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# Design of Boron Nitride/Gelatin Electrospun Nanofibers for Bone Tissue Engineering

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## KEYWORDS

Gelatin, boron nitride, nanosheets, bionanocomposites, electrospinning, mechanical properties, tissue engineering

## **ABSTRACT**

Gelatin is a biodegradable biopolymer obtained by collagen denaturation, which shows poor mechanical properties. Hence, improving its mechanical properties is very essential towards the fabrication of efficient nontoxic material for biomedical applications. For this aim, various methods are employed using external fillers such as ceramics or bioglass. In this report, we introduce boron nitride (BN) reinforced gelatin as a new class of two dimensional biocompatible nanomaterials. The effect of the nanofiller on the mechanical behavior is analyzed. BN is efficiently exfoliated using the biopolymer gelatin as shown through Fourier transform infrared (FTIR) spectroscopy and X-ray diffraction (XRD). The exfoliated BN reinforces gelatin electrospun fibers, which results in an increase in the Young's modulus. The ESM are stable after the glutaraldehyde cross-linking and the fibrous morphology is preserved. The cross-linked gelatin/BN ESM is highly bioactive in forming bone like hydroxyapatite as shown by scanning electron microscopy. Due to their enhanced mineralization ability, the cross-linked ESM have been tested on human bone cells (HOS osteosarcoma cell line). The cell attachment, proliferation and biocompatibility results show that the ESM are nontoxic and biodegradable. The analysis of osteoblast gene expression and the measurement of alkaline phosphatase activity confirm that these materials are suitable for bone tissue engineering.

## INTRODUCTION

Gelatin is denatured collagen, extracted from acidic (Type A) or basic (Type B) treatment of bovine or porcine skin, bone or tendons. Gelatin found broad applications in food and pharmaceutical industry as thickening agent, stabilizer, protective coating material, or in emulsion preparation for ointments and cosmetics<sup>1-2</sup>. The degradable biopolymer gelatin is also used as biomaterial in cardiovascular grafts<sup>3-4</sup>, drug delivery vehicles<sup>5</sup> and scaffolds to form cartilage like extracellular matrix<sup>6</sup>. The hemostatic property of gelatin has attracted interest in surgery as a wound dressing materials and adsorbent pads<sup>7-8</sup>. Gelatin is also studied to be used as a bone grafting material<sup>9</sup>. Zha *et al.* and Lee *et al.* reported that osteoblast cells attach more prominently and proliferate on the surface of gelatin fibers<sup>10-11</sup>. Gelatin possesses poor bioactivity which is improved by the addition of hydroxyapatite or bioglasses<sup>12-14</sup>. However, addition of bioactive hydroxyapatite to the gelatin electrospun does not significantly improve the mechanical properties of the nanofibers<sup>15-17</sup>. Hence, synthesizing new class of biocompatible and bioactive material is essential for the fabrication of biomaterials which allow osteoblast proliferation for orthopedic implant applications.

Hexagonal boron nitride is an isoelectronic analogue of graphite which displays numerous interesting properties<sup>18</sup>. It will be denoted as BN in the following sections, the hexagonal polymorph susceptible to lead to BN sheets by exfoliation being the subject of this paper. The exfoliated single layers of graphene found wide applications due to their high mechanical strength<sup>19</sup>. Hence, preparing composites using exfoliated BN will be a new approach to improve mechanical properties of gelatin biopolymers<sup>20</sup>. Monolayer of BN is prepared using top down<sup>21-22</sup> and bottom up<sup>23-24</sup> approach methods. The exfoliation of BN can be achieved through polar-polar interaction with N,N dimethyl formamide<sup>25</sup>. Other frequently used methods are based on Lewis bases such as amine molecules which complex with BN and

trigger exfoliation<sup>26</sup>. BN nanosheets are also prepared by bottom up approach using chemical vapor deposition technique, and top down methods like scotch tap method and ball milling<sup>27</sup>.

The boron nitride based nanostructures has been widely studied as a nano vector to carry drug and biological molecules, in tissue engineering constructs and in boron neutron capture therapy<sup>28-30</sup>. The BN nanosheets and BN nanotubes (BNNT) are non-toxic to cells and polymer functionalized BN are highly dispersive in aqueous and organic solvents<sup>26, 31-32</sup>. Ciofani *et al.* reported that poly-L lysine functionalized BNNT effectively conjugates fluorescent markers (quantum dots) and targeting moieties (folic acid) thus exhibiting good dispersing ability and cytocompatibility<sup>33-35</sup>. Li *et al.* proved that BNNT enhances the differentiation of mesenchymal stem cells into osteoblasts<sup>36</sup>. Biomolecules like DNA and ferritin are also effectively immobilized on BNNT through non covalent interaction<sup>37</sup>. The quantity of linked biomolecules and drugs can be improved by covalent functionalization using amine grafted BNs<sup>38</sup>. *In vivo* studies of intravenously injected polymer functionalized (Glycol chitosan, Gum arabic) BNNT also evidenced its biocompatibility<sup>39-40</sup>. Hence, the literature clearly evidences that BN nanosheets and BN grafted with various functional groups are cytocompatible and can be employed for biomedical application<sup>32, 41</sup>. Conjugation of BN with bio polymers also found application other than biomedical; for instance, Biscarat *et al.* reported that BN/gelatin composites possess also excellent gas barrier property<sup>20</sup>.

The fibers produced from electrospinning exhibits porous structure and high surface area that allow the proteins adsorption. These properties are essential to create favorable microenvironment for the cell attachment. It possesses also suitable architecture for cell binding, proliferation and tissue regeneration<sup>42</sup>. Hence, synthesizing scaffolds mimicking the extracellular matrix (ECM) is very essential to enhance biological response. For example, the presence of highly organized fibers of collagen type I ECM is essential to differentiate osteo-progenitor cells into mature osteoblasts<sup>43</sup>. The electrospinning is an advantageous technique

for the fabrication of ECM-like fibrous mats. Schindler *et al.* reported that the polyamide electrospun nanofibers promote the *in vivo*-like organization of NIH3T3 fibroblasts and rat kidney cells<sup>44</sup>. Yoshimoto *et al.* reported that polycaprolactone electrospun nanofibers possess enhanced mineralization and collagen type I deposition after seeding with mesenchymal stromal cells<sup>45</sup>. *In vivo* studies of PCL electrospun nanofibers also possess multilayers of osteoblast like cells, woven bone like appearance, osteocyte like cells embedded in mineralized matrix<sup>46</sup>. Kim *et al.* reported that gelatin-hydroxyapatite nanofibers improve bone derived cellular activity<sup>17</sup>. The electrospun fibers of gelatin and polyaniline are found to be a prominent biomaterial for cardiac and neuronal tissue engineering application<sup>47</sup>. Hence this literature clearly evidences that electrospun nanofibers are excellent substitutes to restore and improve the biological activity.

The goal of this paper is to improve the mechanical properties of gelatin electrospun fibers using BN reinforcement without compromising the biocompatibility of gelatin. This is the first study which (i) reports the synthesis of BN reinforced gelatin nanofibers with bioactive properties, (ii) analyses the biological properties of exfoliated BN incorporated in gelatin nanofibers, (iii) quantifies the concentration of BN suitable to enhance the mechanical properties of gelatin ESM and (iv) analyzes cell attachment and proliferation on these ESM and (v) studies the influence of BN concentration on osteoblast gene expression.

## **EXPERIMENTAL**

### **1. Materials**

Type A Gelatin (48722-500G-F) obtained from porcine skin (gel strength 170-195 g Bloom), glacial acetic acid, ethanol and 25% glutaraldehyde (GTA), phosphate buffered saline (PBS) (P44717) tablets, and MTT ((3-(4, 5-Dimethylthiazol-2-yl)-2, 5-diphenyl tetrazolium

bromide) (Sigma -M2128), are purchased from Sigma Aldrich. BN powder (95294, 4-7  $\mu$ m) is purchased from HC STARK. The following chemicals; Dimethyl sulphoxide (DMSO) (BDH prolab 23486.297), fetal bovine serum (FBS) (Eurobio CVFSVF00-01), penicillin/Streptomycin (Gibco 15140-122), 0.05% Trypsin-EDTA (Gibco 25300-054), are used without further purification.

## **2. Preparation of spinning solution**

A solvent with high vapor pressure at room temperature is essential for electrospinning. Indeed, to avoid fiber coalescence, the solvent must evaporate from the fibers before it reaches the collector plate. Since water shows poor vapor pressure, acetic acid is used as a solvent. The gelatin (20% W/V) solution is prepared by dissolving gelatin in glacial acetic acid under magnetic stirring at 60°C for 2 hours. BN powder (0.1, 1 and 5 % (w/v)) is added to gelatin solutions and treated with an ultrasonic probe system (SONOPULS HD 3100) for one hour (1 second pulse “on” and 0.5 second “off”) with 60% amplitude to prepare a gelatin/BN dispersion. The resultant dispersion is centrifuged at 500 rpm for 30 minutes to obtain the stable dispersion of gelatin/BN in the supernatant.

## **3. Electrospinning of gelatin/BN fibers**

The stable gelatin/BN dispersion obtained from the centrifugation is loaded in a 10 mL syringe. A syringe pump (KDS 100) is used to feed the solution through the needle (0.7 mm in diameter). The solution flow rate is controlled to 0.5 mL h<sup>-1</sup> and 25 kV supplied using HPx 600 605 generator (Physical Instruments)<sup>48</sup>. A piece of aluminum foil wrapped around the rotating machine is used to collect the fibers at 400 rpm speed. The electrospinning is carried out 35°C at ambient air atmosphere. The fibers prepared using 20% gelatin are denoted as 0G

and the nanofibers prepared using various BN/gelatin weight fraction such as 0.1, 1 and 5% are referred as 0.1G, 1G, 5G respectively.

#### **4. Cross-linking of ESM**

The collected mats are cut into uniform size of 5x5 cm<sup>2</sup>, and dipped into a 1% glutaraldehyde (GTA) solution at 37°C for 24 h. The 1% GTA solution is prepared from 25 % (V/V) GTA stock solution using 96 % ethanol as a solvent. The cross-linked fibers are separated out after 24 h, washed with PBS (phosphate buffered saline) and the neutralization of the unreacted GTA is performed with 10% glycine solution. The washed samples are dried at 37°C for 24 h. The cross-linked 0G, 0.1G, 1G and 5G are denoted as 0GC, 0.1GC, 1GC and 5GC respectively.

#### **5. Characterization of uncross-linked and cross-linked ESM**

The FTIR spectrum of cross-linked ESM is recorded by NEXUS instrument, equipped with an attenuated total reflection (ATR) accessory in the frequency range of 600-4000 cm<sup>-1</sup>. The FTIR spectrum is scanned at 1 cm<sup>-1</sup> resolution and the signals are averaged from 32 scans. The XRD patterns of gelatin/ BN mats are recorded using CuK $\alpha$  radiation, 2 $\theta$  range of 3-70° and a step rate of 0.02° per second, using the PANalytica Xpert powder XRD system'.

The stability of the cross-linked ESM is tested by equilibrating the 2x2 cm<sup>2</sup> films under 0.01M phosphate buffered saline (PBS) (pH 7.4 10 mL) for 24 hours. Then, the ESM are removed from the PBS, washed with deionized water and dried at 37°C. The morphology of the uncross-linked and cross-linked ESM is observed using HITACHI S4800 scanning electron microscopy system. The mats are coated with platinum using an ion sputter coater



before SEM observations. Transmission electron microscopy (TEM) study of the samples is performed using HRTEM JEOL ARM 200F electron microscope, working in scanning mode at an accelerating voltage of 200 kV. Fibers are electrospun on a commercially available Cu grid with 300 mesh and cross-linked using 1% GTA.

Mechanical properties of the cross-linked BN reinforced ESM are characterized using the dynamic mechanical analysis system 'Metravib 50N' at controlled force mode. Fiber mats of dimensions 3 cm length, 1 cm breadth, 0.15 mm thick are clamped between the tension film clamps. Stress-strain graph is recorded at 0.05 N/s ramp rate. Linear elastic region from the stress-strain graphs is used to calculate the Young's modulus from at least 4 trials and the average results are reported.

The cross-linked ESM is analyzed using a differential scanning calorimeter DSC (TA Instruments 2920), equipped with a RCS90 cooling system. The samples are accurately weighed ( $\approx 3$  mg) in an aluminum TA pan and sealed. An empty sealed pan is used as a reference. The samples are cooled at  $-40$  °C and then heated up to  $250$  °C with a heating rate of  $100$  °C  $\text{min}^{-1}$  following the procedure described by Biscarat *et al.*<sup>20</sup>. The percentage renaturation level of the gelatin triple helix is calculated from the ratio between the denaturation enthalpies of both cross-linked gelatin ESM and pure porcine skin collagen ( $\Delta H_{\text{denatured collagen}} = 47.8$  J/g)<sup>49-50</sup>. The thermogravimetric analysis (TGA) of the samples is performed using TA instruments TGA G500 under oxygen flow of  $60$  mL  $\text{min}^{-1}$ . The samples are heated up to  $700$  °C at a heating rate of  $10$  °C  $\text{min}^{-1}$  and equilibrated for 60 minutes at  $700$  °C for complete degradation of gelatin.

Biom mineralization is carried out using the 1.5x simulated body fluid (SBF). The latter solution is prepared using the standard protocol described by Kokubo *et al.*<sup>51</sup>. The ESM is cut into 2x2

cm<sup>2</sup> and equilibrated to SBF solution. The new SBF solution around the vicinity of the samples is introduced periodically after 24 hours.

Biodegradation of the cross-linked ESM is recorded using collagenase I enzyme at various concentrations (0.02, 0.5, 5 or 50 units.mL<sup>-1</sup>). A control experiment is performed to understand the degradation of the ESM in PBS. The collagenase solutions are prepared from a 0.2 M PBS at pH 7.4. Accurately weighed samples are placed in a vial containing 1 mL of collagenase solution. The collagenase solution is replaced with fresh collagenase solution at 3 days interval. The samples are recovered at different times, washed, dried at 50 °C and weighted. The percentage of ESM degradation is calculated by comparing the initial weight before degradation and the final weight of the sample after degradation.

## **6. Cell culture**

Human HOS osteosarcoma cells are cultured using DMEM (Dulbecco's modified eagle medium) (Gibco 31331-028) medium with 10 % (V/V) fetal bovine serum (FBS) (Eurobio CVFSVF00-01) and 1% (V/V) penicillin/streptomycin (Gibco 15140-122) at 37°C in 5% CO<sub>2</sub>. The cells are maintained in 10 cm diameter Petri dishes and passages with 0.05% Trypsin-EDTA (Gibco 25300-054).

## **7. Cell viability assay and fluorescent staining**

The ESM (6 mm diameter circle) is exposed to UV light ( $\lambda=254$  nm for 1 hour) for sterilization. It is washed with PBS and the ESMs are allowed in DMEM for 30 mins. The HOS cells are plated at  $2 \times 10^3$  cells/well . It is allowed to attach on the plates for 24 hours and the samples are positioned on top of the cells after 24 hours. The cell viability at different

periods (1, 4 and 8 days) is investigated using MTT ((3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) assay. Shortly, 100  $\mu\text{L}$  of the 0.05 mg  $\text{mL}^{-1}$  of MTT solution is added onto the cells and incubated for 4h. The purple colored formazan crystals are dissolved using 100 $\mu\text{L}$  of DMSO (BDH Prolab 23486.297) and the absorbance is recorded at 560 nm using multiskan plate reader (thermos, USA).

Proliferation of cells on the ESM is recorded by epifluorescence imaging microscope system (DM6000 Leica). The concentrated cells ( $2 \times 10^3$  cells/sample) are allowed to attach for 30 minutes and then complete DMEM is supplied. The proliferation of the cells on the ESM at various periods is monitored using fluorescent imaging. For imaging, the cells attached on the samples are fixed by 4% formaldehyde for 20 minutes and stained with Hoechst 33342. Cells attached on the ESM are recorded using SEM. Cells cultured for four days on the ESM are fixed using 2.5% of GTA for 2 hours. Samples are washed and dehydrated using serial concentration of ethanol (30%, 50%, 70%, 90% and absolute ethanol). The samples are further treated with 1 mL of hexamethyldisilazane to dry the sample. Samples are further coated with platinum using an ion sputter coater and the morphology of the cells attached on the ESM is recorded using SEM.

## **8. Real-time qPCR**

RNAs were extracted from cells attached and cultured on the samples with Quick RNA<sup>TM</sup> Miniprep kit (Zymo Research) according to the manufacturer's instructions. The extracted RNA (1 $\mu\text{g}$ ) and qScript cDNA master mix (Quanta Bioscience) is used to synthesis cDNA. The quantitative mRNA expression is determined by real time PCR (qPCR), SYBR Green Mix (VWR) with the LightCycler 480 (Roche Applied Science). The final reaction volume is 10  $\mu\text{L}$ , in which 0.5  $\mu\text{L}$  of each primer (25  $\mu\text{M}$ ), 2.5  $\mu\text{L}$  of  $\text{H}_2\text{O}$ , 5  $\mu\text{L}$  of the SYBR green master

mix, and 2  $\mu\text{L}$  of the template at a 1:5 dilution. The mixtures are preincubated at 95°C, the experiment consists of 40 cycles in which each cycle consists of 15 seconds each at 95°C, 45 seconds at 60°C, and 30 seconds at 72°C, followed by melting curves analysis to eliminate non-specific amplification products. Relative mRNAs expression levels for the interested gene are quantified through normalization with TBP housekeeping gene. The primer sequences are listed in the Table 1.

**Table 1.** The primers sequence of osteoblast genes

<b>Primer</b>	<b>Forward sequence</b>	<b>Reverse sequence</b>
<b>Osteopontin (OSPN)</b>	GTGCCATACCAGTTAAACAG	CTTACTTGGAAGGGTCTGTG
<b>hRunX2</b>	CCTAAATCACTGAGGCGGTC	CAGTAGATGGACCTCGGGAA
<b>hSPARC</b>	GAAAGAAGATCCAGGCCCTC	CTTCAGACTGCCCGGAGA

### **9. Alkaline phosphatase (ALP) activity**

HOS cells ( $8 \times 10^3$  cells/well) are cultured on the cross-linked ESM for 8 days. The cells are washed with PBS and lysed using 50  $\mu\text{L}$  of Tris HCl (0.1M, pH 8) with 0.1% (v/v) triton X 100. The ALP activity is analyzed by quantifying the concentration of p-nitrophenol from p-nitrophenyl phosphate (PNPP). The cell lysate (10  $\mu\text{L}$ ) is incubated with PNPP (50  $\mu\text{L}$ ) (Sigma N1891) at 37°C for 60 min and the amount of p-nitrophenol is estimated by recording the absorbance at 405 nm. The percentage ALP activity of the cells cultured along the ESM is calculated by comparing the ALP activity of cells cultured on tissue culture plates.

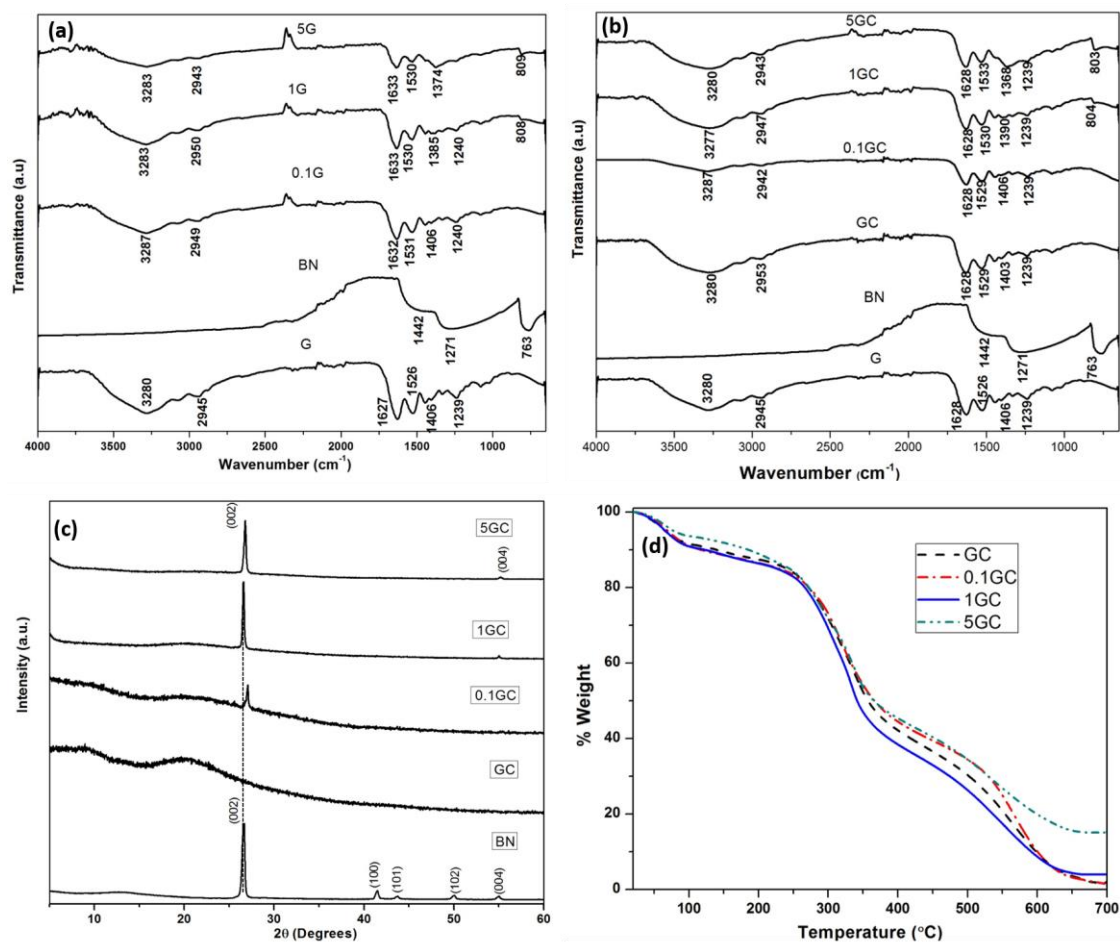
## RESULTS AND DISCUSSION

### 1. Chemical and structural characterization of ESM

Various ESMs (elaborated with 0.1, 1 and 5 % BN weight fraction and referred as uncross-linked 0.1G, 1G and 5G respectively) is generated as described in the Materials and Method section. The synthesized ESM are highly soluble in PBS. Hence, to improve the stability of the electrospun fibers, the fibers are cross-linked with 1% GTA and referred as GC, 0.1GC, 1GC and 5GC respectively.

In order to understand the gelatin chains organization and the interactions of BN with gelatin during the ESM fabrication, FTIR spectra are first recorded. The FTIR spectra of uncross-linked and cross-linked electrospun fibers are depicted in the Figure 1 (a-b) respectively. The peak at about 3280-3287  $\text{cm}^{-1}$  is due to the presence of amide A. The characteristic peaks of amide I, II and III are observed at 1627-1632, 1527-1534 and 1236-1241  $\text{cm}^{-1}$  respectively in synthesized and cross-linked ESM. The low intensity peak observed at 2945  $\text{cm}^{-1}$  shows asymmetric  $\text{CH}_2$  stretching modes of gelatin, whereas the asymmetric and symmetric stretching modes of carboxylates group of gelatin are observed at 1332 and 1406  $\text{cm}^{-1}$  respectively<sup>52</sup>. The shift in the symmetric stretching band of carboxylates groups towards the lower wavenumber from 1406 to 1385  $\text{cm}^{-1}$  in 1G, 5G and the broadening of the band observed at 1406, 1385, 1374  $\text{cm}^{-1}$  in 0.1G, 1G, 5G respectively, suggest that carboxylates have strong interaction with the boron nitride nanosheets and such an interaction facilitates the exfoliation of BN. The presence of amide I and II peaks provides evidences for the presence of gelatin chains. The two major peaks of BN observed at 1271 and 763  $\text{cm}^{-1}$  as

depicted in Figure 1 (a) are assigned to the B-N stretching and out of plane bending of B-N in hexagonal ring respectively<sup>53</sup>.



**Figure 1.** FTIR spectrum of (a) uncross-linked, (b) cross-linked ESMs, (c) XRD graphs and (d) TGA graphs of cross-linked ESM.

The effect of gelatin on the exfoliation of BN and the crystallinity of the exfoliated boron nitride are analyzed using the X-ray diffraction (XRD) patterns of the cross-linked ESM. The XRD patterns are depicted in the Figure 1 (c). The 0.1 GC, 1GC and 5GC nanofibers shows 2 peaks at  $2\theta = 26.6^\circ$  and  $55^\circ$  corresponds to (002) and (004) planes of BN respectively. The peaks observed at  $2\theta = 41.5^\circ$ ,  $43.7^\circ$  and  $50^\circ$  correspond to (100), (101) and (102) planes of BN respectively disappear, evidencing the efficient exfoliation of BN in 0.1GC, 1GC and

5GC and the reinforcement of the gelatin <sup>20</sup>. The broadband centered to  $2\Theta=20^\circ$  is also due to the presence of random coiled conformation of gelatin.

## **2. Thermal analysis**

The effects of the preparation route and BN addition on the renaturation level of gelatin, *i.e.* the triple-helix content, are evaluated through differential scanning calorimeter (DSC) analyses. The helix-coil transition is characterized by the peak temperature of a first endotherm ( $T_m$ ) and by the melting enthalpy ( $\Delta H_m$ ), as the area over the same peak. At temperatures above  $130^\circ\text{C}$  a second endotherm is observed and corresponds to the degradation of the protein. This second peak temperature is called degradation temperature ( $T_d$ ).

It has been previously demonstrated <sup>54</sup> that the triple helix level greatly affects mechanical properties of gelatin films. Higher values of Young's modulus are obtained when the triple-helix content increases. Besides, it is believed that the renaturation of random coil gelatin strands to the collagen-like triple helical structure can only exist if water molecules are included in the crystals formed. The structural water, directly bound to the gelatin strands by hydrogen bonds is responsible of the stabilization of the triple helical gelatin structure. In this study, gelatin is first dissolved in anhydrous solvent, glacial acetic acid, before BN addition, sonication, centrifugation and electrospinning. Thus, the renaturation of gelatin is not favored in those conditions. The triple helix percentage of the gelatin is calculated from the melting enthalpy. The thermal properties of type A gelatin powder (G powder), ESM fabricated using unsonicated gelatin solution (UG), uncross-linked and cross-linked ESM and the corresponding calculated triple helix levels are listed in Table 2.

**Table 2.** Denaturation ( $T_m$ ) and degradation ( $T_d$ ) temperatures, and triple helix percentage of gelatin extracted from DSC data

<b>Sample</b>	<b><math>T_m</math>(°C)</b>	<b><math>T_d</math> (°C)</b>	<b><math>\Delta H_m</math> (J/g)</b>	<b>Triple helix level %</b>
G powder	$80 \pm 1$	$160 \pm 1$	$4.0 \pm 0.4$	$8.4 \pm 0.8$
Un sonicated G	$87 \pm 3$	$161 \pm 5$	$4.3 \pm 0.1$	$8.9 \pm 0.2$
G	$86 \pm 1$	$143 \pm 3$	$0.6 \pm 0.05$	$1.3 \pm 0.1$
0.1G	$93 \pm 1$	$156 \pm 3$	$1.0 \pm 0.3$	$2.0 \pm 0.6$
1G	$81 \pm 1$	$144 \pm 12$	$2.0 \pm 0.1$	$4.2 \pm 0.2$
5G	$92 \pm 1$	$140 \pm 7$	$2.1 \pm 0.2$	$4.5 \pm 0.3$
GC	$87 \pm 1$	$161 \pm 3$	$1.6 \pm 1.2$	$3.3 \pm 2.5$
0.1GC	$94 \pm 3$	$146 \pm 4$	$2.4 \pm 0.1$	$5.0 \pm 0.2$
1GC	$75 \pm 2$	$148 \pm 8$	$2.7 \pm 0.1$	$5.6 \pm 0.2$
5GC	$87 \pm 9$	$134 \pm 6$	$3.0 \pm 0.1$	$6.3 \pm 0.2$

Denaturation and degradation temperature ranges are from 80 to 94°C, and from 134 to 161°C respectively. Kruskal-Wallis tests are performed using the computer program R with a significance level set at  $p \leq 0.05$  for BN reinforced and gelatin samples. They show that neither the formulation nor the process has a significant influence on both degradation and denaturation temperature. However, increase in denaturation enthalpy with rising BN content compare with GC ( $p < 0.05$ ) demonstrates the influence of the preparation route and the formulation on the obtained values.



Firstly, very low triple helix levels could be calculated, meaning the presence of insignificant proportion of the helical conformation of gelatin in the anhydrous solvent involved in this study. Only a small part of gelatin chains are organized in helical conformation. Indeed, 9% triple helix level is determined for ESM fabricated using unsonicated gelatin solution (UG). This value is very close to the results obtained on gelatin powder, and has to be related with the residual water content still present with the gelatin and acting as structural water. Indeed no preliminary drying step is conducted on gelatin raw material. The dry matter determination on type A gelatin reveals that actually there is 8 % residual water in the powder. This can explain the existence of a part of gelatin chains in triple helix conformation. Secondly, results display that sonication drastically decreases triple helix level compare to ESM fabricated without sonicated gelatin solution. This observation is consistent and has to be related to the fragmentation of gelatin chains in ultrasonic bath<sup>18</sup>. The ultrasonic waves act on the solvent and generate the rapidly growing micro bubbles. The disruption of these bubbles generates strong shear forces which damage inter and intra molecular forces of polymers and are responsible for the low degradation temperature of sonicated gelatin ( $143 \pm 3$  °C) compare to non-sonicated gelatin ( $161 \pm 5$  °C) (Table 2)<sup>55</sup>. Finally, increasing the BN concentration enhances the triple helix level of collagen (1.3 to 4.5%). This is due to the intercalation of gelatin between BN nanosheets. The nanosheets restrict the gelatin and the movement segment is controlled<sup>20</sup>. The cross-linking step increases  $\Delta H_m$ . This may be attributed to ESM washes with PBS, allowing the absorption of some water molecules inside the gelatin matrix. To conclude, the as-prepared ESMs display a very low level of gelatin triple helix, which could also affect their mechanical properties and degradation temperature.

The thermal stability and the influence of the concentration of BN on the cross-linked ESM are quantified using the TGA analyses (Figure 1d)). The weight loss (9-10 %) observed below 108°C is due to the solvents (residual acetic acid and structural water of gelatin). The second

major weight loss observed between 200-400°C shows the degradation of gelatin molecules. The third major weight loss observed from around 520°C is due to the thermochemical decomposition of remaining organic content. The 1GC samples exhibits strong interaction with the BN, which results to the high thermal stability of the ESM, whereas, high concentration of non-exfoliated BN shows poor interaction. The gelatin undergoes complete decomposition below 700 °C, the remaining residues are due to BN, which are quantified gravimetrically and listed in the Table 3. It is evidenced that the BN added at higher concentrations (5 and 20%) is not completely dispersed in the gelatin. During the centrifugation step, the remaining unexfoliated BN is separated out.

**Table 3.** Composition of gelatin, solvent and boron nitride present in the cross-linked ESM extracted from the TGA graphs

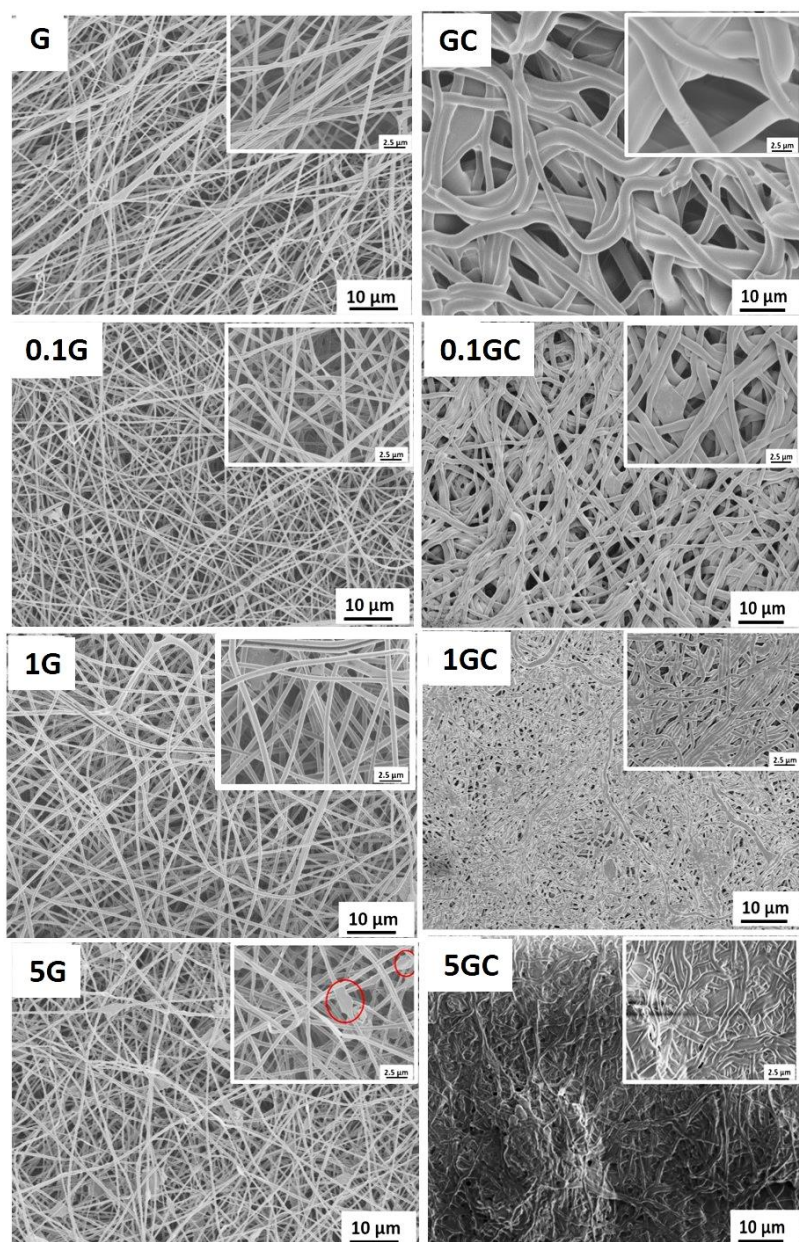
<b>Sample</b>	<b>Gelatin (%)</b>	<b>Solvent (*) (%)</b>	<b>Residues after combustion (%)</b>	<b>% BN added in solution</b>
<b>GC</b>	89 ± 1	9 ± 1	2 ± 1	0
<b>0.1GC</b>	89 ± 1	10 ± 1	1 ± 0.1	0.5 ± 0.1
<b>1GC</b>	86 ± 1	10 ± 1	4 ± 1	5 ± 1
<b>5GC</b>	79 ± 1	7 ± 1	14 ± 1	20 ± 1

(\*) acetic acid and structural water

### 3. Morphological studies of the ESM

Scanning electron microscopy is used to analyze the morphology of uncross-linked fibers, the stability of fibrous network on PBS, the effect of cross-linking on the morphology of uncross-

linked fibers and the size of the electrospun fibers (Figure 2 insert images are higher magnification images of the electrospun fibers). The diameter of the fibers is listed in Table 4.

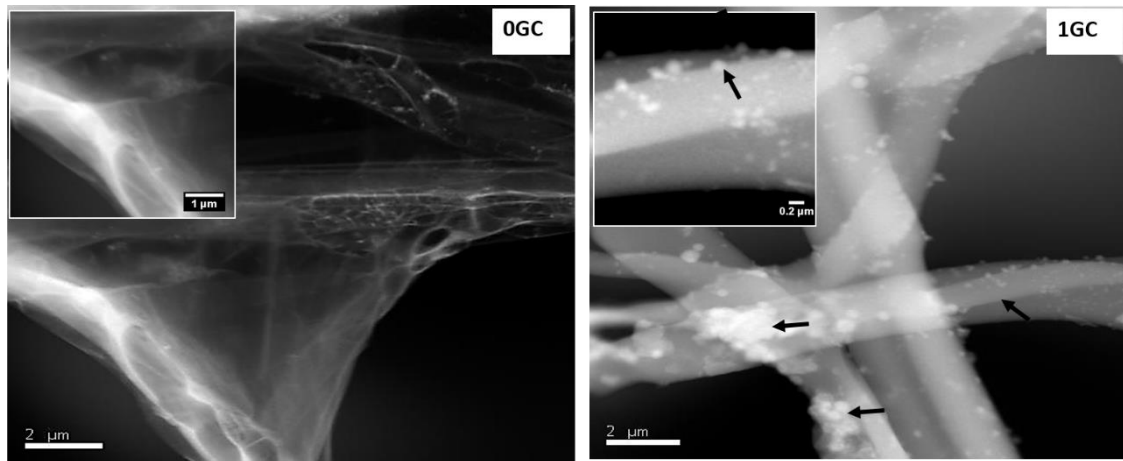


**Figure 2.** Scanning electron micrograph of uncross-linked ESMs (left panel) and cross-linked (right panel) ESM.

**Table 4.** Size of uncross-linked and cross-linked electrospun fibers

<b>Sample</b>	<b>Size of the fibers before cross-linking (nm)</b>	<b>Sample</b>	<b>Size of the fibers after cross-linking (nm)</b>
<b>G</b>	1173 ± 160	<b>GC</b>	2300 ± 111
<b>0.1G</b>	410 ± 49	<b>0.1GC</b>	822 ± 75
<b>1G</b>	483 ± 50	<b>1GC</b>	447 ± 56
<b>5G</b>	346 ± 37	<b>5GC</b>	581 ± 60

The SEM micrograph shows the fibrous morphology of the ESM. The size of the uncross-linked gelatin fibers decreases from 1173 nm ( $\pm 160$ ) to 346 nm ( $\pm 37$ ) when the concentration of BN increases as depicted in Figure 2. The absence of visible grain BN sheets in 0.1G and 1G as shown in the Figure 2 clearly evidences that BN are reinforced and well exfoliated in the gelatin matrix. TEM image is recorded for cross-linked ESM to understand the BN reinforcement. It clearly proved that the BN nano sheets are distributed in gelatin electrospun fibrous matrix as shown in Figure 2 (1GC). TEM images depict the smooth fibrous surface whereas BN plates are oriented along electrospun fibers in 1GC samples. Higher magnification images of electrospun fibers are shown in Figure 3 (inset images). BN nanosheets are easily distinguished inside the Gelatin matrix proving the inclusion of BN inside the gelatin electrospun fibers.

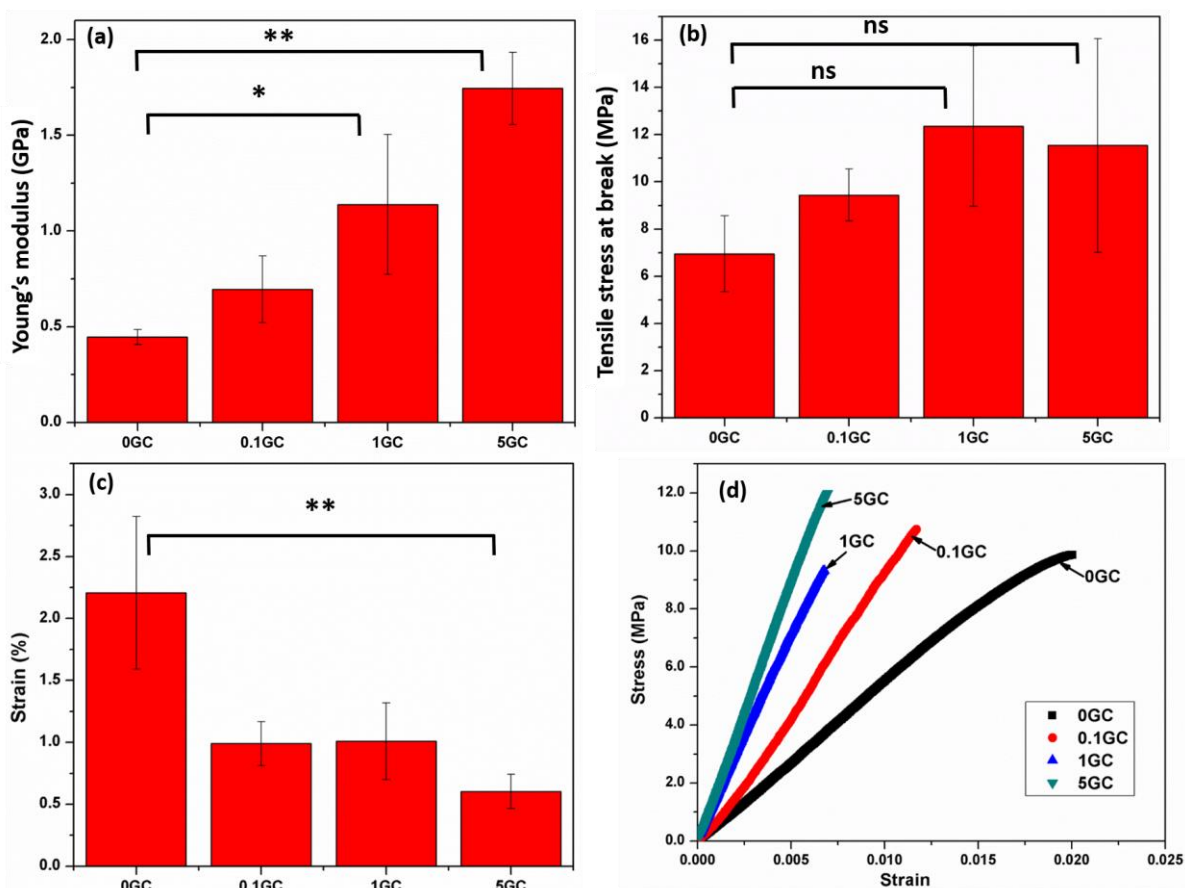


**Figure 3.** TEM images of the cross-linked ESM (The black arrow indicates the distribution of BN). The insets are higher magnification images.

#### 4. Mechanical properties of the electrospun nanofibers

The composite used for the biomedical implant application should withstand high loads. Hence understanding the mechanical properties of the composite is essential for implants application. Mechanical properties of gelatin-based ESM are shown in Figure 4. Siimon *et al.*, reported that the fibers are ordered straight after elastic region, oriented to particular direction and do not slide on each other on applied force. In case of cross-linked fibers, cross-linking bonds are established between gelatin molecules in each nanofiber but also at the intersection of different fibers as evidenced in Figure 2<sup>56</sup>. Hence the cross-linked fibers cannot orient to the applied force, structural changes greatly influence at plastic region of cross linked ESMs. So the study is focused on elastic region as well as on understanding the effect of BN addition on elastic modulus. Young's modulus obtained for the 0GC (470 MPa) ESM is in good agreement with the literature data on cross-linked gelatin ESM.<sup>57</sup> The Young's modulus of the electrospun nanofibers increases with the BN concentration from 0.1GC to 5GC. The Young's modulus of 5GC increases 3 fold in comparison to 0GC. Indeed, no significant

change is observed for tensile stress at break with BN inclusion. The improved stiffness of the BN/Gelatin ESM in comparison to cross-linked gelatin ESM evidenced the reinforcement of fibers with BN. BN reinforcement with gelatin improves the stiffness of the electrospun fibers in comparison to gelatin electrospun fibers fabricated with synthetic, biopolymers or inorganic material like hydroxyapatite<sup>17, 58-60</sup>. Hence, the BN reinforced ESM are promising fillers for the improvement of the mechanical properties of the biopolymer gelatin. In addition, the incorporation of BN can counterbalance the effects of low level of gelatin triple helix on mechanical properties caused by the use of a non-aqueous solvent.

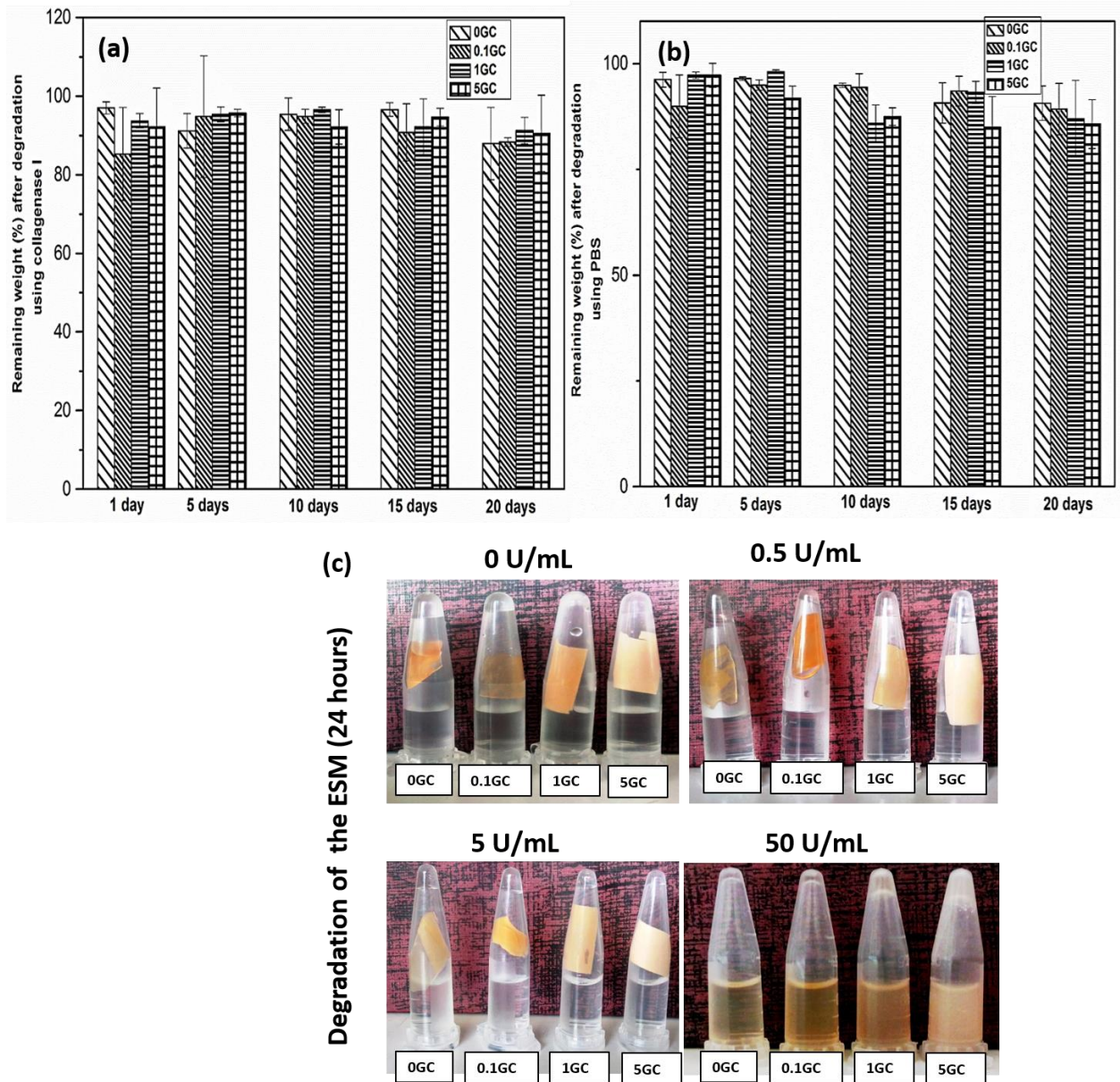


**Figure 4.** Mechanical property of the ESM with (a) Young's modulus, (b) tensile stress at break, (c) strain at break and (d) stress vs strain graph of the cross-linked ESM (ns = no significance,  $p < 0.05 = *$ ,  $p < 0.005 = **$ )

## 5. Biodegradation of the ESM

Designing biodegradable scaffolds is essential for tissue engineering application. Degradation of the polymer scaffolds is necessary to avoid additional surgery for removing the implant matrix. Hence, the stability and the degradation properties of the ESM in PBS are analyzed using collagenase I enzyme and shown in Figure 5. First, the stability of the ESM is analyzed for 20 days in PBS without collagenase (Figure 5a). The results show that the mats are highly stable in PBS solution. The stability of the different ESM is also observed when low concentration of collagenase I ( $0.02 \text{ U/mL}^{-1}$ ) is used for up to 20 days (Figure 5b). These data indicate that the ESMs are suitable scaffolds for cell attachment. However, sustained degradation of the ESM is required for prolonged cell growth on the ESM. In order to demonstrate that the different ESMs are degradable *in vitro*, increasing concentrations of enzyme have been used on a 24hr period. Full degradation of the different ESMs is obtained at a concentration of 50U/ml of collagenase (Figure 5c). Altogether, these results show that the synthesized ESMs are stable in aqueous solution (required to facilitate cell attachment) and enzymatically degradable which are an indispensable property for tissue engineering application.





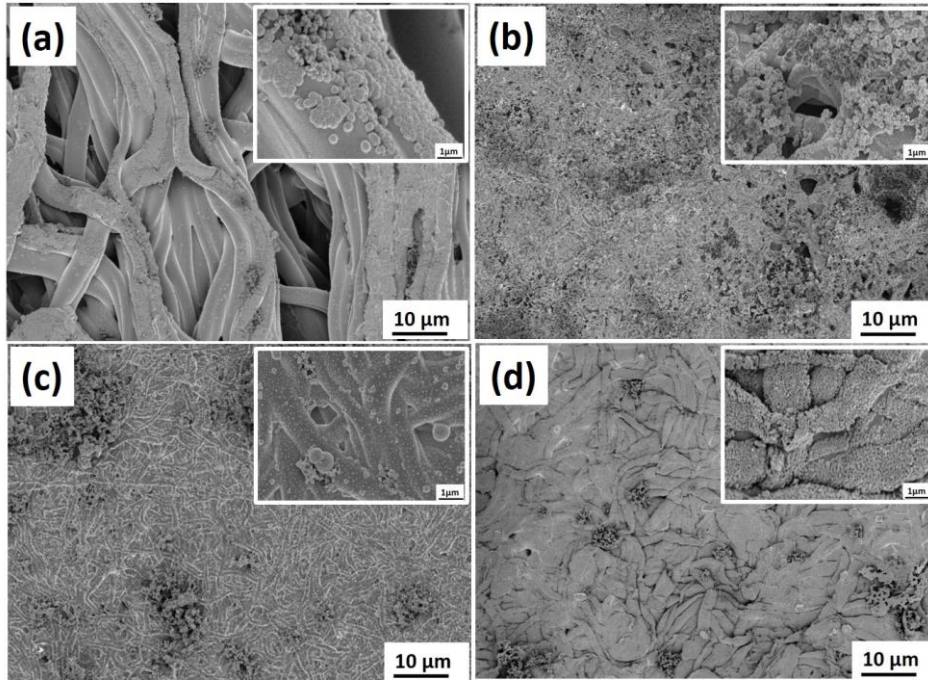
**Figure 5.** Biodegradation of ESM in (a) PBS, (b) time-course analysis with low concentration of collagenase and (c) dose-response of collagenase.

## 6. Biom mineralization of the ESM

Composites showing bioactive properties are essential for bone implant application. The bioactive surfaces develop a hydroxyapatite layer at the surface of the implant materials. Thus the hydroxyapatite layer acts as an intermediate layer between the bone tissue and the

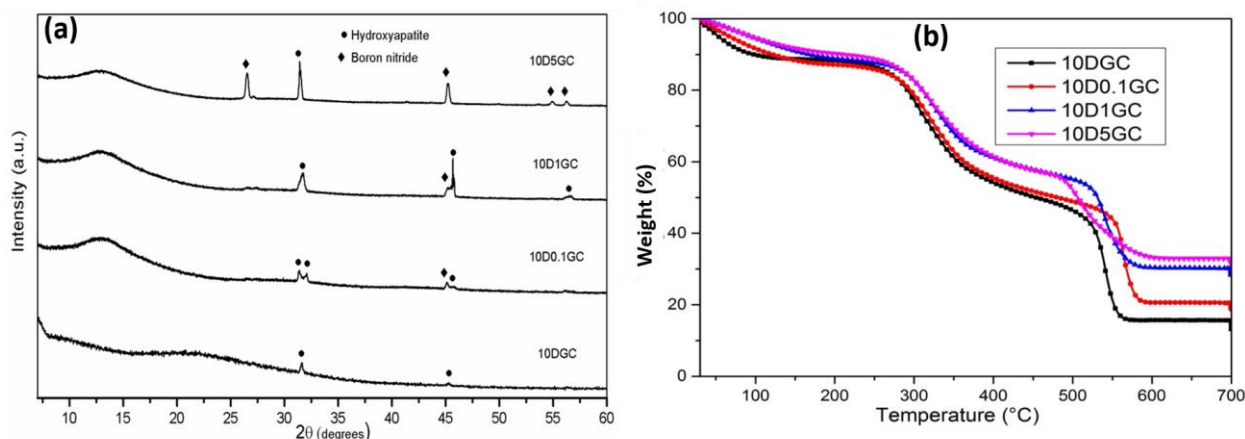


implant<sup>61</sup>. The intermediate hydroxyapatite layer controls the fibrous capsule formation which induces a negative effect during osteo-integration. In order to study the formation of bioactive layers on the cross-linked ESMs GC, 0.1GC, 1GC and 5GC, the mats are mineralized for 10 days. After the mineralization, mats are denoted as 10DGC, 10D0.1GC, 10D1GC, 10D5GC respectively. The SEM images of the mineralized ESM are shown in the Figure 6. The 10DGC is found to have random mineralization of hydroxyapatite on gelatin fibers (Figure 6a); the smooth surfaces are non-mineralized. The presence of hydroxyapatite is confirmed by XRD analysis and shown in the Figure 7a. The 10D0.1GC and 10D1GC possess the uniform nucleation of hydroxyapatite on the fibers. The hydroxyapatite minerals completely cover the surface of the fibers 10D5GC as evidenced in the insert image of Figure 6d. It is clearly evidenced that increasing the concentration of BN increases the bioactivity of the gelatin mats significantly. The vacant “p” orbital of the boron atom present in boron nitride is responsible for the Lewis acid nature. Garcia *et al.* reported that hydroxyapatite is found to possess Lewis base nature<sup>62</sup>. The hydroxyapatite nanocrystals formed from the SBF interact with the BN through Lewis acid-base interaction. Hydroxyapatite mineralization increases along with BN enhancement substantiate the Lewis acid-base interaction. Hence this results shows that BN is highly bioactive and suitable for mineralization of hydroxyapatite.



**Figure 6.** Scanning electron micrograph of cross-linked, SBF equilibrated ESM (a) 10DGC, (b) 10D0.1GC, (c) 10D1GC and (d) 10D5GC

The XRD patterns of the 10 days mineralized GC, 0.1GC, 1GC and 5GC are recorded and shown in Figure 7a. The XRD patterns are in good agreement with the standard JCPDS (09-0432) data for hydroxyapatite. The formation of hydroxyapatite is induced by the carboxylates groups of gelatin through the interaction of calcium ions in the SBF. The major plane of BN observed at unmineralized samples (Figure 1c, 0.1GC and 1GC) disappeared on the 10 days mineralized samples (10D0.1GC and 10D1GC). It shows that mineralized hydroxyapatite (Figure 6 (b-c) crystals affect the X-ray penetration, and the BN diffraction disappeared. The appearance of BN peaks in 10D5GC is due to the flanked un-exfoliated sheets of BN in gelatin fibers as shown in Figure 2 (5G).



**Figure 7.** (a) XRD patterns and (b) TGA graphs of cross-linked gelatin and gelatin/BN electrospun fibers with different BN concentrations after mineralizing for 10 days.

Hydroxyapatite is the major inorganic content found in hard tissues like bone and teeth. Hence observing mineralization ability of the ESM and quantifying the concentration of inorganic content is essential. The hydroxyapatite mineralized in GC, 0.1GC, 1GC and 5GC are quantified using TGA analysis and listed in the Table 5 and the graphs are shown in the Figure 7b. The major weight losses observed from 100 °C to 600 °C in mineralized ESM are due to the degradation of gelatin. The presence of 15 % insoluble residues in GC depicts the mineralized hydroxyapatite. It is found that mineralization of hydroxyapatite increases with the BN nanosheets concentration. The uncertainty in the concentration of hydroxyapatite at 5GC is due to the release of unexfoliated BN in SBF solution, flanked between the fibers as depicted in Figure 2 (5G).

**Table 5.** Concentration of hydroxyapatite mineralized in ESM

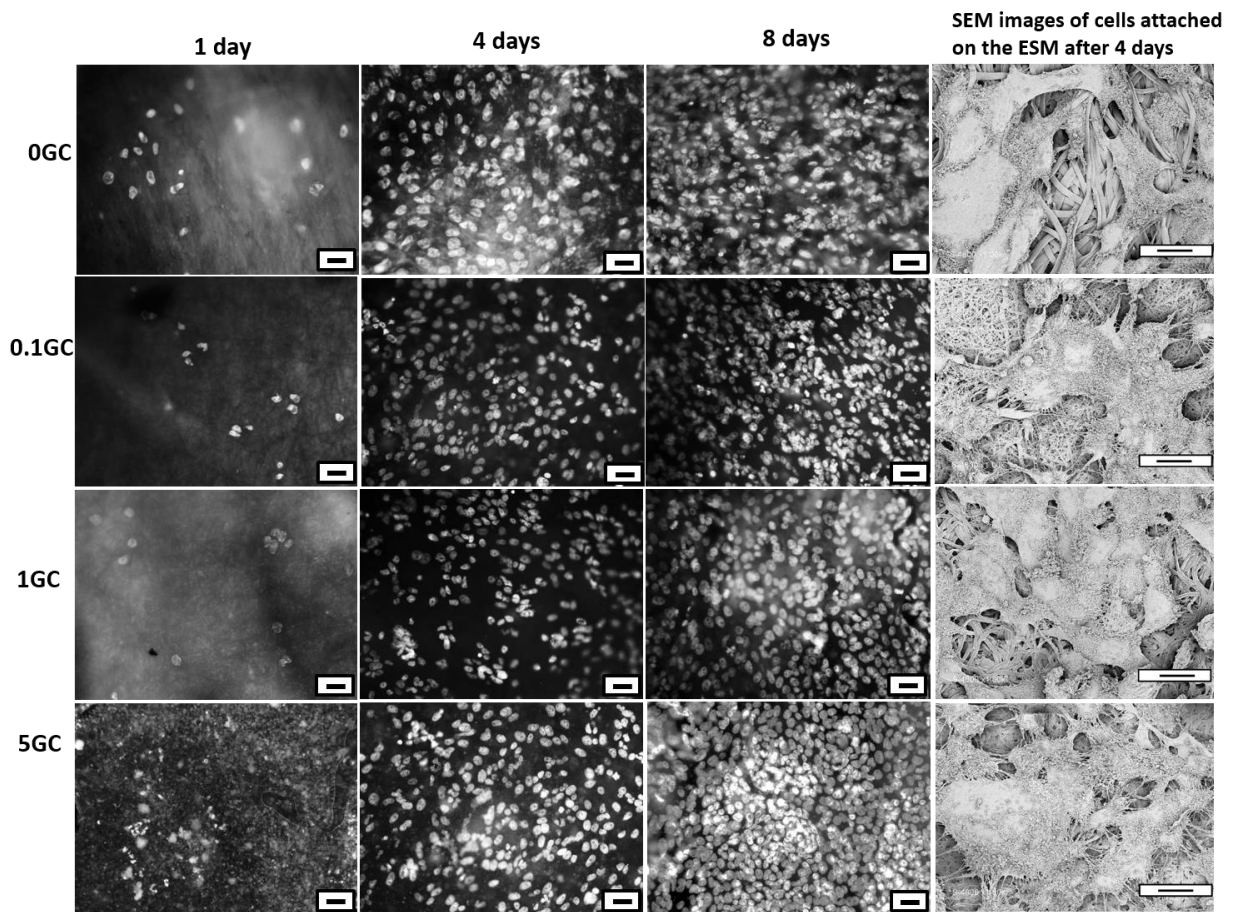
<b>Sample</b> <b>(%)</b>	<b>Gelatin and solvent</b> <b>(%)</b>	<b>BN (%)</b>	<b>Hydroxyapatit</b> <b>e</b> <b>%</b>
<b>GC</b>	85 ±1	0	15
<b>0.1GC</b>	80 ±1	0.5 ±0.1	19.5 ±0.1
<b>1GC</b>	70 ±1	2.1 ±0.1	27.9 ±0.1
<b>5GC</b>	67 ±1	13.2 ±0.1	19.8 ±0.1

## 7. Cell viability and osteoblast gene expression

The cross-linked gelatin/BN ESMs show enhanced biomineralization in SBF thus indicating that they might be suitable to increase the mineralization and may represent potent material for bone tissue engineering. Due to high degree of cross-linking, the GTA has been widely used for cross-linking polymer materials<sup>63</sup>. The aldehyde groups of GTA undergo nucleophilic substitution reaction with amine, thiol and imidazole groups of proteins in cells and induce the toxicity. The aldehyde groups of GTA are utilized for cross-linking the gelatin. However, the unreacted aldehyde terminals are neutralized with 10% glycine and the biocompatibility of cross-linked ESM is investigated using human HOS bone cells, which exhibit osteogenic potential<sup>63-64</sup>. Cell attachment on the ESM is observed by staining the nuclei of the cells since the strong autofluorescence of cross-linked gelatin limits the observation of the actin staining of the cells<sup>65</sup>. The cells attached at 24 hours proliferate on the ESM and reach confluence after 8 days as shown in the Figure 8. Cell attachment and proliferation on the ESM is not affected by the BN concentration and by the direct contact with unexfoliated BN in 5GC. Cells attached on the ESM are cultured for four days and fixed

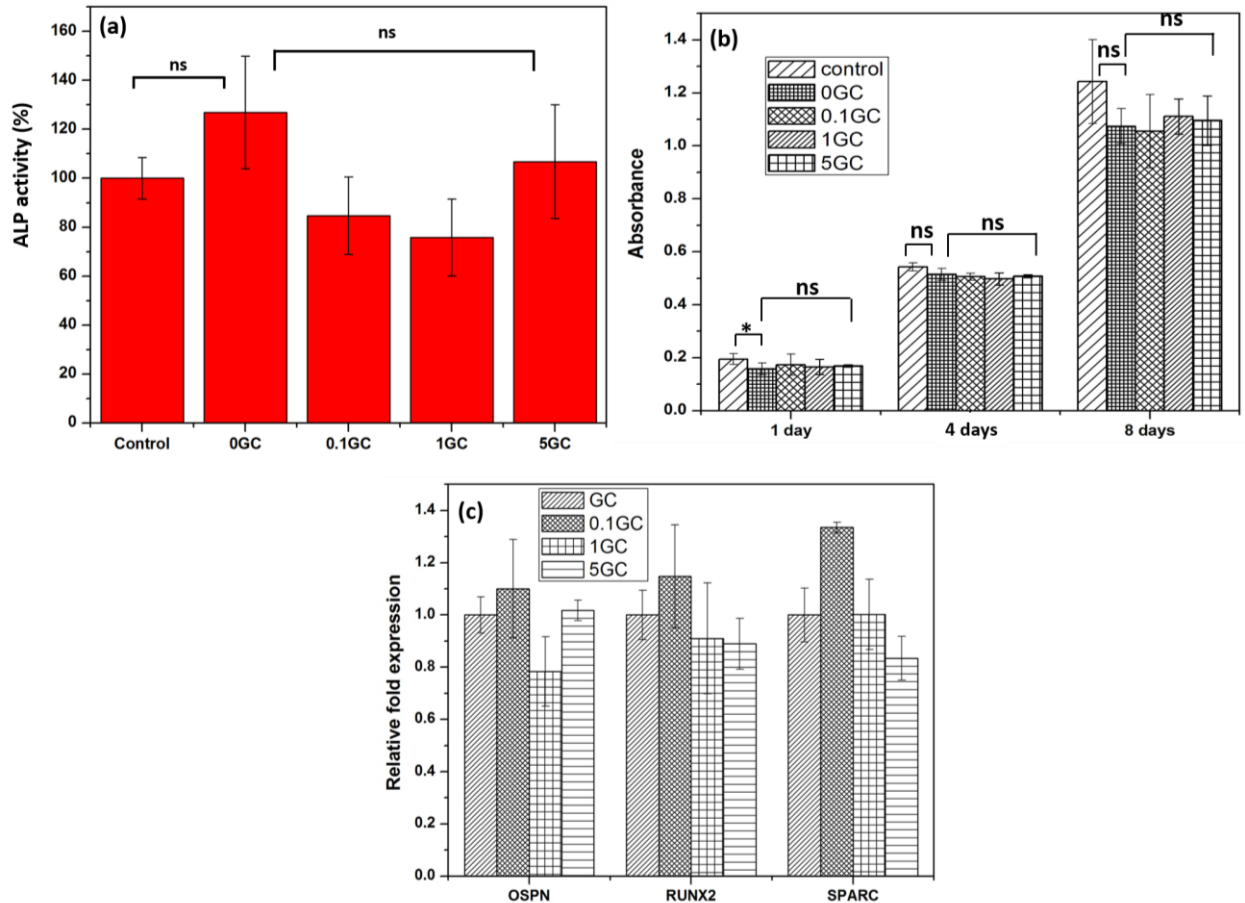
with GTA. The morphology is then recorded using SEM analysis and shown in Figure 8. SEM images show that the ESM facilitates cell attachment and proliferation irrespective of the BN concentration. Cells exhibit enhanced spreading and larger area on the 0GC and BN reinforced GC evidence that the ESM are highly biocompatible material suitable for tissue engineering application.

The alkaline phosphatase (ALP) is the dephosphorylating hydrolase enzyme, which hydrolyses pyrophosphate into phosphate. The free phosphate ions increase calcium phosphate mineralization. Understanding the impact of cross-linked BN reinforced gelatin ESM on HOS cell ALP activity is important for bone tissue engineering. The ALP activity of cells cultured on gelatin and gelatin/BN ESM is measured and the results are shown in Figure 9a. No significant decrease in the ALP activity of cells cultured on 5GC ESM is observed in comparison to the control. These results prove that BN is biocompatible and does not affect ALP activity in HOS cells.



**Figure 8.** The fluorescent images of the Hoechst 33342 stained cells and SEM images of cells attached on the ESM. Scale bar indicates 30  $\mu\text{m}$

The effect of ESMs on cell viability are recorded and shown in the Figure 9b. Cells grown on tissue culture plate without material is considered as a control. Cell viability slightly decreases in the presence of the GC ESMs in comparison to control samples (1 day). However, BN concentration does not influence cell viability. These results suggest that the cross-linking affects cell viability and that boron nitride is biocompatible. Interestingly, by comparing 1, 4 and 8 days of culture, cell proliferation also increases with time in the presence of material thus strengthening the conclusion that BN is biocompatible.



**Figure 9.** (a) Alkaline phosphatase activity, (b) cell viability and (c) quantification of osteoblast gene expression of cells grown on cross-linked ESM (ns = no significance,  $p < 0.05 = *$ ,  $p < 0.005 = **$ ).

Finally, the influence of BN concentration on osteoblast gene expression has been monitored by qPCR analysis (Figure 9c). Osteopontin (OSPN) gene expression is essential for bone mineralization. The presence of phosphorylation, negative charge from the aspartic acid and glutamic acid and calcium chelating sites in the OSPN protein facilitates bone mineralization. It is also proved that high degree of OSPN phosphorylation controls the calcium phosphate crystal growth<sup>66-67</sup>. The major role of RunX2 is to improve osteoblast differentiation and bone formation<sup>68</sup>. SPARC (Osteonectin) is a bone specific protein which plays important roles in linking bone mineral (calcium phosphate) with collagen<sup>69</sup>. Quantification of

osteoblast-specific gene expression is a suitable approach to verify the osteogenic potential of HOS cells as previously used by others to monitor the “bone phenotype” of such cells<sup>32, 70</sup>. The gene expression data are analyzed using Kruskal-Wallis ANOVA statistical test. The expression of OSPN and RunX2 is not significantly affected by the BN concentration ( $p>0.05$ ). SPARC expression increases in cells cultured on ESMs with highly exfoliated BN (0.1GC) in comparison to 0GC ( $p<0.05$  T-test), however 1GC and 5GC do not significantly affect SPARC expression in comparison to 0GC ( $p>0.05$ (*T-test*)). The exfoliated BN has no effect on osteoblast gene expression. However the mechanical properties are significantly improved by exfoliated BN. The GO is found to be toxic at high concentration<sup>71</sup>. The influence of BN on osteoblast expression reported here evidences that BN is a biocompatible alternate to GO<sup>72</sup>. The ESM is highly biodegradable using the collagenase enzyme. On the basis of these results, it can thus be concluded that BN is biocompatible, biodegradable and represents a promising material for bone tissue engineering application.

## CONCLUSIONS

BN reinforced gelatin ESMs are synthesized and characterized. The mechanical properties and stability under PBS are also investigated. DSC results show that gelatin structure displays only a small part of collagen-like triple helical structure meaning that fabrication of ESM with the presented method and using acetic acid as solvent favors coil structure formation. However, the exfoliated BN effectively reinforces the gelatin matrix and increases the Young's modulus. The cross-linked ESMs are highly stable in PBS and keep a fibrous morphology. The exfoliated BN/gelatin ESMs are highly bioactive since they form densely packed hydroxyapatite layers during mineralization in SBF. Cell viability assays, fluorescence imaging, osteoblast gene expression and ALP activity using HOS cells indicate that BN does



not influence cell attachment or proliferation and that the BN reinforced gelatin ESMs are highly biocompatible and suitable for bone tissue engineering. The improved Young's modulus and mineralization of dense hydroxyapatite crystals clearly suggest that the exfoliated BN reinforces electrospun gelatin nanofibers. Regarding these results, BN reinforced gelatin ESMs are promising nontoxic, biocompatible materials for orthopedic applications.

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