a single 1 m<sup>3</sup> tank supplied with well water at 25±1°C under a natural photoperiod and fed commercial feed daily at 2% body mass day<sup>-1</sup>, for 6 weeks. Animals were then tagged (with a passive integrated transponder or PIT) into the dorsal epaxial muscle under mild anaesthesia

(0.1 g l<sup>-1</sup> benzocaine), for individual identification, after which they recovered in routine holding conditions for 1 week before experiments. All experiments were performed at 25°C. Experiments were approved by the Ethics in Animal Experimentation Committee (CEUA) of the Federal University of São Carlos, according to Federal Law 11.794 (authorisation no. 067/2011).

**Bimodal respirometry**

Intermittent stopped-flow respirometry (Steffensen, 1989) modified for bimodal breathers (Lefevre et al., 2016) was used to measure RMR and respiratory partitioning over 24 h in both normoxia and aquatic hypoxia (4 kPa  $P_{O_2}$ , ~20% air saturation). The tagged catfish were exposed to aquatic conditions on separate occasions with half of the animals exposed to normoxia first, half to hypoxia, with at least 96 h recovery between exposure, during which they were returned to standard holding conditions. Catfish were fasted for 24 h prior to measurements in either condition, then individuals were transferred gently to respirometers without air exposure, to minimise effects of handling. There were four respirometer chambers immersed in well-aerated water in an outer bath (1 m<sup>2</sup> surface area, 21 cm water depth) with the entire setup screened behind tarpaulin so that routine air-breathing behaviour was not inhibited by fear of human presence (McKenzie et al., 1991, 2007; Shingles et al., 2005). Fish were placed in the respirometer in the evening between 18:00 h and 20:00 h and fish were then allowed 16 h to recover from handling. Collection of respirometry data started the following morning and continued with the animals undisturbed for 24 h.

The bimodal respirometers were partially immersed in the outer bath, with an underwater water phase of volume 5.72 l and an air phase that projected above the water surface with a volume of ~200 ml. For each respirometer, the exact volume of the air phase was derived from the effects on  $O_2$  partial pressure of a 20 ml bolus of 100%  $N_2$  delivered into the sealed space. The intermittent stopped-flow technique alternated two periods within a 15 min cycle. In the first, water and air chambers were closed to the exterior for 10 min, so that  $O_2$  uptake from both phases could be recorded (see below). The alternate period was when the two phases, aquatic and aerial, were flushed simultaneously for 5 min to replenish  $O_2$  levels. Aquatic hypoxia was created and controlled as described previously, with corrections made for rates of passive diffusion of  $O_2$  from air to water (McKenzie et al., 2007, 2012). Details of the system are provided in Lefevre et al. (2016).

Oxygen levels in the water and air were measured with an optode system (OXY-10 Micro, Pre-Sens Precision Sensing GmbH, www.presens.com), using eight channels such that four individuals were measured for their metabolic rate at any one time. Optodes were positioned to sample water and air for each respirometer, with all data stored on a PC using the manufacturer’s software. Absolute rates of  $O_2$  uptake from air ( $\dot{M}_{O_2,air}$ ) and water ( $\dot{M}_{O_2,water}$ ) were calculated (in mmol  $O_2$  kg<sup>-1</sup> h<sup>-1</sup>) for each 15 min respirometry cycle, based upon the decline in  $O_2$  content in each phase during the closed period, as described previously (McKenzie et al., 2007, 2012). The  $\dot{M}_{O_2,water}$  and  $\dot{M}_{O_2,air}$  were summed to calculate RMR. The proportion of RMR derived from air breathing was calculated as the percentage of  $\dot{M}_{O_2,air}$ . The  $\dot{M}_{O_2,air}$ ,  $\dot{M}_{O_2,water}$  and the resultant RMR and  $\% \dot{M}_{O_2,air}$  measures, were averaged over four sequential respirometry cycles to provide hourly means. These were averaged for the experimental population to reveal general diurnal patterns over 24 h.

**Individual variation in air-breathing behaviour**

The hourly values for the absolute rates of oxygen uptake from air, and the proportion of metabolism derived from aerial respiration, were averaged for each individual from 07:00 h to 18:00 h for daylight and from 19:00 h to 06:00 h for hours of darkness. This was repeated for normoxia and hypoxia.

**Standard metabolic rate**

The SMR was estimated by the quantile method (Dupont-Prinet et al., 2010; Chabot et al., 2016). This assumes that a certain proportion of all measured rates of RMR are below true SMR because of temporal variability and possible measurement errors. The quantile splits the dataset into the

$q$  smallest and the  $1-q$  largest values, where  $q$  is a proportion chosen by the experimenter. Here,  $q$  was fixed at 0.12 such that 12% of values fell below true SMR (Chabot et al., 2016). All measures of RMR were considered for each individual, both in normoxia and hypoxia, hence four measures per hour over 48 h, or 192 measures.

### Boldness

Boldness was evaluated as the time required to resume air breathing ( $T_{\text{res}}$ ) after a fearful stimulus, delivered when air breathing was especially risky but not physiologically essential: daylight normoxia. Following the respirometry in normoxia, the screen covering the respirometers was lifted for 1 min and each fish was disturbed by knocking sharply on the lid of their box 10 times, causing them to retreat to the bottom of the water phase. The  $T_{\text{res}}$  was taken as the time, in minutes, for the individual to resume oxygen uptake from air (Chapman et al., 2011). This represents the propensity to resume a risk-taking behaviour in a potentially dangerous environment (Huntingford et al., 2010; Killen et al., 2011, 2012). This was always performed between 14:00 h and 15:00 h to avoid confounding effects of diurnal rhythms in activity or air breathing.

$T_{\text{res}}$  was evaluated for evidence of a bimodal distribution, indicative of ‘bold’ versus ‘shy’ phenotypes (Frost et al., 2013; Huntingford et al., 2010; Thomson et al., 2012) because such typing into personalities (or ‘coping styles’) can be used to inform interpretation of individual variation in phenotypic traits in fishes (MacKenzie et al., 2009; Rey et al., 2013, 2015). The  $T_{\text{res}}$  data were inspected visually for evidence of two groups, one with relatively short  $T_{\text{res}}$  indicating bold individuals, and a second with relatively longer  $T_{\text{res}}$  indicating shy individuals (Frost et al., 2013; Huntingford et al., 2010; Thomson et al., 2012).

### Data analysis and statistics

Statistics were performed with SPSS Statistics v17.0 (www.ibm.com/software/analytics/spss). The level of significance for all tests was  $\alpha=0.05$ . To understand the variables that influenced air-breathing behaviour, linear mixed-effect models (LMEMs) were constructed with either absolute ( $\dot{M}_{\text{O}_2,\text{air}}$ ) or proportional ( $\% \dot{M}_{\text{O}_2,\text{air}}$ ) rates of air breathing as dependent variables, and fish mass,  $\text{O}_2$  availability (normoxia versus hypoxia), time of day (day versus night), SMR and  $T_{\text{res}}$  as fixed effects. Because each fish was tested under multiple conditions, fish identity was included as a random effect in the models, with time of day and oxygen availability treated as repeated factors. Initial models included all two-way interactions between SMR/ $T_{\text{res}}$  and each context of  $\text{O}_2$  availability and time of day. Non-significant interactions were sequentially dropped and the models re-run. Model assumptions of homogeneity, linearity and normality of residuals were verified by inspection of residuals–fits plots.  $T_{\text{res}}$  was log transformed prior to use in the model. When apposite, dependence of the air breathing upon SMR or  $T_{\text{res}}$  was demonstrated visually using ordinary least-squares regression under each time of day and level of oxygenation.

Following classification of individuals by their  $T_{\text{res}}$ , namely resuming air breathing either relatively fast (bold) or relatively slowly (shy), a separate LMEM was constructed to focus on daylight  $\dot{M}_{\text{O}_2,\text{air}}$  and  $\% \dot{M}_{\text{O}_2,\text{air}}$  as dependent variables. Fish mass,  $\text{O}_2$  availability (normoxia versus hypoxia) and  $T_{\text{res}}$  type (bold versus shy) were fixed effects. Once again, fish identity was included as a random effect in the models, with oxygen availability treated as a repeated factor.

## RESULTS

Data were collected for 29 individuals in normoxia, with a mean ( $\pm$ s.d.) mass of  $292 \pm 60$  g. Among these, data were collected for 24 fish in aquatic hypoxia, with a mean mass of  $300 \pm 58$  g.

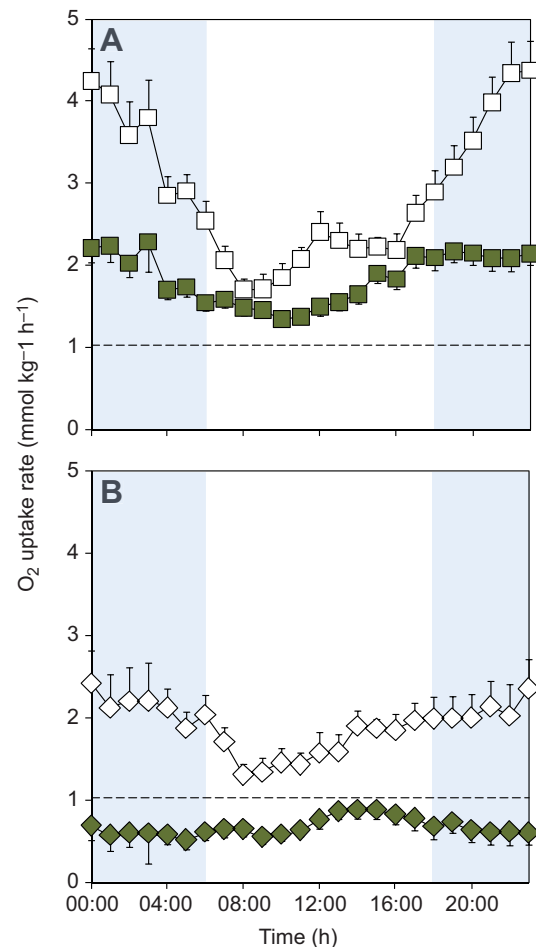
### General patterns of respiratory partitioning

In normoxia, the respirometry data showed a clear diurnal pattern in routine metabolism. Mean hourly RMR was relatively low during the day and much higher during the night (Fig. 1A). There were

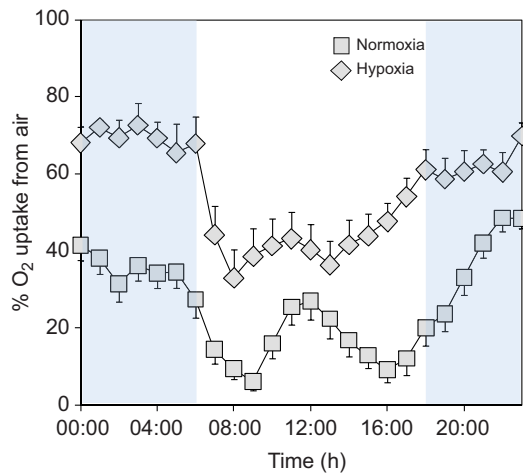
measurable rates of mean  $\text{O}_2$  uptake from air ( $\dot{M}_{\text{O}_2,\text{air}}$ ) throughout the diurnal cycle, but these increased markedly in darkness, contributing to a marked increase in RMR. The mean rates of uptake from water ( $\dot{M}_{\text{O}_2,\text{water}}$ ) were considerably less variable over the diurnal cycle. The mean SMR of the population was always below mean  $\dot{M}_{\text{O}_2,\text{water}}$  in normoxia (Fig. 1A). This presumably indicates that the catfish could meet their  $\text{O}_2$  demands for SMR from gill ventilation alone (Belão et al., 2011).

In aquatic hypoxia, there was still a clear diurnal rhythm in routine metabolism (Fig. 1B). The mean RMR was, however, visibly lower than in normoxia; this was linked to very low mean hourly rates of aquatic respiration, and  $\dot{M}_{\text{O}_2,\text{w}}$  varied very little over the diurnal cycle (Fig. 1B). Furthermore, mean  $\dot{M}_{\text{O}_2,\text{w}}$  was always below mean SMR in aquatic hypoxia, which indicates that the catfish would need to resort to air breathing to meet their basal  $\text{O}_2$  demands.

Calculation of diurnal patterns of the percentage of routine metabolism that was due to  $\text{O}_2$  uptake from air ( $\% \dot{M}_{\text{O}_2,\text{air}}$ ) in normoxia and hypoxia (Fig. 2) confirmed that this showed a large increase at night, especially in normoxia, and also that there was an overall much greater proportional reliance on air breathing in hypoxia.



**Fig. 1. Routine metabolic rate and aquatic oxygen uptake in juvenile *Clarias gariepinus*.** Routine metabolic rate (white symbols) and aquatic oxygen uptake (green symbols) measured over 24 h in (A) normoxia (squares,  $N=29$ ) or (B) aquatic hypoxia at 20% air saturation (diamonds,  $N=25$ ). The difference between aquatic oxygen uptake and RMR is due to oxygen uptake from air. Shaded blue areas represent night-time hours. The dotted line is mean standard metabolic rate. Values are means  $\pm$  s.e.



**Fig. 2.** The percentage of routine metabolic rate derived from air ( $\% \dot{M}_{O_2,air}$ ) over 24 h in juvenile *Clarias gariepinus*, in either normoxia ( $N=29$ ) or aquatic hypoxia at 20% air saturation ( $N=25$ ). The shaded blue areas represent night-time hours. Values are means  $\pm$  s.e.

Table 1 carries descriptive statistics for the two measures of risk-taking to breath air, namely  $\dot{M}_{O_2,air}$  and  $\% \dot{M}_{O_2,air}$ , which were calculated separately for daylight hours (07:00 h to 18:00 h) and night-time darkness (19:00 h to 06:00 h), in normoxia and then hypoxia. It also shows mean SMR. These traits all varied at least two-fold within the sample population.

#### Boldness as $T_{res}$

Table 1 carries descriptive statistics of the measure of boldness,  $T_{res}$ , which varied widely within the experimental population. There was evidence of a bimodal distribution (Fig. S1), whereby individual catfish either resumed air breathing in less than 75 min ( $N=13$ ) or in more than 115 min ( $N=16$ ). Two groups, with mean ( $\pm$ s.e.)  $T_{res}$  of  $37 \pm 5$  versus  $181 \pm 14$  min, were therefore designated as bold versus shy phenotypes, respectively (Frost et al., 2013; Huntingford et al., 2010; Rey et al., 2013; Thomson et al., 2012).

#### Associations between SMR and $T_{res}$

There was no correlation between body mass ( $M_b$ ) and SMR or  $T_{res}$ , over the small range in mass of the experimental population. Individual SMR and  $T_{res}$  were, however, negatively correlated

**Table 1.** Descriptive statistics of rates of oxygen uptake from air and percentage total metabolic rate derived from air by juvenile *Clarias gariepinus*

	Statistic			
	N	mean $\pm$ s.d.	CV	Range
Normoxia				
Day $\dot{M}_{O_2,air}$	29	0.52 $\pm$ 0.41	0.791	0.00–1.43
Night $\dot{M}_{O_2,air}$	29	1.59 $\pm$ 0.85	0.537	0.59–3.66
Day $\% \dot{M}_{O_2,air}$	29	16.3 $\pm$ 10.9	0.664	0.0–34.6
Night $\% \dot{M}_{O_2,air}$	29	37.6 $\pm$ 12.1	0.320	15.4–64.1
Hypoxia				
Day $\dot{M}_{O_2,air}$	25	0.98 $\pm$ 0.52	0.528	0.36–2.84
Night $\dot{M}_{O_2,air}$	25	1.49 $\pm$ 0.39	0.257	0.71–2.31
Day $\% \dot{M}_{O_2,air}$	25	46.5 $\pm$ 17.6	0.378	14.5–83.3
Night $\% \dot{M}_{O_2,air}$	25	66.1 $\pm$ 12.4	0.187	42.5–91.6
SMR	29	1.02 $\pm$ 0.32	0.308	0.48–1.85
$T_{res}$	29	117 $\pm$ 83	0.715	7–325

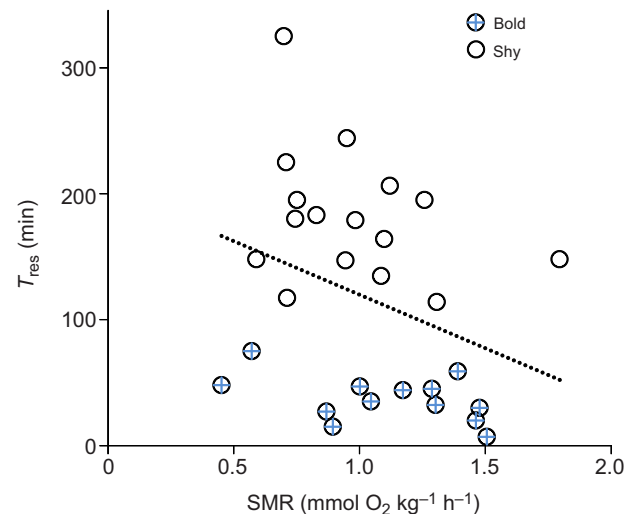
$\dot{M}_{O_2,air}$  and SMR are in  $\text{mmol O}_2 \text{ kg}^{-1} \text{ h}^{-1}$ ;  $T_{res}$  in minutes. CV, coefficient of variation.

(Spearman Rank  $R=-0.391$ ,  $P=0.036$ ,  $N=29$ ), such that catfish with higher basal  $\text{O}_2$  demands resumed risky air breathing relatively more quickly after the fearful stimulus (Fig. 3). Interestingly, the bold and shy phenotypes did not distribute at opposing ends of this relationship. In fact, SMR did not differ between them, being  $1.11 \pm 0.10$  versus  $0.97 \pm 0.08 \text{ mmol O}_2 \text{ kg}^{-1} \text{ h}^{-1}$ , for bold and shy fish, respectively, with a very similar range of values (Fig. 3). It is noteworthy, however, that all of the fish with bold phenotypes had a  $T_{res}$  that fell below the line describing the least-squares linear correlation, while almost all of the fish with shy phenotypes were above it (Fig. 3). The mean residuals for bold fish were  $-73 \pm 8$  min, significantly less than the residuals for shy fish, which were  $60 \pm 13$  min ( $t$ -test  $P < 0.00001$ ). That is, the bold fish were resuming air breathing an average of 73 min faster than would be predicted from their SMR, whereas the shy fish resumed air breathing an average of 60 min slower.

#### Influence of individual SMR and $T_{res}$ on variation in risk-taking to breathe air

The LMEMs did not reveal that SMR and  $T_{res}$  co-varied in their influence upon  $\dot{M}_{O_2,air}$  or  $\% \dot{M}_{O_2,air}$  (Tables 2 and 3). In terms of absolute rates of oxygen uptake from air, there was a significant dependence of  $\dot{M}_{O_2,air}$  on time of day, water  $\text{O}_2$  availability and SMR, but not upon  $T_{res}$  (Table 2). There were no interactions among the factors. The estimates of effects confirmed that the catfish breathed significantly less air during the daytime, with a significantly lower  $\dot{M}_{O_2,air}$  than at night. In hypoxia, there was a general increase in  $\dot{M}_{O_2,air}$  compared with normoxia. Across all these contexts, there was a positive dependence of  $\dot{M}_{O_2,air}$  on SMR: individuals with higher basal  $\text{O}_2$  demands had higher rates of aerial respiration (Table 2).

In terms of the percentage of total metabolic rate that was met by air breathing, the model revealed no significant main effects of time of day, oxygen availability, SMR or  $T_{res}$  on  $\% \dot{M}_{O_2,air}$  (Table 3). There were, however, significant interactions amongst these explanatory variables. Thus, there was a significant interaction between effects of  $\text{O}_2$  availability and SMR, whereby an effect of



**Fig. 3.** Relationship between individual standard metabolic rate and time to resume air breathing after a simulated predator attack in juvenile *Clarias gariepinus*. Scatterplot of the relationship between SMR and time to resume air breathing after a simulated predator attack ( $T_{res}$ ). There was a significant negative Spearman rank correlation ( $R=-0.391$ ,  $P=0.036$ ,  $N=29$ ). The line is a least-squares linear fit. Fish with bold  $T_{res}$  phenotypes resumed air breathing in less than 75 min (see text for further details).

**Table 2. Estimates of fixed effects from a linear mixed-effect model to evaluate the dependence of individual variation in rates of oxygen uptake from the air**

A. Fixed effects				
Source	Numerator d.f.	Denominator d.f.	F	P
Intercept	1	21.7	2.17	0.155
$M_b$	1	20.0	0.22	0.643
Water O <sub>2</sub> level	1	58.8	12.52	<b>&lt;0.001</b>
Time of day	1	46.8	56.06	<b>&lt;0.001</b>
SMR	1	22.2	13.58	<b>0.001</b>
log $T_{res}$	1	67.0	0.54	0.466

## B. Estimates of fixed effects

Parameter	Estimate±s.e.	d.f.	t	P
Intercept	0.19±0.43	44.0	0.45	0.656
$M_b$	0.87±0.85	22.7	1.02	0.321
Water oxygen=hypoxia*	0.35±0.10	73.6	3.38	<b>0.001</b>
Time=day*	-0.66±0.09	48.9	-7.35	<b>&lt;0.001</b>
SMR	0.61±0.17	22.2	3.68	<b>0.001</b>
log $T_{res}$	0.08±0.11	67.0	0.734	0.466

Rates of oxygen uptake from air were measured with bimodal respirometry for every hour over 24 h, either in normoxia ( $N=29$  individuals) or in aquatic hypoxia (20% of air saturation,  $N=25$  individuals). For each oxygen level, averages were calculated for each individual for either daylight hours (07:00 h–18:00 h) or darkness (19:00 h–06:00 h). SMR was assessed from the respirometry,  $T_{res}$  as the time in minutes to resume air breathing after a simulated predator attack in daylight normoxia. See text for further details.

\*Alternative parameter is set to zero because it is redundant. Parameters with significant effects are bold.  $M_b$ , fish body mass.

individual basal O<sub>2</sub> demand on  $\% \dot{M}_{O_{2,air}}$  was only statistically significant in aquatic hypoxia (Table 3). Regressions of  $\% \dot{M}_{O_{2,air}}$  against SMR for the four context combinations (Fig. 4A,B) demonstrate a significant positive dependence of  $\% \dot{M}_{O_{2,air}}$  on SMR in hypoxia only (Fig. 4B), for both times of day. That is, in hypoxia, individuals with higher basal O<sub>2</sub> demands tended to obtain proportionally more of their O<sub>2</sub> from the air.

The LMEM also revealed a significant interaction between  $T_{res}$  and time of day, whereby a significant dependence of  $\% \dot{M}_{O_{2,air}}$  on individual  $T_{res}$  was found, but only during the daytime (Table 3). Regressions of  $\% \dot{M}_{O_{2,air}}$  against  $T_{res}$  for the four different context combinations (Fig. 5A,B) revealed significant negative dependence of  $\% \dot{M}_{O_{2,air}}$  on  $T_{res}$  during the day but not the night, in both normoxia and hypoxia. That is, during the daytime, individuals that had shorter  $T_{res}$  tended to obtain proportionally more of their O<sub>2</sub> from the air.

### Influence of $T_{res}$ phenotype on variation in risk taking to breathe air

The model did not reveal a significant dependence of daylight  $\dot{M}_{O_{2,air}}$  on  $T_{res}$  phenotype (data not shown), but did reveal a dependence for daylight  $\% \dot{M}_{O_{2,air}}$  (Table 4). The proportion of O<sub>2</sub> obtained from air was significantly greater in bold compared with shy phenotypes during the day, with no interaction with level of oxygen availability (Table 4). That is, further to the complex influences of individual SMR and  $T_{res}$  on  $\% \dot{M}_{O_{2,air}}$ , bold fish breathed proportionally more air than shy fish during daylight hours, in both normoxia and hypoxia.

## DISCUSSION

The results support the general hypothesis that the tendency to take risks to gulp air depends upon both basal O<sub>2</sub> demand and intrinsic boldness in a fish with bimodal respiration. Context itself, namely

**Table 3. Estimates of fixed effects from a linear mixed-effect model to evaluate the dependence of individual variation in percentage total metabolic rate derived from air**

A. Fixed effects				
Source	Num. d.f.	Den. d.f.	F	P
Intercept	1	23.7	5.88	<b>0.023</b>
$M_b$	1	22.5	1.06	0.314
Water O <sub>2</sub> level	1	66.4	3.30	0.074
Time of day	1	48.6	0.39	0.537
SMR	1	20.9	3.45	0.077
log $T_{res}$	1	24.0	0.63	0.436
Water O <sub>2</sub> ×SMR	1	66.5	5.18	<b>0.026</b>
Time of day×log $T_{res}$	1	48.4	7.23	<b>0.010</b>

## B. Estimates of fixed effects

Parameter	Estimate±s.e.	d.f.	t	P
Intercept	20.9±14.0	32.7	1.50	0.144
$M_b$	23.5±22.8	22.5	0.62	0.537
Water O <sub>2</sub> =hypoxia*	13.4±7.4	66.3	1.81	0.074
Time=day*	6.3±10.1	48.6	0.62	0.537
SMR	1.0±5.2	32.2	0.19	0.853
log $T_{res}$	4.0±4.4	35.9	0.92	0.366
Hypoxia×SMR*	15.7±6.9	66.5	2.28	<b>0.026</b>
Day×log $T_{res}$ *	-13.8±5.1	48.4	-2.69	<b>0.010</b>

Rates of oxygen uptake from air and water were measured with bimodal respirometry for every hour over 24 h, either in normoxia ( $N=29$  individuals) or in aquatic hypoxia (20% of air saturation,  $N=24$  individuals), to derive  $\% \dot{M}_{O_{2,air}}$ . For each oxygen level, averages were calculated for each individual for either daylight hours (07:00 h–18:00 h) or darkness (19:00 h–06:00 h). SMR was assessed from the respirometry,  $T_{res}$  as the time in minutes to resume air breathing after a simulated predator attack in daylight normoxia. The LMEM tested for interactions between water oxygen level and SMR, and between time of day and  $T_{res}$ . See text for further details.

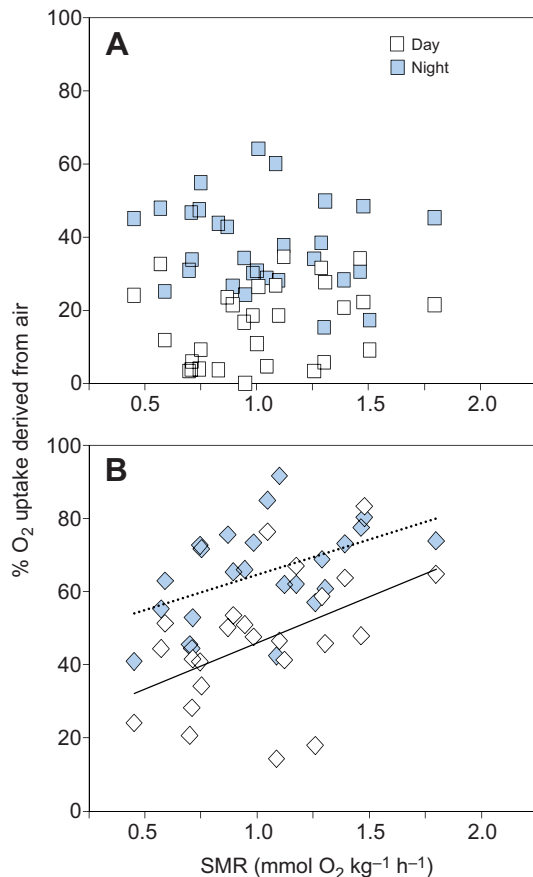
\*Alternative parameter is set to zero because it is redundant. Significant effects are in bold.  $M_b$ , fish body mass.

O<sub>2</sub> availability (normoxia versus hypoxia) or level of perceived risk (day versus night) had profound effects on air-breathing behaviour. The results provided apparently paradoxical support for both specific hypotheses because, although SMR and  $T_{res}$  were correlated, there was clear evidence of independent influences that varied in strength according to the context.

### General patterns of respiratory metabolism and boldness

The marked circadian rhythm in RMR and air-breathing behaviour by *C. gariepinus* is consistent with reports of diurnal variation in activity and a pronounced nocturnal habit in this species (Britz and Pienaar, 1992; Bruton, 1979; Willoughby and Tweddle, 1978). In the wild, juvenile catfish emerge from cover at night, to forage (Bruton, 1979; Willoughby and Tweddle, 1978). Therefore, the overall increase in RMR and air breathing at night presumably reflects an increase in activity by the animals (Lefevre et al., 2014a, b), which is at least partly due to a reduced fear of detection by predators (Britz and Pienaar, 1992).

The fact that SMR was below  $\dot{M}_{O_{2,air}}$  in aquatic normoxia confirms that, in well-aerated water, the species is able to meet and exceed basal O<sub>2</sub> demands by gill ventilation alone and is, indeed, a facultative air breather (Belão et al., 2011). In aquatic hypoxia, the overall increase in air breathing was expected (Belão et al., 2011) and the data demonstrate that the catfish needed to breathe air because aquatic respiration could no longer support their basal metabolic demands, mean  $\dot{M}_{O_{2,water}}$  was consistently below mean SMR. The overall decline in RMR that was also visible in aquatic hypoxia, over the entire daily cycle when compared with normoxia,

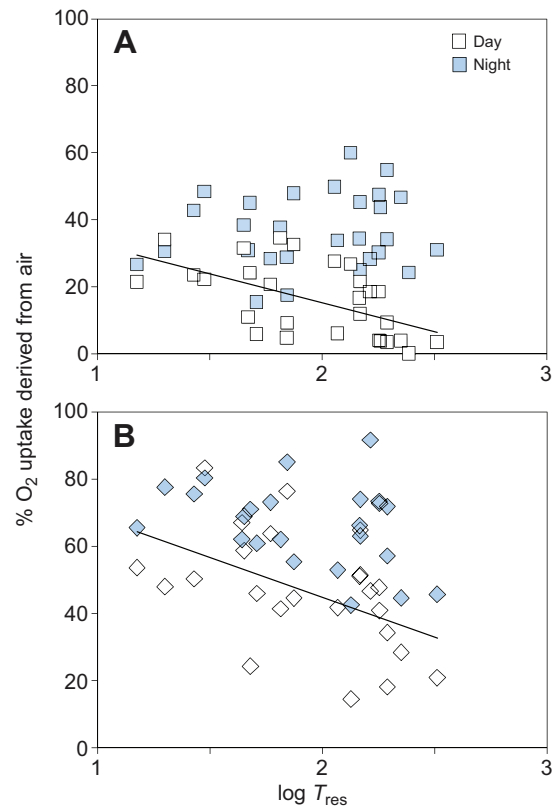


**Fig. 4. The dependence of individual percentage of oxygen uptake obtained from air on standard metabolic rate of juvenile *Clarias gariepinus*.** Scatterplots of the relationship between individual SMR and  $\% \dot{M}_{O_2,air}$  in (A) normoxia ( $N=29$ ) and (B) aquatic hypoxia at 30% air saturation ( $N=24$ ). For normoxic daytime,  $\% \dot{M}_{O_2,air} = 7.64 + 8.45 \times SMR$ ,  $R^2 = 0.063$ ,  $P = 0.187$ ; for normoxic night-time,  $\% \dot{M}_{O_2,air} = 43.90 - 6.11 \times SMR$ ,  $R^2 = 0.030$ ,  $P = 0.395$ ; for hypoxic daytime,  $\% \dot{M}_{O_2,air} = 20.86 + 25.22 \times SMR$ ,  $R^2 = 0.318$ ,  $P = 0.017$  (B, solid line); for hypoxic night-time,  $\% \dot{M}_{O_2,air} = 45.42 + 19.27 \times SMR$ ,  $R^2 = 0.231$ ,  $P = 0.017$  (B, dotted line).

has been observed previously in facultative air-breathing fishes and may reflect a decline in spontaneous activity levels (Lefevre et al., 2012, 2014a,b; McKenzie et al., 2012).

Our measure of boldness – the time to resume air breathing after a fearful stimulus – is similar to previous studies that have, for example, evaluated boldness as the time required to emerge from cover into a potentially threatening environment (Huntingford et al., 2010; Killen et al., 2011, 2012; MacKenzie et al., 2009).  $T_{res}$  should be a valid measure of intrinsic boldness because our respirometry data confirm that the catfish is a facultative air breather in aquatic normoxia. By definition, therefore, the animals should not, in theory, have had any physiological drive to take risks to obtain oxygen from air.

The evidence of a bimodal distribution in  $T_{res}$ , with bold and shy phenotypes, is consistent with previous studies on fishes, which have used various tests, such as emergence from cover or response to novel objects, to classify phenotypes as bold or shy (Frost et al., 2013; Huntingford et al., 2010; Rey et al., 2013; Thomson et al., 2012). Consideration of this apparent difference in personality improved our ability to interpret other elements of individual variance in our data (MacKenzie et al., 2009; Rey et al., 2013, 2015). In particular and as discussed below, the apparent paradox



**Fig. 5. The dependence of individual percentage of oxygen uptake obtained from air on time to resume air breathing after a simulated predator attack of juvenile *Clarias gariepinus*.** Scatterplots of the relationship between  $\log T_{res}$  and  $\% \dot{M}_{O_2,air}$  in (A) normoxia ( $N=29$ ) or (B) aquatic hypoxia at 25% air saturation ( $N=24$ ) in the day or night. For normoxic daytime,  $\% \dot{M}_{O_2,air} = 49.83 - 17.28 \times \log T_{res}$ ,  $R^2 = 0.285$ ,  $P = 0.002$  (A, solid line); for normoxic night-time,  $\% \dot{M}_{O_2,air} = 27.08 + 4.91 \times \log T_{res}$ ,  $R^2 = 0.013$ ,  $P = 0.431$ ; for hypoxic daytime,  $\% \dot{M}_{O_2,air} = 92.23 - 23.71 \times \log T_{res}$ ,  $R^2 = 0.206$ ,  $P = 0.015$  (B, solid line); for hypoxic night-time,  $\% \dot{M}_{O_2,air} = 83.46 - 9.03 \times \log T_{res}$ ,  $R^2 = 0.028$ ,  $P = 0.211$ .

whereby SMR and  $T_{res}$  were correlated but also exerted independent and context-dependent effects on risk taking.

#### **$T_{res}$ is correlated with SMR but also possesses an independent element of boldness**

The significant negative correlation between SMR and  $T_{res}$  is consistent with proposals that individuals with higher basal oxygen demands should also be bolder (Biro and Stamps, 2010; Careau et al., 2008; Réale et al., 2010). It would be predicted, therefore, that catfish with the shortest  $T_{res}$ , with bold phenotypes, would have a significantly higher mean SMR than the shy fish. Instead, they had a similar overall range of values for basal metabolism. Within the overall correlation between SMR and  $T_{res}$  there was, however, clear evidence that the bold fish were indeed intrinsically more risk-prone than shy fish, because the former resumed air breathing relatively sooner than would be predicted from their SMR.

One way of interpreting these results is that there are two mechanisms at work. The general correlation of SMR and  $T_{res}$  may, in fact, be a consequence of a chemoreflexive respiratory drive to gulp air in the catfish. As discussed in detail below, the overall dependence of  $\dot{M}_{O_2,air}$  on SMR reveals such a chemoreflexive drive, whereas the context-dependent influence of SMR on  $\% \dot{M}_{O_2,air}$  in hypoxia confirms it. The second mechanism appears to involve an independent and intrinsic difference in boldness, causing individuals to resume air breathing either relatively faster or



**Table 4. Estimates of fixed effects from a linear mixed-effect model to evaluate the dependence of individual variation in percentage total metabolic rate derived from air during daylight hours**

A. Fixed effects				
Source	Numerator d.f.	Denominator d.f.	<i>F</i>	<i>P</i>
Intercept	1	27.3	13.91	<b>&lt;0.001</b>
$M_b$	1	24.1	0.04	0.950
Water O <sub>2</sub> level	1	24.1	64.14	<b>&lt;0.001</b>
$T_{res}$ phenotype	2	25.1	4.29	<b>0.025</b>
B. Estimates of fixed effects				
Parameter	Estimate±s.e.	d.f.	<i>t</i>	<i>P</i>
Intercept	11.3±8.4	24.1	1.35	0.191
$M_b$	1.8±28.1	24.1	0.06	0.950
Water O <sub>2</sub> =hypoxia*	30.2±3.7	24.1	8.01	<b>&lt;0.001</b>
$T_{res}$ phenotype=bold*	9.8±3.5	24.6	2.78	<b>0.010</b>

Rates of oxygen uptake from air and water were measured with bimodal respirometry for every hour over 24 h, either in normoxia ( $N=29$  individuals) or in aquatic hypoxia (20% of air saturation,  $N=24$  individuals), to derive % $\dot{M}_{O_2,air}$ . For each oxygen level, averages were calculated for each individual for daylight hours (07:00 h–18:00 h).  $T_{res}$  is the time in minutes to resume air breathing after a simulated predator attack in daylight normoxia, where fish with bold phenotypes resumed in less than 75 min ( $N=13$ ), shy fish in greater than 115 min ( $N=16$ ). The LMEM tested for interactions between water oxygen level and boldness phenotype. See text for further details.

\*Alternative parameter is set to zero because it is redundant. Significant effects are bold.  $M_b$ , fish body mass.

slower than their SMR and respiratory drive would predict. This difference in personality was then linked to clear differences in risk-taking during the daytime, as discussed in detail further below.

### Individual standard metabolic rate exerts a pervasive influence on air breathing

The fact that the absolute rates of O<sub>2</sub> uptake from air were dependent upon individual SMR, across all contexts, was unexpected. A facultative air-breathing fish should not, in theory, have a physiological drive to take risks to gulp air in normoxia. One possible explanation for this apparent inconsistency would be that the surfacing responses are, at least in part, inescapable neurophysiological reflexes (Lefevre et al., 2014a). In freshwater fishes, air breathing is believed to have evolved as a response to aquatic hypoxia (Graham, 1997; Johansen, 1968, 1970; Randall et al., 1981). Hypoxic surfacing responses are reflexes, stimulated by oxygen-sensitive chemoreceptors in the gills, which monitor water and blood O<sub>2</sub> levels (Chapman and McKenzie, 2009, 1991; Milsom, 2012). Although these reflex circuits may have evolved because they allow bimodal fishes to increase air breathing and regulate their metabolic rate in aquatic hypoxia, it is conceivable that they also cause them to perform surfacing responses at irregular intervals in normoxia (Lefevre et al., 2014a). It has been proposed that the aperiodic gulping of air by animals with bimodal respiration is indeed a chemoreflex, whereby blood oxygen levels decline progressively over time after an air breath, this is monitored by oxygen-sensitive chemoreceptors in the vasculature that eventually stimulate the animal to surface and take another gulp (Milsom, 2012; Shelton et al., 1984). The overall positive relationship between SMR and  $\dot{M}_{O_2,air}$  could be explained by such a physiological mechanism: if the decline in blood oxygen occurs more rapidly in individuals with higher basal metabolic demands, this will stimulate more reflex air-breathing responses.

This physiological basis to air-breathing responses was almost certainly the mechanism underlying the dependence of % $\dot{M}_{O_2,air}$  on

SMR in aquatic hypoxia. It seems logical to presume that, the higher an individual's SMR, the more their metabolic equilibrium was challenged in aquatic hypoxia (Killen et al., 2012, 2013). Chemoreflexive responses would presumably be stimulated relatively more profoundly and frequently in individuals that were consuming oxygen from their blood at the highest rates.

This evidence of a pervasive chemoreflexive drive to breathe air, even in normoxia, could then explain why SMR and  $T_{res}$  were correlated. Individuals with higher SMR would have a stronger chemoreflexive drive to breathe air and, therefore, would exhibit a reflex response more rapidly after the fearful stimulus, irrespective of any influence of their personality.

### Individual boldness exerts a context-dependent influence on air breathing

The current study provides the first evidence that personality can influence risky air-breathing behaviour by an aquatic vertebrate. It is particularly interesting that this effect was context dependent and independent of any respiratory drive. Our measure of intrinsic boldness,  $T_{res}$ , was clearly linked to the proportion of O<sub>2</sub> obtained from air at a time of day when, in the wild, surfacing would be especially risky, because of the danger of visual detection by predators (Bruton, 1979; Willoughby and Tweddle, 1978). The effect of intrinsic boldness on risk taking was confirmed by the fact that bold  $T_{res}$  phenotypes breathed proportionally more air than fish with shy phenotypes during the daytime.

It is conceivable that choosing to surface and tap O<sub>2</sub>-rich air may allow individuals to complete metabolic activities such as digesting meals more rapidly, and so sustain higher rates of foraging, energy flux and growth in their environment. This would be analogous to previous reports of bolder animals achieving faster growth via increased foraging rates (Biro et al., 2004). It is also conceivable that these air-breathing responses provided other benefits to bolder animals, such as improved maintenance of buoyancy (Hedrick and Jones, 1993) or even improved auditory sensitivity (Shao et al., 2014).

### Conclusions and perspectives

The current data contribute to the ongoing debate regarding the relationships between physiological and behavioural traits, although they indicate that these are not simple. Although our finding that SMR and  $T_{res}$  were correlated is consistent with suggestions that selection should co-adapt physiology and temperament (Biro and Stamps, 2010; Careau and Garland, 2012; Réale et al., 2010; Stamps, 2007), in the current study, the correlation probably reflects our method of measuring boldness. Although we were able to show separate independent influences of metabolic demand and intrinsic boldness on risky air-breathing behaviour, future experiments in air-breathing fishes should utilise measures of boldness that are not confounded by chemoreflexive respiratory drive. This first attempt to disentangle the influences of physiology and personality on air breathing in an aquatic vertebrate highlights the complexities of designing experiments to evaluate causal relationships between energy metabolism and personality (Killen et al., 2013).

Chemoreflexes can be invoked to explain why individuals with high SMR had higher absolute rates of aerial respiration in all situations. This became most prominent during aquatic hypoxia, when the need to obtain atmospheric O<sub>2</sub> constrained risk-taking behaviour, forcing individuals with higher basal metabolic demands to surface relatively more frequently. The question remains as to why, if individual energy metabolism has such a pervasive

influence on risky air-gulping, animals with high SMR did not all have bold  $T_{res}$  phenotypes. Instead, independent effects of intrinsic boldness were observed during the daytime, including in normoxia when surfacing was seemingly non-essential, but potentially more risky. The relative influences of SMR and boldness were much more complex than might be predicted from correlational selection on a similar life history trade-off.

While the relative costs and benefits of foraging activity in relation to predation risk have been well studied (Biro and Stamps, 2008; Biro et al., 2004; Sih et al., 2012; Stamps, 2007), the trade-offs involved in access to other vital resources, such as oxygen for fishes, are little understood and require further investigation. The complex associations among energy demand, personality and the emergent tendency to take risks, indicate that cause–effect links between physiology and behaviour are dynamic and dependent upon context.

#### Acknowledgements

The authors are grateful to two anonymous referees for comments on a previous version of this manuscript. In particular, for valuable suggestions regarding analysis of the behavioural data. The authors are also grateful to Cesar Poletini and Piscicultura Poletini, Mogi Mirim, Brazil, for providing the experimental animals.

#### Competing interests

The authors declare no competing or financial interests.

#### Author contributions

D.M., T.B. and F.R. conceived and designed the experiments; D.M. and T.B. performed the experiments and compiled the raw data; D.M., T.B. and S.K. analyzed the data; D.M., T.B., S.K. and F.R. wrote the paper.

#### Funding

This study was funded in part by the Brazilian National Council for Scientific and Technological Development - CNPq (400177/2011-0 and 301921/2009-1). S.S.K. was supported by NERC Advanced Fellowship NE/J019100/1. Deposited in PMC for immediate release.

#### Supplementary information

Supplementary information available online at <http://jeb.biologists.org/lookup/suppl/doi:10.1242/jeb.122903/-DC1>

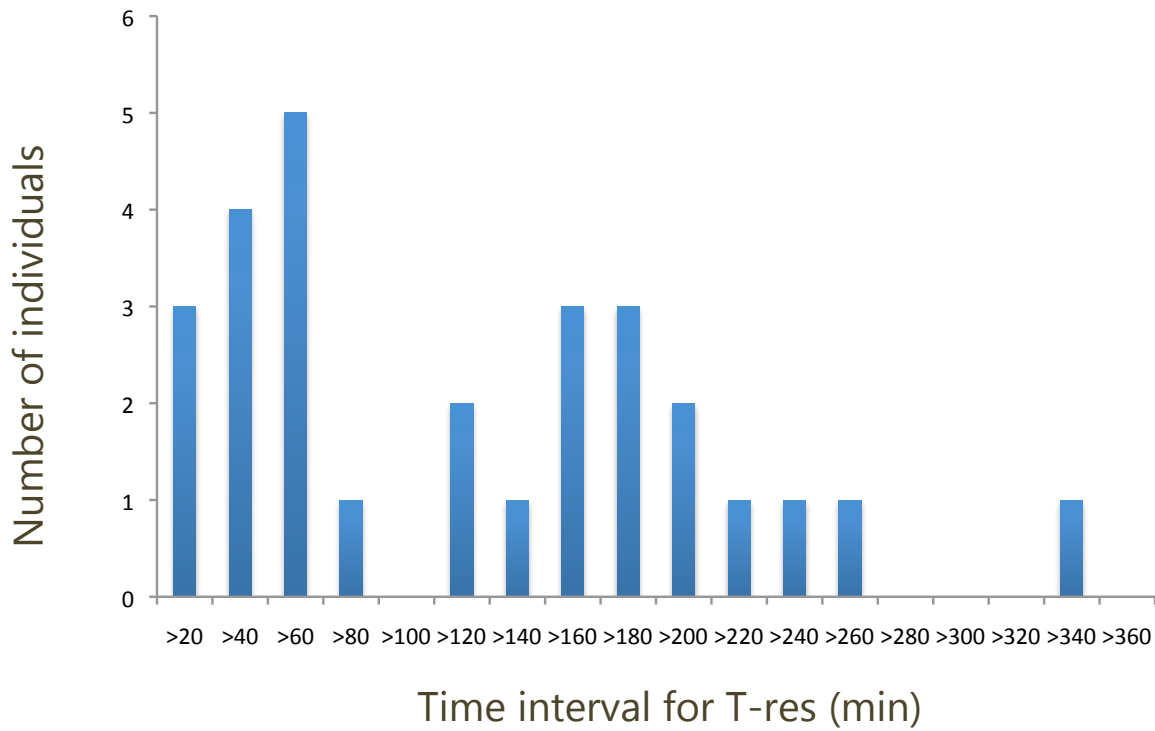
#### References

- Anholt, B. R. and Werner, E. E. (1998). Predictable changes in predation mortality as a consequence of changes in food availability and predation risk. *Evol. Ecol.* **12**, 729–738.
- Belão, T. C., Leite, C. A. C., Florindo, L. H., Kalinin, A. L. and Rantin, F. T. (2011). Cardiorespiratory responses to hypoxia in the African catfish, *Clarias gariepinus* (Burchell 1822), an air-breathing fish. *J. Comp. Physiol. B.* **181**, 905–916.
- Biro, P. A. and Stamps, J. A. (2008). Are animal personality traits linked to life-history productivity? *Trends Ecol. Evol.* **23**, 361–368.
- Biro, P. A. and Stamps, J. A. (2010). Do consistent individual differences in metabolic rate promote consistent individual differences in behavior? *Trends Ecol. Evol.* **25**, 653–659.
- Biro, P. A., Abrahams, M. V., Post, J. R. and Parkinson, E. A. (2004). Predators select against high growth rates and risk-taking behaviour in domestic trout populations. *Proc. R. Soc. B Biol. Sci.* **271**, 2233–2237.
- Britz, P. J. and Pienaar, A. G. (1992). Laboratory experiments on the effect of light and cover on the behaviour and growth of African catfish, *Clarias gariepinus* (Pisces: Clariidae). *J. Zool.* **227**, 43–62.
- Bruton, M. N. (1979). The food and feeding behaviour of *Clarias gariepinus* (Pisces: Clariidae) in Lake Sibaya, South Africa, with emphasis on its role as a predator of cichlids. *Trans. Zool. Soc. London* **35**, 47–114.
- Burton, T., Killen, S. S., Armstrong, J. D. and Metcalfe, N. B. (2011). What causes intraspecific variation in resting metabolic rate and what are its ecological consequences? *Proc. R. Soc. B Biol. Sci.* **278**, 3465–3473.
- Careau, V. and Garland, T. (2012). Performance, personality, and energetics: correlation, causation, and mechanism. *Physiol. Biochem. Zool.* **85**, 543–571.
- Careau, V., Thomas, D., Humphries, M. M. and Réale, D. (2008). Energy metabolism and animal personality. *Oikos* **117**, 641–653.
- Chabot, D., Steffensen, J. F. and Farrell, A. P. (2016). Measuring standard metabolic rate in fishes. *J. Fish Biol.* in press.
- Chapman, L. J. and McKenzie, D. J. (2009). Behavioral responses and ecological consequences. In *Hypoxia, Fish Physiology*, Vol. 27 (ed. J. G. Richards, A.P. Farrell and C.J. Brauner), pp. 25–77. San Diego: Elsevier Inc.
- Chapman, B. B., Hulthén, K., Blomqvist, D. R., Hansson, L.-Å., Nilsson, J.-Å., Brodersen, J., Anders Nilsson, P., Skov, C. and Brönmark, C. (2011). To boldly go: individual differences in boldness influence migratory tendency. *Ecol. Lett.* **14**, 871–876.
- Conrad, J. L., Weinersmith, K. L., Brodin, T., Saltz, J. B. and Sih, A. (2011). Behavioural syndromes in fishes: a review with implications for ecology and fisheries management. *J. Fish Biol.* **78**, 395–435.
- Diaz, R. J. and Breitburg, D. L. (2009). Hypoxia. In *Hypoxia, Fish Physiology*, Vol. 27 (ed. J. G. Richards, A. P. Farrell and C. J. Brauner), pp. 1–23. San Diego: Elsevier.
- Dupont-Prinet, A., Chatain, B., Grima, L., Vandeputte, M., Claireaux, G. and McKenzie, D. J. (2010). Physiological mechanisms underlying a trade-off between growth rate and tolerance of feed deprivation in the European sea bass (*Dicentrarchus labrax*). *J. Exp. Biol.* **213**, 1143–1152.
- Frost, A. J., Thomson, J. S., Smith, C., Burton, H. C., Davis, B., Watts, P. C. and Sneddon, L. U. (2013). Environmental change alters personality in the rainbow trout, *Oncorhynchus mykiss*. *Anim. Behav.* **85**, 1199–1207.
- Graham, J. B. (1997). *Air Breathing Fishes: Evolution, Diversity, and Adaptation*. San Diego: Academic Press.
- Hecht, T. and Pienaar, A. G. (1993). A Review of cannibalism and its implications in fish larviculture. *J. World Aquac. Soc.* **24**, 246–261.
- Hedrick, M. S. and Jones, D. R. (1993). The effects of altered aquatic and aerial respiratory gas concentrations on air-breathing patterns in a primitive fish (*Amia calva*). *J. Exp. Biol.* **94**, 81–94.
- Huntingford, F. A., Andrew, G., Mackenzie, S., Morera, D., Coyle, S. M., Pilarczyk, M. and Kadri, S. (2010). Coping strategies in a strongly schooling fish, the common carp *Cyprinus carpio*. *J. Fish Biol.* **76**, 1576–1591.
- Jenjan, H., Mesquita, F., Huntingford, F. and Adams, C. (2013). Respiratory function in common carp with different stress coping styles: a hidden cost of personality traits? *Anim. Behav.* **85**, 1245–1249.
- Johansen, K. (1968). Air-breathing fishes. *Sci. Am.* **219**, 102–111.
- Johansen, K. (1970). Air breathing in fishes. In *Fish Physiology*, Vol. 4 (ed. W. S. Hoar and D. J. Randall), pp. 361–411. New York: Academic Press.
- Killen, S. S., Marras, S. and McKenzie, D. J. (2011). Fuel, fasting, fear: routine metabolic rate and food deprivation exert synergistic effects on risk-taking in individual juvenile European sea bass. *J. Anim. Ecol.* **80**, 1024–1033.
- Killen, S. S., Marras, S., Ryan, M. R., Domenici, P. and McKenzie, D. J. (2012). A relationship between metabolic rate and risk-taking behaviour is revealed during hypoxia in juvenile European sea bass. *Funct. Ecol.* **26**, 134–143.
- Killen, S. S., Marras, S., Metcalfe, N. B., McKenzie, D. J. and Domenici, P. (2013). Environmental stressors alter relationships between physiology and behaviour. *Trends Ecol. Evol.* **28**, 651–658.
- Kramer, D. L., Manley, D. and Bourgeois, R. (1983). The effect of respiratory mode and oxygen concentration on the risk of aerial predation in fishes. *Can. J. Zool.* **61**, 653–665.
- Lefevre, S., Huang, D. T. T., Phuong, N. T., Wang, T. and Bayley, M. (2012). Effects of hypoxia on the partitioning of oxygen uptake and the rise in metabolism during digestion in the air-breathing fish *Channa striata*. *Aquaculture* **364–365**, 137–142.
- Lefevre, S., Domenici, P. and McKenzie, D. J. (2014a). Swimming in air-breathing fishes. *J. Fish Biol.* **84**, 661–681.
- Lefevre, S., Wang, T., Jensen, A., Cong, N. V., Huang, D. T. T., Phuong, N. T. and Bayley, M. (2014b). Air-breathing fishes in aquaculture. What can we learn from physiology? *J. Fish Biol.* **84**, 705–731.
- Lefevre, S., Bailey, M. and McKenzie, D. J. (2016). Measuring oxygen uptake in fishes with bimodal respiration. *J. Fish Biol.* In press.
- Mackenzie, S., Ribas, L., Pilarczyk, M., Capdevila, D. M., Kadri, S. and Huntingford, F. A. (2009). Screening for coping style increases the power of gene expression studies. *PLoS ONE* **4**, e5314.
- Mangel, M. and Stamps, J. (2001). Trade-offs between growth and mortality and the maintenance of individual variation in growth. *Evol. Ecol. Res.* **3**, 583–593.
- McKenzie, D. J., Burleson, M. L. and Randall, D. J. (1991). The effects of branchial denervation and pseudobranch ablation on cardioventilatory control in an air-breathing fish. *J. Exp. Biol.* **161**, 347–365.
- McKenzie, D. J., Campbell, H. A., Taylor, E. W., Micheli, M., Rantin, F. T. and Abe, A. S. (2007). The autonomic control and functional significance of the changes in heart rate associated with air breathing in the jeju, *Hoplerythrinus unitaeniatius*. *J. Exp. Biol.* **210**, 4224–4232.
- McKenzie, D. J., Steffensen, J. F., Taylor, E. W. and Abe, A. S. (2012). The contribution of air breathing to aerobic scope and exercise performance in the banded knifefish *Gymnotus carapo* L. *J. Exp. Biol.* **215**, 1323–1330.
- Milsom, W. K. (2012). New insights into gill chemoreception: receptor distribution and roles in water and air breathing fish. *Respir. Physiol. Neurobiol.* **184**, 326–339.
- Quinn, J. L., Cole, E. F., Bates, J., Payne, R. W. and Cresswell, W. (2012). Personality predicts individual responsiveness to the risks of starvation and predation. *Proc. R. Soc. B Biol. Sci.* **279**, 1919–1926.

- Rahn, H. (1966). Aquatic gas exchange: theory. *Respir. Physiol.* **1**, 1–12.
- Randall, D. J. (1982). The control of respiration and circulation in fish during exercise and hypoxia. *J. Exp. Biol.* **100**, 275–288.
- Randall, D. J., Burggren, W. W., Farrell, A. P. and Haswell, M. S. (1981). *The Evolution of Air Breathing in Vertebrates*. New York: Cambridge University Press.
- Réale, D., Garant, D., Humphries, M. M., Bergeron, P., Careau, V. and Montiglio, P.-O. (2010). Personality and the emergence of the pace-of-life syndrome concept at the population level. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **365**, 4051–4063.
- Rey, S., Boltana, S., Vargas, R., Roher, N. and Mackenzie, S. (2013). Combining animal personalities with transcriptomics resolves individual variation within a wild-type zebrafish population and identifies underpinning molecular differences in brain function. *Mol. Ecol.* **22**, 6100–6115.
- Rey, S., Digka, N. and MacKenzie, S. (2015). Animal personality relates to thermal preference in wild-type zebrafish, *Danio rerio*. *Zebrafish* **12**, 243–249.
- Shao, Y. T., Chen, I.-S. and Yan, H. Y. (2014). The auditory roles of the gas bladder and suprabranchial chamber in walking catfish (*Clarias batrachus*). *Zool. Stud.* **53**, 1.
- Shelton, G. S., Jones, D. R. and Milsom, W. K. (1984). Control of breathing in lower vertebrates. In *Handbook of Physiology*, Section 3, The Respiratory System (ed. S. R. Geiger, A. P. Fishman, N. S. Cherniac and J. G. Widdicome), Vol. II, Part 2, pp. 857–909. Bethesda, MD: American Physiological Society.
- Shingles, A., McKenzie, D. J., Claireaux, G. and Domenici, P. (2005). Reflex cardioventilatory responses to hypoxia in the flathead gray mullet (*Mugil cephalus*) and their behavioral modulation by perceived threat of predation and water turbidity. *Physiol. Biochem. Zool.* **78**, 744–755.
- Sih, A., Cote, J., Evans, M., Fogarty, S. and Pruitt, J. (2012). Ecological implications of behavioural syndromes. *Ecol. Lett.* **15**, 278–289.
- Smith, R. S. and Kramer, D. L. (1986). The effect of apparent predation risk on the respiratory behavior of the Florida gar (*Lepisosteus platyrhincus*). *Can. J. Zool.* **64**, 2133–2136.
- Stamps, J. A. (2007). Growth-mortality tradeoffs and “personality traits” in animals. *Ecol. Lett.* **10**, 355–363.
- Steffensen, J. F. (1989). Some errors in respirometry of aquatic breathers: how to avoid and correct for them. *Fish Physiol. Biochem.* **6**, 49–59.
- Thomson, J. S., Watts, P. C., Pottinger, T. G. and Sneddon, L. U. (2012). Plasticity of boldness in rainbow trout, *Oncorhynchus mykiss*: do hunger and predation influence risk-taking behaviour? *Horm. Behav.* **61**, 750–757.
- Willoughby, N. G. and Tweddle, D. (1978). The ecology of the catfish *Clarias gariepinus* and *Clarias ngamensis* in the Shire Valley, Malawi. *J. Zool.* **186**, 507–534.
- Wolf, M. and Weissing, F. J. (2012). Animal personalities: consequences for ecology and evolution. *Trends Ecol. Evol.* **27**, 452–461.

**Figure 1 Supplementary Material.**

Evidence for two response phenotypes in time to resume air-breathing after a fearful stimulus (T-res) in a group of juvenile *Clarias gariepinus* ( $n = 29$ ). Graph is a frequency distribution of response times, classified into 20 min bins, of the time intervals within which animals resumed air breathing. Individuals either resumed air breathing in less than 75 min (mean  $\pm$  SE =  $37 \pm 5$  min,  $n = 13$ ) or in greater than 115 min (mean =  $182 \pm 14$  min,  $n = 16$ ). Raw data are carried below the graph.



Bold T-res Phenotypes n = 13		Shy T-res phenotypes n = 16	
Fish ID	T-res	FISH ID	T-res
34	7	15	115
8	15	7	117
1	20	11	135
6	27	21	147
5	30	22	148
10	32	23	148
2	35	25	164
28	44	30	179
26	45	31	180
33	47	13	183
27	48	17	195
29	59	20	195
32	75	18	206
mean	<b>37,26</b>	19	225
SE	<b>5,34</b>	14	244
		24	325
		mean	<b>181,65</b>
		SE	<b>13,64</b>