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QUANTIFYING VARIATION IN SPECIATION AND EXTINCTION RATES WITH CLADE DATA

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1 High-level phylogenies are very common in evolutionary analyses, though they are of-
2 ten treated as incomplete data. Here we provide statistical tools to analyze what we
3 name 'clade data', that are the ages of clades together with their numbers of species.
4 We develop a general approach for the statistical modeling of variation in speciation
5 and extinction rates, including temporal variation, unknown variation, and linear and
6 nonlinear modeling. We show how this approach can be generalized to a wide range of
7 situations, including testing the effects of life-history traits and environmental variables
8 on diversification rates. We report the results of an extensive simulation study to assess
9 the performance of some statistical tests presented here as well as of the estimators of
10 speciation and extinction rates. These latter results suggest the possibility to estimate
11 correctly extinction rate in the absence of fossils. An example with data on fish is
12 presented.

13 **KEY WORDS:** birth–death models, extinction, maximum likelihood, speciation, stem
14 ages.

15 The study of the tempo and mode of evolution has experienced a new wave
16 of interest from evolutionists using new mathematical and statistical tools
17 to analyze molecular phylogenies (Sanderson and Donoghue 1996; Ricklefs
18 2007). Following some initial breakthrough (e.g., Nee et al. 1992, 1994), sig-
19 nificant progress has been achieved in biologically relevant statistical mod-
20 eling of diversification, such as quantifying temporal variation in diversifi-
21 cation (Paradis 2011; Hallinan 2012) or assessing the effects of biological
22 traits on speciation and extinction rates (Maddison et al. 2007; FitzJohn
23 et al. 2009; FitzJohn 2010). Recent advances have also been accomplished
24 in integrating molecular and fossil data (e.g., Morlon et al. 2011; Didier et al.
25 2012).

26 Most of these recent statistical developments have focused on analyz-
27 ing complete phylogenies. Incomplete phylogenies are often treated as a
28 separate case in order to take missing data into account (Pybus et al. 2002;
29 FitzJohn et al. 2009; Stadler 2011). The most common form of such data is
30 a phylogeny resolved at a high level accompanied by the number of species
31 associated to each tip of the tree. On the other hand, the ages of clades
32 together with the numbers of species (named here ‘clade data’) have been
33 a neglected source of data in the analysis of diversification. Magallón and
34 Sanderson (2001) provided some methods for the analysis of such data and
35 applied them to angiosperms. They particularly developed various estima-
36 tors of the (net) rate of diversification of a clade giving its age and number
37 of species.

38 The relative lack of interest towards clade data may come from the fact
39 that, for a given clade, its complete phylogeny contains more information
40 than the pair of values ‘age + number of species’. However, for a collection
41 of clades, such data are a valuable source of information for several reasons.

42 First, clades defined by higher-level taxa (e.g., families, orders) are clearly
43 identified for almost all groups of living beings and their numbers of species
44 are in many cases already known. Second, phylogenetic relationships among
45 higher-level taxa have been much more studied than within them, so it is
46 more straightforward to date the age of a clade rather than the divergences
47 among its species. Third, the fossil record is generally more informative on
48 the origin of higher-level taxa compared to species or other low-level taxa.
49 Fourth, it is easier to examine the impact of the species concept on the
50 definition of clade data rather than on a phylogeny since, in the former the
51 species concept will mostly affect the number of species while in the latter it
52 will be often hard to infer different phylogenies under those distinct species
53 definitions. Clade data have also some disadvantages: the inherent lack
54 of temporal resolution within each clade makes it impossible to study the
55 variation in diversification within them.

56 In the present paper, we extend the approach presented by Magallón
57 and Sanderson (2001) and present statistical tools for the inference of di-
58 versification patterns and processes with clade data. Our approach assumes
59 that each clade, instead of having its own speciation and extinction rates,
60 comes from a ‘statistical population of clades’ so that maximum likelihood
61 inference is straightforward. With this rationale, we show how to make
62 inference on variation in diversification parameters among clades using dif-
63 ferent modeling tools, including testing the effects of life-history traits and
64 environmental variables and the case where variation is a priori unknown.
65 We also present the results of a simulation study in order to assess the sta-
66 tistical performance of several tests and estimators presented in this paper,
67 and finally we apply our approach on a data set of fish.

68 *Statistical Modeling Approach*

69 Throughout this paper we assume that diversification proceeds with speci-
70 ation (λ) and extinction (μ) rates which are the probabilities that a species
71 splits into two daughter-species or goes extinct during a very short time.
72 We denote as X_t the number of species in a clade of age t where this may
73 be either the stem age of the clade (divergence time of the clade from its
74 sister-clade) or its crown age (time to the most common recent ancestor
75 of the species belonging to the clade). Specifically, using equation 8 from
76 Kendall (1948), we can write the probability that X_t takes a specific integer
77 value x :

$$\Pr(X_t = x|\theta, X_0 = 1) = \eta_t(1 - \eta_t)^{x-1} \quad x \geq 1, \quad (1)$$

78 where θ is a vector of parameters specifying how speciation and extinction
79 rates vary through time and η_t is a function of these parameters. The
80 conditioning on $X_0 = 1$ emphasizes that in this paper we consider stem
81 groups. For the case of crown groups ($X_0 = 2$), the probabilities must
82 be summed on all possible combinations. In most applications, stem groups
83 are considered because the origin of a group is inferred from its relationships
84 with its sister-group. On the other hand, deriving the crown age of a group
85 requires to estimate the age of the most recent common ancestor of its species
86 which is usually more complicated because it requires to sample all species
87 in the clade. On the other, inferring stem ages requires one species from the
88 clade and one from its sister-clade.

89 Various forms exist for these probabilities depending on the parameteri-
90 zation of θ and whether we wish to condition them on survival of the lineage
91 until present or not. For instance, if extinction rate is zero and speciation
92 rate is constant, then $\eta_t = e^{-\lambda t}$. This is the Yule (1924) model. Models with

93 a non-null extinction rate are called birth–death models (Kendall 1948).

94 The point of conditioning on no extinction is important when analyzing
95 data on actual groups because total extinction of these groups did not occur.
96 Thus the probabilities must be modified accordingly, otherwise this would
97 result in underestimated extinction rates (Rabosky et al. 2007).

98 Let us consider for the moment the simple Yule model. The expected
99 number of species at time t is given by $E(X_t) = e^{\lambda t}$. From this expectation,
100 a simple estimator of λ based on the method of moments is $\hat{\lambda} = \ln(x)/t$
101 (Magallón and Sanderson 2001). When considering a single clade, and in
102 the absence of more detailed information, it does not seem possible to go
103 further in the inference. When considering more than one group (e.g., the
104 families within an order or a class), researchers usually estimate λ separately
105 for each group, then proceed with standard statistics (e.g., McPeck 2008).
106 This approach assumes that each clade is characterized by its own speciation
107 rate. On the other extreme, one may assume that speciation rate is the same
108 in all groups so that the observed data are independent outcomes of the
109 same diversification process. Thus, it is possible to use maximum likelihood
110 inference using equation 1. The likelihood function is:

$$\prod_i \Pr(x_i|\lambda), \quad (2)$$

111 where $\Pr(x|\lambda)$ is a simplified notation of equation 1. We may expect less
112 bias in the estimates from this approach, but also the possibility to test
113 hypotheses based on fitting alternative models.

114 The assumption of equal speciation rates among clades is, certainly in
115 most cases, unrealistic (Purvis et al. 1995; Paradis 2005; Alfaro et al. 2009).
116 However, since we have several observations we may model the variation
117 in this parameter with a statistical modeling approach. We explore several

118 such approaches below. Firstly, we consider approaches based on determin-
119 istic variation between two or more groups of clades. Secondly, we consider
120 how temporal variation in speciation and extinction rates can be modeled
121 and assessed. Thirdly, we develop an approach handling unknown variation
122 based on mixture modeling, including the combination of mixtures with a
123 linear modeling of the speciation rate. Finally, we attack the problem of
124 estimating extinction rates.

125 **Variation Among Clades**

126 A simple way to model variation in diversification among clades is to assume
127 that there are two categories: some clades diversify with speciation rate λ_1
128 and the others with rate λ_2 . The data are made of n_1 and n_2 clades in each
129 category, respectively. The likelihood function is:

$$\prod_{i_1=1}^{n_1} \Pr(x_{i_1}|\lambda_1) \prod_{i_2=1}^{n_2} \Pr(x_{i_2}|\lambda_2).$$

130 Note that each clade is assigned to a category a priori, although there is
131 no assumption on whether λ_1 is greater, or smaller, than λ_2 . The null
132 hypothesis $\lambda_1 = \lambda_2$ can be tested by fitting this model and the null model
133 whose likelihood is given by equation 2: the likelihood-ratio test (LRT)
134 comparing these two models follows a χ^2 distribution with $df = 1$. An
135 alternative is to use the Akaike information criterion (Akaike 1973).

136 The present approach is easily generalized to more than two categories:
137 let us denote the number of categories as K , then the likelihood function
138 would become the product of K products:

$$\prod_{j=1}^K \prod_{i_j=1}^{n_j} \Pr(x_{i_j}|\lambda_j),$$

139 where n_j is the number of clades in the j th category. The LRT comparing
140 this model with the null model of homogeneous diversification follows a χ^2
141 with $df = K - 1$.

142 These models assume, mostly for simplicity, that there is no extinction
143 ($\mu = 0$); however, variation in extinction rate can be incorporated in a
144 straightforward way. For instance a model with two categories diversify-
145 ing with the same λ but with different extinction rates has the following
146 likelihood function:

$$\prod_{i=1}^{n_1} \Pr(x_{i_1} | \lambda, \mu_1) \prod_{i=1}^{n_2} \Pr(x_{i_2} | \lambda, \mu_2),$$

147 which could be compared with the null model with $\mu > 0$ whose likelihood
148 is:

$$\prod_{i=1}^N \Pr(x_i | \lambda, \mu),$$

149 with $N = n_1 + n_2$. This test is related, but not identical, to the tests
150 of equal diversification using sister-clades where the ages of clades are not
151 needed (Paradis 2012b).

152 The supplementary materials provide annotated R code explaining how
153 to build and fit any model following the present approach.

154 **Linear Modeling**

155 Following the previous section, two extreme models can be defined: the sim-
156 plest one where all clades diversify at the same rate, and the most complex
157 one where each clade has its own parameter(s). This second model will be
158 overparameterized for a likelihood approach. Nevertheless, it is possible to
159 model variation in diversification parameters with linear models. For in-

160 stance, we may know a priori some variables that are likely to affect the
161 value of speciation rate (e.g., body size), and a model that relates such ‘co-
162 variates’ to speciation rate may be an appropriate candidate to model the
163 variation in diversification among clades. We use here a standard strategy
164 to model variation in a rate with respect to a covariate z :

$$g(\lambda_i) = \beta z_i + \alpha,$$

165 where λ_i is the speciation rate in clade i , g is a function used to transform the
166 rate in order to linearize the relationship, and β and α are two parameters.
167 Here β controls the effect of z on λ : if $\beta > 0$ then species with large values
168 of z will speciate faster than those with small values of z (and inversely if
169 $\beta < 0$). It is possible to consider more than one predictor in which case
170 the number of parameters is equal to the number of predictors plus one.
171 Nonlinear models can also be considered. Each clade has its own speciation
172 rate given by (with g^{-1} being the inverse transformation of g):

$$\lambda_i = g^{-1}(\beta z_i + \alpha), \tag{3}$$

173 which is used to calculate the likelihood defined by equation 2: the likelihood
174 function is then maximized to estimate β and α (see code in the Supplemen-
175 tary Material). A common choice for g is the logit function, $\ln(\lambda_i/(1 - \lambda_i))$,
176 so g^{-1} gives:

$$\lambda_i = \frac{1}{1 + e^{-(z_i\beta + \alpha)}},$$

177 The null model is defined by fixing $\beta = 0$ in which case $\lambda = 1/(1 + e^{-\alpha})$ for
178 all clades. The logit function is well suited for parameters varying between

179 0 and 1 which is the case for speciation rates considered on geological time
180 scales (million of years). However, speciation rates may be larger than one
181 on shorter scales. Other transformations can be used such as the one used
182 below.

183 It must be noted that the variation among clades as modeled in the
184 previous section is a special case of linear models where the membership of
185 a clade to a category is coded with a discrete variable and this variable is
186 entered as a predictor into the linear model after coding it into binary 0/1
187 variable(s) (see appendix in Paradis 2005, for details). Therefore, continuous
188 and categorical predictors can be combined in the linear model.

189 **Temporal Variation**

190 Kendall (1948) studied the birth–death model in a very general way, in-
191 cluding the cases where λ and μ vary through time. Thus it is possible to
192 derive the probability density of the distribution of the x_i 's when diversifi-
193 cation changed through time. The likelihood can be defined and fit in the
194 same way as above. Such a temporal model can be compared with the null
195 model of constant diversification with a χ^2 test whose df will be equal to
196 the number of additional parameters in the first model. As before, tempo-
197 ral variation may reflect speciation and/or extinction rate(s). The simplest
198 temporal model has two rates before and after a given time point in the past,
199 so it has one additional parameter than the null model. Note that if the
200 time point is unknown, it could be estimated from the data so there would
201 be two additional parameters. However, a wide variety of temporal models
202 can be defined in `ape` (Paradis et al. 2004) using the function `dbdTime` where
203 the temporal variation is defined by the user with a standard R function.

204 **Unknown Variation**

205 The above models assume that diversification parameters vary in relation
206 to some known variables, either categorical or continuous. On the other
207 hand, it is possible that these variables are not observable. Such unknown
208 variation can be modeled with two approaches depending on whether we
209 assume that the diversification parameters vary in a discrete or continuous
210 manner.

211 A mixture of distributions is based on the assumption that observations
212 come from two or more categories each characterized by its own distribution,
213 but the assignment of an observation to a particular category is unknown
214 (see Flury et al. 1992, for a biological example). As a simple example,
215 consider a mixture of two Yule processes, then the likelihood function will
216 be:

$$\prod_{i=1}^N f \Pr(x_i|\lambda_1) + (1 - f) \Pr(x_i|\lambda_2), \quad (4)$$

217 where f is the proportion of clades in the first category. This model has
218 three parameters (λ_1 , λ_2 and f) and can be compared with the null model
219 of homogeneous speciation with a LRT with $df = 2$. The idea is easily
220 generalized to more than two mixtures: a mixture with K Yule models
221 would have $2K - 1$ parameters. As above, the mixture may involve speciation
222 and/or extinction rate(s). By contrast to the situation above where clades
223 were assigned to categories a priori, there is here no assignment a priori.
224 On the other hand, assignment a posteriori is possible by calculating the
225 relative contributions to the likelihood function.

226 The idea may even be further generalized to include mixtures of linear
227 models. Suppose we know that one variable, say body size, has a significant
228 effect on speciation rate but there is some other, unknown, variation in this

229 parameter that we want to model with a mixture. Then it is possible to
 230 calculate the λ_i 's with equation 3 and use them to compute the likelihood
 231 with eq. 4. Each category would have its own parameters β and α , so a
 232 model with K categories has $3K - 1$ parameters.

233 The second approach assumes that, in the case of a Yule model, λ varies
 234 continuously across clades following a specified distribution whose parame-
 235 ters are estimated from the data. A transformation of λ is useful so that it
 236 follows a normal distribution: $g(\lambda) \sim \mathcal{N}(\mu_\lambda, \sigma_\lambda^2)$. A useful transformation
 237 here is the complementary log-log transformation: $g(\lambda) = \ln(-\ln(\lambda))$. As
 238 above we do not know the value of λ for a given clade, but this time instead
 239 of a discrete sum we have to do a continuous integration. The likelihood
 240 function is thus:

$$\prod_{i=1}^N \int_{-\infty}^{\infty} f_{\mathcal{N}}(u|\mu_\lambda, \sigma_\lambda^2) \Pr(x_i|g^{-1}(u))du,$$

241 where $f_{\mathcal{N}}$ is the density function of the normal distribution. A graphical
 242 representation of the variation in λ is obtained with the inverse transforma-
 243 tion $g^{-1}(u) = \exp(-e^u)$ with the density of u computed with the normal
 244 distribution and the estimates $\hat{\mu}_\lambda$ and $\hat{\sigma}_\lambda^2$.

245 **Estimating Extinction Rates**

246 The estimation of extinction rates in the absence of fossil data has appeared
 247 to be a complicated issue (Paradis 2004, 2011; McPeck 2008; Aldous et al.
 248 2011; Morlon et al. 2011; Didier et al. 2012; Hallinan 2012). To try to tackle
 249 this problem, we implemented a procedure which fits a birth–death model
 250 estimating λ and μ simultaneously. These estimates are denoted as $\hat{\lambda}_{\text{BD}}$ and
 251 $\hat{\mu}_{\text{BD}}$.

252 *Simulation Study*

253 The present statistical modeling approach offers many possibilities and it
254 would take a large number of simulations to assess the statistical proper-
255 ties of all of them. Instead, we focus on a few key questions. What is the
256 statistical power to detect a difference in diversification between two groups
257 of clades? How powerful is the test to detect temporal variation in diver-
258 sification? What is the statistical power to detect unknown variation in
259 diversification between two groups of clades using mixtures? Finally, what
260 is the precision of the λ and μ estimators?

261 To address these four questions, we ran four sets of simulations. First, we
262 considered a simple two-category scenario with n_1 and n_2 clades simulated
263 with rates λ_1 and μ_1 and λ_2 and μ_2 , respectively. The times of evolution
264 were drawn from a uniform distribution: $t_i \sim \mathcal{U}(10, 20)$. A phylogeny was
265 simulated under a birth–death process during a time t_i using `ape` starting
266 from a single species. The number of species surviving at time t_i , x_i , was
267 extracted and the pairs (x_i, t_i) were analyzed as described above using a Yule
268 model. The LRT testing the null hypothesis of homogeneous diversification
269 was computed, and the rejection rate was assessed under different sets of
270 parameter values: $n_1 = n_2 = \{1, 3, 5, 10, 20\}$, $\lambda_1 = \{0.1, 0.15, 0.2\}$, $\lambda_2 = 0.1$,
271 $\mu_1 = \{0, 0.05\}$, $\mu_2 = \{0, 0.05\}$.

272 Second, we performed simulations under three scenarios with different
273 values of diversification rates before and after 30 time units. We first gen-
274 erated 100 values of t from a uniform distribution between 10 and 50. We
275 then simulated clades with constant, increasing, or decreasing diversification
276 rate. The number of species was extracted as before, and two models were
277 fitted: the null Yule model of constant diversification, and an alternative
278 model assuming different speciation rates before and after 30 time units (as

279 above $\mu = 0$ was assumed). The rejection rates of the LRTs comparing both
280 models were computed.

281 Third, a scenario similar to the first one was considered: the difference
282 is that the simulated clades were not identified to a particular category so
283 the data were analyzed with a mixture of Yule models. We used $K = 2$,
284 $n_1 = n_2 = \{10, 20, 50\}$, and $t_i \sim \mathcal{U}(10, 20)$. Four combinations of speciation
285 and extinction rates were used: (i) the null hypothesis is true and there is
286 no extinction: $\lambda_1 = \lambda_2 = 0.1$, $\mu_1 = \mu_2 = 0$; (ii) the null hypothesis is false
287 and there is no extinction: $\lambda_1 = 0.1$, $\lambda_2 = 0.2$, $\mu_1 = \mu_2 = 0$; (iii) the null
288 hypothesis is false but only μ varies: $\lambda_1 = \lambda_2 = 0.2$, $\mu_1 = 0$, $\mu_2 = 0.1$; and
289 (iv) same than before with stronger variation in μ : $\lambda_1 = \lambda_2 = 0.2$, $\mu_1 = 0$,
290 $\mu_2 = 0.15$.

291 Finally, we performed an assessment of the precision of the estimators of
292 speciation and extinction rates using five combinations of λ and μ : (0.1, 0),
293 (0.1, 0.03), (0.1, 0.06), (0.2, 0.1), and (0.2, 0.15). Here $t_i \sim \mathcal{U}(10, 30)$ and
294 $n = 100$.

295 The simulations were replicated 1000 times. Annotated R (R Devel-
296 opment Core Team 2012) code is available in the Supplementary Material
297 with guidelines on how to run these simulations so that the readers can
298 adapt them to their own problems. Besides, we did not attempt to compare
299 our method with previous ones because some scenarios considered here can-
300 not be analyzed by the latter (e.g., the third scenario does not seem to be
301 tractable with Magallón and Sanderson's method).

302 *Application to Fish Data*

303 We used the data from Vega and Wiens (2012) who compiled the number
304 of species, stem age, and percentage of marine fish species for 22 orders and

305 super-orders and for 97 families. They also provided a phylogeny for the
306 22 higher taxa which allowed to compare our estimates with those obtained
307 from the combined analysis of phylogeny and species richness data (Paradis
308 2003). All data were unmodified from the original publication and are avail-
309 able at <http://dx.doi.org/10.1098/rspb.2012.0075>. With this data set, we
310 explored the variation in diversification using different mixtures of Yule and
311 birth–death models. We also tried to assess whether this variation is due to
312 differences in the speciation or in the extinction rates.

313 *Results*

314 **Simulation Study**

315 The first set of simulations showed that, overall, the LRT testing for dif-
316 ferent diversification rates between two categories of clades had satisfactory
317 statistical properties (Table 1). The type I error rate (rejection rate when
318 the null hypothesis is true, i.e., $\lambda_1 = \lambda_2$ and $\mu_1 = \mu_2$) was, as expected,
319 close to 5% (first and seventh lines in Table 1). However, when $\lambda - \mu$ was
320 the same in both categories, the rejection rate was greater than 5% (eighth
321 line in Table 1) showing that the present test does not test for equal di-
322 versification rate. In the cases where the null hypothesis was not true, the
323 rejection rate varied as expected: it was greater for larger sample sizes (n_1)
324 and for larger contrast in the speciation or extinction rate. Interestingly, if
325 one category of clades had smaller μ while λ was the same, then the test was
326 able to detect this difference; however, the statistical power was less than
327 when the same contrast in diversification was due to different λ (compare
328 the second and third lines in Table 1).

329 In the second set of simulations, the test for temporal variation rejected
330 the null hypothesis in more than 90% when $\mu = 0$ and λ varied, whether this

331 was an increase or a decrease (third to sixth lines in Table 2). On the other
332 hand, the results were contrasted when $\mu > 0$. When there was no temporal
333 variation in the parameters, the type I error rates were inflated in relation
334 to the value of μ (seventh and eighth lines in Table 2). When μ varied
335 through time, the test behaved very differently depending on the direction
336 of this variation: it did not reject the null hypothesis in most cases when
337 μ increased (ninth line in Table 2) while it rejected it in 68% of the cases
338 when μ decreased (tenth line in Table 2). To further investigate this point,
339 we repeated some of these simulations but this time the null model was a
340 birth–death model with λ and μ constant through time, and the alternative
341 model was with λ constant and μ allowed to vary before and after 30 time
342 units. In this situation, the test behaved as expected: the rejection rate
343 was less than 5% when μ was constant, whereas it varied between 8% and
344 31% when the null hypothesis was false (Table 3). It is noteworthy that the
345 present test to detect time-dependent extinction rate is not very powerful: it
346 was necessary to simulate a strong contrast in μ to reach a statistical power
347 greater than 0.2.

348 The third set of simulations showed that the mixture-based LRT was
349 able to detect heterogeneous diversification among two unknown categories
350 of clades (Table 4). The test was more powerful when the contrast was
351 due to different λ compared to different μ . Otherwise, the test showed
352 satisfactory statistical performance: its power increased with sample size
353 and/or contrast in the parameters.

354 The distribution of the estimates of speciation rate under the Yule model,
355 $\hat{\lambda}_{\text{Yule}}$, shows that this estimator appeared unbiased when $\mu = 0$ (Fig. 1A).
356 On the other hand, when $\mu > 0$ it was negatively biased though it can be ob-
357 served that $\hat{\lambda}_{\text{Yule}} > \lambda - \mu$ so this cannot be actually taken as an estimator of

358 the net diversification rate. The estimator based on the birth–death model,
359 $\hat{\lambda}_{\text{BD}}$, appears less biased, even though the presence of extinctions seems
360 to induce a slightly more dispersed distribution of the estimates (Fig. 1B).
361 The estimates of extinction rate based on the birth–death model, $\hat{\mu}_{\text{BD}}$, were
362 almost unbiased (Fig. 1C).

363 **Application to Fish Data**

364 The fit of the Yule model to the fish data at the higher level ($N = 22$)
365 resulted in a global estimate $\hat{\lambda}_{\text{Yule}} = 0.058$ (SE = 0.002; AIC = 456). We
366 tried to fit a birth–death model which led to a much improved fit (AIC =
367 376); however, the likelihood function had a pronounced ridge on the line
368 $\lambda = \mu$ (not shown). The fit of mixtures of Yule models with increasing
369 number of categories (K) showed that the best fit was with three categories
370 (Table 5). The parameter estimates were: $\hat{\lambda}_1 = 0.041$, $\hat{\lambda}_2 = 0.080$, $\hat{\lambda}_3 =$
371 0.013 , $\hat{f}_1 = 0.65$, and $\hat{f}_2 = 0.10$. The analysis of the combined taxonomic
372 and phylogenetic data (Paradis 2003) gave $\hat{\lambda} = 0.056$ and $\hat{\mu} = 1.83 \times 10^{-7}$.

373 The analysis at the level of the families ($N = 97$) gave for the Yule
374 model $\hat{\lambda}_{\text{Yule}} = 0.0756$ (SE = 0.0016; AIC = 1483). Like above, the fit of
375 the birth–death model resulted in a likelihood surface with a ridge on the
376 line $\lambda = \mu$. The mixture of Yule models with the best fit had two categories
377 (Table 5); the parameter estimates were: $\hat{\lambda}_1 = 0.099$, $\hat{\lambda}_2 = 0.036$, $\hat{f} = 0.42$.

378 The analysis with a model assuming continuous variation in λ across
379 clades gave close results for both taxonomic levels. In both cases, the model
380 fitted well and the AIC values were smaller than for any of the previous
381 models (Table 6). Figure 2 shows the distribution of λ inferred with the
382 estimated parameters. Trying to introduce μ did not result in successful fits
383 and the estimates of this parameter were close to zero.

384 Vega and Wiens (2012) reported the percentage of marine and freshwater
385 species at both taxonomic levels. This was distributed very asymmetrically
386 with most orders and families having only marine or freshwater species.
387 Thus, we split the data into two groups whether they had more or less
388 than 50% of marine species. A test of different speciation rates between
389 these groups was performed. For orders, the difference was significant (LRT:
390 $\chi_1^2 = 28.09$, $P < 0.001$) with a larger estimate for marine orders ($\hat{\lambda} = 0.063$,
391 $SE = 0.002$) compared to the freshwater ones ($\hat{\lambda} = 0.046$, $SE = 0.002$).
392 An examination of the data suggested that this result was dependent on
393 Percomorpha which is one of the youngest clades in this data set and includes
394 16,625 species (Fig. 3A). Removing this clade resulted in a non-significant
395 test ($\chi_1^2 = 2.10$, $P = 0.147$, $N = 21$). For families, an analogous result
396 was found with a significant test (LRT: $\chi_1^2 = 5.58$, $P = 0.018$) comparing
397 marine families ($\hat{\lambda} = 0.079$, $SE = 0.002$) and freshwater ones ($\hat{\lambda} = 0.071$,
398 $SE = 0.002$). This result was dependent on two families older than 200 Myr
399 (Fig. 3B): the Amiidae (one species) and Polypteridae (12). Removing these
400 two families led to a non-significant test: $\chi_1^2 = 2.02$, $P = 0.155$ ($N = 95$).

401 *Discussion*

402 The analysis of phylogenetic diversification with molecular data is enjoying
403 a remarkable success in the literature. Some spectacular results have been
404 accomplished using complete phylogenies (e.g., Goldberg et al. 2010; Hugall
405 and Stuart-Fox 2012; Penney et al. 2012). Though complete phylogenies,
406 possibly supplemented with fossil data, are probably the best way to inves-
407 tigate evolutionary diversification, the goal of our study was to show the
408 merit of an alternative approach based on the analysis of clade data.

409 Our modeling approach is based on the assumption that each clade is

410 characterized by its diversification parameters and variation among these pa-
411 rameters can be quantified in a statistical way. Bokma (2003) and Paradis
412 (2003) developed a method to combine information from high-level phylo-
413 genies with clade data: both authors considered the simple constant-rate
414 birth–death model. Alfaro et al. (2009) used similar combined data to as-
415 sess variation among clades of vertebrates using a stepwise procedure (see
416 details in Paradis 2012a). Thus the approach in the present paper comple-
417 ments previous methodological developments. The possibility to quantify
418 variation among clades with linear models seems a fruitful way to avoid
419 overparameterization. Future applications will reinforce the relative merits
420 of this approach.

421 Recently, Stadler and Bokma (2013) developed alternative likelihood
422 functions with respect to the way higher taxa are defined. They showed
423 that the estimation of speciation and extinction rates vary substantially de-
424 pending on these definitions. While they considered only the constant-rate
425 birth–death model, it seems possible and interesting to include their sam-
426 pling scheme into the developments presented in the present paper.

427 Our modeling approach ignores the background phylogeny of the clades,
428 the set of branches that link the clades together to make a higher-level
429 phylogeny. There are two reasons for this. First, using information from
430 the background phylogeny is straightforward when the rates of speciation
431 and extinction are constant and homogenous, but when this assumption
432 is relaxed it is not simple how one must assume changes in rates in the
433 background tree. It is clear that if a well-supported background phylogeny
434 is available, this might give additional information which can be combined
435 with clade data (e.g., Paradis 2003). However, this extra information will
436 in most cases require its own model since it relates to older diversification

437 events compared to clade data. On the other hand, ignoring backbone phy-
438 logeny and assuming that the clades are independent units simplifies the
439 definition of alternative models as done in this paper. Second, though some
440 higher-level phylogenies are available (mammals, birds), we believe these are
441 still exceptions rather than the rule. For instance, the basal relationships
442 of reptiles, amphibians, or fishes are still debated. Therefore, having the
443 possibility to analyze their clade data without the need of a background
444 phylogeny is of some general application. Furthermore, the present ap-
445 proach can be used when analyzing sets of clades across different phyla, for
446 instance arthropods, echinoderms, vertebrates, etc., where the background
447 phylogeny would not be very informative since this would branch at the
448 origin of Metazoa.

449 The use of mixtures as an approach to analyze heterogeneity in diver-
450 sification rates is not limited to clade data. For instance, one could model
451 speciation and extinction rates on a fully-resolved phylogeny assuming that
452 these parameters vary among its branches though we do not know a pri-
453 ori which sections of the tree evolved fast and which others evolved slowly.
454 Furthermore, the mixture approach can also be used to model variation in
455 rates of trait evolution along a phylogeny. In that case, the variation may be
456 among branches (as in the previous example), or among traits where some
457 traits are assumed to evolve faster but we do not know which ones.

458 Some subtle but important facts come from the results of the simulation
459 study. Even though most of the tests considered here assumed $\mu = 0$, they
460 appeared not to be tests of equal diversification. If the net diversification
461 rates ($\lambda - \mu$) were equal among clades, the tests rejected the null hypothesis
462 in more than 5% (see eighth row of Table 1). On the other hand, if λ was
463 equal among clades, the tests detected differences in μ . It is clear that results

464 based only on the Yule model must be interpreted with caution.

465 The tests of temporal variation showed some contrasted but interesting
466 results. When the extinction rate was zero, these tests performed very well
467 and were able to detect either a decrease or an increase in speciation rate.
468 However, when extinction rate was not null, the tests based on the Yule
469 model showed poor performance with an increased type I error rate and a
470 high type II error rate (frequency of accepting the null hypothesis when it is
471 false) when μ decreased through time. These poor performances were cor-
472 rected if the assumption $\mu = 0$ was relaxed (i.e., if a null birth–death model
473 was used in place of the Yule one), though the test had low power. Some
474 of these results make sense: the increased type I error rate obtained with
475 the Yule model is clearly due to the fact that a pattern of accelerated spe-
476 ciation can be created under a diversification process with extinction, when
477 old lineages are mostly extinct (e.g., Paradis 2011). On the other hand, the
478 high type II error rate of the same model when extinction rate increased
479 through time is somehow surprising considering the widely reported results
480 of slowing-down diversification (Rabosky and Lovette 2008b,a; Morlon et al.
481 2011; Etienne and Haegeman 2012, among others). Obviously, the same test
482 was not used in these studies, so this clearly requires further investigation.
483 Besides, the result that the test based on a birth–death model shows statis-
484 tically consistent results (i.e., the null hypothesis was rejected in less than
485 5% when μ was constant and in more than 5% when this parameter varied
486 through time) is encouraging and will also be further investigated. Interest-
487 ingly, this test was more powerful when the extinction rate increased through
488 time.

489 A particularly interesting result comes from the precision of the estimator
490 of extinction rate, $\hat{\mu}_{\text{BD}}$, which appears to have a very small bias, even when

491 the data were simulated with a relatively large value of μ . This contrasts
492 with previous studies showing that the estimator of extinction rate based
493 on complete phylogenies is, overall, inaccurate except if it is small compared
494 to the speciation rate (Paradis 2004; Didier et al. 2012). This result is
495 important because several authors have cast doubt on the possibility to
496 estimate with some precision extinction rates without fossils (Paradis 2011;
497 Aldous et al. 2011).

498 The analysis with the fish data were essentially illustrative, but the re-
499 sults call for several comments. The present method seems successful in
500 quantifying variation in diversification rates from a sample of clades. The
501 difference in the results from both taxonomic levels makes sense since we
502 expect more variation among families than among orders. The AIC values
503 evidence that the model assuming continuous variation in λ across clades
504 fits better than a model with discrete variation in this parameter. Since
505 similar tests have not been done with other data, this clearly calls for fur-
506 ther analyses before concluding whether diversification varies continuously
507 or discretely across clades.

508 The apparent failure to estimate the extinction rate, μ , of fishes is disap-
509 pointing since our simulation study showed that this parameter can be es-
510 timated correctly with the present approach. The fossil record shows many
511 episodes of radiations, extinctions, and turn-over during the evolutionary
512 history of fishes (Friedman and Sallan 2012). So the reality is very differ-
513 ent from the homogeneous scenario used in our simulations. Our results
514 combined with previous studies (e.g., Aldous et al. 2011) suggest that the
515 estimators of μ are far more complex when rate heterogeneity is present
516 which is likely the case with most real data set.

517 Vega and Wiens (2012) addressed the paradox of equivalent species di-

518 versity between marine and freshwater fishes despite the fact that freshwater
519 environments occupy a considerably smaller fraction of the Earth's surface
520 than oceans. In particular they wondered whether this could be related to
521 differences in diversification rates. Our results are in agreement with these
522 authors' who tested their hypothesis by correlating the proportion of marine
523 species in a clade with the method-of-moment estimator from Magallón and
524 Sanderson (2001). We found significant differences in λ between marine and
525 freshwater clades from the raw data; however, the small difference in $\hat{\lambda}$ be-
526 tween both groups suggested the influence of one or two clades. Hopefully,
527 the analysis of a more comprehensive data set with the statistical tools intro-
528 duced in this paper will help to solve the paradox of less biological diversity
529 in the ocean (Mora et al. 2011).

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634 sions of Dr. J. C. Willis, F.R.S. *Phil. Trans. R. Soc. Lond. B* 213:21–87.

Table 1. Rejection rate for the test of equality of diversification rate between two categories with n_1 and $n_2 (= n_1)$ clades.

λ_1	μ_1	λ_2	μ_2	n_1				
				1	3	5	10	20
0.1	0	0.1	0	0.044	0.060	0.066	0.054	0.056
0.1	0	0.1	0.05	0.038	0.085	0.108	0.140	0.203
0.15	0	0.1	0	0.094	0.177	0.232	0.418	0.711
0.15	0	0.1	0.05	0.112	0.297	0.446	0.759	0.958
0.2	0	0.1	0	0.174	0.497	0.707	0.943	0.998
0.2	0	0.1	0.05	0.236	0.643	0.855	0.993	1.000
0.1	0.05	0.1	0.05	0.040	0.054	0.048	0.054	0.061
0.15	0.05	0.1	0	0.050	0.075	0.083	0.123	0.209
0.15	0.05	0.1	0.05	0.069	0.129	0.201	0.387	0.653
0.2	0.05	0.1	0	0.119	0.240	0.384	0.693	0.928
0.2	0.05	0.1	0.05	0.143	0.407	0.618	0.878	0.995

Table 2. Rejection rate for the test of temporal variation in diversification. The null model was a Yule model with constant rate, and the alternative model was a Yule model with λ allowed to take different values before and after 30 time units. The first two pairs of columns give the parameter values used for the simulations (Ancient and Recent: values before and after 30 time units).

Ancient		Recent		Rejection rate
λ	μ	λ	μ	
0.01	0	0.01	0	0.029
0.1	0	0.1	0	0.038
0.1	0	0.05	0	0.917
0.1	0	0.01	0	1.000
0.05	0	0.1	0	0.923
0.01	0	0.1	0	1.000
0.1	0.025	0.1	0.025	0.105
0.1	0.05	0.1	0.05	0.248
0.1	0.025	0.1	0.075	0.057
0.1	0.075	0.1	0.025	0.682

Table 3. Same than in Table 2 but the null model was a birth–death model with constant rates, and the alternative model was a model with λ constant and μ allowed to take different values before and after 30 time units.

Ancient		Recent		Rejection rate
λ	μ	λ	μ	
0.1	0.05	0.1	0.05	0.019
0.1	0.075	0.1	0.025	0.080
0.1	0.025	0.1	0.075	0.121
0.1	0	0.1	0.08	0.313
0.1	0.08	0.1	0	0.211

Table 4. Rejection rate for the test of equality of diversification rate between two unknown categories using mixtures with n clades in each category.

λ	μ	n		
		10	20	50
0.1	0	0.011	0.006	0.011
(0.1, 0.2)	0	0.235	0.548	0.929
0.2	(0, 0.1)	0.057	0.143	0.369
0.2	(0, 0.15)	0.199	0.423	0.829

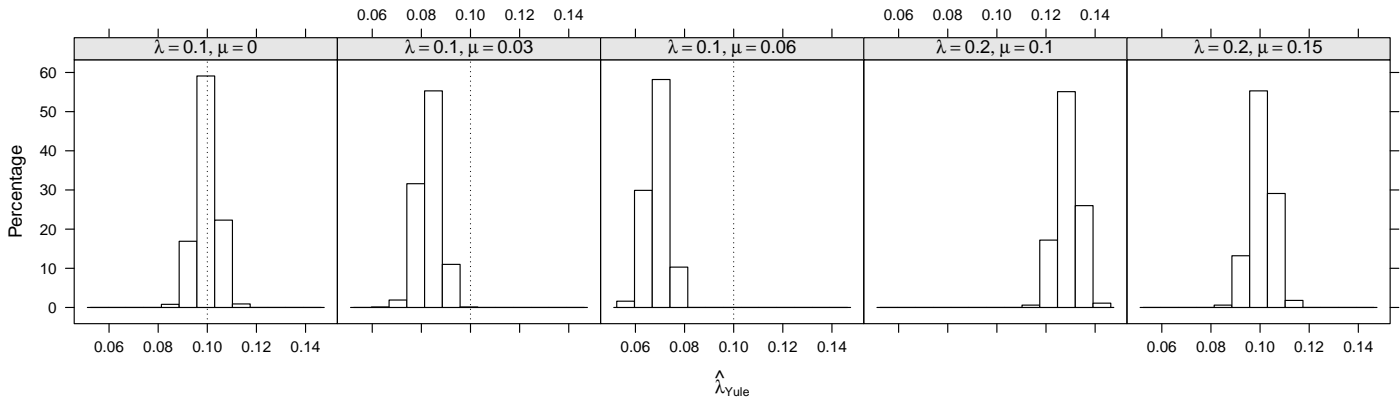
Table 5. Results of fitting models to the fish data using mixtures of Yule processes with K from two to seven.

K	Orders		Families	
	$\ln L$	AIC	$\ln L$	AIC
2	-171.207	348.414	-599.846	1205.691
3	-158.381	326.763	-599.846	1209.691
4	-171.207	356.414	-599.846	1213.691
5	-171.207	360.414	-599.846	1217.691
6	-171.207	364.414	-599.846	1221.691
7	-171.207	368.414	-599.846	1225.691

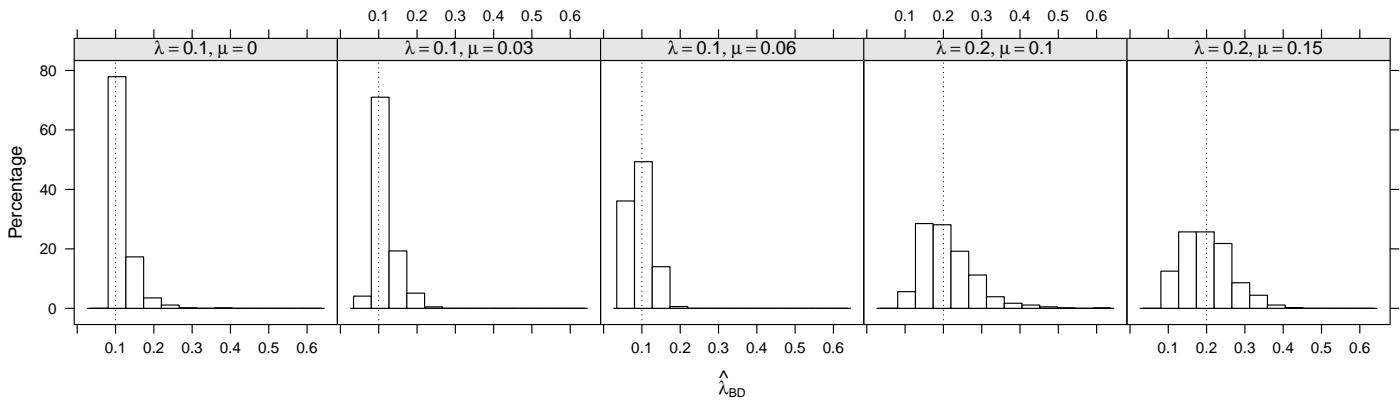
Table 6. Results of fitting a model of continuous variation in speciation rate across orders ($N = 22$) and families ($N = 97$) of fish.

	AIC	$\hat{\mu}_\lambda$ (SE)	$\hat{\sigma}_\lambda$ (SE)
Orders	320.396	1.221 (0.039)	0.163 (0.032)
Families	1137.066	1.086 (0.026)	0.224 (0.022)

A)



B)



C)

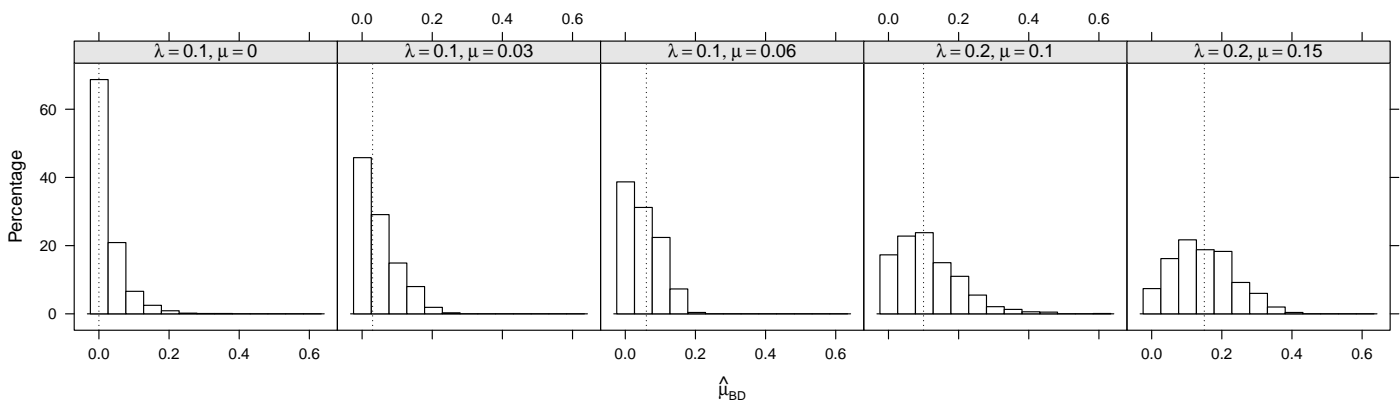


Figure 1. Distribution of the estimates of λ and μ with (A) the Yule model ($\hat{\lambda}_{\text{Yule}}$) and (B and C) the birth–death model ($\hat{\lambda}_{\text{BD}}$ and $\hat{\mu}_{\text{BD}}$) under five sets of parameters (values are given in the strips). Note the different scales of the x -axes. The vertical dotted lines indicate the values of λ (A and B) or μ (C) used in the simulation (not visible if outside the range of the x -axis). In all cases $n = 100$ clades.

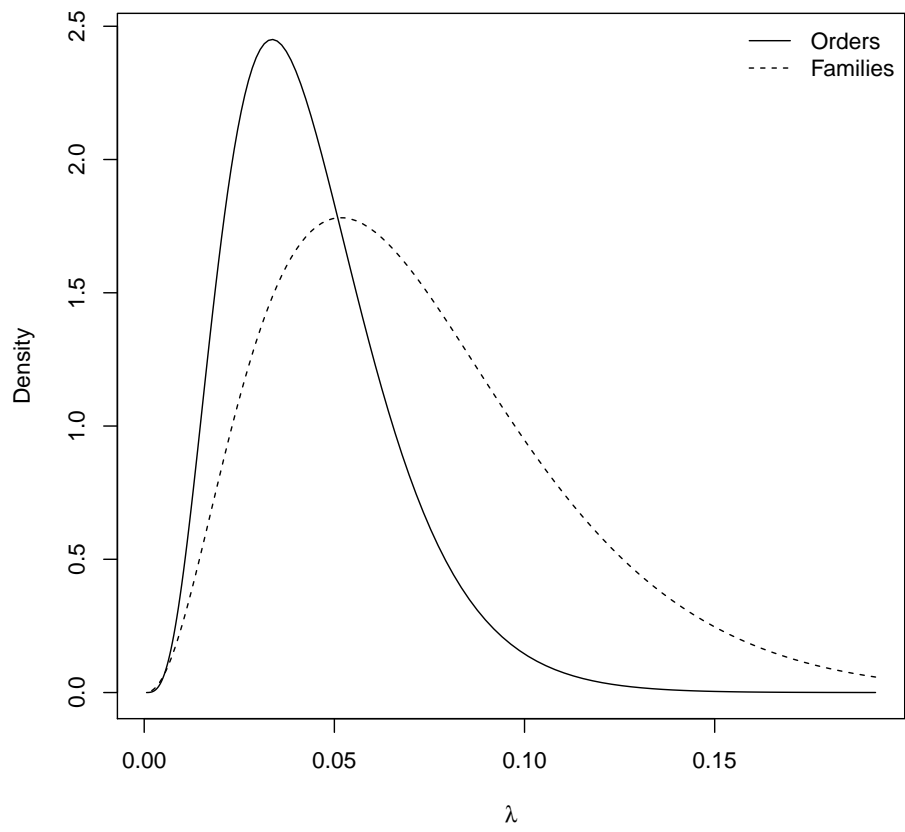


Figure 2. Inferred distribution of speciation rate among orders and families of fish.

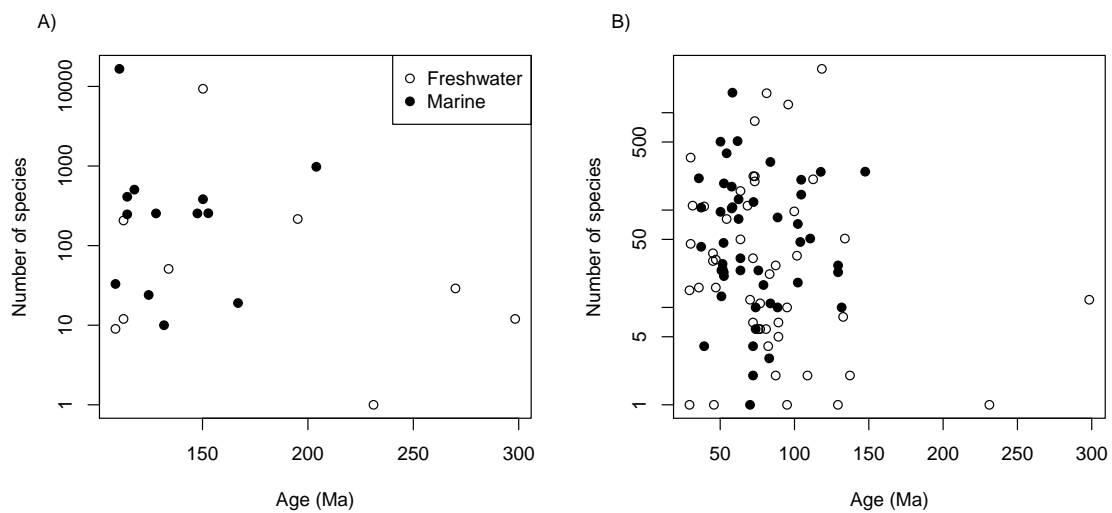


Figure 3. Number of species with respect to stem clade age for (A) orders and some super-orders and (B) families of fish (the legend is the same for both plots).