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To cite this version:
Daisy Bougard, Maxime Belondrade, Charly Mayran, Lilian Bruyère-Ostells, Sylvain Lehmann, et al.. Diagnosis of Methionine/Valine Variant Creutzfeldt-Jakob Disease by Protein Misfolding Cyclic Amplification. Emerging Infectious Diseases, Centers for Disease Control and Prevention, 2018, 24 (7), pp.1364 - 1366. 10.3201/eid2407.172105. hal-01842307

HAL Id: hal-01842307
https://hal.umontpellier.fr/hal-01842307
Submitted on 6 Jan 2020
Diagnosis of Methionine/Valine Variant Creutzfeldt-Jakob Disease by Protein Misfolding Cyclic Amplification

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A patient with a heterozygous variant of Creutzfeldt-Jakob disease (CJD) with a methionine/valine genotype at codon 129 of the prion protein gene was recently reported. Using an ultrasensitive and specific protein misfolding cyclic amplification–based assay for detecting variant CJD prions in cerebrospinal fluid, we discriminated this heterozygous case of variant CJD from cases of sporadic CJD.

Variant Creutzfeldt-Jakob disease (vCJD) is a neurodegenerative infectious disease caused by transmission of a cattle prion disease (bovine spongiform encephalopathy) to humans (1). Most vCJD cases have occurred in the United Kingdom, where an estimated 1 in 2,000 persons is potentially asymptomatically infected, although there is some uncertainty about interpretation of detection of abnormal prion protein (PrP<sup>TSE</sup>) in appendix tissues on which this incidence is based (2) (Public Health England, https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/546883/hpr2616.pdf).

Until recently, all clinical cases of vCJD for which the prion protein gene has been analyzed have been shown to be methionine homozygous at codon 129, a genotype present in almost 40% of Caucasian populations. The report of the first definite heterozygous methionine/valine vCJD patient who died in 2016 (3) underlined previous concern about a possible second wave of vCJD cases (4). This patient was diagnosed during an autopsy; florid plaques were observed by histologic examination of the brain and peripheral detection of PrP<sup>TSE</sup> in lymphoid tissues. Western blot analysis of brain tissue confirmed a type 2B molecular profile of PrP<sup>TSE</sup>, which is characteristic for vCJD.

A diagnostic test to identify methionine/valine heterozygous vCJD cases is urgently needed to enable discrimination between heterozygous vCJD and sCJD and in view of the potential reservoir of methionine/valine heterozygous asymptomatic vCJD carriers in the blood donor population. We developed a highly sensitive and specific assay that accurately detects vCJD prions in blood even before the occurrence of clinical signs (5). We adapted this assay, which was based on protein misfolding cyclic amplification (PMCA) (6), for specific detection of vCJD in cerebrospinal fluid (CSF) and confirmed the ability of this assay to differentiate patients with atypical heterozygous vCJD from patients with sCJD.

The Study

We blindly analyzed 98 CSF samples provided by the National CJD Research and Surveillance Unit (Edinburgh, Scotland, UK) and the Centre Hospitalier Universitaire de Montpellier (Montpellier, France) after obtaining appropriate consent. Clinicians distributed CSF samples into blinded panels from the United Kingdom and France; 41 from patients with vCJD; 23 from patients with sCJD; 1 from a patient with genetic CJD; and 33 from patients with non-CJD, including samples from patients with Alzheimer’s disease and patients with nonneurodegenerative diseases.

CSF samples were thawed at room temperature and used directly in PMCA. We performed PMCA amplification by using brains from humanized transgenic mice as substrate for normal prion protein. After successive rounds of 160 cycles of PMCA for 15 min and sonication for 20 s, we detected PrP<sup>TSE</sup> by using Western blot after digestion with proteinase K (6).

Of the 98 CSF samples analyzed, our assay identified 40 of 41 cases of clinical vCJD, including the methionine/valine heterozygous patient, thus showing a diagnostic sensitivity of 97.6% (95% CI 87.1%–99.9%) (Table). One CSF sample from a probable case of vCJD showed a negative result. After decoding by clinicians, we retested this sample in duplicate; it showed a positive result.
Our assay also showed high analytical specificity; 0 of 57 potentially cross-reacting CSF specimens from patients with sCJD, gCJD, Alzheimer’s disease, and other nonneurodegenerative diseases showed a positive result (specificity 100% [95% CI 93.7%–100%]) (Table). The case-patient with methionine/valine heterozygous vCJD was specifically discriminated from the 12 methionine/valine heterozygous neuropathologically confirmed sCJD case-patients tested.

We then compared by using Western blot the PrP$^{\text{TSE}}$ molecular signature obtained for the clinical vCJD amplified samples from classical methionine homozgyous cases and the new methionine/valine heterozygous vCJD case with that of the reference brain sample from a patient with vCJD (Figure). As expected, the profile obtained after PMCA amplification of the CSF from the methionine/valine heterozygous vCJD patient was similar to those obtained for methionine homozygous vCJD patients. The characteristic type 2 mobility and clear predominance of the diglycosylated isoform was obtained for all vCJD patients before or after amplification.

**Table.** Analysis of CSF samples from patients with CJD and controls by PMCA*

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>CSF and codon 129 genotype/no. tested</th>
<th>No. patients with positive detection of PrP$^{\text{TSE}}$</th>
<th>Analytical performance, % (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Total</td>
<td>MM</td>
<td>MV</td>
</tr>
<tr>
<td>Clinical CJD</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Variant CJD</td>
<td></td>
<td>40/41†</td>
<td>37/38</td>
</tr>
<tr>
<td>Probable</td>
<td></td>
<td>29/29</td>
<td>28/28</td>
</tr>
<tr>
<td>Possible</td>
<td></td>
<td>10/11</td>
<td>8/9</td>
</tr>
<tr>
<td>Sporadic CJD</td>
<td></td>
<td>1/1</td>
<td>1/1</td>
</tr>
<tr>
<td>Probable</td>
<td></td>
<td>0/2</td>
<td>0/2</td>
</tr>
<tr>
<td>Genetic CJD</td>
<td></td>
<td>0/1</td>
<td>0/1</td>
</tr>
<tr>
<td>Non-CJD</td>
<td></td>
<td>0/12</td>
<td>ND</td>
</tr>
<tr>
<td>Alzheimer’s disease</td>
<td></td>
<td>0/21</td>
<td>ND</td>
</tr>
</tbody>
</table>

* CJD, Creutzfeldt-Jakob disease; CSF, cerebrospinal fluid; MM, methionine homozygous; MV, methionine/valine heterozygous; NA, not available; ND, not determined; PMCA, protein misfolding cyclic amplification; PrP$^{\text{TSE}}$, abnormal prion protein; VV, valine homozygous.
† The protease-resistant protein subtype was available for 3 patients with definite sporadic CJD and showed an equal distribution of MM1, MM2a, and VV2a.

![Western blot analysis of vCJD prions obtained after amplification by protein misfolding cyclic amplification (PMCA) of cerebrospinal fluid (CSF) from 2 patients with vCJD (MM and MV) and 3 control patients and a crude reference brain homogenate from a vCJD patient (National Institute for Biological Standards and Control [Ridge, UK] no. NHBY0/0003). Abnormal prion protein patterns were assessed by using antibody 3F4 after digestion of samples with protease K. A total of 20 µL of each sample was subjected to electrophoresis on a 12% polyacrylamide gel. Lane NBH, negative control brain homogenate from a person without CJD and no digestion with protease K (National Institute for Biological Standards and Control no. NHBZ0/0005); lane –3na, Western blot control (10$^{-3}$ dilution of vCJD reference brain sample without amplification); lane TP, positive control for amplification (10$^{-4}$ dilution of vCJD reference brain sample after 1 round of PMCA); lane vCJD-MM, CSF from a patient with MM vCJD after 3 rounds of PMCA; lane vCJD-MV, CSF from a patient with MV vCJD after 3 rounds of PMCA; lane sCJD-MM, CSF from a patient with MM vCJD after 5 rounds of PMCA; lane gCJD, CSF from a patient with gCJD after 5 rounds of PMCA; lane AD, CSF from a patient with Alzheimer’s disease after 5 rounds of PMCA. CJD, Creutzfeldt-Jakob disease; gCJD, genetic CJD; MM, methionine homozygous; MV, methionine/valine heterozygous; Rd, round (1 round indicates 80 cycles of PMCA); sCJD, sporadic CJD; vCJD, variant CJD.**

**Figure.** Western blot analysis of vCJD prions obtained after amplification by protein misfolding cyclic amplification (PMCA) of cerebrospinal fluid (CSF) from 2 patients with vCJD (MM and MV) and 3 control patients and a crude reference brain homogenate from a vCJD patient (National Institute for Biological Standards and Control [Ridge, UK] no. NHBY0/0003). Abnormal prion protein patterns were assessed by using antibody 3F4 after digestion of samples with protease K. A total of 20 µL of each sample was subjected to electrophoresis on a 12% polyacrylamide gel. Lane NBH, negative control brain homogenate from a person without CJD and no digestion with protease K (National Institute for Biological Standards and Control no. NHBZ0/0005); lane –3na, Western blot control (10$^{-3}$ dilution of vCJD reference brain sample without amplification); lane TP, positive control for amplification (10$^{-4}$ dilution of vCJD reference brain sample after 1 round of PMCA); lane vCJD-MM, CSF from a patient with MM vCJD after 3 rounds of PMCA; lane vCJD-MV, CSF from a patient with MV vCJD after 3 rounds of PMCA; lane sCJD-MM, CSF from a patient with MM vCJD after 5 rounds of PMCA; lane gCJD, CSF from a patient with gCJD after 5 rounds of PMCA; lane AD, CSF from a patient with Alzheimer’s disease after 5 rounds of PMCA. CJD, Creutzfeldt-Jakob disease; gCJD, genetic CJD; MM, methionine homozygous; MV, methionine/valine heterozygous; Rd, round (1 round indicates 80 cycles of PMCA); sCJD, sporadic CJD; vCJD, variant CJD.
methionine prion protein (9). However, PMCA analysis should be performed in a Biosafety Level 3 laboratory and requires highly experienced personnel.

Iatrogenic transmission of vCJD by blood transfusion has been documented in 3 recipients of nonleukodepleted erythrocyte concentrates from blood donors during development of disease (10). One additional probable case of vCJD transmission by blood transfusion was identified during an autopsy of a methionine/valine heterozygous patient who died from a nonneurologic disorder and in whom vCJD prion protein was detected in the spleen (11). The presence of infectivity in blood of the definite methionine/valine heterozygous vCJD patient involved in our study is uncertain and requires further investigation.

From a clinical point of view, prion amplification technologies, such as PMCA and real-time quaking-induced conversion (RT-QuIC), have already shown their sensitive detection of disease-related prion protein in biologic fluids (6,12–14). Independent studies have shown that detection of prion protein seeding activity in CSF by RT-QuIC might have a specificity of 99%–100% for diagnosis of sCJD (13,15). Application of RT-QuIC and PMCA for CSF samples might represent a suitable strategy for premortem discrimination between sCJD and vCJD including methionine/valine heterozygous case-patients, particularly for cases with a heterozygous codon 129 genotype in which clinical distinction between sCJD and vCJD is problematic.

Acknowledgments
We thank Maria-Teresa Alvarez and Jacques-Damien Arnaud for providing helpful assistance in the L3 facility at the Etablissement Confiné d’Expérimentation–Centre d’Elevage et de Conditionnement Expérimental des Modèles Animaux of the University of Montpellier.

This study was supported by Etablissement Français du Sang and the UK Department of Health Policy Research Programme (grant PRST061400008).

About the Author
Dr. Bougard is a research scientist in charge of the Prion Group at Etablissement Français du Sang of Montpellier, France. Her primary research interests include development of innovative tools for the prevention of transfusion risk associated with nonconventional agents.

References

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