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Incisor enamel microstructure of hystricognathous and anomaluroid rodents from the earliest Oligocene of Dakhla, Atlantic Sahara (Morocco)

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3 **Incisor enamel microstructure of hystricognathous and anomaluroid**
4 **rodents from the earliest Oligocene of Dakhla, Atlantic Sahara (Morocco)**
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Abstract

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3 Seven hystricognaths and five anomaluroids have been recently described from the earliest
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5 Oligocene of the Dakhla (DAK C₂) region of Morocco, based primarily on isolated cheek
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7 teeth. Here, we analyzed the enamel microstructure of thirty associated isolated fragments of
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9 incisors. Among these specimens, only three display an early stage of uniserial Hunter-
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11 Schreger bands (HSBs), with mostly a single prism per band, but also occasionally two prisms
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13 per band (in two specimens), and a thin interprismatic matrix (IPM) that runs parallel to the
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15 prism direction, thereby documenting incisors of anomaluroids. All other sampled incisors
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17 display an enamel with multiseriate HSBs, thereby documenting hystricognaths. For these
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19 latter, we recorded primarily an IPM crystallite arrangement describing the subtype 2 of
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21 multiseriate HSBs, but with variation including a wide amplitude in the angle (acute) formed
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23 between the crystallites of IPM and those of the prisms, some variations in the frequency of
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25 the IPM sheet anastomoses, in the number of prisms per HSBs, and variations in the
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27 inclination of the HSBs. The absence of the subtypes 2-3 and 3 of multiseriate HSBs in DAK
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29 C₂ suggests that African hystricognathous rodents had still not achieved these most resistant
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31 multiseriate HSBs at that time. The drier, cooler climatic regime of the early Oligocene, having
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33 increased the fragmentation and opening of habitats, might have played a role in the
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35 subsequent selection of taxa having acquired a more resistant incisor enamel.
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Keywords

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51 Africa; Paleogene; Rodentia; uniserial; multiseriate
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Introduction

In the framework of our paleontological research program in North Africa, since 2009 we have concentrated some of our field efforts in the westernmost part of the Sahara, notably on the geological outcrops of the upper Samlat Formation, exposed on the mainland shoreline of the Rio de Oro, east of the Dakhla peninsula, Morocco (Fig. 1). Our paleontological surveys have allowed for the discovery of the first Paleogene record of rodents from the Atlantic margin of North Africa (Marivaux et al. 2017a,b). The fossils come from a single stratigraphic level (Dakhla C₂ [DAK C₂]; Adnet et al. 2017; Benammi et al. 2017; Marivaux et al. 2017a,b), which is particularly well exposed and accessible in two sections (Porto Rico [Pto] and El Argoub [Arg]; Fig. 1), situated in a limited area, about 20 km east of the Dakhla city. In both sections, this fossil-bearing level corresponds to estuarine deposits, which were dated to the earliest Oligocene by chemostratigraphy (Benammi et al. 2017; Noiret et al. 2017). Wet-screening of several tons of sediment (Pto C₂: ca. 1,700 kg; Arg C₂: ca. 1,500 kg) have yielded in the two localities similar fossil assemblages of marine and estuarine invertebrates and vertebrates, together with terrestrial mammals including rodents, primates, and afrotherians. Rodents are documented by isolated teeth and few jaw and bone fragments, which illustrate members of two phylogenetically distinct groups: Anomaluroidea and Hystricognathi (the only rodent clades to be known/present in Africa at that time). More than a dozen isolated cheek teeth document five anomaluroid species (*Paranomalurus riodeoroensis*, *Argouburus minutus*, *Oromys zenkerellinopsis*, *Nonanomalurus parvus*, and *Dakhlamys ultimus*; see Marivaux et al. 2017a), and several tens of isolated cheek teeth plus three jaw fragments document at least seven species of hystricognaths (*Gaudeamus* cf. *G. aslius*, *Gaudeamus* cf. *G. hylaeus*, *Phenacophiomys occidentalis*, *Birkamys* aff. *B. korai*, *Mubhammys atlanticus*, *Neophiomys minutus*, and ?*Phiocricetomys* sp.; see Marivaux et al.

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2017b). Whereas anomaluroid taxa provide an unprecedented taxonomic record of that rodent
group in Africa at that time, hystricognaths primarily document close relatives of taxa that are
known from Egyptian localities of the Jebel Qatrani Formation (Fayum Depression), either
dating from the latest Eocene (L-41; [Sallam et al. 2011](#); [Sallam and Seiffert 2016](#)) or from the
early Oligocene ([Wood 1968](#); see also [Coster et al. 2012](#), for the report of earliest Oligocene
hystricognaths from Libya, Zallah [Z5R locality]). This discovery provides a remarkable
snapshot regarding the paleodiversity of anomaluroid and hystricognathous rodents at that
time, which was so far only documented by fossil assemblages from northern and
northeastern Africa (Libya, Egypt) and from Arabia (Dhofar in Oman). This new earliest
Oligocene rodent fauna from the northern Atlantic margin of Africa (Atlantic Sahara) is
therefore particularly critical for balancing the early Oligocene rodent fossil record of North
Africa, and for a better understanding of the paleobiogeography of mammals at that time
([Marivaux et al. 2017a,b](#)). Furthermore, this unique assemblage demonstrates that rodents
were particularly diversified near the global cooling recorded at the Eocene/Oligocene
transition, thereby suggesting that this tropical region of North Africa was seemingly less
affected by the climatic changes recorded at that time (e.g., [Berggren and Prothero 1992](#);
[Coxall et al. 2005](#); [Lear et al. 2008](#); [Zachos et al. 2008](#); [Hren et al. 2013](#); [Tramoy et al. 2016](#)).

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In this paper, we continue the study of the DAK C₂ rodent fauna, in analyzing the enamel
microstructure of several isolated incisors (lower and upper specimens). For Paleogene
African (and Asian) hystricognaths and anomaluroids, this kind of analyses has so far been
carried out only on a few specimens, primarily by Martin ([1992, 1993, 1995](#); see also Martin
in [Coster et al., 2010](#)), and on a more systematic basis by Marivaux et al. ([2000, 2011, 2012,](#)
[2014, 2015, 2017a](#); see also [Marivaux 2000](#) [unpublished PhD]), when rodent taxa were
newly described or re-studied. Basically, the incisor enamel in rodents is primarily formed by
two layers ([Korvenkontio, 1934](#)), the Portio interna (PI), which includes decussating prism

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layers, named Hunter-Schreger bands (HSBs), and the Portio externa (PE), which consists of radial enamel (prisms are oriented in the same direction). From what is currently documented (extinct and extant species), all hystricognathous rodents (i.e., Old and New Worlds species [stem hystricognaths, phiomorphs, and caviomorphs]; ESM 1) display an incisor enamel characterized by a PI exhibiting decussating multi-prism layers (i.e., multiseriate HSBs). In contrast, the incisor enamel of anomaluroids (ESM 2) is characterized by a PI having decussating one (to two, in primitive species) prism layers (i.e., uniseriate HSBs). Initially, Martin (1992, 1993, 1994, 1997) distinguished three subtypes of multiseriate HSBs, especially on the basis of the angle formed by the direction of the interprismatic crystallites (IPM) with respect to the prism long axes (see also Vieytes 2003; Vucetich and Vieytes 2006; Boivin et al. submitted). In the subtype 1, the IPM crystallites run parallel to those of the prisms, or form a very low angle with them, and anastomose very regularly (sheet to sheath-like IPM). In the subtype 2, the IPM crystallites form an acute angle and anastomose regularly, while in the subtype 3, the IPM shows a few or no anastomoses, and its crystallites run at a right angle to those of the prisms (plate-like IPM). The same is true for the uniseriate HSBs, where different subtypes are observed in function of the angle of the IPM crystallites with respect to the prism long axes (e.g., Martin 1997). It has been argued that an increasing angulation of the IPM strengthens the enamel in the third dimensions, and that a rectangular IPM arrangement is the most derived and specialized condition as a result (better resistance to the crack propagations; Martin 1992, 1993, 1994, 1997). This biomechanical assumption is otherwise corroborated by the successive stratigraphic occurrences of the different subtypes (Martin 1994, 1997; see also Boivin et al. submitted and references herein).

Here, we selected, prepared, and analyzed some thirty isolated upper and lower rodent incisors of different sizes from DAK C₂ (Pto and Arg), in order to document a maximum of incisor enamel microstructural patterns recorded in this area at that time. With this large

1 sample analyzed, we expect to document as thoroughly as possible the evolutionary degrees
2 characterizing the incisor enamel microstructural complexity in these earliest Oligocene
3 hystricognathous and anomaluroid rodents. These degrees of complexity will be compared
4 with those of their more ancient, coeval, or more recent counterparts. This study will further
5 our understanding of the early evolutionary history of the incisor enamel microstructure
6 within these two distinct groups of rodents from Africa.
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18 **Material and Methods**

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25 Several rodent incisors (complete or fragmentary) were collected from the coarse (mesh \geq
26 2 mm) and thin ($1 \text{ mm} \leq \text{mesh} < 2 \text{ mm}$) residues of screening of the DAK-Arg and DAK-Pto
27 sediments (level C₂ [DAK C₂]). They correspond to upper (small radius of curvature) or lower
28 (high radius of curvature) incisors of different sizes, showing a continuous range of width,
29 primarily between 0.5 to 1.3 mm (Figs. 2 and 3). Although certain incisor specimens are
30 complete and compatible in size with molars of different hystricognath and anomaluroid
31 species from the same localities, a formal specific assignation of these incisors remains
32 uncertain in the absence of well-preserved and complete mandibles and skulls having incisors
33 associated with premolars and molars. Furthermore, several species of hystricognaths are very
34 close in size (somewhat overlapping), a condition which makes difficult or even impossible a
35 species attribution of the incisors. Two exceptions are the very large DAK-Pto-003 and DAK-
36 Arg-073 upper incisors (width $\geq 1.6 \text{ mm}$), which can be confidently attributed to the largest
37 rodent taxon recorded at DAK-Arg C₂ and DAK-Pto C₂ localities, the anomalurid
38 *Paranomalurus riodeoroensis* (see [Marivaux et al. 2017a](#)). Otherwise, despite the lack of
39 formal assignation for most of the specimens, we have nonetheless analyzed the enamel
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1 microstructure of a set of incisors from each locality. We have selected 15 specimens (lower
2 or upper incisor fragments) of different sizes in each locality. The specimens were embedded
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4 in artificial resin, then cut transversally in order to observe the enamel layer in cross section
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6 (Figs 2 and 3), and subsequently polished longitudinally. The polished sections were etched
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8 for 30 sec with phosphoric acid (H₃PO₄) at 37%, to make the enamel microstructural details
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10 visible. Each specimen was in fine examined by scanning electron microscope (SEM) at
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12 different magnifications (Figs. 4-8). The nomenclature of the enamel microstructure follows
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14 that of Koenigswald and Sander (1997) and Martin (1992, 1993). The measures carried out on
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16 the enamel layer observed in longitudinal section (Tables 1 and 2) follow those proposed by
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18 Martin (1992, 1993) and Boivin et al. (submitted). The datasets (SEM photographs) generated
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20 and analyzed during the current study are available from the corresponding author on
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22 reasonable request. In contrast, all prepared specimens during the current study are
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24 temporarily housed in the paleontological collections of the Université de Poitiers (Institut de
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26 Paléoprimatologie, Paléontologie Humaine : Évolution et Paléoenvironnements [iPHEP]).
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28 They will be subsequently housed permanently at the Université IBN Tofail, Kénitra
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43 **Results**

44 45 46 47 48 49 **Types of incisor enamel microstructures recorded and potential species attribution**

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55 Among the 30 incisors analyzed, only three specimens display an enamel characterized
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57 by uniserial Hunter-Schreger bands (HSBs). All the other specimens have an enamel
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1 exhibiting multiseriate HSBs (Tables 1 and 2). This low rate of recording of incisors with
2 uniseriate HSBs, corresponding to anomaluroid incisors, could have been in fact expected
3 given the low number of cheek teeth (15) documenting the five anomaluroid species from
4 DAK C₂. It was somewhat a chance to document at least five species with so little dental
5 material (Marivaux et al. 2017a). As mentioned above, given the very large size (width \geq 1.6
6 mm) of at least two incisors displaying an enamel with uniseriate HSBs (DAK-Pto-003 and
7 DAK-Arg-073; Tables 1 and 2; Fig. 4A-D), we can confidently attribute these two specimens
8 to the largest taxon of the DAK C₂ rodent fauna, the anomaluroid *Paranomalurus*
9 *riodeoroensis*, the molars of which are compatible in size with these two incisors. The third
10 incisor displaying an enamel with uniseriate HSBs (DAK-Pto-043; Table 1; Fig. 4E-F) is a
11 medium-sized specimen, and could either be attributed to the medium-sized nonanomaluroid
12 *Nonanomalurus parvus* or ?zegdoumyid *Dakhlamys ultimus*, the cheek teeth of these two
13 anomaluroid taxa being similar in size (Marivaux et al. 2017a).

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32 Regarding hystricognaths from DAK C₂, except for the tiny ?*Phiocricetomys* sp., which
33 is known by a single lower molar, the six other species recorded are documented by much
34 more dental material (see Marivaux et al. 2017b). This is consistent with the high rate of
35 recording of incisors with multiseriate HSBs. The dental material of *Mubhammys atlanticus*
36 and *Phenacophiomys occidentalis* is particularly abundant. These two taxa are medium-sized
37 species, *Phenacophiomys* being slightly larger than *Mubhammys*. Therefore, it might be
38 expected that the few largest incisors ($1.1 \text{ mm} \leq \text{width} \leq 1.3 \text{ mm}$; Table 1 and 2) displaying
39 an enamel characterized by multiseriate HSBs (Figs. 5A-B, 7A-B and 8A-B) could have been
40 those of *Phenacophiomys*. The abundant slightly smaller incisors ($0.8 \text{ mm} \leq \text{width} \leq 1.0 \text{ mm}$;
41 Tables 1 and 2) could then have been those of *Mubhammys*. However, although abundant, the
42 molars of *Mubhammys* are roughly similar in size to those of the two species of *Gaudeamus*
43 (*Gaudeamus* cf. *G. hylaeus* and *Gaudeamus* cf. *G. aslius*) from DAK C₂, a dental size overlap
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1 which then precludes any precise species attribution of these medium-sized incisors
2 displaying an enamel with multiseriate HSBs (Figs. 5C-F, 6A-B and 8C-F). The same is true
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4 regarding the smallest incisors ($0.5 \text{ mm} \leq \text{width} \leq 0.7 \text{ mm}$; Table 1 and 2) displaying an
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6 enamel with multiseriate HSBs (Figs. 6C-F and 7C-F), which could either belong to the tiny
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8 species from DAK C₂, *Neophiomys minutus* or *?Phenacophiomys* sp., or eventually to the
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10 slightly larger *Birkamys* aff. *B. korai*. It is worth noting that in the absence of complete
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12 mandibles and skulls having incisors associated with premolars and molars, our assessment of
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14 the taxonomic identification of the isolated incisors using for each enamel category ('uniseriate
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16 HSBs-anomaluroids' and 'multiseriate HSBs-hystricognaths'), the criterion of size
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18 compatibility between incisors and cheek teeth remains only tentative.
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28 **Incisor enamel with uniseriate HSBs**

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34 We have sampled only three upper incisors (no lower incisor) displaying an enamel with
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36 uniseriate HSBs. Among them, the two largest specimens (DAK-Pto-003 and DAK-Arg-073;
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38 Tables 1 and 2; Fig. 4A-D) exhibit roughly similar microstructural arrangements. Despite a
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40 difference in the total enamel thickness between the two incisors (enamel layer slightly
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42 thinner in DAK-Arg-073, but that specimen is also slightly smaller than DAK-Pto-003), both
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44 specimens are characterized by a strong development (i.e., thick) of the Portio externa (PE).
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46 The Portio interna (PI) of their enamel layer consists of decussating HSBs, among which
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48 about 80% display one prism per band, and punctually (not regularly) two prisms per band.
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50 The HSBs are straight and faintly inclined (max. 10°). The prisms are strongly flattened,
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52 thereby appearing oval and inclined in cross section. The inter-prismatic matrix (IPM) in the
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54 PI is thin and does not form a sheath surrounding the prisms. The IPM crystallites rather form
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1 thin sheets that run nearly parallel to the prism direction and anastomose continuously
2 between the prisms. This generates an IPM sheet of transition between two adjacent
3 decussating HSBs. Such an IPM crystallite arrangement in PI associated with the occasional
4 presence of HSBs having two prisms per band testify of an early stage toward the
5 achievement of a fully uniserial condition (Korvenkontio 1934; Martin 1993, 1997).
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12 The third upper incisor displaying an enamel with uniserial HSBs is a medium-sized
13 specimen (DAK-Pto-043; Table 1; Fig. 4E-F). As the two other large upper incisors described
14 above, the enamel layer of this specimen is moderately thin (compared with large-, medium-,
15 or small-sized incisors with an enamel characterized by multiserial HSBs) and with a thick
16 PE. The latter is however slightly thinner than in the two other anomaluroid incisors. In the PI
17 of the enamel layer of DAK-Pto-043, 100% of the decussating HSBs have only one prism per
18 band. The HSBs are also much more inclined ($\sim 56^\circ$) than in the enamel of the two other
19 incisors. The prisms are less flattened, appearing round or slightly oval in cross section. The
20 IPM crystallites in the PI form thin sheets that run nearly parallel to the prism direction,
21 without frequent anastomoses. Such an IPM crystallite arrangement in PI associated with
22 HSBs having only one-prism layers, testify of an early stage of the uniserial condition.
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43 **Incisor enamel with multiserial HSBs**

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49 Of the 30 incisor fragments of different sizes selected from the two localities of the
50 Dakhla level C₂ (DAK-Pto and DAK-Arg), we have in fact primarily sampled upper and
51 lower incisors of hystricognaths, inasmuch as 27 specimens exhibit an enamel with multiserial
52 HSBs (Tables 1 and 2). Whatever the size (large-sized [$1.1 \text{ mm} \leq \text{width} \leq 1.3 \text{ mm}$], medium-
53 sized [$0.8 \text{ mm} \leq \text{width} \leq 1.0 \text{ mm}$], or small-sized [$0.5 \text{ mm} \leq \text{width} \leq 0.7 \text{ mm}$] specimens) or
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1 the jaw position (upper or lower) of the incisors, we do not observe strong variation in the
2 enamel crystallite arrangements. The majority of incisors displays an enamel with multiseriate
3 HSBs, the IPM crystallites of which appear as thin sheets that anastomose regularly, and their
4 direction form an acute angle to the prism direction. Following Martin (1992, 1993, 1994,
5 1997, 2007) (but also Vieytes 2003; Vucetich and Vieytes 2006), this kind of crystallite
6 arrangement typifies the subtype 2 of multiseriate HSBs. Despite that unique subtype record,
7 we observe nonetheless some variations across incisors, principally in the thickness of the
8 enamel layer, in the value of the acute angle (wide amplitude) formed between the direction
9 of the prism crystallites and those of the IPM sheets, and to a lesser extent, in the number of
10 prisms per HSBs, the inclination of the HSBs, and in the frequency of the IPM sheet
11 anastomoses (Tables 1 and 2). Therefore, instead of describing exhaustively the enamel
12 microstructure for each incisor sampled (all exhibiting a subtype 2 of multiseriate HSBs), here
13 after we focus on the main microstructural characters that can differ between incisors of
14 different sizes and between upper and lower incisors (of similar size).

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38 **Large-sized incisors:** Among lower incisors of similar size (equal width), the enamel
39 thickness strongly varies (+ 60%; e.g., DAK-Pto-042 vs DAK-Pto-037; Table 1; Fig. 5A-B).
40 Such a marked difference in enamel thickness could indicate the presence of distinct species
41 equivalent in size. However, despite this thickness distinction, the crystallite arrangements are
42 roughly similar, notably regarding the IPM configuration, which shows a comparable range in
43 the values of the acute angle formed between the IPM sheets and the prism long axes. The
44 unique upper incisor analyzed for this size category (DAK-Arg-074; Table 2; Fig. 8A-B)
45 display a thick enamel layer, similar to that of the lower DAK-Pto-037 incisor (Table 2; Fig.
46 5A-B). In contrast, that upper incisor displays an enamel with thicker HSBs, having 4-5
47 prisms per band, contra 3-4 prisms per band in the other large specimens. In this upper
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2 incisor, the value of the acute angle between the crystallites of IPM and the prism direction is
3 also often less marked (Table 2; Fig. 8B).
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9 **Medium-sized incisors:** This size category also exhibits a wide spectrum of enamel
10 thickness among and within the lower and upper specimens. Two upper incisors (DAK-Arg-
11 076 and DAK-Arg-085; Table 2; Fig. 8E-F) display particularly thick HSBs, including 5-6
12 prisms per band, and a moderate value of the acute angle between the crystallites of IPM and
13 those of the prisms. Other upper incisors of this size category have rather 3-4 prisms per HSB
14 but similar values of the IPM/prism crystallite angles (Tables 1 and 2). Lower incisors display
15 also 3-4 prisms per HSB, and generally slightly higher values of the IPM/prism crystallite
16 angles. Practically all specimens display prisms of transitional zone between two adjacent
17 HSBs. One upper incisor (DAK-Pto-038; Table 1; Fig. 6A-B), among the smallest one of this
18 size category (i.e., width = 0.8 mm), exhibits prisms having a section appearing more rounded
19 (circular) than that observed in the other upper or lower specimens (prisms being oval to
20 flattened in section). The IPM crystallites deviate only by 23° from those of the prisms, and
21 anastomose very frequently (very regularly), more than on the other specimens. However, the
22 IPM crystallites do not form a sheath-like IPM (as the condition describing the subtype 1 of
23 multiserial HSBs), but form sheets. Given the latter characteristics, this specimen could
24 illustrate a case of transitional subtype 1-2 of multiserial HSBs.
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51 **Small-sized incisors:** We have analyzed the enamel microstructure of several lower and
52 upper incisors of this size category (Tables 1 and 2; Figs. 6C-F and 7C-F). Practically all
53 specimens have 3-4 prisms per HSB (rarely up to 5 prisms per band), a similar HSB
54 inclination, no prisms of transitional zone between two adjacent decussating HSBs, and a
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1 relatively similar enamel thickness, except for one specimen (DAK-Arg-079; Table 2; not
2 figured). The latter, an upper incisor, indeed shows a markedly thinner enamel layer than that
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4 of the other upper and lower incisors of similar size (width = 0.7 mm). Within this size
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6 category, two upper incisors (DAK-Pto-046 and DAK-Pto-050; Table 1; Fig. 6C-D) have
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8 moderately low values ($\sim 25^\circ$) of the angle formed between the crystallites of IPM and those
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10 of the prisms. On these two specimens, the sheets of IPM also anastomose very frequently,
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12 nearly forming a sheath-like IPM arrangement (Fig. 6D). In contrast, some incisors of this
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14 size category, notably the lower incisors (e.g., DAK-Arg-077 [Fig. 7C-D], DAK-Arg-082
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16 [Fig. 7E-F], and DAK-Pto-049) exhibit markedly higher values of the IPM/prism crystallite
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18 angles (50° up to 60° ; Tables 1 and 2), with less frequent to rare anastomoses of the IPM
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20 sheets. With such an IPM arrangement, the latter incisors illustrate an enamel microstructure
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22 characterized by an advanced subtype 2 of multiseriate HSBs.
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33 **Discussion and Conclusion**

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39 These earliest Oligocene estuarine deposits from Dakhla (DAK C₂) yield the only
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41 Paleogene rodent assemblage from the Atlantic margin of North Africa (Atlantic Sahara). The
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43 great rodent diversity recorded in that area, associating in sympatry at least seven
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45 hystricognaths and five anomaluroids was somewhat unexpected given the unfavorable global
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47 climatic conditions characterizing this time window (i.e., global cooling recorded at the
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49 Eocene/Oligocene transition). Such a rodent diversity from DAK C₂, including several
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51 arboreal species, rather indicates that this tropical region of North Africa was seemingly less
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53 affected by these climatic changes (Marivaux et al. 2017a,b). This new northwestern African
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55 fauna hence represents a remarkable snapshot regarding the paleodiversity of rodents at that
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1 time, and balances the early Oligocene rodent fossil record of North Africa, which was so far
2 only documented by fossil assemblages from northern and northeastern Africa (Libya, Egypt)
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4 and from Arabia (Dhofar in Oman). Wet-screening of the sediments of the two rodent-bearing
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6 localities from DAK C₂ has allowed collecting many isolated cheek teeth but also many
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8 fragments of rodent incisors. Despite these fragments are isolated, and as such not formally
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10 (or firmly) attributed to a given species, we thought it worthwhile to analyze the enamel
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12 microstructure of several specimens of different sizes in order to have, for the first time, an
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14 overview of the enamel evolutionary degrees recorded for an African rodent community of
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16 that age. Indeed, only few analyses of the rodent incisor enamel microstructure have been
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18 carried out for African fossil rodents. For the Oligocene of Africa, the incisor enamel
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20 microstructure is only documented for *Phiomys andrewsi*, *Metaphiomys schaubi*, *Gaudeamus*
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22 *aegyptius* (see [Martin 1992](#)), *G. lavocati* ([Martin in Coster et al. 2010](#)) and *Turkanamys*
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24 *hexalophus* (see [Marivaux et al. 2012](#)) (ESM 1). These taxa occur in stratigraphic levels more
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26 recent than DAK C₂ (for a review, see [Marivaux et al. 2017b](#): fig. 10). The latest Eocene L-41
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28 locality of the Fayum, Egypt, very close in age to DAK C₂, and that yields a great
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30 hystricognath diversity ([Holroyd 1994](#); [Sallam et al. 2011, 2012](#); [Sallam and Seiffert 2016](#)),
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32 has not hitherto been the subject of incisor enamel microstructure analyses. The same is true
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34 for the BQ-2 locality (Fayum) dating from the early late Eocene, the Dur at-Talah localities
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36 (Libya) dating from the late Eocene, or for the Taqah locality (Oman) dating from the early
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38 Oligocene, localities which yield sympatric anomaluroid and hystricognathous rodents
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40 ([Thomas et al. 1989, 1992](#); [Sallam et al. 2009, 2010a,b](#); [Jaeger et al. 2010](#); [Coster et al. 2015](#)).
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42 The enamel microstructure for Eocene African rodents is only documented for the late middle
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44 to early late Eocene hystricognath *Protophiomys tunisiensis* (Tunisia; see [Marivaux et al.](#)
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46 [2014](#)) and ?*P. algeriensis* (Algeria; see [Coiffait et al. 1984](#); [Martin 1992, 1993](#)) (see ESM 1),
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48 and for the late early to early middle Eocene zegdoumyid anomaluroids (Algeria and Tunisia;
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Martin 1993; Marivaux et al. 2011, 2015) and early late Eocene nementchamyid anomaluroids (Algeria; Martin 1993) (see ESM 2). With the study of the incisor enamel of the rodent fauna from Dakhla (DAK C₂), we then contribute to a better knowledge of the diversity and evolutionary degree of the enamel microtextural pattern developed at that time for the two emblematic African rodent groups: anomaluroids and hystricognaths.

The numerous works of Martin (e.g., 1992, 1993, 1994, 1995, 1997) concerning the analysis of the incisor enamel microstructure in rodents, have shown that all hystricognaths display an enamel characterized by multiseriate HSBs (ESM 1), whereas anomaluroids exhibit an enamel incisor with uniseriate HSBs (or transitional pauciseriate/uniseriate HSBs for the stem groups; ESM 2). These clearly distinct enamel patterns allow therefore for a proper supra-familial attribution of the isolated incisors from DAK C₂. In contrast, their generic or specific attributions remain much more delicate in the absence of preserved incisor-premolar-molar association, and the criterion of size compatibility between incisors and cheek teeth in each enamel category remains only tentative (see results). Despite the large number of incisors sampled for this study, we found only three incisors having an enamel with uniseriate HSBs. This small number of anomaluroid incisors sampled is consistent with the small number of anomaluroid cheek teeth recovered in DAK-Arg C₂ and DAK-Pto C₂, and documenting the five species (Marivaux et al. 2017a). Two large incisors are likely attributable to *Paranomalurus riodeoroensis*, the largest rodent from DAK C₂. The enamel microstructure of these two incisors is characterized by a thinning of the HSBs, with generally a single flattened prism per band, but also occasionally two prisms per band, and a thin IPM that runs parallel to the prism direction, thereby typifying an early stage toward the achievement of a fully uniseriate condition. The third anomaluroid incisor (slightly smaller in size than the two others) has an enamel displaying 100% of uniseriate HSBs (no double-layered HSBs observed), but with a similar thin IPM sheet arrangement (i.e., parallel to the prism direction) and with prism

1 having a round section, thereby testifying of an early stage of the uniserial condition. The
2 incisor enamel microstructure of modern anomaluroids is only documented by the genus
3 *Anomalurus* (Anomalurinae, Anomaluridae). The incisor enamel of this taxon primarily
4 shows single flattened prism layers, but also, only rarely, double-layered HSBs, and a thin
5 IPM that runs parallel to the prism direction (Martin 1993, 1997), an enamel microstructural
6 pattern that is in fact strongly reminiscent to that observed in *Paranomalurus riodeoroensis*
7 (Anomalurinae) from Dakhla. Unfortunately, nothing is known regarding the incisor enamel
8 microstructure of the other modern anomaluroids, *Anomalurops* (Anomalurinae,
9 Anomaluridae), *Idiurus* (Idiurinae, Anomaluridae) and *Zenkerella* (Zenkerellidae), or of the
10 Miocene taxa such as the different species of *Paranomalurus* (*P. walkeri*, *P. bishopi*;
11 Anomalurinae, Anomaluridae), and *Nonanomalurus soniae* (Nonanomaluridae). This lack of
12 enamel description for Miocene and modern anomaluroids limits *i*) our comparisons and
13 therefore the precise assessment of the enamel evolutionary degree observed in anomaluroids
14 from Dakhla, and *ii*) our understanding of the evolutionary trends of the enamel
15 microstructure in this rodent group. The incisor enamel microstructure is somewhat better
16 documented for stem anomaluroids, notably for the Eocene zegdoumyids and
17 nementchamyids (Martin 1993; Marivaux et al. 2005, 2011) (ESM 2). These two groups have
18 an incisor enamel showing the tendency towards a thinning of the HSBs (one or one-two
19 prisms per band), a trend observed as early as the late early Eocene (in zegdoumyids), but the
20 IPM crystallite arrangement in that enamel is not as advanced as in the anomalurine enamel
21 incisors from Dakhla. In zegdoumyids and nementchamyids, the IPM is indeed thicker,
22 parallel to the prism direction, and generally forms a sheath surrounding the prisms (not
23 appearing as thin sheets). Such an IPM arrangement typifies an enamel transitional from the
24 primitive pauciserial to the uniserial condition (Martin 1993; Marivaux et al. 2005, 2011).
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The other DAK C₂ sampled incisors display an enamel with multiseriate HSBs. Despite the large number of incisors with this enamel type, we recorded primarily an IPM crystallite arrangement describing the subtype 2 of multiseriate HSBs (Martin 1992, 1993, 1994, 1997; Vieytes 2003). However, we observed many microstructural variations across incisors (i.e., differences in the degree of crystallite arrangements), likely reflecting different species. We recorded indeed a wide amplitude in the angle (acute) formed between the crystallites of IPM and those of the prisms, some variations in the frequency of the IPM sheet anastomoses, in the number of prisms per HSBs, and variations in the inclination of the HSBs (Tables 1 and 2). The thickness of the enamel layer also strongly varies across incisors of different sizes and within a size category. Only two incisors show an enamel with an IPM crystallite arrangement that describes a case of transitional subtype 1-2 of multiseriate HSBs, and three others have an enamel characterized by an advanced subtype 2 of multiseriate HSBs (see results). The subtype 2 of multiseriate enamel is widespread among modern and extinct New World hystricognaths (i.e., caviomorphs, notably chinchilloids, cavioids, and erethizontoids, with the exception of most octodontoids; Martin 1992, 1993, 1994; 2004, 2005; Vieytes 2003; Vucetich and Vieytes 2006; Vucetich et al. 2010; for a review see Boivin et al. submitted), among early Old World hystricognaths (e.g., Asian tsaganomyids, South Asian baluchimyines, early African protophiomysines, phiomysids, and gaudeamurids; Martin 1992; Marivaux 2000; Marivaux et al. 2000, 2012; Martin in Coster et al. 2010; see ESM 1), but also in Asian “ctenodactylids” (e.g., early diatomyids and early ctenodactylids; Martin 1992, 1995, 2007; Marivaux et al. 2004; see ESM 1). It is worth noting that from the large set of incisor fragments analyzed from DAK C₂, we did not record any enamel documenting a subtype 1 of multiseriate HSBs, as that observed in the oldest hystricognath rodent to be known in Africa thus far (*Protophiomys tunisiensis*, 39.5 Ma; see Marivaux et al. 2014), but also in certain early Oligocene baluchimyine hystricognaths from South Asia (Martin 1992, 1995), in the middle Eocene

1 tamquammyid ctenodactyloids from Mongolia (Martin 1993, 1995, 2007), and in the early
2 Miocene diamantomyid taxon (*Diamantomys*; hystricognaths) from Kenya (Martin 1992)
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4 (ESM 1). The subtype 1 is also characteristic of the incisor enamel of modern hystricids
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6 (Martin 1992), and it is particularly widespread among several extinct and extant
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8 caviomorphs, notably in erethizontoids, cavioids, and chinchillids (Martin 1992, 2004, 2005;
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10 Vиейtes 2003; for a review see Boivin et al. submitted). Also, we did not record in DAK C₂
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12 any enamel documenting the transitional subtype 2-3 and the subtype 3 of multiserial HSBs,
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14 as that characterizing some more recent and evolutionarily advanced phiomorphs (ESM 1),
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16 such as *Metaphiomys schaubi* (early Oligocene) or *Paraphiomys pigotti* (early Miocene). All
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18 modern phiomorphs (i.e., Thryonomyoidea; ESM 1) have an incisor enamel characterized
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20 either by the subtype 2-3 of multiserial HSBs (Thryonomyidae and Bathyergidae) or by the
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22 subtype 3 of multiserial HSBs (Petromuridae). The latter enamel microstructure also
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24 characterizes incisors of modern and most of extinct South American octodontoid
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26 caviomorphs (Martin 1992, 1994, 2005; Vиейtes 2003; Vиейtes et al. 2007; Morgan et al.
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28 2017; for a review see Boivin et al. submitted), as well as some Miocene and modern
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30 “ctenodactyloids” (ctenodactylids and diatomyids; Martin 1992, 1995; Dawson et al. 2006)
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32 (ESM 1). The subtype 3 of multiserial HSBs is characterized by IPM crystallites running at a
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34 right angle to those of the prisms (plate-like IPM with a few or no anastomoses). Given that
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36 an increasing angulation of the IPM sheets is considered as strengthening the enamel in the
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38 third dimensions, multiserial HSBs with rectangular IPM (subtype 3) provide to the incisor
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40 enamel a better resistance to the crack propagations (Martin 1992, 1993, 1994, 1997, 2007).
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42 On the basis of this biomechanical consideration and the stratigraphic occurrences of taxa, the
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44 subtype 3 is considered as the most derived and specialized multiserial condition.
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46 Accordingly, considering the IPM arrangement, the subtype 1 is the most primitive (and the
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48 less resistant) condition of multiserial HSBs, and the subtype 2 is intermediary in terms of
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resistance and evolutionary degree, between the subtypes 1 and 3 (Martin 1992, 1993, 1994, 1997). The subtypes 1 and 2 (and the transitional subtype 1-2) are therefore preserved in several modern hystricognaths, and the subtype 3 (and the transitional subtype 2-3) seems to have been acquired iteratively in several extinct and extant groups (Martin 1992, 1994; see also Vucetich and Vieytes 2006; Boivin et al. submitted).

In DAK C₂, dating from the earliest Oligocene, we record almost exclusively the intermediary subtype 2 and no subtype 1 and 3 (or 2-3) of multiseriate HSBs. Given the absence of the subtypes 2-3 and 3 in DAK C₂, we might then question whether African hystricognathous rodents had already achieved the most resistant multiseriate HSBs at that time. But, the possibility exists that this unique enamel subtype 2 sampling from Dakhla is the result of a taphonomic bias, or that taxa displaying the subtypes 2-3 or 3 of multiseriate HSBs were not present in the westernmost part of Africa at that time. A fully achieved subtype 3 of multiseriate HSBs is observed shortly after the DAK C₂ record, notably in the early Oligocene localities of the Fayum in Egypt, with *Metaphiomys schaubi* (Martin 1992). Recent analyses of the enamel microstructure of Eocene and Oligocene rodent incisors from Peruvian Amazonia (Boivin et al. submitted) have shown that the subtypes 2-3 and 3 of multiseriate HSBs are only recorded from early Oligocene localities (i.e., Shapara and Santa Rosa; see also Martin 2004, 2005 for Santa Rosa, the age of which is uncertain: ?latest Eocene/early Oligocene), and not in older Eocene localities (Contamana), where only the subtypes 1 and 2 are recorded. Boivin et al. (submitted) have suggested that the derived subtypes 2-3 and 3 conditions were secondarily but rapidly achieved in caviomorphs, “and seemingly evolved iteratively but only in the octodontoid clade” (see also Vucetich and Vieytes 2006), the subtypes 1 and 2 being “conserved in most of caviomorph superfamilies through time” (Boivin et al. submitted). For Ctenohystrica in general, it would be then interesting to identify the ecological (and perhaps genetic?) and paleoenvironmental factors, that have driven the

1 achievement and selection of more resistant incisor enamel microstructures (subtypes 2-3 and
2 3) in certain groups only, while other taxa, in seemingly similar environments, conserved less
3 resistant microstructures (subtypes 1, 2, or 1-2) through time. For Africa, we might question
4 whether the drier, cooler climatic regime of the early Oligocene, having probably increased
5 the fragmentation and opening of habitats (with more abrasive and harder food items), have
6 played a role in the selection of taxa having acquired a more resistant incisor enamel in these
7 region of North Africa. But a similar scenario could be also advocated for explaining the
8 Oligocene achievement of a subtype 3 of multiseriate HSBs in Old World “ctenodactylids”
9 (i.e., ctenodactylids and diatomyids) and in some New World hystricognaths (i.e.,
10 octodontoids), inasmuch as the climatic deterioration of the early Oligocene was recorded at
11 the global scale (e.g., [Coxall et al. 2005](#); [Lear et al. 2008](#); [Zachos et al. 2008](#); [Hren et al. 2013](#);
12 [Tramoy et al. 2016](#)). Only a broader analysis of the incisor enamel microstructure in
13 hystricognathous rodents (and more widely Ctenohystrica) documenting the late Eocene –
14 early Oligocene time interval would allow to assess the timing of achievement of the subtypes
15 2-3 and 3 of multiseriate HSBs, and the possible relationships with the paleoenvironmental
16 changes.

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Fig. 1 Geographic locations of the Dakhla peninsula, northwestern Africa (Atlantic Sahara), and of the fossil rodent-bearing localities. The geological outcrops of interest (El Argoub [Arg] and Porto Rico [Pto]) are exposed on the mainland shoreline of the *Rio de Oro* inlet, east of the Dakhla peninsula (photographs by L. Marivaux). The rodent-bearing localities, El Argoub level C₂ (DAK-Arg C₂) and Porto Rico level C₂ (DAK-Pto C₂), are situated directly southeast of the Dakhla city. On the two photographs, the dashed white lines indicate the fossiliferous level C₂, which consists of a ca. 30 cm thick sandy and unconsolidated microconglomerate.

Fig. 2 Incisor cross-sections of Dakhla C₂ rodents from Porto Rico. **a**, DAK-Pto-043. **b**, DAK-Pto-037. **c**, DAK-Pto-038. **d**, DAK-Pto-039. **e**, DAK-Pto-044. **f**, DAK-Pto-041. **g**, DAK-Pto-042. **h**, DAK-Pto-046. **i**, DAK-Pto-048. **j**, DAK-Pto-040. **k**, DAK-Pto-050. **l**, DAK-Pto-051. **m**, DAK-Pto-047. **n**, DAK-Pto-045. **o**, DAK-Pto-049. **a**, **c-d**, **h-i** and **k-l**: upper incisors. **b**, **e-g**, **j** and **m-o**: lower incisors. Scale bar = 1 mm.

Fig. 3 Incisor cross-sections of Dakhla C₂ rodents from El Argoub. **a**, DAK-Arg-074. **b**, DAK-Arg-076. **c**, DAK-Arg-073. **d**, DAK-Arg-072. **e**, DAK-Arg-085. **f**, DAK-Arg-075. **g**, DAK-Arg-086. **h**, DAK-Arg-078. **i**, DAK-Arg-079. **j**, DAK-Arg-080. **k**, DAK-Arg-077. **l**, DAK-Arg-081. **m**, DAK-Arg-082. **n**, DAK-Arg-083. **o**, DAK-Arg-084. **a-c**, **e**, **h-i** and **n**: upper incisors. **d**, **f-g**, **j-m** and **o**: lower incisors. Scale bar = 1 mm.

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Fig. 4 Scanning electron photomicrographs of Dakhla C₂ upper incisors of anomaluroids from Porto Rico and El Argoub, for which the enamel microstructure (uniserial HSBs) is shown in longitudinal section and at different magnifications. The left pictures illustrate an overview of the enamel layer for each specimen, and the associated right ones a detail of the microstructure in PI. **a-b**, DAK-Pto-003, very large-sized incisor (already figured in [Marivaux et al., 2017a, fig. 7](#)). **c-d**, DAK-Arg-073, large-sized incisor. **e-f**, DAK-Pto-043, medium-sized incisor.

Fig. 5 Scanning electron photomicrographs of Dakhla C₂ lower incisors of hystricognaths from Porto Rico, for which the enamel microstructure (multiserial HSBs) is shown in longitudinal section and at different magnifications. The left pictures illustrate an overview of the enamel layer for each specimen, and the associated right ones a detail of the microstructure in PI. **a-b**, DAK-Pto-037, large-sized incisor; **c-d**, DAK-Pto-041, medium-sized incisor. **e-f**, DAK-Pto-044, medium-sized incisor.

Fig. 6 Scanning electron photomicrographs of Dakhla C₂ upper incisors of hystricognaths from Porto Rico, for which the enamel microstructure (multiserial HSBs) is shown in longitudinal section and at different magnifications. The left pictures illustrate an overview of the enamel layer for each specimen, and the associated right ones a detail of the microstructure in PI. **a-b**, DAK-Pto-038, medium-sized incisor. **c-d**, DAK-Pto-046, small-sized incisor. **e-f**, DAK-Pto-048, small-sized incisor.

Fig. 7 Scanning electron photomicrographs of Dakhla C₂ lower incisors of hystricognaths from El Argoub, for which the enamel microstructure (multiserial HSBs) is shown in

1 longitudinal section and at different magnifications. The left pictures illustrate an overview of
2 the enamel layer for each specimen, and the associated right ones a detail of the
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4 microstructure in PI. **a-b**, DAK-Arg-075, large-sized incisor. **c-d**, DAK-Arg-077, medium-
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6 sized incisor. **e-f**, DAK-Arg-082, small-sized incisor.
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12 **Fig. 8** Scanning electron photomicrographs of Dakhla C₂ upper incisors of hystricognaths
13 from El Argoub, for which the enamel microstructure (multiserial HSBs) is shown in
14 longitudinal section and at different magnifications. The left pictures illustrate an overview of
15 the enamel layer for each specimen, and the associated right ones a detail of the
16 microstructure in PI. **a-b**, DAK-Arg-074, large-sized incisor. **c-d**, DAK-Arg-078, medium-
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18 sized incisor. **e-f**, DAK-Arg-085, medium-sized incisor.
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Table captions

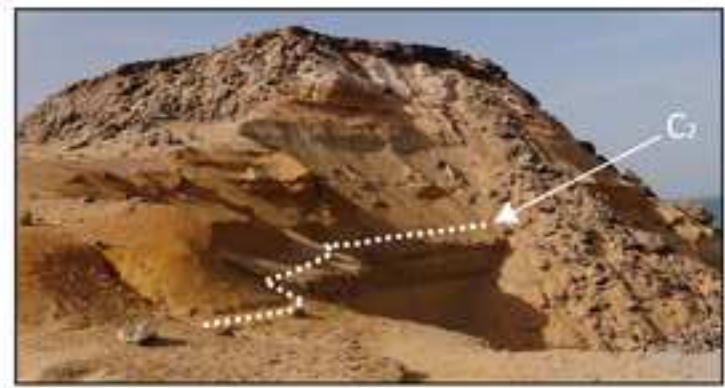
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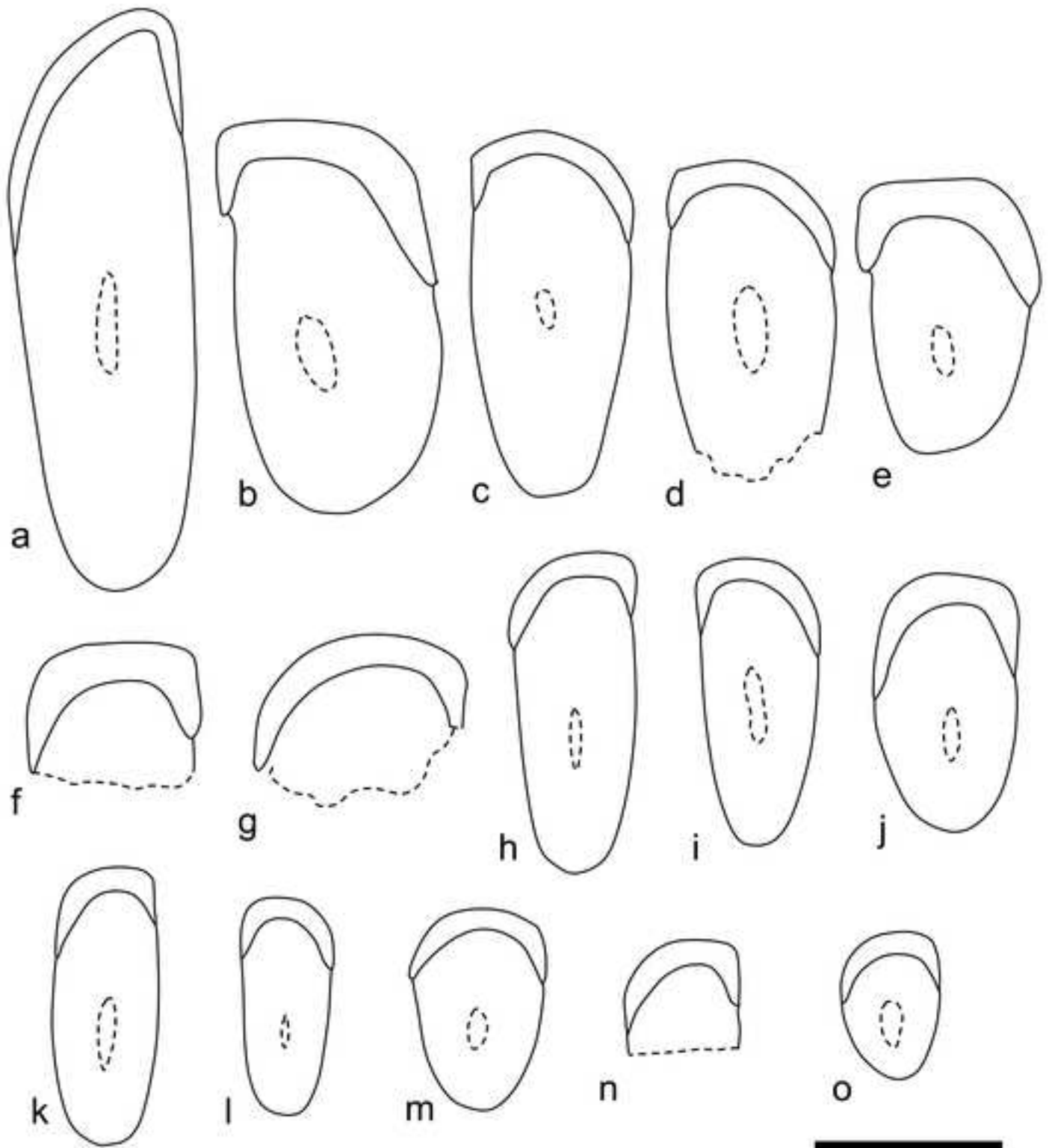
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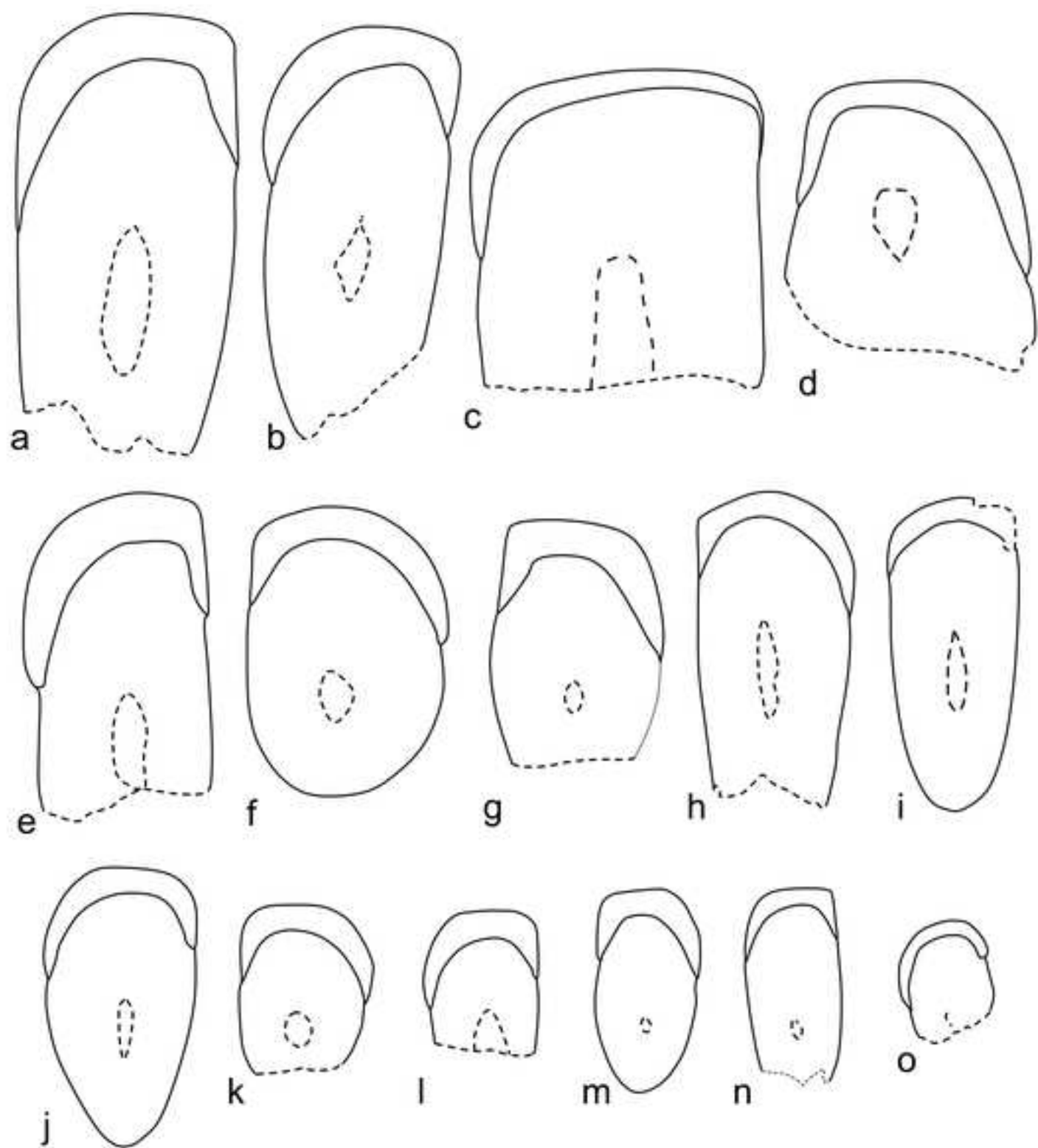
Supplementary Material

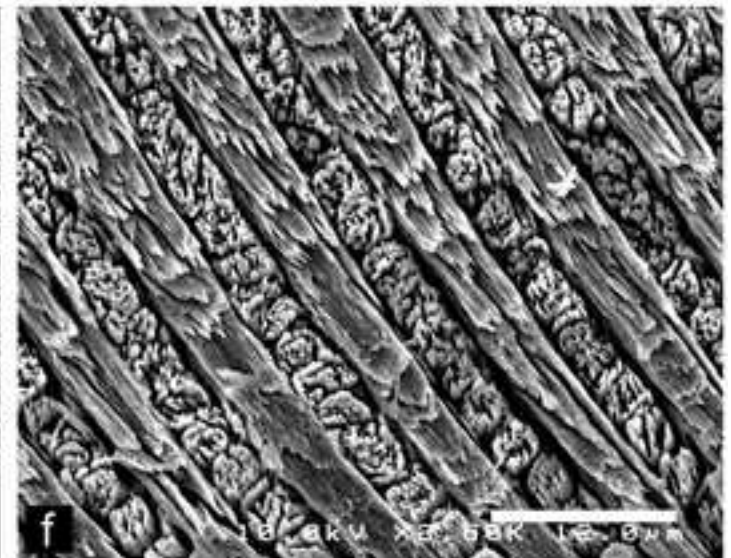
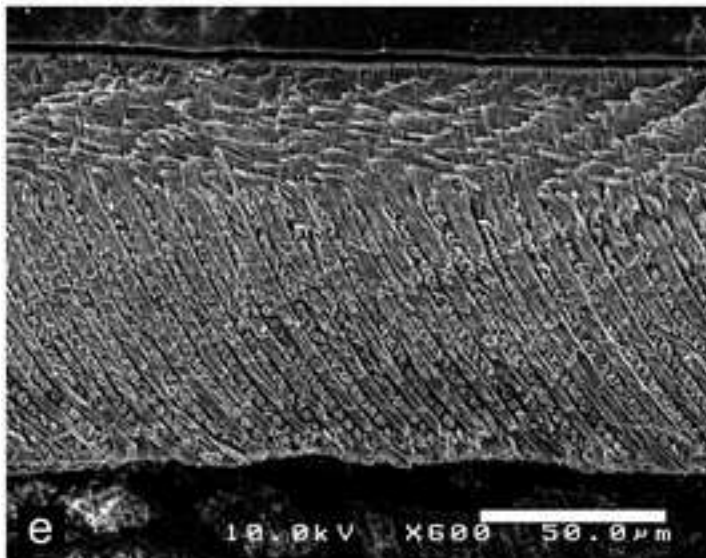
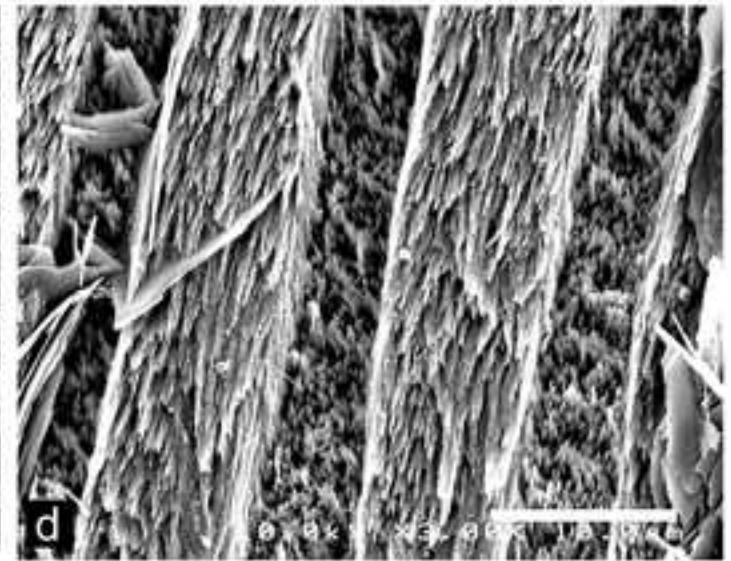
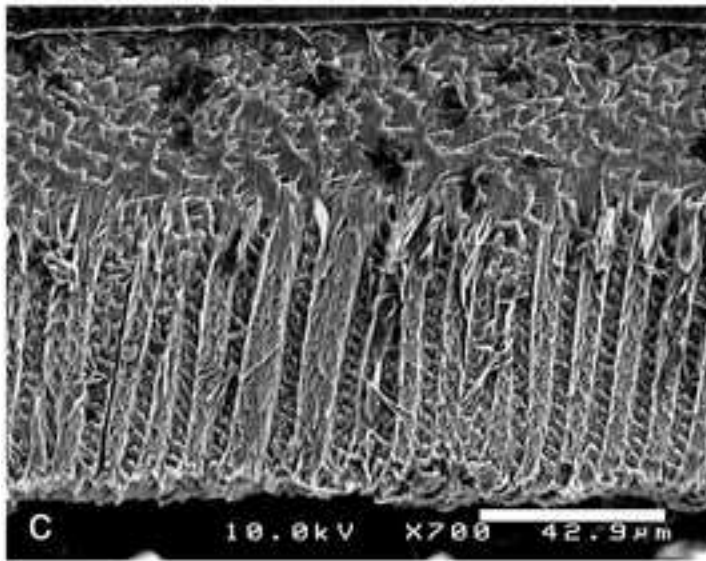
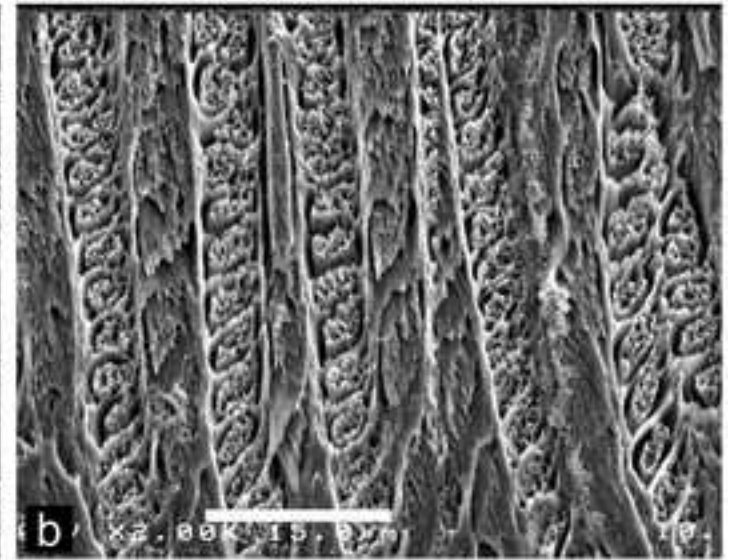
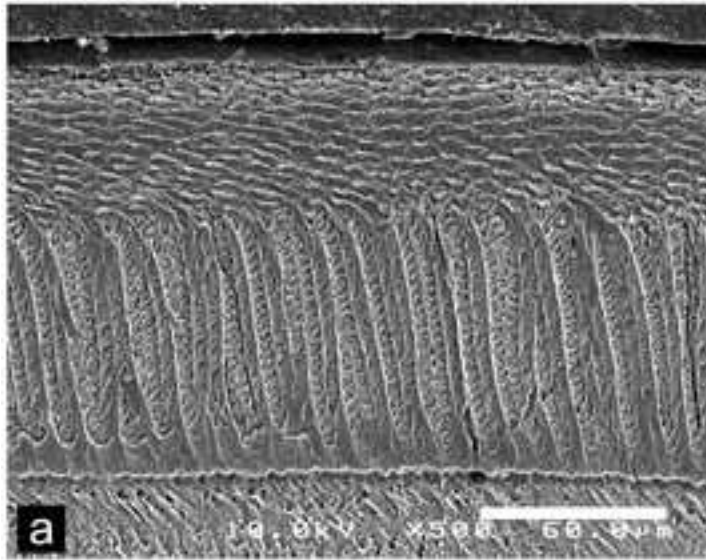
ESM 1 Incisor enamel microstructure for extant and extinct Ctenohystrica. The bibliographic references are those of the main text.

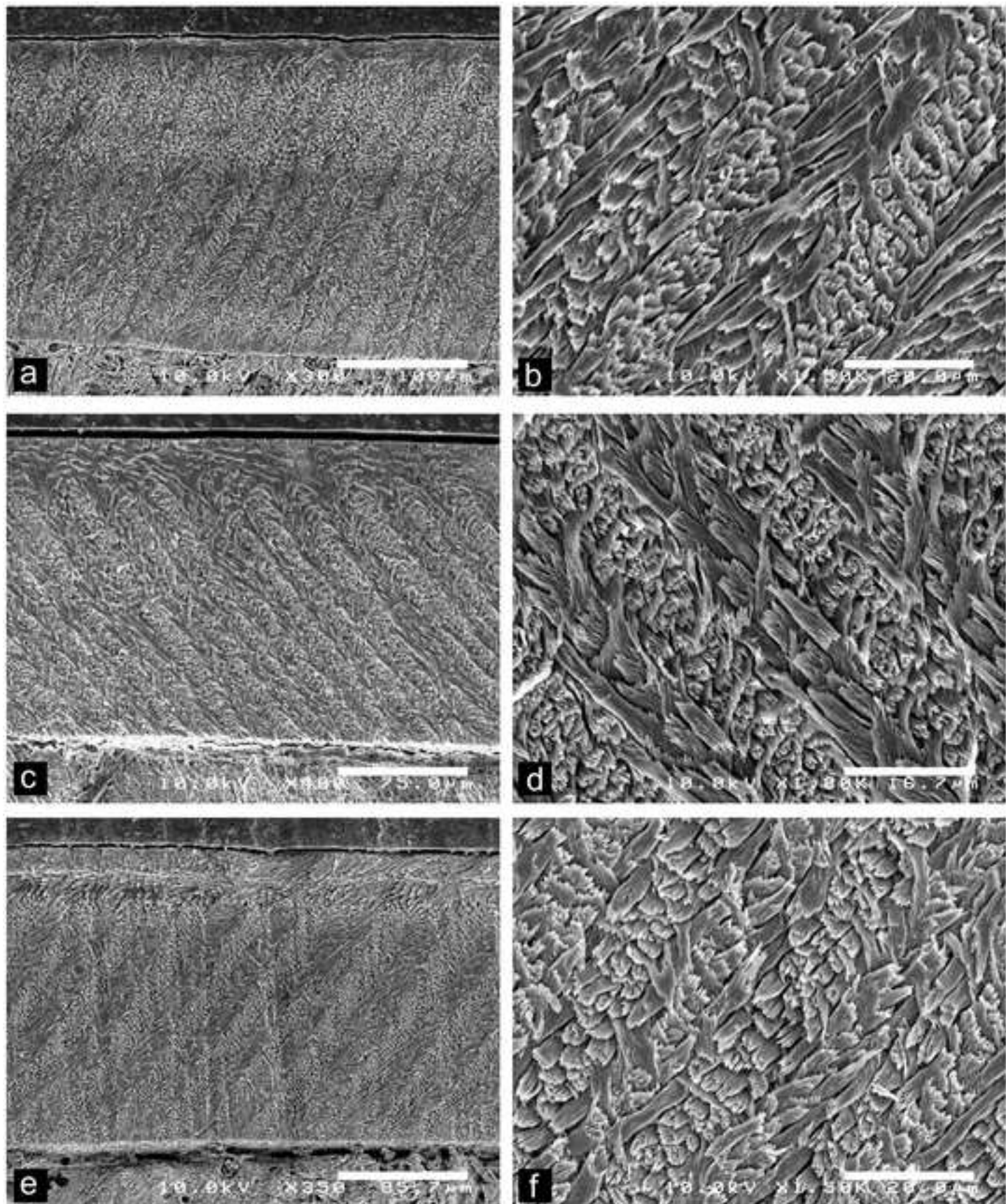
ESM 2 Incisor enamel microstructure for extant and extinct Anomaluroidea. The bibliographic references are those of the main text.

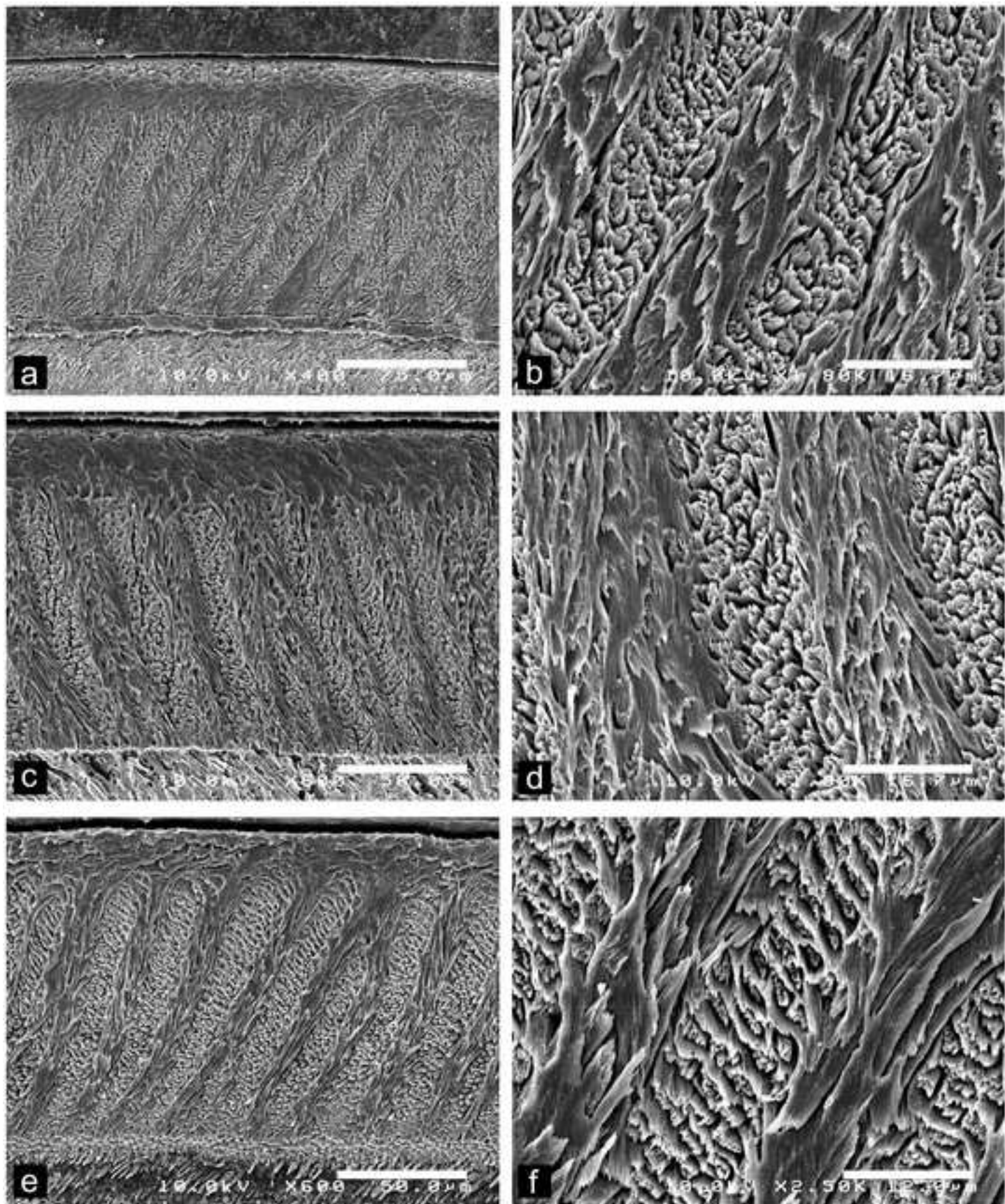


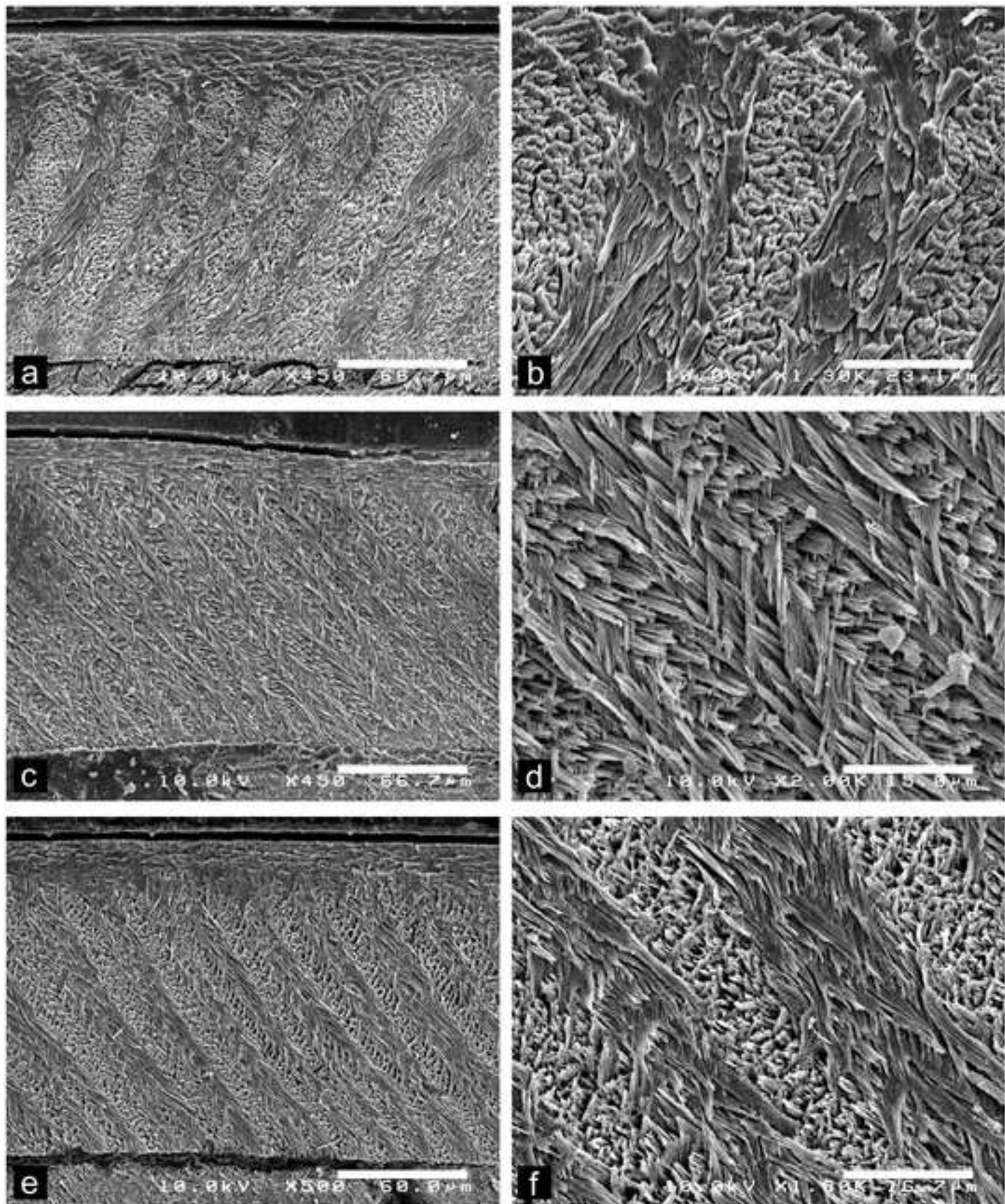


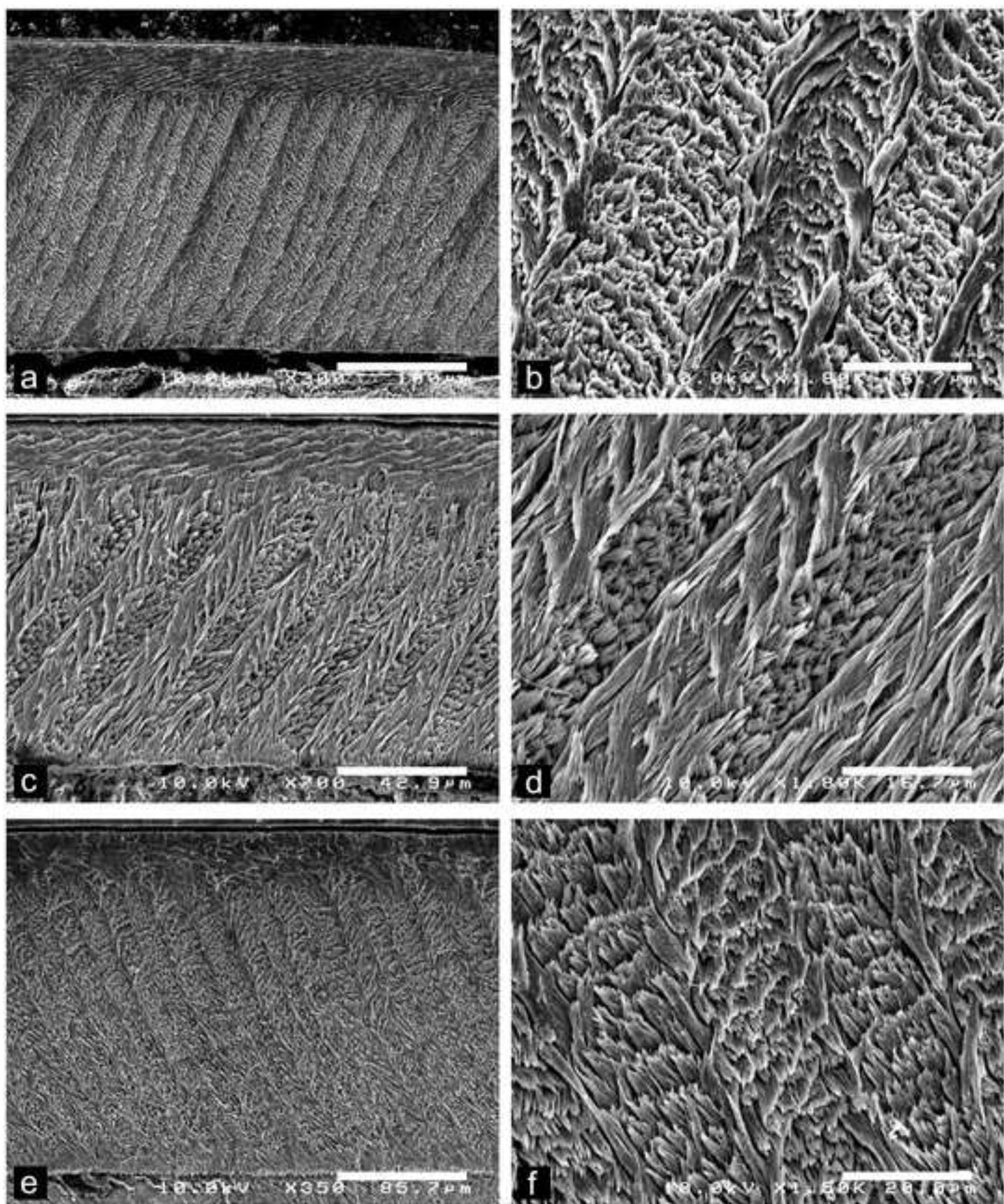






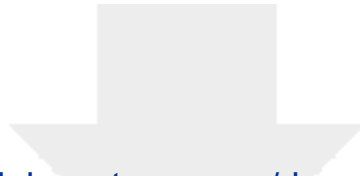






Specimen number	Incisor type	Incisor width (mm)	Enamel thickness (μm)	Percentage of PE	Inclination of prisms in PE	Inclination of HSBs	Prisms per HSB	Prisms of transitional zone	Division HSB	Prism cross section in PI	IPM configuration in PI	IPM anastomose	Angle crystallites of IPM and prism	Enamel type	Subtype	Figure
DAK-Pto-003	Upper	1.7	134	35	68-70°	5-10°	1-(2)	IPM sheet of transition	Possible, but infrequent	Oval to flattened	Sheet-like to sheath-like	Permanent	Parallel (0°)	Uniserial	1	4a-b
DAK-Pto-037	Lower	1.1	257	9	70°	30°	3-4	Well-marked	Absent	Oval	Sheet-like	Frequent and regular	Acute (50-53°)	Multiserial	2	5a-b
DAK-Pto-042	Lower	1.1	161	13	71°	33°	4	Well-marked	Absent	Flattened	Sheet-like	Frequent and regular	Acute (40-42°)	Multiserial	2	-
DAK-Pto-039	Upper	0.9	126	19	57°	18°	3-4	Absent	Possible, but infrequent	Flattened	Sheet-like	Frequent and regular	Acute (23-30°)	Multiserial	2	-
DAK-Pto-041	Lower	0.9	176	18	64°	36°	4	Well-marked	Absent	Oval	Sheet-like	Frequent and regular	Acute (43-48°)	Multiserial	2	5c-d
DAK-Pto-043	Upper	0.9	112	32	77°	55-57°	1	Absent	Absent	Round	Sheet-like to sheath-like	Not frequent and irregular	Parallel (0°)	Uniserial	1	4e-f
DAK-Pto-044	Lower	0.9	196	15	62°	34°	3-4	Present, but not well-marked	Possible, but infrequent	Oval	Sheet-like	Rare	Acute (50-55°)	Multiserial	2	5e-f
DAK-Pto-038	Upper	0.8	147	23	60°	23-31°	3-4	Absent	Absent	Round	Sheet-like	Very frequent and regular	Acute (23°)	Multiserial	(1)-2	6a-b
DAK-Pto-040	Lower	0.7	195	15	80-86°	25°	3-4	Absent	Frequent	Flattened	Sheet-like	Frequent and regular	Acute (32-42°)	Multiserial	2	-
DAK-Pto-047	Lower	0.7	145	16	90°	32-37°	3-5	Absent	Frequent	Flattened	Sheet-like	Frequent and regular	Acute (36-43°)	Multiserial	2	-
DAK-Pto-045	Lower	0.6	162	16	79°	36°	3-4	Absent	Possible, but infrequent	Round to oval	Sheet-like	Frequent and regular	Acute (34-39°)	Multiserial	2	-
DAK-Pto-046	Upper	0.6	125	23	50-55°	21°	4	Absent	Possible, but infrequent	Oval to flattened	Sheet-like to sheath-like	Very frequent and regular	Acute (23-33°)	Multiserial	2	6c-d
DAK-Pto-048	Upper	0.6	125	20	63°	25-31°	3-4	Absent	Possible, but infrequent	Flattened	Sheet-like	Very frequent and regular	Acute (28-35°)	Multiserial	2	6e-f
DAK-Pto-049	Lower	0.5	108	19	90°	33-34°	4	Well-marked	Absent	Flattened	Sheet-like	Rare	Acute (60°)	Multiserial	2	-
DAK-Pto-050	Upper	0.5	112	15	90°	23°	?4	Present, but not well-marked	Absent	Flattened	Sheet-like to sheath-like	Very frequent and regular	Acute (23°)	Multiserial	(1)-2	-
DAK-Pto-051	Upper	0.5	98	14	72°	23°	(3)-4	Absent	Absent	Flattened	Sheet-like	Frequent and regular	Acute (30-35°)	Multiserial	2	-

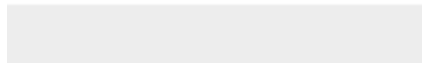
Specimen number	Incisor type	Incisor width (mm)	Enamel thickness (μm)	Percentage of PE	Inclination of prisms in PE	Inclination of HSBs	Prisms per HSB	Prisms of transitional zone	Division HSB	Prism cross section in PI	IPM configuration in PI	IPM anastomose	Angle crystallites of IPM and prism	Enamel type	Subtype	Figure
DAK-Arg-073	Upper	1.6	109	41	90°	6°	1-(2)	Absent	Absent	Flattened	Sheet-like to sheath-like	Permanent	Parallel (0°)	Uniserial	1	4c-d
DAK-Arg-072	Lower	1.3	117	31	38°	7-15°	3-4	Absent	Possible, but infrequent	?	?	?	?	Multiserial	?	-
DAK-Arg-074	Upper	1.1	240	17	70°	24°	4-5	Well-marked	Possible, but infrequent	Flattened	Sheet-like	Frequent and regular	Acute (19-35°)	Multiserial	2	8a-b
DAK-Arg-075	Lower	1.1	170	20	76°	23°	3-4	Absent	Possible, but infrequent	Oval	Sheet-like	Frequent and regular	Acute (33-40°)	Multiserial	2	7a-b
DAK-Arg-076	Upper	1.0	217	14	84°	25°	5-6	Present, but not well-marked	Possible, but infrequent	Oval	Sheet-like	Frequent and regular	Acute (23-36°)	Multiserial	2	-
DAK-Arg-085	Upper	0.9	229	17	70°	27°	5-6	Well-marked	Absent	Oval to flattened	Sheet-like	Frequent and regular	Acute (25-30°)	Multiserial	2	8e-f
DAK-Arg-086	Lower	0.9	185	14	66°	40°	4	Well-marked	Absent	Flattened	Sheet-like	Frequent and regular	Acute (39-50°)	Multiserial	2	-
DAK-Arg-078	Upper	0.8	115	17	71°	36-37°	3-4	Infrequent and scarce	Absent	Oval	Sheet-like	Very frequent and regular	Acute (34-38°)	Multiserial	2	8c-d
DAK-Arg-080	Lower	0.8	131	17	55°	30°	3-4	Well-marked	Absent	Oval	Sheet-like	Frequent and regular	Acute (40°)	Multiserial	2	-
DAK-Arg-077	Lower	0.7	161	10	90°	38-40°	3-4	Absent	Absent	?	Sheet-like	Rare	Acute (43-56°)	Multiserial	2	7c-d
DAK-Arg-079	Upper	0.7	110	20	66°	32-34°	3-5	Well-marked	Possible, but infrequent	Oval	Sheet-like	Frequent and regular	Acute (30-40°)	Multiserial	2	-
DAK-Arg-081	Lower	0.6	168	14	?	30-35°	3-4	Absent	Possible, but infrequent	Flattened	Sheet-like	Rare	Acute 40-45°)	Multiserial	2	-
DAK-Arg-082	Lower	0.5	148	16	84°	34-36°	(3)-4	Absent	Frequent	Flattened	Sheet-like	Frequent and regular	Acute (35-53°)	Multiserial	2	7e-f
DAK-Arg-083	Upper	0.5	90	20	63°	25°	3-4	Absent	?	Oval to flattened	Sheet-like	Frequent and regular	Acute (34°)	Multiserial	2	-
DAK-Arg-084	Lower	0.5	98 (but eroded)	NA	?	28°	3-4	Absent	Absent	?	Sheet-like	Frequent and regular	Acute (40-50°)	Multiserial	2	-



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